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Original article

Prevalence and clinical significance of respiratory viruses and bacteria detected in tuberculosis patients compared to household contact controls in Tanzania: a cohort study[☆]

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ABSTRACT

Objectives: To describe the prevalence of respiratory pathogens in tuberculosis (TB) patients and in their household contact controls, and to determine the clinical significance of respiratory pathogens in TB patients.

Methods: We studied 489 smear-positive adult TB patients and 305 household contact controls without TB with nasopharyngeal swab samples within an ongoing prospective cohort study in Dar es Salaam, Tanzania, between 2013 and 2015. We used multiplex real-time PCR to detect 16 respiratory viruses and seven bacterial pathogens from nasopharyngeal swabs.

Results: The median age of the study participants was 33 years; 61% (484/794) were men, and 21% (168/794) were HIV-positive. TB patients had a higher prevalence of HIV (28.6%; 140/489) than controls (9.2%; 28/305). Overall prevalence of respiratory viral pathogens was 20.4% (160/794; 95%CI 17.7–23.3%) and of bacterial pathogens 38.2% (303/794; 95%CI 34.9–41.6%). TB patients and controls did not differ in the prevalence of respiratory viruses (Odds Ratio [OR] 1.00, 95%CI 0.71–1.44), but respiratory bacteria were less frequently detected in TB patients (OR 0.70, 95%CI 0.53–0.94). TB patients with both respiratory viruses and respiratory bacteria were likely to have more severe disease (adjusted OR [aOR] 1.6, 95%CI 1.1–2.4; *p* 0.011). TB patients with respiratory viruses tended to have more frequent lung cavitations (aOR 1.6, 95%CI 0.93–2.7; *p* 0.089).

Conclusions: Respiratory viruses are common for both TB patients and household controls. TB patients may present with more severe TB disease, particularly when they are co-infected with both bacteria and viruses. **F. Mhimbira, Clin Microbiol Infect 2019;25:107.e1–107.e7**

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Introduction

Tuberculosis (TB) caused by the bacterium *Mycobacterium tuberculosis* affected an estimated 10.4 million new cases and caused 1.7 million deaths in 2016, making TB the leading global cause of death from an infectious disease [1]. Influenza pandemics have selectively caused higher mortality among persons with TB

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compared to the general population [2,3]. For instance, the 1918 influenza pandemic brought about a sharp decline in TB burden, possibly because of the higher mortality among TB patients co-infected with influenza viruses [3,4].

In sub-Saharan Africa, HIV has been the most important risk factor driving the TB epidemic in recent decades [5]. The efforts towards understanding other risk factors in TB—such as respiratory viruses, helminths [6], and bacteria [7]—are becoming increasingly important [8]. Evidence from experimental mouse models suggests that respiratory viruses such as influenza viruses may play a pathogenic role in individuals with tuberculous disease by negatively affecting immunity against *M. tuberculosis* [9]. The effect of respiratory viruses on TB may mimic the development of bacterial pneumonia immediately after an infection with respiratory viruses [10]. Studies of the lung microbiota (the community of microorganisms in the airways), which have focused on respiratory bacterial species populations among patients with and without TB, among new and recurrent TB patients, as well as among those in whom TB treatment has failed [7].

The differences in airway microorganism populations could indicate that respiratory pathogens can be involved in TB pathogenesis [11]. However, little is known about the prevalence of respiratory pathogens, whether viral or bacterial, and their role in clinical presentation in TB. We therefore studied the prevalence of respiratory pathogens in TB patients and household contact controls, and assessed the associations between both respiratory viruses and bacterial pathogens and the clinical presentation of TB patients who were prospectively recruited in an area of Dar es Salaam (Tanzania) with a high TB burden.

Methods

Study setting and study population

This is a prospective cohort study conducted in the densely populated Temeke district of Dar es Salaam, Tanzania; it is a study within a previously described ongoing prospective cohort study of TB patients and household contact controls in Dar es Salaam (TB-DAR) [6]. Between November 2013 and October 2015, we recruited smear-positive TB patients diagnosed at Temeke district hospital and household contact controls who lived in the same household as the index TB cases [6].

Assuming (a) a prevalence of respiratory viruses of 25% in TB patients and of 12.5% in household contact controls [12], based on the prevalence of respiratory viruses in similar settings, and assuming that respiratory viruses are more frequent in TB patients than in controls, (b) a cluster correlation of 0.2, and (c) a non-participation rate of 20%, we estimated that 175 case–control pairs would provide 85% power to observe a statistically significant difference in the prevalence at the 5% level of significance.

Study procedures

At the time of recruitment of study participants, we collected a single sample of nasopharyngeal swabs from TB patients and controls using flexible nasopharyngeal flocked swabs (Copan Diagnostics, CA, USA) [13]. For TB patients, the nasopharyngeal swabs were taken immediately after diagnosis of TB and prior to initiation of anti-TB treatment. The nasopharyngeal swab samples were then added to 1-mL eNAT tubes for transportation at temperatures between 2° and 8°C to the Ifakara Health Institute (IHI) research laboratory at Bagamoyo where they were stored at –80°C pending analysis.

Laboratory investigations

Detection of respiratory pathogens by multiplex PCR

We used the validated multiplex real-time PCRs from Seegene (www.seegene.com/) for detection of a broad panel of respiratory viral and bacterial pathogens in accordance with the manufacturer's instructions, as previously published [14]. The nasopharyngeal swab samples were also analysed using Anyplex II RV16 simultaneously which detects 16 respiratory viruses, and the Allplex Respiratory Panel 4 assay which detects seven respiratory bacterial pathogens (Table S1). Sample processing and analysis to detect respiratory pathogens were all done at the IHI research laboratory in accordance with the manufacturer's instructions and Standard Operating Procedures (SOP).

Other laboratory procedures

TB confirmation was by positive Löwenstein–Jensen (LJ) solid media mycobacterial culture (done at the IHI Research laboratory in Bagamoyo). We ruled out TB in controls with both a negative Gene Xpert MTB/RIF result and no mycobacterial growth in LJ solid media culture. HIV testing for TB patients and controls was done as per Tanzania HIV testing algorithms using an Alere Determine HIV (Alere, USA) and a Uni-Gold HIV (Trinity Biotech; Wicklow, Ireland) confirmatory test rapid tests [15]. CD4+ T cells and full blood-cell counts were obtained as previously described [6].

Data collection and definitions

Clinical severity of TB was graded as per published clinical TB score [16], but modified to a set of 12 TB score parameters instead of 13, since tachycardia was not systematically measured as previously noted [6]. Diagnosis delay was defined as a cough duration of ≥ 3 weeks as previously published from the same cohort study [17].

Data were captured using electronic case report forms developed from the open source data collection software Open Data Kit (ODK, <https://opendatakit.org/>) on Android PC tablets, and data were then managed using the eManagement tool 'odk_planner' as published previously [18].

Statistical analysis

We compared the baseline characteristics of TB patients and household contact controls using the McNemar test, paired t-test, or Wilcoxon signed rank test, as appropriate. We estimated the prevalence of any respiratory viruses and bacteria using logistic regression models adjusting for clustering at the household level. We used mixed-effects logistic regression models with random household intercepts to assess the risk factors associated with detection of respiratory pathogens in TB patients and controls. The differences in the mean Ct values of respiratory bacteria detected (as a relative measure of the bacterial load) between TB patients and controls were assessed using mixed-effect linear regression models. Logistic regression models adjusting for age, sex, and HIV infection were used to assess the associations between respiratory pathogens and clinical presentations of TB at the time of recruitment among TB patients, with the following outcome variables: severe TB score (score of ≥ 6) versus mild (score of 1–5), high sputum bacterial load (sputum AFB smear microscopy of $\geq 2+$) versus low bacterial load, and presence versus absence of lung infiltrations and cavitations (chest x-ray findings). Associations were expressed as crude odds ratios (ORs) and adjusted ORs (aORs) with their corresponding 95% confidence intervals (95% CIs). All analyses were performed in Stata version 14.0 (Stata Corp, College Station, Texas, USA).

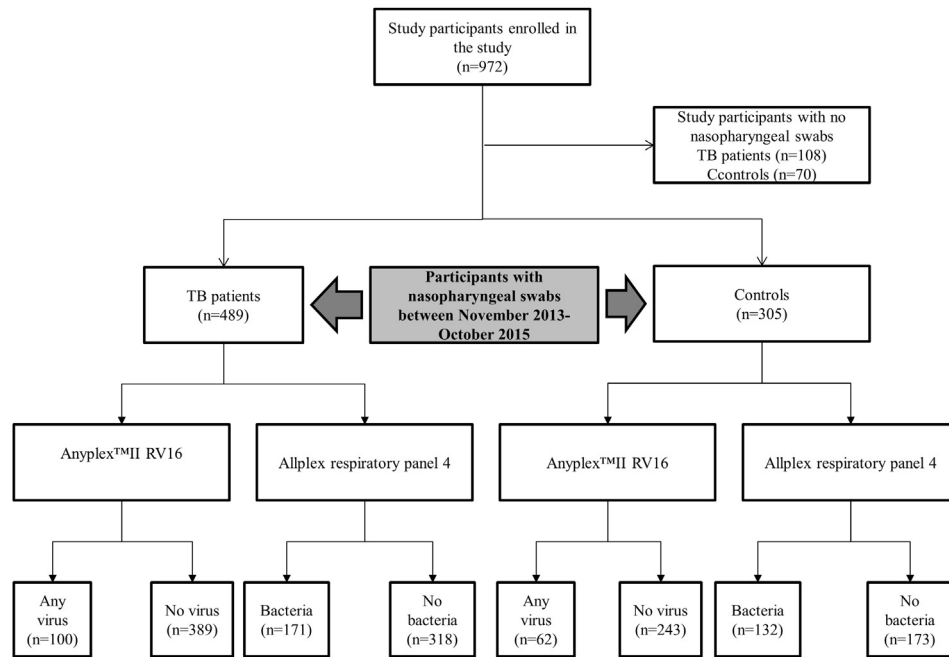


Fig. 1. Flow diagram for participants enrolled in the study.

Ethics approval

The study was approved by the IHI Institutional Review Board (IHI/IRB/No: 04-2015) and the Medical Research Coordinating Committee of the National Institute of Medical Research (NIMR/HQ/R.8c/Vol.I/357) in Tanzania, as well as by the Ethics Committee of the Canton of Basel in Switzerland (EKNZ UBE-15/42). All study participants gave written informed consent. TB patients were managed as per National TB and Leprosy Programme treatment

guidelines [19]. Treatment and care for HIV-positive individuals were as per Tanzania National HIV/AIDS treatment guideline [15].

Results

Characteristics of study participants

Between November 2013 and October 2015, 972 study participants were enrolled in the TB-Dar study. A total of 794 study

Table 1
Baseline characteristics of 489 tuberculosis (TB) patients and 305 household contact controls without TB in Dar es Salaam, Tanzania

Characteristics n (%)	All (n = 794)	TB patients (n = 489)	Controls (n = 305)	p
Age (years), median (IQR)	33.0 (26.1–41.2)	33.0 (27.0–40.0)	32.0 (25.7–42.4)	0.7
Age groups (years)				0.096
<25	153 (19.3)	82 (16.8)	71 (23.3)	
25–34	290 (36.5)	188 (38.4)	102 (33.4)	
35–44	205 (25.8)	132 (27)	73 (23.9)	
≥45	146 (18.4)	87 (17.8)	59 (19.3)	
Male, sex	484 (61.0)	336 (68.7)	148 (48.5)	<0.001
HIV-positive	168 (21.2)	140 (28.6)	28 (9.2)	<0.001
Education level				0.19
No/primary	285 (35.9)	168 (34.4)	117 (38.4)	
Secondary/University	509 (64.1)	321 (65.6)	188 (61.6)	
Occupation				0.25
Employed	509 (64.1)	321 (65.6)	188 (61.6)	
Current smoker				0.053
Yes	104 (13.1)	73 (14.9)	31 (10.2)	
People in the household				0.22
>3 people	204 (25.7)	133 (27.2)	71 (23.3)	
Weight (kg), median (IQR)	54.0 (48.0–61.0)	51.0 (45.5–57.0)	59.0 (53.0–67.0)	<0.001
BMI (kg/m²), median(IQR)	20.1 (17.5–23.4)	18.3 (16.5–20.4)	24.0 (21.7–27.9)	<0.001
BMI categories (kg/m²)				<0.001
Normal/obese ≥18	517 (65.1)	227 (46.4)	290 (95.1)	
Underweight <18	277 (34.9)	262 (53.6)	15 (4.9)	
Body fat percentage (%)	9.92 (7.5–14.4)	9.41 (6.8–13.5)	11 (8.5–15.8)	<0.001
Hb level (g/dL), median (IQR)	12.1 (10.4–13.3)	11.4 (9.9–12.7)	12.8 (11.5–14.2)	<0.001
TB categories				NA
New	477 (97.5)	477 (97.5)	NA	NA
Retreatment	12 (2.5)	12 (2.5)	NA	

BMI, body mass index; IQR, interquartile range; Hb, haemoglobin; HIV, human immunodeficiency virus; NA, not applicable.

participants (81.6%; 794/972) had a nasopharyngeal swab taken, of whom 489 were TB patients and 305 were household contact controls (Fig. 1). The overall median age was 33 years (interquartile range (IQR) 26.1–41.2 years), and 61% (484/794) were men. The overall HIV prevalence was 21.2% (168/794; 95%CI 18.4–24.1%); 140 TB patients (28.6%; 140/489) and 28 controls (9.2%; 28/305) were HIV-positive (Table 1).

Prevalence of respiratory viral and bacterial pathogens

The frequency distributions of the respiratory viruses detected from study participants are summarized in Table 2. The overall prevalence of any respiratory virus among TB patients and controls was 20.4% (161/794; 95%CI 17.7–23.3%), and the odds of detecting any virus was the same in TB patients and controls (OR 1.00, 95%CI 0.71–1.44; *p* 0.98). The most common respiratory species detected was human rhinovirus A/B/C (HRV), which was found in 9.3% (74/794) of the study participants, followed by influenza A in 3.1% (25/794) and respiratory syncytial virus A (RSVA) in 1.9% (15/794). We detected only minor differences between TB patients and household contact controls in the prevalence (Table 2) and the semi-quantitative detection (Fig. S1) of respiratory viruses. We detected respiratory viruses more frequently during the months of March and April, and October to November (Fig. S2).

The prevalence of any bacterial pathogen among study participants was 38.2% (303/794; 95%CI 34.9–41.6%, Table 2). Respiratory bacteria were less likely to be detected in TB patients than in controls (OR 0.70, 95%CI 0.53–0.94; *p* 0.02). The most common bacterial species detected were *Haemophilus influenzae*, found in 26.1% of study participants (207/794), followed by *Streptococcus pneumoniae* in 21.5% (171/794). TB patients were less likely than household contact controls to have *H. influenzae* (OR 0.62, 95%CI 0.45–0.86; *p* 0.004). The mean values of cycle threshold for TB patients were slightly higher (indicating a smaller bacterial load) than those of household contact controls (greater bacterial load), but this did not reach conventional levels of statistical significance (Fig. S3). There were 12 TB patients on a retreatment drug regimen, and in ten of them (83.3%) both respiratory bacteria and respiratory viruses were detected.

The factors associated with detection of respiratory viruses include smoking and households containing three or more people. Men were more likely to have respiratory bacteria (see Table S3).

Associations between respiratory pathogens and clinical presentation

TB patients with both viral and bacterial pathogens had significantly more severe TB disease than TB-only patients (aOR 1.64, 95%

Table 2
Frequencies of virus detection in the tuberculosis (TB) patients and household contact controls in Tanzania, and odds ratios of detection in TB patients compared to controls

Detection of viral species	All (<i>n</i> = 794)	TB patients (<i>n</i> = 489)	Controls (<i>n</i> = 305)	OR (95%CI)	<i>p</i>
Any respiratory virus	162 (20.4)	100 (20.4)	62 (20.3)	1.00 (0.71–1.44)	0.98
Respiratory viral species					
Human rhinovirus A/B/C	74 (9.3)	42 (8.6)	32 (10.5)	0.80 (0.49–1.30)	0.37
Influenza A	25 (3.1)	15 (3.1)	10 (3.3)	0.93 (0.41–2.11)	0.87
RSVA	15 (1.9)	12 (2.5)	3 (1.0)	2.53 (0.71–9.05)	0.15
Adenovirus	14 (1.8)	9 (1.8)	5 (1.6)	1.13 (0.37–3.39)	0.83
RSVB	12 (1.5)	9 (1.8)	3 (1.0)	1.89 (0.51–7.03)	0.34
Parainfluenza virus 4	9 (1.1)	4 (0.8)	5 (1.6)	0.49 (0.13–1.86)	0.3
Coronavirus OC43	9 (1.1)	5 (1.0)	4 (1.3)	0.78 (0.21–2.92)	0.71
Coronavirus NL63	4 (0.5)	3 (0.6)	1 (0.3)	1.87 (0.19–18.12)	0.59
Enterovirus	4 (0.5)	4 (0.8)	0 (0)	NA	
Bocavirus 1/2/3/4	3 (0.4)	2 (0.4)	1 (0.3)	1.24 (0.11–13.83)	0.86
Parainfluenza virus 2	3 (0.4)	1 (0.2)	2 (0.7)	0.31 (0.03–3.44)	0.34
Influenza B	2 (0.3)	1 (0.2)	1 (0.3)	0.62 (0.04–10.0)	0.74
Parainfluenza virus 3	2 (0.3)	2 (0.4)	0 (0)	0.90 (0.41–1.97)	0.80
Metapneumovirus	1 (0.1)	1 (0.2)	0 (0)	NA	
Coronavirus 229E	1 (0.1)	0 (0)	1 (0.3)	NA	
Groups of detected viruses					
Influenza A/B	27 (3.4)	16 (3.3)	11 (3.6)	0.90 (0.41–1.97)	0.80
Influenza-like (influenza and parainfluenza viruses)	40 (5.0)	23 (4.7)	17 (5.6)	0.84 (0.44–1.60)	0.59
Coronaviruses	14 (1.8)	8 (1.6)	6 (2.0)	0.83 (0.28–2.41)	0.73
Parainfluenza 2/3/4	13 (1.6)	7 (1.4)	6 (2.0)	0.72 (0.24–2.17)	0.56
RSV	25 (3.1)	19 (3.9)	6 (2.0)	2.01 (0.80–5.10)	0.14
Groups according to the number of detected viral species					
One species	145 (18.3)	89 (18.2)	56 (18.4)	0.99 (0.69–1.44)	0.96
≥2 species	17 (2.1)	11 (2.2)	6 (2.0)	1.15 (0.42–3.13)	
Respiratory bacterial pathogens					
Any bacterial species	303 (38.2)	171 (35.0)	132 (43.3)	0.70 (0.53–0.94)	0.019
Respiratory bacterial species					
<i>Haemophilus influenzae</i>	207 (26.1)	110 (22.5)	97 (31.8)	0.62 (0.45–0.86)	0.004
<i>Streptococcus pneumoniae</i>	171 (21.5)	99 (20.2)	72 (23.6)	0.82 (0.58–1.16)	0.26
<i>Legionella pneumophila</i>	12 (1.5)	9 (1.8)	3 (1.0)	1.89 (0.51–7.03)	0.34
<i>Bordetella parapertussis</i>	4 (0.5)	3 (0.6)	1 (0.3)	1.88 (0.19–18.12)	0.59
<i>Mycoplasma pneumoniae</i>	0 (0)	0 (0)	0 (0)	NA	N/A
<i>Bordetella pertussis</i>	5 (0.6)	4 (0.8)	1 (0.3)	2.51 (0.28–22.54)	0.41
<i>Chlamydomphila pneumoniae</i>	0 (0)	0 (0)	0 (0)	NA	NA
Groups according to the number of detected bacterial species					
One specie	209 (26.3)	119 (24.3)	90 (29.5)	0.72 (0.52–1.00)	0.062
≥2 species	94 (11.8)	52 (10.6)	42 (13.8)	0.67 (0.43–1.05)	

OR, odds ratio; 95%CI, 95% confidence interval; NA, not applicable.

ORs and *p* calculated from mixed-effects logistic regression models with random household intercepts.

Table 3
Clinical significance of respiratory pathogens among tuberculosis (TB) patients at the time of TB diagnosis

Respiratory pathogens detected	Severe TB score ^a		High sputum bacterial load ^b		Lung cavitation ^c		Lung infiltrations ^c		Diagnostic delay ^d	
	aOR (95%CI)	p	aOR (95%CI)	p	aOR (95%CI)	p	aOR (95%CI)	p	aOR (95%CI)	p
Respiratory viruses										
Any viral species		0.072		0.69		0.089		0.88		0.46
Yes	1.52 (0.96–2.4)		1.10 (0.69–1.76)		1.58 (0.93–2.68)		1.04 (0.60–1.83)		0.80 (0.45–1.43)	
Respiratory bacteria										
Any bacterial species		0.17		0.89		0.65		0.85		0.77
Yes	1.32 (0.89–1.94)		1.03 (0.69–1.53)		0.90 (0.56–1.44)		0.95 (0.58–1.56)		1.07 (0.67–1.71)	
Combined detection of viral and bacterial species										
Yes	1.64 (1.11–2.37)	0.01	1.00 (0.68–1.46)	0.95	1.09 (0.70–1.71)	0.7	0.92 (0.57–1.46)	0.71	0.87 (0.55–1.37)	0.53

aOR, adjusted odds ratios; 95% CI, 95% confidence interval

Logistic regression model adjusted for age-group, sex, and HIV infection.

^a Severe TB score (6 to 12) compared to mild TB score (1 to 5).^b High sputum bacterial load ($\geq 2+$ according to qualitative AFB smear microscopy grading) compared to low load (scanty up to 1+).^c As determined by chest x-ray features.^d Diagnostic delay defined as defined symptoms duration of ≥ 3 weeks.

CI 1.11–2.37; $p < 0.01$; Table 3). Bacterial respiratory pathogens were not significantly associated with the clinical presentation of TB patients at TB diagnosis. Detection of respiratory pathogens was not associated with including diagnostic delay (defined as duration of symptoms of 3 weeks or more) (see Table 3). No association was found between detection of respiratory pathogens and chest x-ray findings among TB patients (Table 4). In addition, the detection of respiratory pathogens was similar for HIV-positive and HIV-negative TB patients (Tables S2 and S3).

Discussion

Both respiratory viruses and respiratory bacteria are commonly detected in a high-TB-incidence setting, and the prevalence of respiratory bacteria was lower in TB patients than in household contact controls. Detection of respiratory viruses and respiratory bacteria in TB patients was associated with more severe disease.

The prevalence of any respiratory viruses was the same (20%) for both TB patients and controls without TB. Similar prevalence of respiratory viruses in controls as compared to TB patients could be due to controls being more active than TB patients, hence having increased social contacts. The prevalence shown in our study was lower than that reported in an Indonesian study of influenza viruses that observed respective prevalences of around 46% and 41% for TB patients and controls, respectively [20]. The difference in the prevalence of influenza in that study compared to ours could be due to

the different study region (Asia versus sub-Saharan Africa) and the use of different diagnostic methods (immunological assay versus molecular detection). We detected respiratory viruses from nasopharyngeal swabs using a highly sensitive and specific molecular technique [21], whereas the study from Indonesia [20] measured influenza virus antibody titres which also detect patients with previous exposure to influenza viruses. The influenza antibody titres were higher in TB patients than in controls, suggesting recent viral infection before the clinical manifestations of TB [20].

We did not find any evidence for an association between HIV infection and detection of respiratory pathogens. In line with our results, a household study on respiratory illness surveillance and HIV testing in Kenya [22] did not find any association between HIV and influenza viruses. However, household contacts of the HIV-infected influenza index cases were twice as likely to develop a secondary case of influenza-like illness [22].

We also found that the presence of both viruses and bacteria could potentially alter the clinical course of TB, and present with a more severe disease as measured by the previously validated clinical TB score [16]. Direct evidence of clinical effects of respiratory viruses on TB have only been demonstrated in experimental mouse models that have exhibited higher mycobacterial loads in the lungs and increased lung inflammation [9]. The immunological pathway responsible for more severe clinical presentation in TB patients may occur either via the type I interferon-receptor-dependent pathway [9] or via decreased MHC II expression on

Table 4
Chest x-ray findings of tuberculosis (TB) patients with and without any respiratory pathogens (viruses and bacteria)

Chest x-ray findings	Total	TB and respiratory pathogen(s)		p ^a
	n (%)	n (%)		
Any respiratory viruses				
Infiltrates	236 (64)	51 (64.6)	185 (63.8)	0.90
Cavitations	125 (33.9)	33 (41.8)	92 (31.7)	0.09
Pleural effusion	44 (11.9)	7 (8.9)	37 (12.8)	0.34
Lymph nodes	31 (8.4)	6 (7.6)	25 (8.6)	0.77
Micronodules	22 (6)	5 (6.3)	17 (5.9)	0.88
Macronodules	5 (1.4)	1 (1.3)	4 (1.4)	0.94
Any respiratory bacteria				
Infiltrations	236 (64)	78 (63.9)	158 (64)	0.99
Cavitations	125 (33.9)	40 (32.8)	85 (34.4)	0.76
Pleural effusion	44 (11.9)	15 (12.3)	29 (11.7)	0.88
Lymph nodes	31 (8.4)	11 (9)	20 (8.1)	0.77
Micronodules	22 (6)	9 (7.4)	13 (5.3)	0.42
Macronodules	5 (1.4)	1 (0.8)	4 (1.6)	0.53

^a p calculated from χ^2 -square test.

dendritic cells [23], which may result in poor clearance of *M. tuberculosis* from the lungs.

We found smoking and living in a household with three or more persons to be risk factors for respiratory viruses. Smoking has been shown, at least in animal models, to inhibit the pulmonary T-cell response to influenza viruses, thus increasing susceptibility to infection [24], and respiratory viruses were more likely to be detected in children living with a smoker [25]. In addition, overcrowding—which we defined as three or more persons in a household—is a common risk factor for most airborne pathogens such as *M. tuberculosis* [26] and respiratory viruses [22].

The prevalence of bacterial respiratory pathogens in our study was lower in TB patients than in household contact controls, suggesting interactions between *M. tuberculosis* and the bacterial populations in the airways. Overall respiratory bacterial load was smaller in TB patients than in controls. This is similar to findings from a microbiota study which reported smaller bacterial loads in TB patients than in controls [27]. The authors argue that the initial phase of *M. tuberculosis* invasion in the lungs may prompt an immune response that could also reduce the commensal flora in the lower respiratory tract [27]. Interestingly, in a mouse model, *M. tuberculosis* infection in the lungs appeared also to have an effect on the gut microbiota, which is part of the collective human microbiota [28]. These findings consistently suggest interactions between *M. tuberculosis* and the communities of microorganisms, and a role for these interactions in TB pathogenesis.

We believe that this is the first study to have looked systematically at a wide range of viral and bacterial species in TB patients and controls, and using sensitive molecular techniques and clinical specimens from a well-defined compartment of the airways. A particular strength of the study is that potentially confounding and unmeasured risk factors were minimized by studying patients and controls who lived in the same households. A limitation of the study is its undifferentiated attention to respiratory viruses because of small numbers which precluded assessment of the clinical effects of individual viruses. However, we presume that all respiratory viruses have similar levels of immunomodulation, and thus we could combine all respiratory viruses together.

In conclusion, respiratory pathogens are common in the high-TB setting of Tanzania for both TB patients and household contact controls without TB. However, respiratory bacterial species were more frequently detected in household contact controls than in TB patients. Our findings suggest that TB patients co-infected with both respiratory viruses and respiratory bacteria have severe TB disease. Further research should focus on the pathogenic role of respiratory pathogens in high-TB-incidence settings and their effects on clinical and treatment outcomes.

Transparency declaration

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Appendix A. Supplementary data

Supplementary data related to this article can be found at <https://doi.org/10.1016/j.cmi.2018.03.019>.

References

- [1] WHO. Global tuberculosis report. Geneva, Switzerland: World Health Organization; 2017.
- [2] Noymer A. Testing the influenza–tuberculosis selective mortality hypothesis with Union Army data. *Soc Sci Med* 2009;68:1599–608. <https://doi.org/10.1016/j.socscimed.2009.02.021>.
- [3] Zürcher K, Zwahlen M, Ballif M, Rieder HL, Egger M, Fenner L. Influenza pandemics and tuberculosis mortality in 1889 and 1918: analysis of historical data from Switzerland. *PLoS One* 2016;11:e0162575. <https://doi.org/10.1371/journal.pone.0162575>.
- [4] Noymer A. The 1918 influenza pandemic hastened the decline of tuberculosis in the United States: an age, period, cohort analysis. *Vaccine* 2011;29:B38–41. <https://doi.org/10.1016/j.vaccine.2011.02.053>.
- [5] WHO. Global tuberculosis report. 2016. p. 2016.
- [6] Mhimbira F, Hella J, Said K, Kamwela L, Sasamalo M, Maroa T, et al. Prevalence and clinical relevance of helminth co-infections among tuberculosis patients in urban Tanzania. *PLoS Negl Trop Dis* 2017;11:e0005342. <https://doi.org/10.1371/journal.pntd.0005342>.
- [7] Wu J, Liu W, He L, Huang F, Chen J, Cui P, et al. Sputum microbiota associated with new, recurrent and treatment failure tuberculosis. *PLoS One* 2013;8:e83445. <https://doi.org/10.1371/journal.pone.0083445>.
- [8] Marais BJ, Lönnroth K, Lawn SD, Migliori GB, Mwaba P, Glaziou P, et al. Tuberculosis comorbidity with communicable and non-communicable diseases: integrating health services and control efforts. *Lancet Infect Dis* 2013;3. [https://doi.org/10.1016/S1473-3099\(13\)70015-X](https://doi.org/10.1016/S1473-3099(13)70015-X).
- [9] Redford PS, Mayer-Barber KD, McNab FW, Stavropoulos E, Wack A, Sher A, et al. Influenza A virus impairs control of *Mycobacterium tuberculosis* coinfection through a type I interferon receptor-dependent pathway. *J Infect Dis* 2013. <https://doi.org/10.1093/infdis/jit424>.
- [10] Rohde G, Wiethage A, Borg I, Kauth M, Bauer TT, Gillissen A, et al. Respiratory viruses in exacerbations of chronic obstructive pulmonary disease requiring hospitalisation: a case–control study. *Thorax* 2003;58:37–42.
- [11] Wood MR, Yu EA, Mehta S. The human microbiome in the fight against tuberculosis. *Am J Trop Med Hyg* 2017;96:1274–84. <https://doi.org/10.4269/ajtmh.16-0581>.
- [12] Bigogo GM, Breiman RF, Feikin DR, Audi AO, Aura B, Cosmas L, et al. Epidemiology of respiratory syncytial virus infection in rural and urban Kenya. *J Infect Dis* 2013;208:S207–16. <https://doi.org/10.1093/infdis/jit489>.
- [13] FLOQSwabs™. COPAN's patented flocced swabs. Copan Diagn Inc n.d. <http://www.copanusa.com/products/collection-transport/floqswabs-flocced-swabs/> (accessed June 5, 2017).
- [14] Cho CH, Chulten B, Lee CK, Nam MH, Yoon SY, Lim CS, et al. Evaluation of a novel real-time RT-PCR using TOCE technology compared with culture and Seeplex RV15 for simultaneous detection of respiratory viruses. *J Clin Virol* 2013;57:338–42. <https://doi.org/10.1016/j.jcv.2013.04.014>.
- [15] NACP. National guidelines for the management of HIV and AIDS. 5th ed. Dar es Salaam: Ministry of Health and Social Welfare; 2015.
- [16] Wejse C, Gustafson P, Nielsen J, Gomes VF, Aaby P, Andersen PL, et al. TB score: signs and symptoms from tuberculosis patients in a low-resource setting have predictive value and may be used to assess clinical course. *Scand J Infect Dis* 2008;40:111–20. <https://doi.org/10.1080/00365540701558698>.
- [17] Said K, Hella J, Mhalu G, Chiryankubi M, Masika E, Maroa T, et al. Diagnostic delay and associated factors among patients with pulmonary tuberculosis in Dar es Salaam, Tanzania. *Infect Dis Poverty* 2017;6:64. <https://doi.org/10.1186/s40249-017-0276-4>.
- [18] Steiner A, Hella J, Grüninger S, Mhalu G, Mhimbira F, Cercamondi CI, et al. Managing research and surveillance projects in real-time with a novel open-source eManagement tool designed for under-resourced countries. *J Am Med Inform Assoc* 2016;ocv185. <https://doi.org/10.1093/jamia/ocv185>.
- [19] NTLP, MoHSW. Manual for the management of tuberculosis and leprosy. 6th ed. Dar es Salaam: Ministry of Health and Social Welfare; 2013.
- [20] de Paus RA, van Crevel R, van Beek R, Sahiratmadja E, Alisjahbana B, Marzuki S, et al. The influence of influenza virus infections on the development of tuberculosis. *Tuberculosis* 2013;93:338–42. <https://doi.org/10.1016/j.tube.2013.02.006>.
- [21] Niang MN, Diop NS, Fall A, Kiori DE, Sarr FD, Sy S, et al. Respiratory viruses in patients with influenza-like illness in Senegal: focus on human respiratory adenoviruses. *PLoS One* 2017;12:e0174287. <https://doi.org/10.1371/journal.pone.0174287>.
- [22] Judd MC, Emukule GO, Njuguna H, McMorrow ML, Arunga GO, Katz MA, et al. The role of HIV in the household introduction and transmission of influenza in an urban slum, Nairobi, Kenya, 2008–2011. *J Infect Dis* 2015;212:740–4. <https://doi.org/10.1093/infdis/jiv106>.
- [23] Flórido M, Grima M, Gillis CM, Xia Y, Turner SJ, Triccas J, et al. Influenza A virus infection impairs mycobacteria-specific T cell responses and mycobacterial clearance in the lung during pulmonary coinfection. *J Immunol* 2013;191:302–11. <https://doi.org/10.4049/jimmunol.1202824>.

- [24] Feng Y, Kong Y, Barnes PF, Huang F-F, Klucar P, Wang X, et al. Exposure to cigarette smoke inhibits the pulmonary T-cell response to influenza virus and *Mycobacterium tuberculosis*. *Infect Immun* 2010;79:229–37. <https://doi.org/10.1128/IAI.00709-10>.
- [25] Nicolai A, Frassanito A, Nenna R, Cangiano G, Petrarca L, Papoff P, et al. Risk factors for virus-induced acute respiratory tract infections in children younger than 3 years and recurrent wheezing at 36 months follow-up after discharge. *Pediatr Infect Dis* 2017;36:179–83. <https://doi.org/10.1097/INF.0000000000001385>.
- [26] Corbett EL, Bandason T, Cheung YB, Makamure B, Dauya E, Munyati SS, et al. Prevalent infectious tuberculosis in Harare, Zimbabwe: burden, risk factors and implications for control. *Int J Tuberc Lung Dis* 2009;13:1231–7.
- [27] Cui Z, Zhou Y, Li H, Zhang Y, Zhang S, Tang S, et al. Complex sputum microbial composition in patients with pulmonary tuberculosis. *BMC Microbiol* 2012;12:276. <https://doi.org/10.1186/1471-2180-12-276>.
- [28] Winglee K, Eloie-Fadrosch E, Gupta S, Guo H, Fraser C, Bishai W. Aerosol *Mycobacterium tuberculosis* infection causes rapid loss of diversity in gut microbiota. *PLoS One* 2014;9:e97048. <https://doi.org/10.1371/journal.pone.0097048>.