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A common *CHRNE* mutation in Brazilian patients with congenital myasthenic syndrome

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Abstract

The most common causes of congenital myasthenic syndromes (CMS) are *CHRNE* mutations, and some pathogenic allelic variants in this gene are especially frequent in certain ethnic groups. In the southern region of Brazil, a study found the c.130dupG *CHRNE* mutation in up to 33% of families with CMS. Here, we aimed to verify the frequency of this mutation among individuals with CMS in a larger cohort of CMS patients from different areas of Brazil and to characterize clinical features of these patients. Eighty-four patients with CMS, from 72 families, were clinically evaluated and submitted to direct sequencing of the exon 2 of *CHRNE*. The c.130dupG mutation was found in 32 patients (23 families), with 26 patients (19 families, 26.3%) in homozygosis, confirming its high prevalence in different regions of Brazil. Among the homozygous patients, the following characteristics were frequent: onset of symptoms before 2 years of age (92.3%), little functional restriction (92.3%), fluctuating symptoms (100%), ocular muscle impairment (96.1%), ptosis (100%), limb weakness (88.4%), response to pyridostigmine (100%), facial involvement (77%), and bulbar symptoms (70.8%). The pretest probability of finding at least one allele

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Ethical Standards

All human studies have been approved by the local ethics committee and have therefore been performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki.

Conflicts of interest

The authors declare that they have no conflict of interest.

harbouring the c.130dupG mutation was 38.1%. Selecting only patients with impaired eye movement together with limb weakness and improvement with pyridostigmine, the probability increases to 72.2%. This clinical pre-selection of patients is likely a useful tool for regions where *CHRNE* mutations have a founder effect. In conclusion, the *CHRNE* mutation c.130dupG leads to fairly benign natural course of the disease with relative homogeneity.

Keywords

Congenital Myasthenic Syndrome; Neuromuscular Junction; *CHRNE*; Acetylcholine receptor; pyridostigmine

Introduction

Congenital myasthenic syndromes (CMS) are a group of genetic disorders in which the safety factor of the neuromuscular transmission is impaired. The phenotype is variable, making diagnosis difficult [1,2]. The increasing number of CMS causative genes adds to the difficulty of establishing the correct molecular diagnosis, and knowing the exact type of CMS has implications for appropriate care of the patients [1,3]. However, *CHRNE* mutations account for up to 50% of all CMS [3,4]. Other genes that more commonly lead to CMS are *DOK7*, *COLQ*, and *RAPSN* [5], but the frequency of the affected genes varies according to the studied geographic region and population [6,7].

Although there are several *CHRNE* mutations described, some pathogenic allelic variants are especially frequent in certain ethnic groups [8,9]. Nevertheless, in these cases the diagnosis can also be difficult due phenotypic variability, as some *CHRNE* mutations can present a high clinical heterogeneity, even when present in homozygous state [10]. In Brazil, a study with patients with CMS from the state of Paraná (southern part of the country) found the c.130dupG mutation in the *CHRNE* gene present in 6 out of 18 families [6]. Those patients frequently had ophthalmoplegia, palpebral ptosis, dysphagia, and proximal muscle weakness with onset before 2 years old, which are expected characteristics, considering the affected gene [1,2,3]. In the present study we aimed to verify the frequency of this mutation among individuals with CMS from different areas of Brazil and to characterize clinical features of these patients.

Patients and Methods

Eighty-four patients with CMS (49 females, 35 males) from 72 unrelated families, aged from 3 to 65 years at the time of clinical evaluation, were recruited. The parents or the patients were invited to participate in the study after informed consent was obtained, and the study was approved by the local ethics committee. The patients were evaluated at our centre in São Paulo, Hospital das Clínicas/USP (n = 61); in Salvador, at Hospital das Clínicas/UFBA (n = 2); and in Ribeirão Preto, at Hospital das Clínicas/USPRP (n = 21).

Patients were evaluated by an experienced neurologist. Data included medical history and general physical and neurological examinations. Results of ancillary exams such as muscle biopsy, nerve conduction studies and electromyography (NCS/EMG), when available, were

noted. To detect the mutation c.130dupG in *CHRNE* (NM_000080.3), a fragment containing whole exon 2 was amplified by PCR for Sanger sequencing. Patients heterozygous for the c.130dupG mutation were screened for mutations in the whole coding *CHRNE* gene using primers covering all exons and splice junctions.

Clinical findings were compared only between the groups of homozygous patients for c.130dupG and the group without the mutation, with the objective to identify clinical clues for the c.130dupG mutation. Fisher's exact test was used for the comparison.

Results

Average age at assessment of all CMS patients was 24.6 years and 24.15 years among the homozygous *CHRNE* c.130dupG patients. The *CHRNE* mutation was found in 32 patients (38.1% of the patients) from 23 different families (31.9% of the families) including 26 patients (19 families) in homozygous state (table 1). From 6 heterozygous patients, another pathogenic mutation was found in 5 (table 1). Families had segregation analysis performed, confirming compound heterozygous status. Clinical data of patients carrying the *CHRNE* mutation are shown in table 1.

Among the homozygous c.130dupG mutation patients ($n = 26$), the following characteristics were very frequent: onset of symptoms before 2 years of age (92.3%); fluctuating symptoms (100%); ocular muscle impairment (96.1%); ptosis (100%); improvement with pyridostigmine (100%); limb weakness (88.4%); facial involvement (77%); and bulbar symptoms (70.8%). Delayed motor milestones were observed in 56.5% of the homozygous c.130dupG patients, neonatal hypotonia in 54.1%, breastfeeding difficulties in 55%, high arched palate in 66.6%, and scoliosis in 38.4%. No *CHRNE* c.130dupG patients presented with any congenital contractures. Twenty-two out of the 26 homozygous patients had available data of Electromyography/Nerve conduction study: 15 (68.1%) showed decremental response to 3 Hz repetitive stimulus (DRRS). Of the seven patients without DRRS, three were submitted to single fibre study, which demonstrated increased jitter in all.

Interestingly, of the CMS patients with ocular movement impairment, most of them presented with partial, instead of complete, impairment. That was the case in 88% of the patients homozygous for the *CHRNE* mutation and 87% of the patients without the mutation, considering patients with ocular movement impairment. In particular, the rectus inferior muscles were the most frequently spared, being not affected or relatively less affected in 73.9% and 76.4% of the patients with and without the c.130dupG mutation, respectively (figure 1).

Comparing the patients with homozygous c.130dupG mutations to those without, the following clinical findings were statistically more frequent in the homozygous c.130dupG mutation group: impaired eye movement ($p = 0.0002$), facial involvement ($p = 0.01$), limb weakness ($p = 0.03$), and good response to pyridostigmine ($p = 0.0001$), type II selective atrophy on muscle biopsy ($p = 0.0089$). Frequency of clinical characteristics of all patients are shown on table 2.

The pretest probability among our cohort of finding at least one allele harbouring the *CHRNE* c.130dupG mutation was 38.1%. Selecting only patients with impaired eye movement who improved with pyridostigmine (n = 47), the probability of finding at least one allele mutated increased to 61.7% (29/47, $p = 0.0001$). In patients with impaired eye movement and limb weakness who improved with pyridostigmine (n = 36), the probability of finding at least one allele with the mutation was 72.2% (26/36, $p = 0.0001$).

Regarding the distribution of the mutation in the country, 27 patients (22 homozygous) of 19 families were from the state of São Paulo (33.9% of São Paulo families), 4 patients (3 homozygous) of 4 families were from the state of Minas Gerais (75% of Minas Gerais families), and 1 patient (homozygous) was from the state of Bahia (incidence of 20% of Bahia families).

Discussion

CHRNE c.130dupG (p.Glu44Glyfs*3) is a known pathogenic variant [13], with an allele frequency on GenomAD of 0.00010, that leads to a lack of adult form acetylcholine receptor (AChR) on the post-synaptic membrane [14,15] and consequently impairs neuromuscular transmission. This mutation has been repeatedly identified in patients with Spain and Portugal origin [16], and was present in 6.4% of CMS patients on a Spanish cohort [17]. Mihaylova et al. showed a higher prevalence of this mutation in CMS Brazilian patients from the state of Paraná [6]. In our cohort, the mutation was found among patients from other regions of the country, confirming the high prevalence of the mutation across different regions of Brazil.

We found a pretest probability of the c.130dupG mutation of 31.9% in at least one allele of CMS patients, and when considering only homozygous patients the percentage is still high (26.4%). The percentages notably increase if we consider only patients with impaired eye movement and improvement of symptoms with pyridostigmine. This may be driven by the likely exclusion of *RAPSN* and *DOK7* cases by this selection, as these genetic entities are more likely to not have those clinical features. In addition, for patients with impaired eye movement as well as with limb weakness and improvement with pyridostigmine, the percentage of patients with the *CHRNE* c.130dupG mutation is even higher (72.2% and 58.3% for one and two alleles). This extra effect can be explained by the exclusion of patients with pure ocular symptoms, which seem to have a low probability to be genetically solved [4].

The clinical characteristics of the c.130dupG patients represent a good prognosis, and are quite similar to those bearing other *CHRNE* mutations that lead to AChR deficiency [8,15] although some clinical features seem to be strongly linked to this specific mutation. In general, the clinical presentation of the homozygous *CHRNE* c.130dupG patients was fairly homogenous, contrary to the great clinical variability shown in other CMS syndromes with other homozygous mutations [10, 18,19]. One possible explanation for this difference could be the fact that the c.130dupG mutation occurs in an early portion of the expressing gene, likely resulting in a null mutation. This might lead to stronger stimulus for compensating factors compared to, for instance, more C-terminal frameshift mutations, which may result

in expression of dysfunctional proteins. Therefore, in this case, patients with dysfunctional epsilon subunit would depend on other mechanisms to trigger compensating factors (which in turn might lead to more variable compensation) in comparison with patients with no epsilon subunit at all.

Interestingly, most of the patients presented with partial instead of complete ocular movement impairment, a characteristic not explored in other clinical studies. The ocular muscle most frequently spared was the rectus inferior, which might indicate that it tends to be the latest affected, once virtually every ocular muscle tends to be eventually paretic. This finding was similar in the groups with and without the c.130dupG mutation, indicating it is probably a general CMS feature rather than specific for *CHRNE*. It would be interesting to compare this feature with other diseases, and see, for instance, if it can help to clinically differentiate CMS from mitochondrial myopathy and congenital myopathy, which sometimes can be difficult to do in clinical practice [20]. Regarding therapeutics, we verified a high rate of response to anticholinesterase medication and an additional benefit to β 2-agonists among the homozygous *CHRNE* c.130dupG patients. Most likely, this may not be specific for the c.130dupG mutation, as β 2-agonists were shown to be beneficial to patients with AChR deficiency as an add-on therapy [21].

The mutation c.130dupG in the *CHRNE* gene seems to have a high prevalence in different regions of Brazil, and to lead to expected *CHRNE*-related characteristics, with relative homogeneity and a fairly benign natural course of the disease. Careful selection of patients with a few simple clinical features can lead to a percentage as high as 72.2% of patients with the mutation, which could be a useful tool for the first screening of CMS patients, including in different countries where other *CHRNE* mutations also have a founder effect.

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References

1. Engel A, Shen X-M, Secen D, Sine S. Congenital myasthenic syndromes: pathogenesis, diagnosis, and treatment. *Lancet Neurology*. 2015; 14:421–434.
2. Finlayson S, Beeson D, Palace J. Congenital myasthenic syndromes: an update. *Practical Neurology*. 2013; 13:80–91. [PubMed: 23468559]
3. McMacken G, Abicht A, Evangelista T, Spendiff S, Lochmüller H. The Increasing Genetic and Phenotypical Diversity of Congenital Myasthenic Syndromes. *Neuropediatrics*. 2017; 48:294–308. [PubMed: 28505670]
4. Abicht A, Dusl M, Gallenmüller C, et al. Congenital myasthenic syndromes: achievements and limitations of phenotype-guided gene-after-gene sequencing in diagnostic practice: a study of 680 patients. *Human Mutation*. 2012; 33:1474–1484. [PubMed: 22678886]
5. Abicht, A, Müller, JS, Lochmüller, H. Congenital Myasthenic Syndromes GeneReviews®. Adam, MP, Ardinger, HH, Pagon, RA. , et al., editors. Seattle (WA): University of Washington, Seattle; 2003. May 9, 1993-2017. [Updated 2016 Jul 14]. [Internet] [accessed 10/10/2017]
6. Mihaylova V, Scola RH, Gervini B, et al. Molecular characterisation of congenital myasthenic syndromes in Southern Brazil. *Journal of Neurology Neurosurgery and Psychiatry*. 2010; 81:973–977.

7. Aharoni S, Sadeh M, Shapira Y, et al. Congenital myasthenic syndrome in Israel: Genetic and clinical characterization. *Neuromuscular Disorders*. 2017; 27:136–140. [PubMed: 28024842]
8. Abicht A, Stucka R, Karcagi V, et al. A common mutation (epsilon1267delG) in congenital myasthenic patients of Gypsy ethnic origin. *Neurology*. 1999; 53:1564–1569. [PubMed: 10534268]
9. Beeson D, Hantai D, Lochmuller H, Engel AG. 126th International Workshop: congenital myasthenic syndromes, 24-26 September 2004, Naarden, the Netherlands. *Neuromuscular Disorders*. 2005; 15:498–512. [PubMed: 15951177]
10. Natera-de Benito D, Domínguez-Carral J, Muelas N, Nascimento A, Ortez C, Jaijo T, Arteaga R, Colomer J, Vilchez JJ. Phenotypic heterogeneity in two large Roma families with a congenital myasthenic syndrome due to CHRNE 1267delG mutation. A long-term follow-up. *Neuromuscul Disord*. 2016; 26(11):789–795. [PubMed: 27634344]
11. Barisic N, Schmidt C, Sidorova OP, et al. Congenital myasthenic syndrome (CMS) in three European kinships due to a novel splice mutation (IVS7-2A/G) in the epsilon acetylcholine receptor (AChR) subunit gene. *Neuropediatrics*. 2002; 33:249–254. [PubMed: 12536367]
12. Ohno K, Wang HL, Milone M, et al. Congenital myasthenic syndrome caused by decreased agonist binding affinity due to a mutation in the acetylcholine receptor epsilon subunit. *Neuron*. 1996; 17:157–70. [PubMed: 8755487]
13. Ohno K, Anlar B, Ozdirim E, Brengman JM, DeBleecker JL, Engel AG. Myasthenic Syndromes in Turkish Kinships Due to Mutations in the Acetylcholine Receptor. *Annals of Neurology*. 1998; 44:234–41. [PubMed: 9708546]
14. Croxson R, Vincent A, Newsom-Davis J, Beeson D. Myasthenia gravis in a woman with congenital AChR deficiency due to epsilon-subunit mutations. *Neurology*. 2002; 58:1563–1565. [PubMed: 12034803]
15. Burke G, Cossins J, Maxwell S, et al. Distinct phenotypes of congenital acetylcholine receptor deficiency. *Neuromuscular Disorders*. 2004; 14:356–364. [PubMed: 15145336]
16. Beeson D, Hantai D, Lochmüller H, Engel AG. 126th International Workshop: congenital myasthenic syndromes, 24-26 September 2004, Naarden, the Netherlands. *Neuromuscul Disord*. 2005; 15(7):498–512. [PubMed: 15951177]
17. Natera-de Benito D, Töpf A, Vilchez JJ, González-Quereda L, Domínguez-Carral J, Díaz-Manera J, Ortez C, Bestué M, Gallano P, Dusl M, Abicht A, et al. Molecular characterization of congenital myasthenic syndromes in Spain. *Neuromuscular Disorders*. 2017; 27:1087–1098. [PubMed: 29054425]
18. Mihaylova V, Müller JS, Vilchez JJ, et al. Clinical and molecular genetic findings in COLQ-mutant congenital myasthenic syndromes. *Brain*. 2008; 131:747–759. [PubMed: 18180250]
19. Müller JS, Mildner G, Müller-Felber W, et al. Rapsyn N88K is a frequent cause of congenital myasthenic syndromes in European patients. *Neurology*. 2003; 60:1805–1810. [PubMed: 12796535]
20. Neto, O Abath; Heise, CO; Moreno, CA; , et al. Nonlethal CHRNA1-Related Congenital Myasthenic Syndrome with a Homozygous Null Mutation. *The Canadian Journal of Neurological Sciences*. 2017; 44:125–127. [PubMed: 27748205]
21. Rodríguez Cruz PM, Palace J, Ramjattan H, Jayawant S, Robb SA, Beeson D. Salbutamol and ephedrine in the treatment of severe AChR deficiency syndromes. *Neurology*. 2015; 85:1043–1047. [PubMed: 26296515]

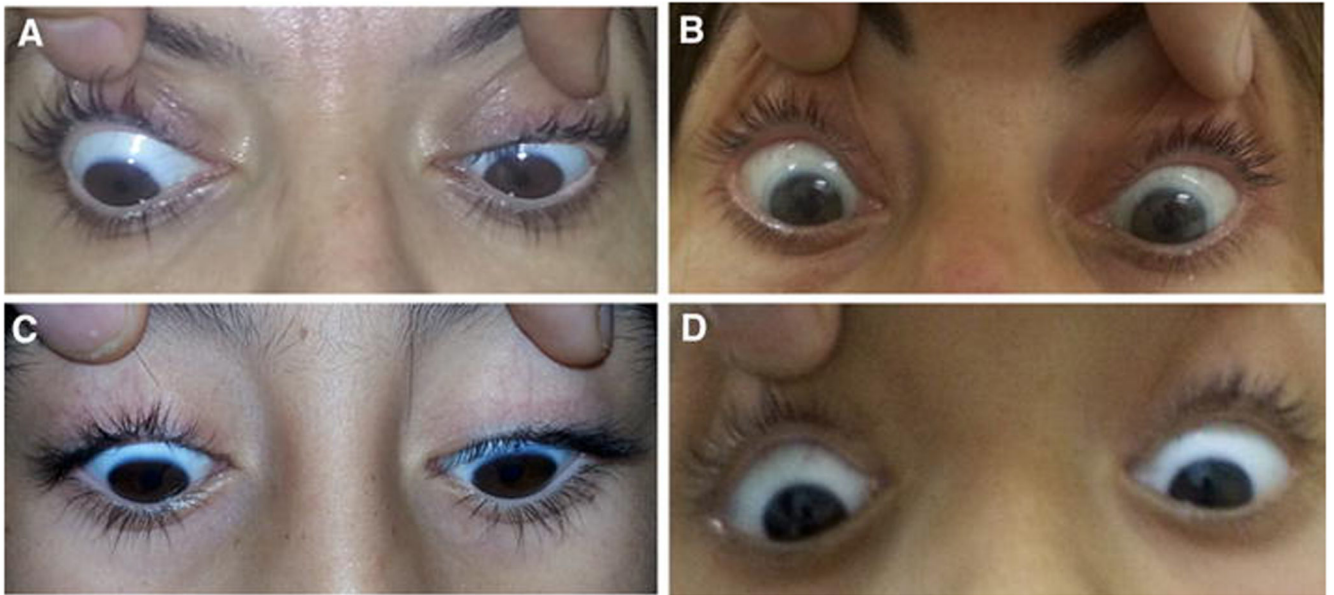


Figure. Relative sparing of the muscle rectus inferior among patients with partial ophthalmoparesis.

Patients presented with different levels of asymmetrical impairment of ocular movements, which are not related to age. A. Patient 6 (28 years old). B. Patient 20 (37 years old). C. Patient 42 (8 years old). D. Patient 21 (10 years old)

Table 1

Main clinical data from patients with the *CHRNE* c.130dupG mutation + = present; - = absent; nd = no data or not tested; EO = early onset; IP = improvement with pyridostigmine; FS = fluctuating symptoms; IEM = impaired eye movement; PIE = partially impaired eye movement; SRI= impaired eye movement sparing rectus inferior muscles; PP = ptosis; LW = limb weakness; BS = bulbar symptoms; FI = facial involvement; NPC = no progressive course; Variants found associated with c.130dupG: c.803-2A>G (previously reported as pathogenic [11]); c.858_859dup (p.leu287ser fs*14, novel); c.422T>A (p.pro141.leu, previously reported as pathogenic [12]).

Patient / Family	Age (years)/Sex	EO	IP	FS	IEM/ PIE/ SRI	PP	LW	BS	FI	NPC	Mutation state
1/1	30/M	+	+	+	+/+/+	+	+	-	+	+	c.130dupG/c.130dupG
2/1	29/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
3/1	12/M	+	+	+	+/+/+	+	+	+	-	+	c.130dupG/c.130dupG
6/3	28/F	+	+	+	+/+/+	+	-	-	-	+	c.130dupG/c.130dupG
7/3	19/F	+	+	+	+/-/-	+	+	-	-	+	c.130dupG/c.130dupG
8/4	13/F	+	+	+	+/+/-	+	+	+	-	+	c.130dupG/c.130dupG
9/5	32/M	+	+	+	+/+/-	+	+	+	+	+	c.130dupG/-
18/14	12/M	+	+	+	+/+/-	+	+	+	+	+	c.130dupG/c.130dupG
20/16	37/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
21/17	10/M	+	+	+	+/+/+	+	+	+	+	-	c.130dupG/c.130dupG
25/21	25/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
27/23	4/F	+	+	+	+/-/-	+	-	+	+	+	c.130dupG/c.130dupG
29/25	34/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
34/29	14/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
40/34	64/M	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.803-2A>G
41/34	54/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.803-2A>G
42/35	8/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
43/36	16/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.858_859dup
44/36	7/M	+	+	+	+/+/+	+	-	-	+	+	c.130dupG/c.858_859dup
49/41	38/F	+	+	+	+/+/+	+	+	-	+	+	c.130dupG/c.130dupG
50/42	21/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
51/42	28/F	+	+	+	+/+/+	+	+	+	+	+	c.130dupG/c.130dupG
60/51	43/F	+	+	+	-/-/-	+	+	+	-	+	c.130dupG/c.130dupG
64/55	34/M	-	+	+	+/nd/nd	+	+	-	+	+	c.130dupG/c.130dupG
65/55	38/F	+	+	+	+/nd/nd	+	+	+	+	-	c.130dupG/c.130dupG
66/56	18/F	+	+	+	+/nd/nd	+	+	-	+	+	c.130dupG/c.130dupG
67/57	27/F	+	+	+	+/nd/nd	+	+	+	+	+	c.130dupG/c.130dupG
68/58	34/M	+	+	+	+/nd/nd	+	+	+	+	+	c.130dupG/c.130dupG
69/58	19/M	+	+	+	+/nd/nd	+	+	-	-	+	c.130dupG/c.130dupG
70/59	19/F	+	nd	+	+/nd/nd	+	-	nd	+	+	c.130dupG/c.130dupG
71/59	34/M	-	nd	+	+/nd/nd	+	+	nd	+	+	c.130dupG/c.130dupG

Patient / Family	Age (years)/Sex	EO	IP	FS	IEM/ PIE/ SRI	PP	LW	BS	FI	NPC	Mutation state
72/60	5/F	+	+	+	+/nd/nd	+	+	+	+	+	c.130dupG/c.422T>A

Table 2

Frequency of clinical features of patients with Congenital Myasthenic Syndrome

Clinical Features	Frequency (%) in the group with homozygous c.130dupG mutation	Frequency (%) in the group without c.130dupG mutation
Early onset of symptoms	92.3	75
Fuctuation of symptoms	100	86.5
Ocular muscle impairment/ partial /sparing muscle rectus inferior	96.1 / 83.3 / 72.2	55.7 / 43.4 / 36.9
Eyelid ptosis	100	86.5
Limb weakness	88.4	63.4
Bulbar symptoms	70.8	50
Facial weakness	77	52
Little functional restriction	92.6	80.7
Exacerbations crisis: At least one episode/more than 2 per year	30.7 / 7.7	21.1 / 7.6
Delayed motor milestones	56.5	53.8
Neonatal hypotonia	54.1	50
High arched palate	66.6	59.6
Breastfeeding difficulties	55	27.5
Scoliosis	38.4	28.8
Congenital Contractures	0	15
Response to pyridostigmine	100	54.1
Response to adding β 2-agonists: Ephedrine/salbutamol	100 / 100	100 / 100
with type II selective atrophy on muscle biopsy	66.6	20
Electromyography and nerve conduction study: decremental response to repetitive stimulus / single fibre study whith increased jitter	68.1 / 100	48.9 / 54.5