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Catalytic Deuterium Incorporation within Metabolically Stable β -Amino C—H Bonds of Drug Molecules

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Abstract

An efficient deuteration process of β -amino C—H bonds in various *N*-alkylamine-based pharmaceutical compounds has been developed. Catalytic reactions begin with the action of Lewis acidic B(C₆F₅)₃ and Brønsted basic *N*-alkylamine, converting a drug molecule into the corresponding enamine. The acid/base catalysts also promote the dedeuteration of acetone-*d*₆ to afford a deuterated ammonium ion. Ensuing deuteration of the enamine then leads to the formation of β -deuterated bioactive amines with up to 99% deuterium incorporation.

Graphical Abstract



Deuterium-labeled pharmaceuticals are pivotal diagnostic tools in research aimed at determination of the corresponding biological outcomes and metabolites.¹⁻⁶ Drugs containing C—D bonds have been prepared through multistep synthesis involving the reduction of unsaturated or halogenated intermediates.³ However, innovations in organometal-catalyzed C—H activation have enabled direct hydrogen isotope exchange (HIE) at C—H bonds for deuterium.⁴⁻⁶ In particular, HIE reaction targeting C(sp³)—H bonds of pharmaceuticals that contain an *N*-alkylamine unit is in high demand because these

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entities constitute over 50% of the top-selling commercial drugs.⁷ The state-of-the-art processes include Beller's *a*- and *β*-amino C—H deuteration of metoclopramide (**A1**) and two other structurally related drug molecules promoted by Ru-based Shvo catalyst (Figure 1A).⁵ MacMillan's photoredox-mediated *a*-deuteration and *a*-tritiation represents a notable strategy for isotopic labelling of a range of *N*-alkylamine-based drugs (Figure 1B).⁶ Still, development of methods for regioselective deuteration of poorly reactive *β*-amino C(sp³) —H bonds of drugs containing Lewis acid- and base-sensitive functional groups with an inexpensive deuterium source and promoted by non-precious metal-based catalysts is a significant challenge.^{8,9} Regioselective deuteration of metabolically stable *β*-amino C—H bonds (vs more labile *a*-amino C—H bonds) is particularly attractive as it minimizes the loss of the label due to exchange.¹⁻³

We began by contemplating a possible way to design a method for deuteration of biologically active compounds that contain an *N*-alkylamine unit (1) with readily available acetone- d_6 **2** as a source of deuterium (Figure 1C). We considered utilizing a combination of Lewis acid and Brønsted base catalysts that would function cooperatively.¹⁰⁻¹² We envisioned that B(C₆F₅)₃ could receive a hydride from an amine (1), generating a borohydride and an iminium ion (I).¹³⁻¹⁹ Subsequently, a Brønsted basic amine catalyst would deprotonate I, furnishing enamine II.¹³⁻¹⁶ Concurrently, the *N*-alkylamine could dedeuterate B(C₆F₅)₃-activated acetone- d_6 **2**, generating an enolate and a deuterated ammonium ion (III).²⁰ Ensuing deuteration (IV) of the enamine II by III gives an iminium ion; subsequent borohydride reduction would afford β -deuterated product **3**. Here, we report the development of a catalyst system for β -amino C—H deuteration of bioactive amines.

We first set out to identify a desirable combination of catalysts. We probed the ability of $B(C_6F_5)_3$ and various Brønsted bases to catalyze the reaction between verapamil 1a and acetone-d₆ 2 (6.8 equivalent), generating 3a (Table 1). Treatment of 1a and 2 with 5.0 mol% B(C₆F₅)₃ and 10 mol% NEt₃, NBn₃, or 1,2,2, 6,6-pentamethylpiperidine (PMP) afforded **3a** in >90% yield (toluene, 125 °C, 1 h); 16-34% of *β*-amino C—H bonds were converted to C-D bonds (entries 1-3). With more Brønsted basic 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), no labeling was observed (entry 4). When the transformation was performed without a Brønsted base co-catalyst, 3a was generated with 35% and 21% deuterium incorporation (entry 5), suggesting that N-alkylamines 1a and/or 3a can promote deprotonation of the iminium ion (I, Figure 1C; $NR_3 = 1a$ and/or 3a). Deuterium incorporation diminished to <10% with 5.0 mol% of B(C_6F_5)₃ and reaction temperature of 100 °C (entry 6), but when the reaction mixture was heated at 150 °C, **3a** was obtained with >80% labeling (entry 7). With 10 mol% $B(C_6F_5)_3$ there was only minor improvement (entry 8, 88% and 92%). However, by reacting **1a** with two batches of 5.0 mol% of $B(C_6F_5)_3$ and **2** (6.8 equivalent), we were able to obtain 3a with >95% deuterium incorporation (entry 9). There was no labeling without $B(C_6F_5)_3$ or when the less hindered BF_3 or the less Lewis acidic BPh₃ were used (entries 10–12). These findings support the notion that strongly acidic $B(C_6F_5)_3$ together with sterically demanding and electron-rich N-alkylamine constitute the most effective combination.¹⁰

Acyclic β -amino C—H bonds in a number of pharmaceuticals (**1a–1j**) underwent efficient deuteration (Table 2). This protocol was found to be compatible with compounds that

contain an array of Lewis acid-sensitive functional groups. In addition to the *N*-alkylamine units of **1a–1j**, cyano (**1a**), ester (**1b**), amide (**1d**, **1e**, **1f**) and ketone (**1j**) were tolerated to give the deuteration products **3a–3j** in 77 to >95% yield after purification by silica gel chromatography. Labeling took place with high regioselectivity for β -amino C—H bonds. In addition, drug molecules that possess acidic *a*-carbonyl C—H bonds also underwent efficient deuteration (**3d**, **3j**) based on analysis of ¹H NMR spectra of unpurified mixtures. Nonetheless, in case of **3d**, *a*-carbonyl C—D bonds of indolin-2-one underwent H—D exchange during purification.

For substrates that possess electronically and sterically disparate β -amino C—H bonds (1a, 1b, 1c, 1d, 1g, 1j), deuterium labeling occurred at varying levels. With verapamil 1a, benzylic C2—H and non-benzylic C4—H bonds were converted to C—D bonds in >95%, but deuteration of non-benzylic C4—H bonds was more efficient (Table 1, entries 5-9). With dicyclomine 1b, while 90% of C2'—H bonds of *N*-ethyl groups was deuterated, only 23% of C1—H bonds adjacent to an ester group were converted to C—D bonds. Similar reactivity was observed with clomiphene 1c: 90% of C2'—H bonds were converted into C2' —D bonds, but 15% of *a*-aryloxy C1—H bonds were deuterated. Ropinirole 1d was labeled at benzylic C1—H bonds (63%), and C2'—H bonds of *N*-propyl group (86%).

Although the catalytic protocol tolerates an array of functional groups, isotopic labeling was more efficient with substrates bearing a protecting group. For instance, 80% of β -C—H bonds of lidocaine **1e**, which possesses acidic amide N—H bonds, was converted to C—D bonds to give **3e**. However, deuteration of *N*-benzyl-protected lidocaine **1f** proceeded more efficiently to afford **3f** with 96% d-incorporation; in addition, deuteration of *a*-carbonyl C—H bonds was also observed (9%). Cinacalcet **1g** containing a secondary amine moiety was a compatible substrate to provide **3g** (63% [C2] and 8% [C2']), and *N*-benzyl-protected cinacalcet **3h** was obtained with >98% of β -amino C2—H bonds selectively converted into C—D bonds. With less sterically hindered secondary amines nortriptyline **1i** and propafenone **1j**, their reaction with acetone-*d*₆ may inhibit labeling. However, with an *N*-benzhydryl group installed, **3i** and **3j** could be readily generated. Silyl protection of the secondary alcohol proved to be effective in the case of propafenone **1j**, giving **3j** (76% [C2] and 0% at more sterically hindered [C2']).

Next, we investigated possible labeling of various pharmaceuticals that contain cyclic β amino C—H bonds (Table 3; 1k–1s). A variety of Lewis acid-sensitive heterocycles such as piperidine (1k–1q), 1,4-diazepane (1r), piperazine (1s), thiophene (1k, 1l), indanone (1m), benzodioxole (1o, 1p), benzothiophene (1q), as well as benzoimidazole (1r) were tolerated to give the corresponding deuteration products in 85 to >95% yield. With clopidogrel 1k, prasugrel 1l, and donepezil 1m, both β -amino C—H bonds and enolizable *a*-carbonyl C—H bonds underwent efficient deuteration to give 3k–3m, but acidic *a*-keto C—D bond of 3l was converted to C—H bond during purification. Less acidic *a*-amide C—H bond of bupivacaine 1n was not deuterated. With bupivacaine 1n and raloxifene 1q that contain acyclic and cyclic β -amino C—H bonds, labeling of the cyclic C—H bond was more efficient (>90% vs 29% for the acyclic C—H). *N*-Benzyl (1o) and *N*-benzhydryl (1p)protected paroxetine gave 3o and 3p, respectively. The level of labeling for the more

hindered **3p** (94%) was superior to **3o** (76%). Furthermore, deuteration of the C5—H bond occurred selectively, while the tertiary C3—H bond remained intact. Deuteration of emedastine **1r** was found to take place at C2—H (33%) and C6—H (60%) bonds. All eight C—H bonds of piperazine ring of *O*-TBS-protected dropropizine **1s** underwent deuteration to afford **3s** (>86%). Using this protocol, *a*-amino C—H deuteration occurred only when these bonds were also alpha to a carbonyl group (**3f**) or beta to a N atom (**3s**).

The method is scalable. Treatment of 1.4 g (3.0 mmol) of verapamil **1a** with 5.0 mol% $B(C_6F_5)_3$, 20 mmol of acetone- d_6 (toluene, 12 h, 150 °C), followed by filtration through a pad of silica gel and repeating the aforementioned procedure afforded **3a** in 95% yield (2.9 mmol, 1.3 g) and >93% deuterium incorporation (Scheme 1).

To summarize, we have designed an efficient and regioselective deuterium labeling of β amino C—H bonds in various bioactive molecules, provided that sufficient steric congestion is present around the reacting amine. By implementing the cooperative action of B(C₆F₅)₃ and *N*-alkylamine catalyst system, we show that it is possible to convert an *N*-alkylaminebased pharmaceutical compound to the corresponding enamine, and that the same catalyst system can generate a labeling agent from acetone- d_6 . The principles outlined herein, entailing conversion of amine containing drugs into enamines and its reaction with in situ generated electrophilic partner, provide a new rational framework for late-stage modification of a drug candidate. Studies along these lines are in progress.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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B: Deuteration and tritiation of α -amino C⁻H bonds by photoredox catalysts







Proposed catalytic cycle:







Scheme 1. Scale-up experiment

Table 1.

Evaluation of Reaction Parameters *a,b,c,d*



entry	Lewis acid	Brønsted base	temperature	d-incorporation (%)	
	(mol%)	(mol%)	(°C)	[C2]	[C4]
1	$B(C_6F_5)_3(5.0)$	NEt ₃ (10)	125	17	26
2	$B(C_6F_5)_3(5.0)$	NBn ₃ (10)	125	20	34
3	$B(C_6F_5)_3(5.0)$	PMP (10)	125	16	26
4	$B(C_6F_5)_3(5.0)$	DBU (10)	125	0	0
5	$B(C_6F_5)_3(5.0)$	none	125	21	35
6	$B(C_6F_5)_3(5.0)$	none	100	<5	7
7	$B(C_6F_5)_3(5.0)$	none	150	80	85
8	$B(C_6F_5)_3(10)$	none	150	88	92
9 ^{<i>c</i>}	B(C ₆ F ₅) ₃ (5.0 x 2)	none	150	95	>98
10	none	none	150	0	0
11	BF3•OEt2 (5.0)	none	150	0	0
12	BPh ₃ (5.0)	none	150	0	0

^aConditions: verapamil (1a, 0.1 mmol), acetone-d₆ (2, 0.68 mmol), organoborane, Brønsted base, toluene (0.4 mL), under N₂, 1 h.

 b Yield and deuterium incorporation level was determined by ¹H NMR analysis of unpurified reaction mixtures with mesitylene as the internal standard.

^cConditions: verapamil (1a, 0.2 mmol), acetone- d_6 (2, 1.4 mmol), B(C₆F₅)₃ (5.0 mol%), toluene (0.8 mL), under N₂, 150 °C, 1 h. Isolated and purified **3a** was reacted with acetone- d_6 (2, 1.4 mmol), B(C₆F₅)₃ (5.0 mol%), toluene (0.8 mL), under N₂, 150 °C, 1 h.

 $d_{\text{Green label indicates sites that are beta to N.}}$

Table 2.

Deuteration of Acyclic β -Amino C—H Bonds ^{*a,b,c*}



^aConditions: *N*-alkylamine (1, 0.2 mmol), acetone-*d*₆ (2, 1.36 mmol), B(C₆F₅)₃ (10 mol%), toluene (0.8 mL), under N₂, 150 °C, 3 h.

 b Yield of isolated and purified product. Deuterium incorporation level was determined by ¹H NMR analysis of the isolated and purified product.

^cGreen label indicates sites that are beta to N. Red label is used for any other sites that undergo deuteration.

 d Conditions: *N*-alkylamine (1, 0.2 mmol), acetone- d_{6} (2, 1.4 mmol), B(C₆F₅)₃ (5.0 mol%), toluene (0.8 mL), under N₂, 150 °C, 3 h. After the filtration of the crude reaction mixture through a pad of silica gel and removal of volatiles, acetone- d_{6} (2, 1.4 mmol), B(C₆F₅)₃ (5.0 mol%), and toluene (1.0 mL) were added under N₂, and then heated at 150 °C, 3 h.

^eThe reaction was carried out in two batches, using 10 mol% of B(C₆F₅)₃ in the first batch, and 5.0 mol% in the second. For details, see the SI.

Table 3.

Deuteration of Cyclic Amino C-H Bonds ^{a,b}



^aConditions: N-alkylamine (1, 0.2 mmol), acetone-d₆ (2, 1.36 mmol), B(C₆F₅)₃ (10 mol%), toluene (0.8 mL), under N₂, 150 °C, 3 h.

 b Yield of isolated and purified product. Deuterium incorporation level was determined by 1 H NMR analysis of the isolated and purified product.

 c Green label indicates sites that are beta to N. Red label is used for any other sites that undergo deuteration.

 d Conditions: *N*-alkylamine (1, 0.2 mmol), acetone- d_{6} (2, 1.4 mmol), B(C₆F₅)₃ (5.0 mol%), toluene (0.8 mL), under N₂, 150 °C, 3 h. After the filtration of the crude reaction mixture through a pad of silica gel and removal of volatiles, acetone- d_{6} (2, 1.4 mmol), B(C₆F₅)₃ (5.0 mol%), and toluene (1.0 mL) were added under N₂, and then heated at 150 °C, 3 h.

^e The reaction was carried out in two batches, using 10 mol% of B(C6F5)3 in the first batch, and 5.0 mol% in the second. For details, see the SI.