



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



## Inhibitory effects of some derivatives of glycyrrhizic acid against Epstein-Barr virus infection: Structure–activity relationships

Jung-Chung Lin<sup>a,\*</sup>, Jaw-Ming Cherng<sup>b</sup>, Man-Shan Hung<sup>a</sup>, Lidia A. Baltina<sup>c,d</sup>, Lia Baltina<sup>c,d</sup>, Rimma Kondratenko<sup>d</sup>

<sup>a</sup> Institute of Microbiology Immunology and Molecular Medicine, Tzu Chi University, 701, Section 3, Chung Yang Road, Hualien 970, Taiwan, ROC

<sup>b</sup> Department of Internal Medicine, China Medical University Peikang Hospital/China Medical University, Taichung 404, Taiwan, ROC

<sup>c</sup> Institute of Organic Chemistry, Ufa Research Center, Russian Academy of Sciences, pr. Oktyabrya 71, Ufa 450054, Russia

<sup>d</sup> Bashkir State Medicinal University, Lenin's Street 3, Ufa 450000, Russia

### ARTICLE INFO

#### Article history:

Received 2 November 2007

Accepted 31 January 2008

#### Keywords:

Glycyrrhizic acid derivatives

Epstein-Barr virus

Antiviral activity

### ABSTRACT

Glycyrrhizic acid (18 $\beta$ -GL or GL) is a herbal drug with a broad spectrum of antiviral activities and pharmacological effects and multiple sites of action. Previously we showed that GL inhibits Epstein-Barr virus (EBV) infection *in vitro* by interfering with an early step of the EBV replication cycle (possibly attachment/penetration). Here we tested the effects of 15 GL derivatives against EBV infection by scoring the numbers of cell expressing viral antigens and quantifying EBV DNA copy numbers in superinfected Raji cells. The derivatives were made either by transformation of GL on carboxyl and hydroxyl groups or by conjugation of amino acid residues into the carbohydrate part. We identified seven compounds active against EBV and all showed dose-dependent inhibition as determined by both assays. Among these active compounds, the introduction of amino acid residues into the GL carbohydrate part enhanced the antiviral activity in three of the seven active compounds. However, when Glu(OH)-OMe was substituted by Glu(OMe)-OMe, its antiviral activity was completely abolished. Introduction of potassium or ammonium salt to GL reduced the antiviral activity with no significant effect on cytotoxicity. The  $\alpha$ -isomer (18 $\alpha$ -GL) of 18 $\beta$ -GL was as potent as the  $\beta$ -form, but its sodium salt lost antiviral activity. The metabolic product of GL, 18 $\beta$ -glycyrrhetic acid (18 $\beta$ -GA or GA), was 7.5-fold more active against EBV than its parental compound GL but, concomitantly, exhibited increased cytotoxicity resulting in a decreased therapeutic index.

© 2008 Elsevier B.V. All rights reserved.

### 1. Introduction

Glycyrrhizic acid (18 $\beta$ -GL or GL) is one of the bioactive compounds of licorice roots (*Glycyrrhiza radix*) and is composed of one molecule of glycyrrhetic acid (18 $\beta$ -GA or GA), which has a steroid-like structure, and two molecules of glucuronic acid. After oral administration or intravenous injection, GL is hydrolyzed by glucuronidase in intestinal bacteria to its active principle aglycone, 18 $\beta$ -GA, which is then absorbed into the blood (Takeda et al., 1996). GL and GA have been shown to possess several beneficial pharmacological activities (Dirnagl et al., 1999; Shibata, 2000; Abe et al., 2003; Armanini et al., 2002; Cherng et al., 2006; Cinatl et al., 2005; Fiore et al., 2004), which include an anti-ulcerative effect, anti-inflammatory activity (Fujisawa et al., 2000), interferon (IFN)- $\gamma$  induction, anti-hepatotoxic effect (Chan et al., 2003; Gumprich et al., 2005; Zheng and Lou, 2003), anti-tumor activity (Hibasami et al., 2006), and it is active against a range of viruses (Lin, 2003;

Cinatl et al., 2003; Crance et al., 2003; Briolant et al., 2004; Cherng et al., 2004; Hoever et al., 2005). Clinically, GL has been used to treat patients with chronic active hepatitis (van Rossum et al., 1999; Bean, 2002; Coon and Ernst, 2004).

In previous years, we have shown that many nucleoside analogs selectively inhibit replication of Epstein-Barr virus (EBV) *in vitro* (Lin et al., 1983, 1984, 1985, 1986, 1987, 1991, 1992; Lin and Machida, 1988; Mar et al., 1995; for a review see Gershburg and Pagano, 2005; Lin, 2006). The molecular target of all nucleoside analogs is the virus-encoded DNA polymerase. Recently, we have reported that GL is active in a dose-dependent fashion against EBV replication in superinfected Raji cells (Lin, 2003). Our results also indicated that GL interferes with an early step of the EBV replication cycle (possibly adsorption and/or penetration) (Lin, 2003). Thus, GL represents a new class of anti-EBV compounds with a mode of action different from that of the nucleoside analogs that inhibit the viral DNA polymerase. In the present studies we extend our previous findings after a search for more potent compounds against EBV. We tested the antiviral activity of 15 GL derivatives synthesized in the laboratory. Here we report the identification of seven active compounds against EBV

\* Corresponding author. Tel.: +886 3 856 5301x7209; fax: +886 3 856 6724.  
E-mail address: [jclin@mail.tcu.edu.tw](mailto:jclin@mail.tcu.edu.tw) (J.-C. Lin).

infection *in vitro* and elucidate their structure–activity relationships.

## 2. Materials and methods

### 2.1. Chemicals

Glycyrrhizic acid (glycyrrhizin, 18 $\beta$ -GL or GL) and 18 $\beta$ -glycyrrhetic acid (18 $\beta$ -GA or GA) were purchased from Sigma-Aldrich Chemie GmbH, Germany. The synthesis of GL derivatives was reported previously (Baltina, 2003; Baltina et al., 2006; Kondratenko et al., 2006). L-Aminoacids were used for the synthesis of GL glycopeptides. Unless otherwise specified, all drugs were dissolved in phosphate-buffered saline, pH 7.5. The chemical structures are shown in Chart 1.

### 2.2. Cell cultures

A latently EBV-infected non-virus producing cell line (Raji) was maintained at between  $4 \times 10^5$  and  $6 \times 10^5$  cells/ml in RPMI 1640 medium containing 10% fetal calf serum (FCS) supplemented with 100 IU penicillin/ml and 100  $\mu$ g streptomycin/ml, as described previously (Lin et al., 1984).

### 2.3. Preparation of virus stocks

A highly productive virus-producer cell line, P3HR-1 (LS), derived by low-serum cloning (Lin, 2000), was maintained in the same culture medium as for Raji cells, except that the serum concentration was reduced to 2%. Viral stocks were prepared from cultures of P3HR-1 (LS) that had been treated with 12-*O*-tetradecanoyl-phorbol-12-acetate (TPA) at a concentration of 30 ng/ml (Lin, 2000). Briefly, the P3HR-1 (LS) cells were induced with TPA for 14 days without additional medium. The cells were removed by centrifugation at  $1200 \times g$  for 10 min, virus was then pelleted from the cell-free supernatant fluids by centrifugation at  $13,000 \times g$  for 90 min in a GS3 rotor (Ivan Sorvall, Inc.). The centrifuged bottles were swabbed to remove residual medium

and the virus pellets were suspended in RPMI 1640 medium. The viral suspension was clarified to remove cellular debris by filtering through 1.2-, 0.8-, and 0.45- $\mu$ m-pore-size filters and stored at  $-80^\circ\text{C}$ .

### 2.4. Drug effect on superinfected Raji cells

As an assay system for drug effects we used Raji cells superinfected with P3HR-1 (LS) virus, which results in reactivation of the latent EBV infection and replication of virus. Raji cells were first propagated in medium containing a sub-effective dose of compound (1  $\mu$ M). For superinfection (Lin, 2003),  $10^6$  Raji cells growing exponentially were pelleted and resuspended in 0.5 ml of RPMI 1640 medium containing 2% FCS and appropriate concentrations of drug as specified. Then 10 units of EBV early antigen (EA)-inducing virus were added to the cells. One unit of virus is defined as the amount of virus capable of inducing 10% of infected Raji cells to express EA. After 2 h adsorption at  $37^\circ\text{C}$  in a  $\text{CO}_2$  incubator, the cells were pelleted and suspended in 1 ml of the medium and incubation continued for 24 h.

### 2.5. Indirect immunofluorescent assay

The levels of EBV early antigen (EA) and viral capsid antigen (VCA) were monitored by indirect immunofluorescent assay (IFA) at 24 h after infection with EBV-positive sera from NPC patients. The specificity of sera for EBV EA/VCA was confirmed by Western blots as reported previously (Lin and Pagano, 1986). The detailed protocol for IFA was described previously (Lin, 2000). Briefly, superinfected Raji cells were smeared on slides and fixed in methanol for 15 min at room temperature. The fixed cells were first reacted with NPC patients' serum, followed by fluorescein-conjugated goat anti-human IgG (H + L) (Santa Cruz Biotechnology, Inc.). After counterstaining with 4',6'-diamidino-2-phenylindole (DAPI) (0.1  $\mu$ g/ml in PBS) for 5 min in the dark, the slides were analyzed under the fluorescent microscope.

### 2.6. Real-time quantitative PCR for EBV DNA

The drug effects on EBV DNA in superinfected Raji cells were determined using a real-time quantitative PCR system toward the BamHI-W fragment region of the EBV genome as described (Lo et al., 1999). Real-time quantitative PCR is based on the continuous optical monitoring of the progress of a fluorogenic PCR reaction. In this system, the amplification primers used in conventional PCR toward the BamHI-W were: W-44F (5'-CCCAACTCCACCACACC-3') and W-119R (5'-TCTTAGGAGCTGTCCGAGGG-3'). The dual-labeled fluorogenic hybridization probe was W-67T[5'-(FAM)CACACTACACACACCCACCCGCTCTC(TAMRA)-3']. The fluorescent probes contained a 3'-blocking phosphate group to prevent probe extension during PCR.

### 2.7. Cell growth and viability assay

The effect of GL derivatives on Raji cell cytotoxicity was carried out by MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide; thiazolyl blue) cell-proliferative Kit I (Roche, Mannheim, Germany). Raji cells were seeded in wells of a 96-well plates and treated with various concentrations of drug for 3 days. Then 10  $\mu$ l of sterile MTT dye were added, and the cells were incubated for 6 h at  $37^\circ\text{C}$ , followed by adding 100  $\mu$ l of acidic isopropanol (0.04 M HCl in isopropanol). Spectrometric absorbance at 595 nm (for formazan dye) was measured with the absorbance at 655 nm for reference.

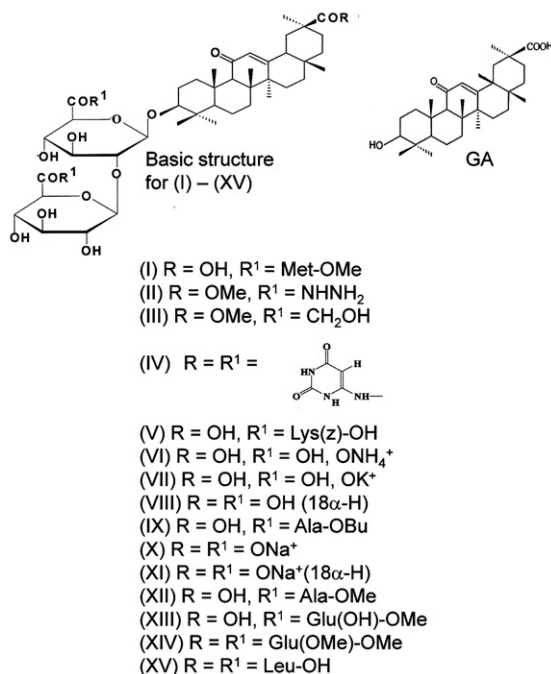
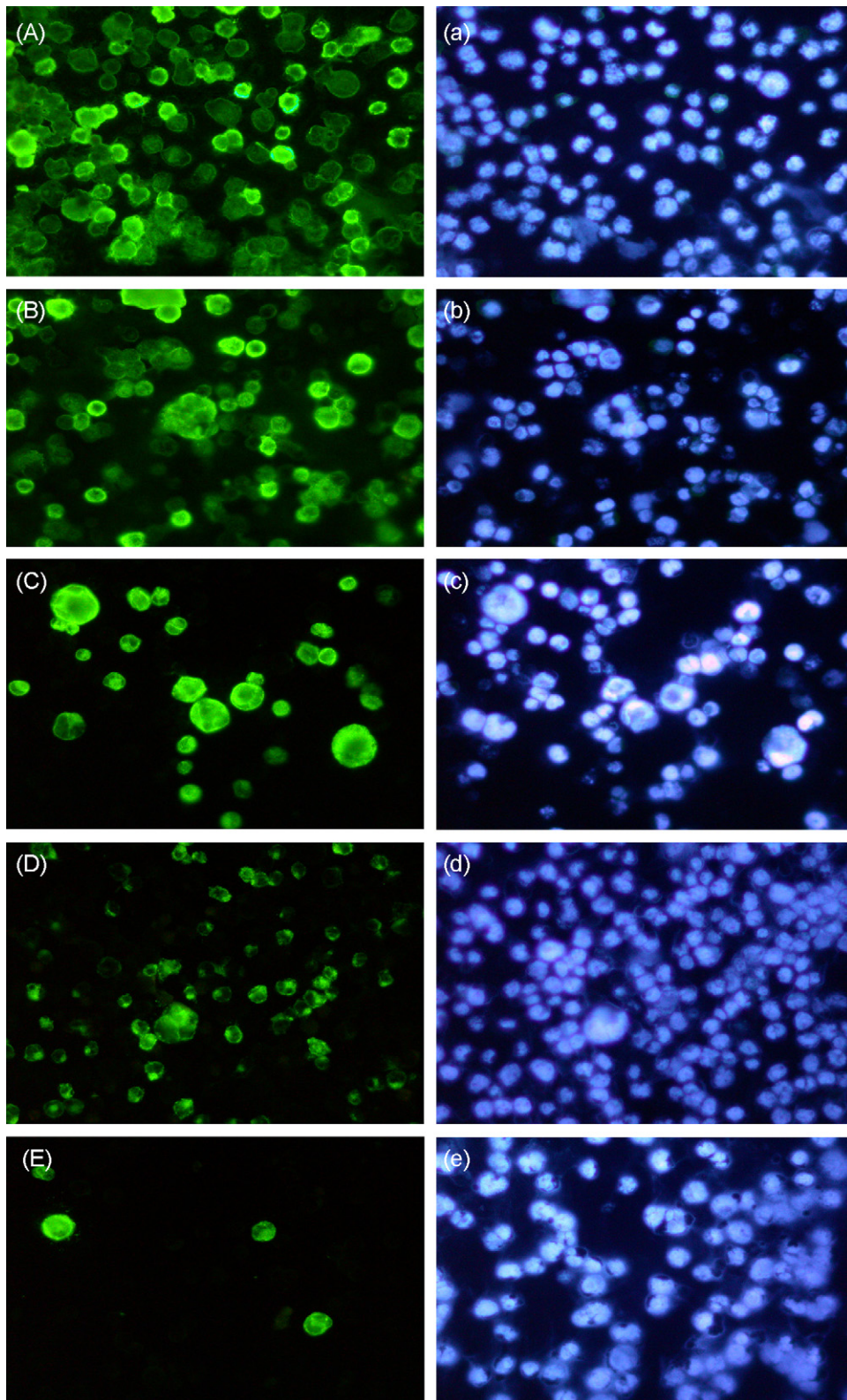


Chart 1.





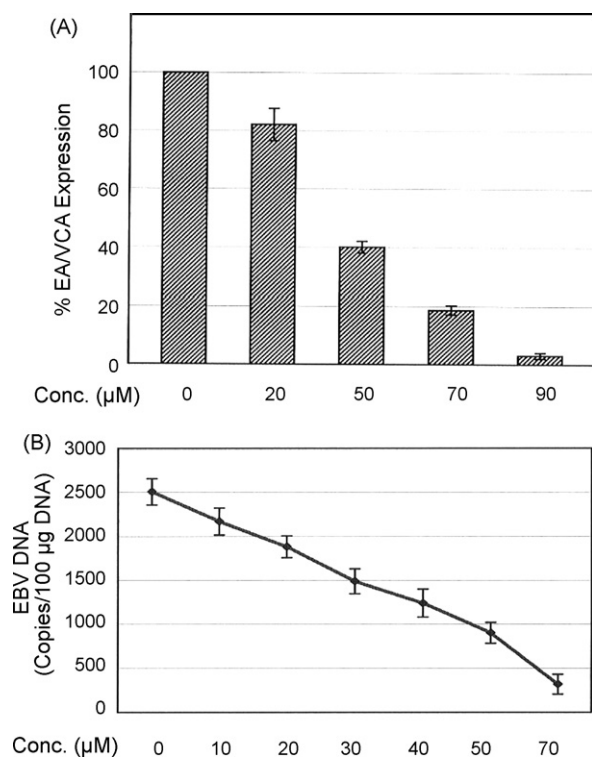
**Fig. 1.** Inhibition of EBV antigen expression in superinfected Raji cells by compound (XV). Raji cells were superinfected with 10 units of EA-inducing virus in the absence and presence of various concentrations of compound (XV). Cells were fixed with cold methanol 24 h after infection and processed for EA/VCA staining using NPC patients' serum. After immunofluorescent staining (panels A–E), the same slides were counterstained with DAPI (panels a–e). Panel A, superinfected cells without drug; panel B, superinfected cells with 20  $\mu\text{M}$  of compound (XV); panel C, superinfected cells with 50  $\mu\text{M}$  of compound (XV); panel D, superinfected cells with 70  $\mu\text{M}$  of compound (XV); panel E, superinfected cells with 90  $\mu\text{M}$  of compound (XV).

### 3. Results

#### 3.1. Effects of GL derivatives on EBV antigen expression and cell proliferation

To determine the dose-dependent effect of GL derivatives, Raji cells were superinfected with P3HR-1 (LS) virus in the presence of various concentrations of compounds. The expression of EA and VCA were monitored by IFA at 24 h after superinfection. A representative result from compound (XV) is shown in Fig. 1. Without drug treatment, approximately 95% of infected cells became positive for viral antigens (Fig. 1, panel A). In the presence of drugs, a dose-dependent inhibition of the expression of viral antigens was observed (Fig. 1, panels B, C, D, and E). The concentration of drug required to inhibit EBV antigen expression by 50% ( $EC_{50}$ ) was determined from the plot of drug concentrations against % of viral antigen-positive cells, assuming no-drug control as 100. Fig. 2A shows graphical quantitation for compound (XV). The  $EC_{50}$  determined from the graph was approximately 42  $\mu$ M. As shown in Table 1, seven GL derivatives were active against EBV infection with  $EC_{50}$  ranging from 42 to 175  $\mu$ M. All these active compounds exhibited a dose-dependent inhibition manner. The parental compound GL was used as a positive drug control. In addition, GA, the metabolite of GL was also tested in parallel with the derivatives. It is interesting to note that GA was the most effective compound to block EBV infection with an  $EC_{50}$  of 5  $\mu$ M, which was 7.6-fold lower than for GL (38  $\mu$ M). However, GA was approximately 60-fold more toxic ( $CC_{50}$  = 75  $\mu$ M) than GL ( $CC_{50}$  = 4500  $\mu$ M).

The cytotoxicity of the GL derivatives was determined in 96-well microtiter plates by the MTT assay and the results are expressed as the concentration of the compound that decreased cell viability to 50% ( $CC_{50}$ ) of the no drug control (Table 1). These compounds exhibited variable cytotoxicity, the  $CC_{50}$  ranging from 1000 to 8000  $\mu$ M.



**Fig. 2.** Dose-dependent inhibition produced by compound (XV) in superinfected Raji cells. Panel A, quantitation of inhibitory effects of compound (XV) on EBV antigen expression determined by IFA; Panel B, quantitation of inhibitory effects of compound (XV) on EBV DNA copy numbers determined by real-time quantitative PCR.

**Table 1**

Effects of GL derivatives on EBV replication in superinfected Raji cells

Compound	$EC_{50}$ ( $\mu$ M) <sup>a</sup>	$CC_{50}$ ( $\mu$ M) <sup>b</sup>	Therapeutic index <sup>c</sup>
(I)	>1000	>8000	ND <sup>d</sup>
(II)	>1000	>8000	ND
(III)	>1000	>8000	ND
(IV)	>1000	>1000	ND
(V)	>1000	>8000	ND
(VI)	75 ± 15 (90 ± 20)	1500 ± 100	20
(VII)	65 ± 10 (75 ± 10)	1200 ± 60	18
(VIII)	30 ± 5 (50 ± 5)	4800 ± 180	160
(IX)	>1000	>8000	ND
(X)	95 ± 25 (130 ± 20)	1400 ± 100	15
(XI)	>1000	>8000	ND
(XII)	30 ± 5 (45 ± 5)	2000 ± 80	67
(XIII)	135 ± 35 (175 ± 20)	4000 ± 160	30
(XIV)	>1000	>8000	ND
(XV)	25 ± 5 (42 ± 6)	5000 ± 150	200
GL (18β-GL)	30 ± 5 (38 ± 4)	4500 ± 160	150
GA (18β-GA)	4 ± 1 (5 ± 1)	75 ± 10	19

<sup>a</sup>  $EC_{50}$ , effective concentration of the compound needed to inhibit EBV genome copy numbers and, in parentheses, EA/VCA expression to 50% of untreated cells.

<sup>b</sup>  $CC_{50}$ , cytotoxic concentration of the compound that decreased cell viability to 50% of untreated cells determined by MTT. Results represent mean values ± S.D. of three independent experiments.

<sup>c</sup> Therapeutic index is defined as the ratio of  $CC_{50}$  to  $EC_{50}$ .

<sup>d</sup> ND, not determined.

#### 3.2. Effect of GL derivative on EBV genome copy numbers

To more precisely determine the drug effects, EBV DNA copy numbers were measured using a real-time quantitative PCR system that amplified a DNA segment in the BamHI-W fragment region of the EBV genome (Lo et al., 1999). A dose-dependent inhibition of viral genome copy numbers by compound (XV) was observed (Fig. 2B). The  $EC_{50}$  obtained by this method is approximately 25  $\mu$ M, in contrast to 42  $\mu$ M determined by IFA (Table 1). Data obtained by PCR and IFA are shown side by side in Table 1. It should be noted that the  $EC_{50}$  values obtained by real-time quantitative PCR is approximately 20–40% lower than that obtained by IFA. However, the inhibitory profile of these compounds remains unchanged, indicating that IFA is as reliable as the quantitative PCR method for initial drug screening.

### 4. Discussion

Evaluation of antiviral agents effective against EBV has been hampered by the lack of a permissive cell system for the replication of this virus. Superinfection of Raji cells with P3HR-1 virus results in shut-down of host cell DNA synthesis and stimulation of viral DNA replication and lytic antigen synthesis and production of virus. We have previously shown that the percentage of EA- and VCA-positive cells is directly proportional to the amount of EBV DNA in the cells (Lin, 2003). Furthermore, we have also demonstrated that GL blocks EBV replication at an early step of the viral replication cycle. However, GL has no effects on EBV DNA in the spontaneously virus-producing cell line P3HR-1 and the latently infected nonvirus-producing Raji cells (Lin, 2003). The lack of effect of GL in P3HR-1 and Raji cells, which are already infected, is consistent with our proposed mechanism of GL action, i.e., inhibition of virus attachment/penetration. Thus, superinfection of Raji cells represents a useful system for evaluation of compounds that block early steps in the EBV replication cycle.

Our results indicate that GL and several of its derivatives are highly selective in their antiviral action in that they inhibit EBV infection at concentrations far below the cytotoxic concentration. As demonstrated previously (Lin, 2003), pretreatment of cells with sub-effective dose of GL markedly reduced cytotoxic effects while



decreasing the amount of drug (approximately 10-fold reduction) needed to reach the same effective level. The marked reduction of  $EC_{50}$  obtained by pretreatment of cells with sub-effective doses of the compounds has clinical implication with regard to prophylactic and therapeutic effects.

Among seven active compounds (Table 1), three of them contain amino acid residues in the GL carbohydrate part. The Ala-OMe containing glycopeptide (XII), the Glu(OH)-OMe containing glycopeptide (XIII), and Leu containing glycopeptides (XV) were active against EBV. However, compounds (XII) ( $EC_{50} = 30 \mu\text{M}$ ) and (XV) ( $EC_{50} = 25 \mu\text{M}$ ) were approximately 5-fold more active against EBV than compound (XIII) ( $EC_{50} = 135 \mu\text{M}$ ). The  $CC_{50}$  values of glycopeptides (XII), (XIII), and (XV) were, respectively, 2000, 4000, and 5000  $\mu\text{M}$ , resulting in a therapeutic index of 67, 30, and 200. In contrast, two other glycopeptide containing compounds such as (I) and (IX) did not exhibit activity against EBV at concentrations up to 1 mM. Interestingly, when Glu(OH)-OMe was substituted by Glu(OMe)-OMe as in compound (XIV), its antiviral activity was completely abolished.

Introduction of a hydroxyl or methoxyl group at the R' position of the aglycon moiety of GL, such as in compounds (I), (II), and (III) did not show any antiviral activity. Compounds (IX) and (XII) both contain a hydroxyl group at the R' position of GA molecule, but with different substitution at the R position of the carbohydrate part of GL. Compound (XII) with an Ala-OMe group at the R position showed good antiviral activity, whereas substitution of compound (IX) with an Ala-OBu group at the same position abolished the antiviral activity. Interestingly, other substitutions at the R' position of GA, such as Lys(z)-OH in compound (V) and L-Glu(OMe)-OMe in compound (XIV), did not confer any antiviral activity. Thus, it appears that the modification of the carbohydrate moiety of GL rather than the R' position of aglycon may affect the antiviral activity. These findings may provide a lead for further development of more active antiviral compounds.

The two commercially available compounds GL and GA were tested in parallel with semisynthetic GL derivatives. Unexpectedly, GA, the metabolic product of GL, was 7.5-fold more active ( $EC_{50} = 4 \mu\text{M}$ ) against EBV than its parental compound ( $EC_{50} = 30 \mu\text{M}$ ), but was 60-fold more toxic ( $CC_{50} = 75 \mu\text{M}$ ) than GL ( $CC_{50} = 4500 \mu\text{M}$ ), resulting in a therapeutic index of 19 and 150, respectively. Introduction of potassium [compound (VI)] or ammonium salt [compound (VII)] to GL reduced the antiviral activity with a significant effect on cytotoxicity. The  $\alpha$ -isomer [18 $\alpha$ -GL, compound (VIII)] derived from 18 $\beta$ -GL was as potent as the  $\beta$ -form, with  $EC_{50}$  of 30 and  $CC_{50}$  of 4800, resulting in a therapeutic index of 160. In contrast, the sodium salt of  $\alpha$ -isomer of 18 $\beta$ -GL [compound (XI)] lost antiviral activity as compared to its  $\beta$ -form [compound (X)]. Previous study indicated that the sugar moiety of GL is essential for anti-SARS-CoV activity (Hoever et al., 2005). The present study clearly demonstrated that compounds (VI), (VII), (VIII), (X), (XII), (XIII), and (XV) are active inhibitors of EBV infection and all contain a sugar moiety. In summary, among the newly identified analogs four showed better or equal activity than the parent GL.

Recently we have demonstrated that GL interferes with an early step of EBV replication cycle (possibly attachment/penetration) (Lin, 2003). However, other possible mechanism of GL may also involve inhibition of EBV replication. Both GL and GA have been shown to elicit a dose-dependent increase in NO production and the level of iNOS mRNA in macrophages (Jeong and Kim, 2002). As a host defense molecule, NO has an antimicrobial activity against a variety of pathogens including viruses. Previous studies indicated that NO is involved in maintaining EBV latency through inhibition of EBV reactivation (Mannick et al., 1994; Gao et al., 1999).

The molecular mechanism underlying the GL inhibition of viral attachment is not well understood. A complement protein called

C3d is a breakdown product of complement C3. Receptors for C3d, called type 2 complement receptors (CR2 or CD21), are expressed on mature B lymphocytes. When B lymphocytes recognize an antigen by their antigen receptors and simultaneously recognize C3d bound to the antigen by the complement receptor, the B cells are activated. The major EBV outer envelope glycoprotein gp350/220 is the CD21 ligand (Nemerow et al., 1987; Tanner et al., 1987). CD21 is the only B-lymphocyte surface protein that binds to gp350/220 (Nemerow et al., 1989), which mediates virus attachment to the EBV/C3dg receptor (CR2) of human B lymphocytes. A recent study indicated that complement C3 is a GL-binding protein (Kawakami et al., 2003). Synthetic peptides corresponding to two regions in gp350/220, which have a similar amino acid sequence with the complement C3dg protein, were used to identify a receptor binding epitope. A peptide corresponding to the N terminus of gp350/220, EDPGFFNVEI, bound to purified CR2 and to CR2 positive but not CR2 negative B and T lymphoblastoid cell lines (Nemerow et al., 1989). This peptide sequence EDPGFFNVEI is similar to the peptide sequence EDPGKQLYNVEA, through which the C3d component of complement binds to CD21 (Nemerow et al., 1989). Synthetic peptides containing the LYNVEA C3d sequence block C3d binding to CD21 (Lambris et al., 1985), whereas peptides containing the EDPGFFNVEA sequence block EBV infection (Nemerow et al., 1989, 1990). Taken together these studies open the possibility that the inhibitory effect of GL could be in part due to the blockade of EBV receptor. The anti-EBV activity of glycopeptides (XII), (XIII), and (XV) could be attributed to the similarity of their amino acid residues with the peptide sequence EDPGKQLYNVA of C3d component of complement that binds to CD21. Ascertaining this possibility would require further study and is currently under investigation.

From the target perspective, the drugs that might be candidates for treatment of EBV infection fall into two groups (for reviews see refs. Gershburg and Pagano, 2005; Lin, 2006). The first group targeting viral DNA polymerase includes acyclic nucleoside analogues (acyclovir, ganciclovir, penciclovir, valaciclovir, valganciclovir and famciclovir); acyclic nucleotide analogues (cidofovir and adefovir); pyrophosphate analogues (phosphonoformic acid and phosphonoacetic acid); possibly 4-oxo-dihydroquinolines (PNU-182171 and PNU-183792). The second group contains compounds of mixed nature that have a distinct structure such as maribavir,  $\beta$ -L-5-iododioxolane uracil and indolocarbazole NIGC-I. Mechanisms of action of these drugs are still under study, but they do not involve inhibition of the viral DNA polymerase.

The molecular target in herpesviruses of all nucleoside analogs are the virus-encoded DNA polymerases from which the mode of action of GL and its derivatives differs. The therapeutic and prophylactic effects of GL on chronic active viral hepatitis (Bean, 2002; Coon and Ernst, 2004; Shibata, 2000; van Rossum et al., 1999) and the inhibitory effect on EBV replication observed in the present study together with its relative lack of toxicity at the cellular level all suggest that GL and selected derivatives may be worth further evaluating for their efficacy and safety in the treatment of active EBV infection.

## Acknowledgments

We thank Joseph Pagano for his critical reading and comments on this manuscript. This work was supported in part by grants from the National Science Council of the Republic of China (NSC94-2320-B-320-011; NSC95-2320-B-320-011) and by an institutional grant (TCIRP 95002-05Y1) of Tzu Chi University. Russian authors thank the Federal Agency on Science and Innovation (Russia) for financial support (contract 02.434.11.7060).

## References

- Abe, N., Akbar, S.M.F., Hasebe, A., Horiike, N., Onji, M., 2003. Glycyrrhizin enhances interleukin-10 production by liver dendritic cells in mice with hepatitis. *J. Gastroenterol.* 38, 962–967.
- Armanini, D., Fiore, C., Mattarello, M.J., Bielenberg, J., Palermo, M., 2002. History of the endocrine effects of licorice. *Exp. Clin. Endocrinol. Diabetes* 110, 257–261.
- Baltina, L.A., 2003. Chemical modification of glycyrrhizic acid as a route to new bioactive compounds for medicine. *Curr. Med. Chem.* 10, 155–171.
- Baltina Jr., L.A., Kondratenko, R.M., Baltina, L.A., Plyasunova, O.A., Galin, F.Z., Tolstikov, G.A., 2006. Synthesis of glycyrrhizic acid conjugates containing L-lysine. *Chem. Nat. Comp.* 42, 543–548.
- Bean, P., 2002. Containing the spread of HIV infection among high-risk groups. *Am. Clin. Lab.* 21, 19–21.
- Briolant, S., Garin, D.N., Scaramozzino, A., Jouan, N., Crance, J.M., 2004. In vitro inhibition of Chikungunya and Semliki Forest viruses replication by antiviral compounds: synergistic effect of interferon-alpha and ribavirin combination. *Antiviral Res.* 61, 111–117.
- Chan, H.T., Chan, C., Ho, J.W., 2003. Inhibition of glycyrrhizic acid on aflatoxin B<sub>1</sub>-induced cytotoxicity in hepatoma cells. *Toxicology* 188, 211–217.
- Cherng, J.-M., Lin, H.J., Hsu, Y.-H., Hung, M.-S., Lin, J.-C., 2004. A quantitative bioassay for HIV-1 gene expression based on UV-activation: effect of glycyrrhizic acid. *Antiviral Res.* 62, 27–36.
- Cherng, J.-M., Lin, H.-J., Hung, M.-S., Lin, Y.-R., Chan, M.-H., Lin, J.-C., 2006. Inhibition of nuclear factor κB is associated with neuroprotective effects of glycyrrhizic acid on glutamate-induced excitotoxicity in primary neurons. *Eur. J. Pharmacol.* 547, 10–21.
- Cinatl, J., Morgenstern, B., Bauer, G., Chandra, P., Rabenau, H., Doerr, H.W., 2003. Glycyrrhizin, an active component of liquorice roots, and replication of SARS-associated coronavirus. *Lancet* 361, 2045–2046.
- Cinatl, J., Michaelis, M., Hoever, G., Preiser, W., Doerr, H.W., 2005. Development of antiviral therapy for severe acute respiratory syndrome. *Antiviral Res.* 66, 81–97.
- Coon, J.T., Ernst, E., 2004. Complementary and alternative therapies in the treatment of chronic hepatitis C: a systematic review. *J. Hepatol.* 40, 491–500.
- Crance, J.M., Scaramozzino, N., Jouan, A., Garin, D., 2003. Interferon, ribavirin, 6-azauridine and glycyrrhizin: antiviral compounds active against pathogenic flaviviruses. *Antiviral Res.* 58, 73–79.
- Dirnagl, U., Iadecola, C., Moskowitz, M.A., 1999. Pathobiology of ischaemic stroke: an integrated view. *Trends Neurosci.* 22, 391–397.
- Fiore, C., Salvi, M., Palermo, M., Sinigaglia, G., Armanini, D., Toninello, A., 2004. On the mechanism of mitochondrial permeability transition induction by glycyrrhizic acid. *Biochim. Biophys. Acta* 1658, 195–201.
- Fujisawa, Y., Sakamoto, M., Matsushita, M., Fujita, T., Nishioka, K., 2000. Glycyrrhizin inhibits the lytic pathway of complement – possible mechanism of its anti-inflammatory effect on liver cells in viral hepatitis. *Microbiol. Immunol.* 44, 799–804.
- Gao, X., Tajima, M., Sairenji, T., 1999. Nitric oxide down-regulates Epstein-Barr virus reactivation in epithelial cell lines. *Virology* 258, 375–381.
- Gershburg, E., Pagano, J.S., 2005. Epstein-Barr virus infections: prospects for treatment. *J. Antimicrob. Chemother.* 56, 277–281.
- Gumprich, E., Dahl, R., Devereaux, M.W., Sokol, R.J., 2005. Licorice compounds glycyrrhizin and 18β-glycyrrhetic acid are potent modulators of bile acid-induced cytotoxicity in rat hepatocytes. *J. Biol. Chem.* 280, 10556–10563.
- Hibasami, H., Iwase, H., Yoshioka, K., Takahashi, H., 2006. Glycyrrhetic acid (a metabolic substance and aglycon of glycyrrhizin) induces apoptosis in human hepatoma, promyelocytic leukemia and stomach cancer cells. *Int. J. Mol. Med.* 17, 215–219.
- Hoever, G., Baltina, L., Michaelis, M., Kondratenko, R., Baltina, L., Tolstikov, G.A., Doerr, H.W., Cinatl Jr., J., 2005. Antiviral activity of glycyrrhizic acid derivatives against SARS-coronavirus. *J. Med. Chem.* 48, 1256–1259.
- Jeong, H.G., Kim, J.Y., 2002. Induction of inducible nitric oxide synthase expression by 18β-glycyrrhetic acid in macrophages. *FEBS Lett.* 513, 208–212.
- Kawakami, F., Shimoyama, Y., Ohtsuki, K., 2003. Characterization of complement C3 as a glycyrrhizin (GL)-binding protein and the phosphorylation of C3α by CK-2, which is potentially inhibited by GL and glycyrrhetic acid in vitro. *J. Biochem.* 133, 231–237.
- Kondratenko, R.M., Baltina Jr., L.A., Baltina, L.A., Baschenko, N.Z., Tolstikov, G.A., 2006. Synthesis and immunomodulating activity of new glycopeptides of glycyrrhizic acid containing residues of L-glutamic acid. *Bioorg. Khim.* 32, 660–666.
- Lambris, J.D., Ganu, Y.S., Hiram, S., Müller-Eberhard, H.J., 1985. Mapping of the C3d receptor (CR2) binding site and a neoantigenic site in the C3d domain of the third component of complement. *Proc. Natl. Acad. Sci. U.S.A.* 82, 4235–4239.
- Lin, J.-C., 2000. Strategies for evaluation of antiviral agents against Epstein-Barr virus in culture. In: Kinchington, D., Schinazi, R.F. (Eds.), *Methods in Molecular Medicine: Antiviral Chemotherapy Protocols*. The Humana Press Inc., New Jersey, pp. 139–150.
- Lin, J.-C., 2003. Mechanism of action of glycyrrhizic acid in inhibition of Epstein-Barr virus replication in vitro. *Antiviral Res.* 59, 41–47.
- Lin, J.-C., 2006. Pathogenesis and therapy of Epstein-Barr virus infection: novel therapeutic approaches. In: Umar, C.S. (Ed.), *New Developments in Epstein-Barr Virus*. Nova Science Publishers, Inc., New York, pp. 1–35.
- Lin, J.-C., Machida, H., 1988. Comparison of two bromovinyl nucleoside analogs, 1-β-D-arabinofuranosyl-E-5-(2-bromovinyl)uracil and E-5-(2-bromovinyl)-2'-deoxyuridine, with acyclovir in inhibition of Epstein-Barr virus replication. *Antimicrob. Agents Chemother.* 32, 1068–1072.
- Lin, J.-C., Pagano, J.S., 1986. Sequential detection of different antigens induced by Epstein-Barr virus and herpes simplex virus in the same Western blot by using dual antibody probes. *J. Virol.* 59, 522–524.
- Lin, J.-C., Smith, M.C., Cheng, Y.C., Pagano, J.S., 1983. Epstein-Barr virus: inhibition of replication by three new drugs. *Science* 221, 578–579.
- Lin, J.-C., Smith, M.C., Pagano, J.S., 1984. Prolonged inhibitory effect of 9-(1,3-dihydroxy-2-propoxymethyl)guanidine against replication of Epstein-Barr virus. *J. Virol.* 50, 50–55.
- Lin, J.-C., Smith, M.C., Pagano, J.S., 1985. Comparative efficacy and selectivity of some nucleoside analogs against Epstein-Barr virus. *Antimicrob. Agents Chemother.* 27, 971–973.
- Lin, J.-C., Nelson, D.J., Lambe, C.U., Choi, E.I., 1986. Metabolic activation of 9([2-hydroxy-1-(hydroxymethyl)ethoxy]methyl)guanidine in human lymphoblastoid cell lines infected with Epstein-Barr virus. *J. Virol.* 60, 569–573.
- Lin, J.-C., De Clercq, E., Pagano, J.S., 1987. Novel acyclic adenosine analogs inhibit Epstein-Barr virus replication. *Antimicrob. Agent Chemother.* 31, 1431–1433.
- Lin, J.-C., De Clercq, E., Pagano, J.S., 1991. Inhibitory effects of acyclic nucleoside phosphonate analogs, including (S)-1-(3-hydroxy-2-phosphonylmethoxypropyl)cytosine, on Epstein-Barr virus replication. *Antimicrob. Agents Chemother.* 35, 2440–2443.
- Lin, J.-C., Reefschlager, J., Herrmann, G., Pagano, J.S., 1992. Structure-activity relationship between (E)-5-(2-bromovinyl)- and 5-vinyl-1-β-D-arabinofuranosyluracil (BV-araU, V-araU) in inhibition of Epstein-Barr virus replication. *Antiviral Res.* 17, 43–52.
- Lo, Y.M.D., Chan, L.Y.S., Lo, K.W., Leung, S.F., Zhang, J., Chan, A.T.C., Lee, J.C.K., Hjelm, N.M., Johnson, P.J., Huang, D.P., 1999. Quantitative analysis of cell-free Epstein-Barr virus DNA in plasma of patients with nasopharyngeal carcinoma. *Cancer Res.* 59, 1188–1191.
- Mannick, J.B., Asano, K., Izumi, K., Kieff, E., Stamler, J.S., 1994. Nitric oxide produced by human B lymphocytes inhibits apoptosis and Epstein-Barr virus reactivation. *Cell* 79, 1137–1146.
- Mar, E.C., Chu, C.K., Lin, J.-C., 1995. Some nucleoside analogs with anti-human immunodeficiency virus activity inhibit replication of Epstein-Barr virus. *Antiviral Res.* 28, 1–11.
- Nemerow, G., Mold, C.V., Keivens Schwend, V., Tollefson, V., Cooper, N.R., 1987. Identification of gp350 as the viral glycoprotein mediating attachment of Epstein-Barr virus (EBV) to the EBV/C3d receptor of B cells: sequence homology of gp 350 and C3 complement fragment C3d. *J. Virol.* 61, 1416–1420.
- Nemerow, G., Houghton, R., Moore, M., Cooper, N.R., 1989. Identification of an epitope in the major envelope protein of Epstein-Barr virus that mediates viral binding to the B lymphocytes Epstein-Barr virus receptor (CR2). *Cell* 56, 369–377.
- Nemerow, G., Mullen, J.J., Dickson, P.W., Cooper, N.R., 1990. Soluble recombinant CR2 (CD21) inhibits Epstein-Barr virus infection. *J. Virol.* 64, 1348–1352.
- Shibata, S., 2000. A drug over the millennia: pharmacognosy, chemistry, and pharmacology of licorice. *Yakugaku Zasshi* 120, 849–862.
- Takeda, S., Ishihara, K., Wakui, Y., Amagaya, S., Maruno, M., Akao, T., Kobashi, K., 1996. Bioavailability study of glycyrrhetic acid after oral administration of glycyrrhizin in rats: relevance to the intestinal bacterial hydrolysis. *J. Pharm. Pharmacol.* 48, 902–905.
- Tanner, J., Weis, J., Fearon, D., Whang, Y., Kieff, E., 1987. Epstein-Barr virus gp350/220 binding to the B lymphocyte C3d receptor mediates adsorption, capping, and endocytosis. *Cell* 50, 203–213.
- van Rossum, T.G., Vulto, A.G., Hop, W.C., Brouwer, J.T., Niesters, H.G., Schalm, S.W., 1999. Intravenous glycyrrhizin for the treatment of chronic hepatitis C: a double-blind, randomized, placebo-controlled phase I/II trial. *J. Gastroenterol. Hepatol.* 14, 1093–1099.
- Zheng, Q.-Z., Lou, Y.-J., 2003. Pathologic characteristics of immunologic injury in primary cultured rat hepatocytes and protective effect of glycyrrhizin in vitro. *Acta Pharmacol. Sin.* 24, 771–777.