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Cane Molasses Graphene Quantum Dots Passivated by PEG Functionalization for Detection of Metal Ions

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ABSTRACT: Poly(ethylene glycol) passivated graphene quantum dots (PEG-GQDs) were synthesized based on a green and effective strategy of the hydrothermal treatment of cane molasses. The prepared PEG-GQDs, with an average size of 2.5 nm, exhibit a brighter blue fluorescence and a higher quantum yield (QY) (up to approximately 21.32%) than the QY of GQDs without surface passivation $(QY = 10.44\%)$. The PEG-GQDs can be used to detect and quantify paramagnetic transition-metal ions including Fe³⁺, Cu^{2+} , Co^{2+} , Ni^{2+} , Pb^{2+} , and Mn^{2+} . In the case of ethylenediaminetetraacetic acid (EDTA) solution as a masking agent, Fe³⁺ ions can be well selectively determined in a transition-metal ion mixture, following the lowest limit of detection (LOD) of 5.77 μ M. The quenching mechanism of Fe^{3+} on PEG-GQDs belongs to dynamic quenching. Furthermore, Fe^{3+} in human serum can be successfully detected by the PEG-GQDs, indicating that the green prepared PEG-GQDs can be applied as a promising candidate for the selective detection of $Fe³⁺$ in clinics.

ENTRODUCTION

GQDs are well known as a distinguished candidate for metalion analysis due to its small size, good biocompatibility, and easy chemical modification of the surface, which implies a good potential application in the fields of environmental detection, biological imaging, drug carrier, and photocatalytic and electrocatalytic technology. $1-5$ $1-5$ $1-5$

Toxicity presents a common practical problem in traditional QDs containing CdS, CdSe, CdTe, and PbS that limit their application in biomedicine.^{[6](#page-8-0)−[8](#page-8-0)} Therefore, it is necessary to choose a green and nontoxic material as a carbon resource to prepare GQDs. Some biomass materials such as durian,^{[9](#page-8-0)} rice husks, 10 etc. are chosen as carbon resources owing to their low price, easy access, and eco-friendliness.

GQDs can be prepared by top-down and bottom-up approaches. The "top-down" synthesis involves stripping large carbon sources into very small quantum dots by physical or chemical means. The "bottom-up" synthesis, as opposed to the top-down synthesis, uses very small carbon materials such as molecular or ionic states to produce quantum dots. $11,12$ The characteristics of luminescent color adjustment, dimensional dependence, pH dependence, excitation wavelength depend-

ence, and solvent dependence associated with these prepara-tion methods have been reported.^{[13](#page-8-0)} Although the research on GQDs has made tremendous progress, many problems need to be solved. The design of high-fluorescence quantum yield (QY) GQDs and the improvement of physicochemical properties of GQDs remain a challenging research task, which severely limits its practical application. To enhance the performance of GQDs, surface treatment is present to revise the GQDs surface with chemicals with functional groups. Surface treatment with various molecules has been developed as a facile and cost-effective approach to manipulate the properties of GQDs in recent years.^{[14](#page-8-0),[15](#page-8-0)} Sun et al.¹⁶ reported that the QY of GQDs modified with amino groups is six times higher than that of unmodified GQDs. The results

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Figure 1. TEM and HRTEM images of (a) GQDs and (b) PEG-GQDs; size distribution of (c) GQDs and (d) PEG-GQDs; and (e) atomic force microscopy (AFM) image of GQDs.

show that only a few defect-free GQDs generate band gap fluorescence in the limited size sp^2 region, and most GQDs inevitably introduce surface defects in the preparation process. These defects on the surface will form many level traps or surface state energy levels, which will become nonradiative recombination centers of electron and hole, leading to a decrease in luminous efficiency. The surface of quantum dots was usually modified to eliminate the defect state luminescence caused by surface defects and improve luminous efficiency.

Ferric ion (Fe^{3+}) , as the main form of iron elements, can be combined with a variety of proteins. However, $Fe³⁺$ is detrimental when present in excess in vivo, causing a variety of diseases. Hence, the monitoring of $Fe³⁺$ is of great significance in life. So far, a variety of methods have been introduced and developed for the detection of $Fe³⁺$, involving REDOX titration, gravimetric method, and spectrophotometric method. However, the shortcomings of these methods, such as long test cycle, multiple analysis steps, wastage of reagents, and interference of impurities in the determination results, limit

Figure 2. (a) FT-IR spectra of GQDs and PEG-GQDs. (b) X-ray photoelectron spectroscopy (XPS) survey scan spectrum of PEG-GQDs. Highresolution XP spectra of (c) carbon and (d) oxygen of PEG-GQDs. (e) UV−vis absorption spectra of GQDs and PEG-GQDs. (f) Fluorescence emission spectra of GQDs and PEG-GQDs and the inset of the fluorescence photographs of GQDs (left) and PEG-GQDs (right). (g) Fluorescence emission spectra of PEG-GQDs recorded for progressively longer excitation wavelengths from 280 to 480 nm. (h) Diameter histogram of the effect of adding an amount of PEG on the fluorescence emission spectra of PEG-GQDs.

their usage. Therefore, effective improvement in terms of detection technology is essential.

In this paper, we report our work on the preparation of GQDs using cane molasses, an environment-friendly as well as economically inexpensive natural waste biomass, as carbon source, and surface modification of PEG (PEG-GQDs) was carried out by a facile and rapid method. The modified GQDs show stronger fluorescence, higher QY, and better stability than the unmodified synthetic GQDs. Moreover, the

fluorescence quenching and the selectivity of PEG-GQDs for the detection of $Fe³⁺$ were also investigated.

■ RESULTS AND DISCUSSION

Morphology and Composition of the PEG-GQDs. Different conditions were tested for the best performance of GQDs, and it was found that a volume ratio of 1:16 $(V_{\text{cane molasses}}/V_{\text{ultrapure water}})$, reaction temperature of 190 °C, and the reaction time of 24 h yielded the best result, so this condition was used throughout the present work ([Figure S1](http://pubs.acs.org/doi/suppl/10.1021/acsomega.0c00098/suppl_file/ao0c00098_si_001.pdf), Supporting Information). Transmission electron microscopy (TEM) images of the synthesized GQDs and PEG-GQDs in [Figure 1](#page-1-0) demonstrate that both GQDs and PEG-GQDs are present with nearly spherical morphology. The average particle size of PEG-GQDs ([Figure 1b](#page-1-0)) appears to be similar to that of GQDs [\(Figure 1a](#page-1-0)), but the dispersion of PEG-GQDs is better than that of GQDs. The GQDs exhibit an average diameter of 2.2 nm when counting about 100 particles ranging from 1.1 to 3.2 nm [\(Figure 1](#page-1-0)c). For PEG-GQDs, the particle size was found to range from 1.5 to 3.5 nm with an average lateral size of about 2.5 nm ([Figure 1](#page-1-0)d). High-resolution TEM (HRTEM) images show that the graphitic lattice of both GQDs (the inset of [Figure 1](#page-1-0)a) and PEG-GQDs (the inset of [Figure 1b](#page-1-0)) possesses a lattice spacing of 0.21 nm, which are assigned to the (100) of graphitic carbon, suggesting a highly uniform crystallinity of the produced GQDs and PEG-GQDs.¹

The AFM image in [Figure 1e](#page-1-0) displays that the maximum thickness of the GQDs is no more than 2.7 nm, corresponding to three to eight layers of graphene.

[Figure 2](#page-2-0)a displays the Fourier transform infrared (FTIR) spectra of GQDs and PEG-GQDs, with assignments of functional groups of the GQDs and PEG-GQDs labeled. The broad band in 3402 and 3302 cm[−]¹ corresponds to the stretching vibration of O−H. The weak peaks at 1631 and 1656 cm⁻¹ are related to C=C skeletal vibrations.^{[18,19](#page-8-0)} Compared to the GQDs FTIR spectrum, three additional peaks at 2875, 1346, and 1079 cm[−]¹ emerge in PEG-GQDs in the FTIR spectrum, which is due to the stretching vibration, bending vibration of C−H, and the stretching vibration of C− O, respectively.^{[20](#page-8-0),[21](#page-8-0)} These results verified that hydroxyl, carbon−hydrogen bond, and carbon−oxygen were present in the PEG-GQDs sample.

The XPS survey scan spectrum of the PEG-GQDs is shown in [Figure 2b](#page-2-0), which displays two binding energy peaks at 285 and 532 eV, corresponding to C 1s and O 1s, respectively. The C 1s spectrum shown in [Figure 2](#page-2-0)c can be deconvoluted into three components at 284.2, 285.3, and 286.6 eV, which are attributed to the C-C/C-H, C-OH, and C=O, respec-tively.^{[22](#page-8-0)} The O 1s spectrum ([Figure 2d](#page-2-0)) shows two major components at 531.5 and 532.2 eV, which are related to the O=C and HO−C groups, respectively.^{[20](#page-8-0)} These functionalities of PED-GQDs are consistent with the infrared spectroscopic analysis results, indicating that PEG was coated effectively on the surface of GQDs.

Optical Properties and Stability of PEG-GQDs. UV−vis absorption and fluorescence spectra of the PEG-GQDs were also collected at room temperature to study the effect of surface modification of PEG on optical properties, and the results are shown in [Figure 2e](#page-2-0). Both GQDs and PEG-GQDs have an absorption peak near 260 nm, which is assigned to the $\pi-\pi$ * transition of the C=C bond.^{[21](#page-8-0)[,23](#page-9-0)} In contrast, the absorption peak at 260 nm of the PEG-GQDs has a more obvious variation tendency, which may be due to the fact that the polarity of water is stronger than that of PEG, since the strong polar solvent may entail the spectrum fine structure from the vibration effect to disappear and produce a wide $\rm{peak.}^{24}$

The excitation and emission fluorescence spectra of the GQDs and PEG-GQDs are displayed in [Figure 2f](#page-2-0). A fluorescence emission peak was observed at 440 nm for GQDs, and a fluorescence emission peak was observed at 445 nm when PEG-GQDs were excited at 376 nm. The emission

peak position of PEG-GQDs appeared to shift to red compared to the GQDs emission peak, which suggests that the electronic state of the GQDs was affected by the coating of PEG. The electrons on the GQDs layer can easily jump to the PEG layer. However, the holes that should have been paired with these electrons can only stay on the surface of the GQDs, resulting in a relatively small separation of electrons from the holes. This small separation reduces the binding energy of excitons formed, leading to the red shift of PEG-GQDs peak position.^{[25](#page-9-0)}

The relationship between the fluorescence emission peak position of GQDs and PEG-GQDs and the excitation wavelength was investigated by exciting at different excitation wavelengths from 280 to 480 nm in an interval of 20 nm. The emission wavelength positions are found to red shift with increasing excitation wavelength, and the fluorescence intensity shows an increasing trend from 280 to 380 nm then decreasing from 380 to 480 nm. The GQDs can reach the maximum fluorescence intensity at an excitation wavelength near 360 nm ([Figure S2a,](http://pubs.acs.org/doi/suppl/10.1021/acsomega.0c00098/suppl_file/ao0c00098_si_001.pdf) Supporting Information); nevertheless, the maximum excitation wavelength of PEG-GQDs is around 380 nm [\(Figure 2g](#page-2-0)). Thus, both GQDs and PEG-GQDs also show a typical excitation wavelength dependence of the luminescence.

The fluorescence intensity of PEG-GQDs was found to increase by 40% in comparison to that of GQDs. PEG is selected to modify the defected surface of GQDs to eliminate the defect state luminescence and to create more exciton luminescence, thereby improving the luminescence efficiency of GQDs. Some studies have shown that different GQDs luminescence colors are related to multitudinous carbon sources (such as carbon fiber, graphite, and graphene oxide).[26](#page-9-0)−[28](#page-9-0) The luminescence of PEG-GQDs derived from cane molasses in the present work is dominated by indigo blue light. The inset in [Figure 2f](#page-2-0) is the fluorescence photographs of the PEG-GQDs on the right side and the GQDs on the left side under the UV lamp ($\lambda = 365$ nm); as can be seen, PEG-GQDs can emit brighter indigo blue fluorescent light in comparison to GQDs. The highest fluorescence QY of the obtained PEG-GQDs was determined to be 21.32%, which is much higher than that of the GQDs (10.44%) and the previous report of C-dots (5.8%) ,^{[29](#page-9-0)} which also focuses on cane molasses as a carbon source. This implied that the effective passivation of GQDs surfaces with PEG significantly enhanced the fluorescence QY of GQDs. Moreover, the PEG capping on the outside of GQDs was found to change the fluorescence decay lifetime of GQDs. The fluorescence lifetime of GQDs is prolonged from 2.09 to 2.25 ns (as shown in Table 1). This

Table 1. Fluorescence Decay Time and Pre-Exponential Factor of GQDs and PEG-GQDs

parameter	τ_1 (ns)	τ_2 (ns)	A1	A2	$\tau_{\rm ave}$ (ns)
GODs	1.10	3.67	18671.88	3499.49	2.09
PEG-GODs	1.80	10.30	11928.71	268.85	2.77
$PEG-GODs-Fe3+$	1.36	4.17	36 188.45	2212.06	1.80

may be the result of the conjugated aromatic structure of GQDs being modified by PEG, forming an electron delocalization field by combining the lowest unoccupied molecular orbital (LUMO) energy levels of PEG ligand with the conductive band-edge energy of GQDs. The excited electron of PEG-GQDs with a more stable environment was

supplied from the electron delocalized field, thus the fluorescence lifetime of PEG-GQDs is prolonged.^{[30](#page-9-0)}

To explore the effect of PEG addition on the luminescence efficiency of PEG-GQDs, different dilution factors were chosen to passivate the surface of GQDs. It was found that the fluorescence intensity was the highest when the GQDs solution was diluted 50 times with PEG ([Figure 2](#page-2-0)h). It seemed that a complete encapsulation of GQDs defects was not achieved when the PEG content is too low, and the excessive amount of PEG may affect the emission of the characteristic peak of GQDs, giving rise to lower fluorescence intensity.

PEG is widely used in the field of biomedicine because of its biocompatibility, nontoxicity, antigen-free, and nonimmune characteristics. 31 PEG comprises two chemical groups: one is hydrophilic polar groups and the other is oily nonpolar groups. 32 When the surface of GQDs is modified by PEG, the nonpolar pro-oil groups are adsorbed to the surface of GQDs by the exchange of coordination bonds, which allows good permeability and good biocompatibility of cells. The polar hydrophilic group is compatible with water; the surface modification of GQDs by PEG can also reduce the surface tension of the GQDs, which changes its surface properties and makes the PEG-GQDs more stable. On the other hand, the long tail end can form spatial resistance to water molecules on the surface of PEG-GQDs, which results in PEG-GQDs in the water to have a large repulsive volume, thus preventing the agglomeration of \tilde{GQDs} .³³ Thus, the surface modification of PEG can also result in good water solubility, good dispersion, and few defects in luminescent materials, which contribute to improving the stability and the fluorescence efficiency of GQDs; the function mechanism of PEG is shown in Figure 3.

Figure 3. Schematic illustration of the function of PEG for GQDs.

ζ-potential is an important index to characterize the stability of product dispersion. The lower the absolute value of the ζpotential, the more molecules or particles in the system coagulate. 34 The ζ -potential plot in [Figure 4a](#page-5-0) shows that PEG-GQDs show two peaks at 30 and 70 mV in the positive electric region, while GQDs display only one peak at −7 mV in the negative electric region. Clearly, PEG-GQDs show a bigger potential than the negatively charged GQDs in terms of absolute value, indicating that PEG-GQDs have better stability compared to that of GQDs. [Figure 4b](#page-5-0) demonstrates the fluorescence intensity change of PEG-GQDs and GQDs within 35 days, the less change in the fluorescence intensity of PEG-GQDs further demonstrates that the stability of PEG-GQDs is significantly better than that of GQDs.

To further test if the above mechanism is rational and universal, methanol (MT), ethanol (EtOH), n-butyl alcohol (NBA), glycerol (GI), ethylene glycol (EG), PEG-400, and PEG-600 were respectively selected for the fluorescence emission spectrum testing experiments. As can be seen in [Figure 4](#page-5-0)c, the fluorescence intensity of GQDs modified by these passivators was found to increase compared to that of the

unmodified GQDs, indicating that the surface of GQDs can be passivated by passivators with a hydroxyl group. Among all the passivators in this experiment, GQDs passivated by PEG-200 show a much higher fluorescence intensity than other alcohols, probably because its long-chain polymer structure can achieve a tighter coating of the inner layer GQDs, which is in contrast to micromolecules without a long-chain structure, such as MT, EtOH, NBA, GI, and EG, and because the hydroxyl values for PEG-400 and PEG-600 are far less than that of PEG-200.³⁵ PEG-200 is thus considered to be the best choice for passivating GQDs in this paper.

Sensitivity Detection of Metal Ions. The characterization of PEG-GQDs promises the feasibility of an application. Here, to develop a fluorescence quenching method for metal ions sensing, the fluorescence emission spectra of PEG-GQDs and GQDs homogeneous solutions containing 400 μ M various single-metal ions (Cr³⁺, Ca²⁺, Al³⁺, Cu²⁺, Pb²⁺ Mn^{2+} , Ag⁺, Ba²⁺, Co²⁺, Cd²⁺, Zn²⁺, Hg²⁺, Fe³⁺, Mg²⁺, Ni²⁺) were scanned with a 376 nm excitation wavelength, respectively. The $(F_0 - F)/F_0$ bar diagram of the PEG-GQDs and the GQDs with the above ions is shown in [Figure](#page-6-0) [5](#page-6-0)a, where F and F_0 refer to the fluorescence intensities in the presence and absence of metal ions, respectively. As can be seen in [Figure 5](#page-6-0)a, the fluorescence of PEG-GQDs is greatly quenched by several paramagnetic transition-metal ions such as Fe³⁺, Cu²⁺, Co²⁺, Ni²⁺, Pb²⁺, and Mn²⁺, among which the quenching degree of $Fe³⁺$ is the highest. [Figure 5a](#page-6-0) also shows that the quenching degree of the PEG-GQDs by all metal ions screened in the present work is greater than that of GQDs.

The fluorescence intensity changes of the GQDs and the PEG-GQDs in the solutions with different concentrations of Fe3+ are displayed in [Figure 5](#page-6-0)b,c, respectively. A continuous decreasing trend with gradual increase in the concentration of $Fe³⁺$ in the solution from 0 to 240 μ M for the GQDs and from 0 to 60 μ M for the PEG-GQDs was observed, suggesting that the fluorescence intensity of the GQDs and the PEG-GQDs is sensitive to $Fe³⁺$. The calibration curves of the degree of fluorescence quenching $(F_0 - F)/F_0$ of both GQDs and PEG-GQDs versus the concentration of $Fe³⁺$ ions are illustrated in [Figure 5d](#page-6-0), which displays a good linear relationship for the GQDs in the concentration range of $40-200$ μ M with a correlation coefficient (R^2) of 0.97, and a good linear concentration range and R^2 value for PEG-GQDs are 24–52 μ M and 0.98, respectively. The limit of detections (LODs) of Fe³⁺ are estimated to be 30 μ M for GQDs and 15 μ M for PEG-GQDs. The $Fe³⁺$ detection limit was defined as LODs = $3\sigma/m$, where σ is the standard deviation and m is the slope of the line in the linear response region of the calibration curve. Similarly, the detection limit of other metal ions are also determined using the above computing method with the PEG-GQDs system. The linear concentration range for the detection of Cu^{2+} , Co^{2+} , Ni^{2+} , Mn^{2+} , and Pb^{2+} by PEG-GQDs in single-metal ion solution is 20−52, 40−70, 60−180, 120−190, and 100−280 μM, respectively, and the corresponding LODs are 15.73, 23, 59.60, 80.69, and 93.95 μM, respectively [\(Figure S2b](http://pubs.acs.org/doi/suppl/10.1021/acsomega.0c00098/suppl_file/ao0c00098_si_001.pdf)−f, Supporting Information).

Selective Detection of $Fe³⁺$ for PEG-GQDs. Iron is an indispensable element for the normal operation of life. Hence, it is very crucial to be able to conduct selective and quantitative detection of Fe^{3+} ions in a solution mixture with multiple metal ions. To avoid interference from other ions, the experiment was carried out by adding EDTA in both PEG-GQDs and GQDs sensing systems with single metal ions, respectively.

Figure 4. (a) ζ-potentials of GQDs and PEG-GQDs. (b) Stability of GQDs and PEG-GQDs for 35 days. (c) Fluorescence emission spectra of passivated GQDs under different modifiers.

EDTA plays the role of a complex agent to mask the effects of other metal ions. [Figure 5](#page-6-0)e shows that in the presence of 4 mM EDTA, all metal ions (0.01 M) in our experiment except for Fe3+ have little effect on the fluorescence of both PEG-GQDs and GQDs, which demonstrates that the sensing systems have high selectivity for Fe^{3+} .

Quantitative analysis results of Fe^{3+} in multifarious metal ion solution $(Cr^{3+}, Ca^{2+}, Al^{3+}, Cu^{2+}, Pb^{2+}, Mn^{2+}, Ag^+, Ba^{2+}, Co^{2+},$ Cd^{2+} , Zn^{2+} , Hg^{2+} , Mg^{2+} , Ni^{2+} , 0.01 M) in the presence of 4 mM EDTA for PEG-GQDs and GQDs are shown in [Figure 5](#page-6-0)f,g, respectively. In the PEG-GQDs system, the linear concentration range for Fe³⁺ detection is 8–24 μ M, with R² of 0.99, and the LODs can reach 5.77 μ M, which is even higher than that in the case of $Fe³⁺$ alone and the previous report by Ananthanarayanan et al. (LODs \approx 7.22 μ M).^{[36](#page-9-0)} For the GQDs system, the linear concentration range is $12-32 \mu M$, with R^2 of 0.97, and the LODs can reach 10.78 μ M, which also shows high selectivity and sensitivity like PEG-GQDs.

Fluorescence Quenching Mechanism of PEG-GQDs. To study whether the above-mentioned fluorescence quenching mechanism of PEG-GQDs by $Fe³⁺$ is dynamic or static, the effect of temperature on the quenching constant, fluorescence lifetime, and UV−vis absorption spectra were also studied in the present work. Dynamic quenching is associated with diffusion, the viscosity of the solution decreases, and the motion of the molecule accelerates when the temperature increases, resulting in an increase in diffusion coefficient and thus an increase in the molecule quenching constant. According to the Stern–Volmer equation $F_0/F = 1 + K_{SV}\tau_0$ [C] (where F_0 and F are the fluorescence intensities of PEG-GQDs in the absence and presence of Fe^{3+} , respectively, K_{SV} is the quenching constant, τ_0 is the lifetime of PEG-GQDs

without Fe³⁺, [C] is the concentration of Fe³⁺), K_{SV} increases with temperature if it is dynamic quenching. Besides, since the collision of fluorescence molecules and quenching agent only affects the excitation state of fluorescence molecules, the absorption spectrum of fluorescence molecules will not change, while the lifetime of the excitation state of fluorescence molecules will change in the presence of quenching agent. 37 [Figure 5h](#page-6-0) shows that the slope K_{SV} rises from 0.009 to 0.027 when the temperature is elevated from 25 to 40 °C. The weighted average fluorescence lifetime calculated from the fluorescence decay curves ([Figure S3a](http://pubs.acs.org/doi/suppl/10.1021/acsomega.0c00098/suppl_file/ao0c00098_si_001.pdf), Supporting Information) of PEG-GQDs and PEG-GQDs-Fe³⁺ systems is 2.25 and 1.80 ns [\(Table 1\)](#page-3-0), respectively. So, the characteristics of the increase of K_{SV} , the decrease of fluorescence lifetime and the almost invariant UV−vis absorption spectra of PEG-GQDs ([Figure S3b,](http://pubs.acs.org/doi/suppl/10.1021/acsomega.0c00098/suppl_file/ao0c00098_si_001.pdf) Supporting Information) can demonstrate that the mechanism of quenching of PEG-GQDs by $Fe³⁺$ is dynamic.

In the case of a dynamic quenching process, the excited-state molecule of the fluorescent material loses its excitation energy by the mechanism of energy transfer or charge transfer through the collision with the quenching agent molecule and returns to the ground state. Hence, this fluorescence quenching phenomenon may be ascribed to the energy transfer caused by the collision of the PEG-GQDs and metal ions, during which the energy of PEG-GQDs is transferred from the conduction band to the metal ions, leading to a decrease in the fluorescence intensity, 38 as shown in [Figure 6.](#page-7-0) In addition, the −OH groups on the surface of PEG-GQDs are able to coordinate with metal ions forming metal hydroxides, $39,40$ which results in aggregation between PEG-GQDs, giving rise to decrease in fluorescence intensity. These features of PEG-

Figure 5. (a) Fluorescence responses of GQDs and PEG-GQDs to the different metal ions in the absence of ethylenediaminetetraacetic acid (EDTA). (b) Fluorescence emission spectra of GQDs quenched by different concentrations of Fe³⁺ (from 0 to 240 μ M) in a homogeneous solution. (c) Fluorescence emission spectra of PEG-GQDs quenched by different concentrations of Fe³⁺ (from 0 to 60 μ M) in a homogeneous solution. (d) Calibration curves of the degree of fluorescence quenching $[(F_0 - F)/F_0]$ of GQDs and PEG-GQDs versus Fe³⁺ ions concentration. (e) Fluorescence responses of GQDs and PEG-GQDs to different metal ions in the presence of EDTA. Calibration curves of the degree of fluorescence quenching $[(F_0 - F)/F_0]$ of (f) PEG-GQDs and (g) GQDs versus Fe³⁺ ions concentration in the presence of EDTA, respectively. (h) Stern-Volmer curve for fluorescence quenching of PEG-GQDs by Fe³⁺ at different temperatures.

GQDs could contribute to higher sensitivity in the detection of metal ions compared with GQDs.

Detection of Fe³⁺ in Healthy Human Serum. To explore the feasibility of the PEG-GQDs as a tool for the quantitative analysis of $Fe³⁺$, healthy human serum was used for verification in this experiment under the above-mentioned optimal conditions. The average recovery of adding standard

sample is close to 100% and the RSD is less than 5%, as shown in [Table 2,](#page-7-0) indicating the synthesized PEG-GQDs can meet the requirements of detecting $Fe³⁺$ in serum.

■ **CONCLUSIONS**

In summary, the blue fluorescent PEG-GQDs were smoothly synthesized using cane molasses as the source of carbon by the

Figure 6. Quenching mechanism of metal ions of PEG-GQDs.

Table 2. Detection of $Fe³⁺$ in Healthy Human Serum by PEG-GQDs

sample		added $(\mu \text{mol/L})$ found $(\mu \text{mol/L})$ recovery $(\%)$		RSD(%)
healthy	Ω	17.45		
human	2	19.59	107.07	
serum	2	19.38	96.40	4.34
	2	19.52	103.59	

hydrothermal method, followed by passivation. PEG-GQDs showed more uniform and dispersed spherical morphology, higher stability, longer fluorescence lifetime, and higher fluorescence intensity and quantum yield than GQDs. The PEG-GQDs can selectively detect $\mathrm{Fe^{3+}}$ in a mixture solution with multiple metal ions in the presence of EDTA, which can be realized by the detection of $Fe³⁺$ in serum. The GQDs passivated by PEG with high QY can meet the requirements for the multifunctional applications.

EXPERIMENTAL SECTION

Materials. Cane molasses was provided by Ganhua Group Co., Ltd., Guigang, Guangxi, China. Poly(ethylene glycol) (PEG) including PEG-200, PEG-400, and PEG-600, methanol (MT), ethanol (EtOH), n-butyl alcohol (NBA), glycerol (GI), and ethylene glycol (EG) were purchased from Xilong Scientific, Guangdong, China. Analytical grade EDTA and AgCl, AlCl₃, BaCl₂, FeCl₃, CdCl₂, Co(NO3)₂, CrCl₃, CuCl₂, FeCl₃, Hg(NO₃)₂, MgCl₂, MnCl₂, Ni(NO₃)₂, Zn(NO₃)₂, and $(CH_3COO)_2$ Pb were also purchased from Xilong Chemical, Guangdong, China. All chemicals in our work were used strictly according to regulations. All experiments used ultrapure water (UP water). PEG-200 was used in all experiments except those in which the type of poly(ethylene glycol) was specified. Blood sample (provided by the inspection department of Guilin University of Technology Hospital) was centrifuged at high speed (3500 r/min) for 10 min, and serum was collected for $Fe³⁺$ analysis.

Synthesis of GQDs and PEG-GQDs. Typically, GQDs were prepared by the hydrothermal method. Cane molasses (1.8 mL) was dissolved in 28.2 mL of ultrapure water to form a homogeneous aqueous solution and then ultrasonicated and centrifuged, each for 10 min, respectively. Twenty milliliters of cane molasses supernatant was then transferred to a 30 mL steel high-pressure reactor and heated in a hot-air oven at 190 °C for 24 h. After completion of the hydrothermal process, the reactor was allowed to cool down, and the obtained carbon slag residue was separated from the initial product by filtration using a 0.2 μ m filter paper to acquire GQDs primary liquid. GQDs solution was then obtained by diluting the GQDs primary liquid with UP water for 50 times. PEG-GQDs solution with high-fluorescence QY for metal ions detection was produced by diluting the GQDs primary liquid with PEG-200 for 50 times.

Characterization Methods. Various physicochemical techniques were used to determine the surface morphology, chemical component, and optical properties of the synthesized GQDs and PEG-GQDs. Transmission electron microscopy (TEM) was executed on a JEM-2100F TEM at 200 kV. Atomic force microscopy (AFM) observation was executed with Ntegra Prima SPM. Fourier transform infrared (FTIR) spectra were collected using a Thermo Nexus 470FT-IR spectrometer (KBr disk). X-ray photoelectron spectroscopy (XPS) was carried out using ESCALAB250Xi. The fluorescence lifetime of the synthetic samples was collected using an FLS980 fluorometer. UV−vis absorption and fluorescence spectra were recorded by a UV-3600 UV−vis−NIR spectrophotometer and a VARIAN Cary 100 fluorospectrophotometer (the voltage is set to 550 V and the slit widths of excitation and emission are both set to 5 nm). Fluorescence quantum yield (QY) was detected using quinine sulfate as the standard (QY = 54% in 0.1 M H_2SO_4). The ζ -potential of GQDs and PEG-GQDs was collected on a Malvern Zetasizer Nano ZS (ZS 90, UK).

Fluorescence Detection of Metal Ions. In the experiment of sensing capability of PEG-GQDs toward metal ions, 100 $μ$ L GQDs primary liquid was mixed with 200 $μ$ L of metal ions $(Cr^{3+}$, Ca^{2+} , Al^{3+} , Cu^{2+} , Pb^{2+} , Mn^{2+} , Ag^+ , Ba^{2+} , Co^{2+} , Cd^{2+} , $\rm Zn^{2+},$ $\rm Hg^{2+},$ $\rm Fe^{3+},$ $\rm Mg^{2+},$ $\rm Ni^{2+})$ solutions (0.01 M), respectively. Then, the solution was fixed to 5 mL with PEG as the diluent with vigorous shaking at room temperature. Fluorescence emission spectra of each mixture solution were collected with an excitation wavelength at 376 nm to verify the responses of the above ions to the fluorescence of PEG-GQDs after 10 min. As a reference, the sensing capability of GQDs toward $Fe³⁺$ was also investigated using UP water as the diluent following the same procedure.

For the masking experiment phase, the difference from the experiment of sensing capability of PEG-GQDs and GQDs toward metal ions is the addition of 200 μ L of EDTA (0.1 M) to prove the masking effect before adding different metal ions. One hundred microliters of healthy human serum was diluted to 5 mL with PEG after adding 200 μ L of EDTA (0.1 M) at room temperature to quantify the amount of $Fe³⁺$ by the standard addition method, which means that a certain amount of $Fe³⁺$ is added to the serum. More detailed experimental procedures can be seen in the [Supporting Information.](http://pubs.acs.org/doi/suppl/10.1021/acsomega.0c00098/suppl_file/ao0c00098_si_001.pdf)

■ ASSOCIATED CONTENT

³ Supporting Information

The Supporting Information is available free of charge at [https://pubs.acs.org/doi/10.1021/acsomega.0c00098.](https://pubs.acs.org/doi/10.1021/acsomega.0c00098?goto=supporting-info)

> GQDs fluorescence probe for the detection of metal ions at different conditions; optimum synthesis conditions of GQDs; excitation wavelength dependence of GQDs, the degree of fluorescence quenching varies with the concentration of Cu^{2+} , Co^{2+} , Ni^{2+} , Mn^{2+} , and Pb^{2+} for PEG-GQDs; fluorescence decay curve of the GQDs, PEG-GQDs, and PEG-GQDs in the presence of Fe³⁺, UV−vis absorption spectra of PEG-GQDs in the presence of Fe³⁺, and the superposition of the UV–vis absorption spectra of PEG-GQDs and Fe³⁺ solution ([PDF](http://pubs.acs.org/doi/suppl/10.1021/acsomega.0c00098/suppl_file/ao0c00098_si_001.pdf))

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Notes

The authors declare no competing financial interest.

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