

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

# The virulence of mouse hepatitis virus 3, as evidenced by permissivity of cultured hepatic cells toward escape mutants

J.P. Martin<sup>(1)</sup><sup>(\*)</sup>, W. Chen<sup>(1)</sup>, F. Koehren<sup>(1)</sup> and C.A. Pereira<sup>(1, 2)</sup>

<sup>(1)</sup> Unité INSERM U74 et Institut de Virologie de la Faculté de Médecine, Université Louis Pasteur, 67000 Strasbourg (France), and

<sup>(2)</sup> Instituto Butantan, Laboratorio de Immunologia Viral, Sao Paulo (Brasil)

It has been shown that the spike (S) glycoprotein from the *Coronaviridae* is a determining factor in the cell fusion properties as well as in the neuropathogenic effects of some isolated murine hepatitis viruses (MHV) (Fleming *et al.*, 1986; Makino *et al.*, 1987; Daziel *et al.*, 1988; Wege *et al.*, 1988). It therefore seems to play a major role in the pathogenicity of the virus and influences its replication in tissue cultures and its ability to invade various organs in the animal (Daziel *et al.*, 1986; Wege *et al.*, 1988). The role of S glycoprotein in determining MHV3 virulence has been studied superficially, although the nucleotide fingerprints of hepatotropic MHV3 and neurotropic MHVA59 were very similar (Lai *et al.*, 1981).

We prepared monoclonal antibodies (mAb) for the S glycoprotein and used them to select attenuated MHV3 escape mutants for further investigations into the viral pathogenicity and hepatotropism of MHV3, which produces an acute fulminating hepatitis with extensive necrosis of the liver in susceptible BALB/c mice (Lucchiari *et al.*, 1991; Martin *et al.*, 1988; 1990).

Among the 60 mAb obtained by the method of Fazekas de St Groth and Scheidegger (1980), 15

were able to neutralize the subcloned MHV3 (MHV3 wt) at a dilution of over 1/5,000 (table I). All fifteen of them recognized the S protein in its precursor form (120 kDa), its glycosylated uncleaved form (180 kDa) and its cleaved mature form (90 kDa). Among these antibodies, 12 were also able to inhibit cell fusion, if they were added 2 h after viral infection. The 15 above mentioned mAb were used to select escape mutants. In each,  $1.5 \times 10^5$  infectious particles were incubated with an individual mAb at a 1/100 dilution. The number of mutants isolated varied for each mAb. The mutation frequency was about  $10^{-4}$  to  $10^{-5}$ . Initially, 283 escape mutants were isolated and, after recloning in presence of the specific mAb, 100 PFU of each were injected by the intraperitoneal route into five 8- to 12-week-old sensitive BALB/c mice (IFFA Credo, L'Arbresle, France) free of coronavirus antibodies. With MHV3 wt, the mice died 3 to 4 days after infection. The infection of mice with MHV3 51.6 and MHV3 Cl12 obtained with two different mAb (A51 and A37, respectively) identified by agar gel diffusion as being an IgG1 and an IgA isotype, provoked a delay in mortality (fig. 1A). The MHV3 51.6 was of medium virulence, with mice dying 5 to 9 days

(\*) Corresponding author.

Submitted February 7, 1994, accepted April 25, 1994.

### J.P. MARTIN ET AL.

mAb	Neutrali- zation	Inhibition	Protection of mice	Isotype	Number of mutants
A2	>10.000	1,000	9/10	IgG1	2
A6	5,000	0	6/10	IgG2b	23
A15	10,000	0	6/10	ĬgG1	29
A34	5,000	0	4/10	IgG2a	35
A37	>10.000	400	0/10	ĬgA	21
A42	10,000	400	3/10	IgG2a	23
A45	>10,000	1,000	7/10	IgG3	1
A46	5.000	100	10/10	IgG2a	19
A48	10,000	100	7/10	IgG2a	26
A50	>10.000	1.000	5/10	IgG1	0
A51	10,000	100	0/10	IgG1	24
A53	>10,000	1,000	9/10	IgG1	1
A55	>10.000	400	7/10	IgG2a	15
A56	5,000	0	5/10	IgG2a	19
A60	>10,000	400	4/10	IgG2a	45

Table I. Biochemical and biological characterization of mAb.

The titre of neutralization and inhibition of cell fusion was the highest dilution able to halve the number of plaques. Protection in vivo was tested by injection of 0.2 ml of an ascites fluid into mice 1 day before the injection of 100 PFU of MHV3 wt.

after infection; MHV3 Cl12 was less virulent, with mice dying 8 to 10 days post-infection.

Virus replication in the livers of BALB/c mice infected with MHV3 wt, MHV3 51.6 or MHV3 Cl12 (fig. 1B) corresponded to the degree of virulence shown by these viruses in BALB/c mice. In MHV3-wt-infected mice, the virus titres reached a peak 2 days after infection. In MHV3-51.6-infected mice, the virus titre increased gradually to reach a peak 8 days after infection. In MHV3-Cl12-infected mice, virus titres were not detectable until 4 days after infection, when they started to increase.

To correlate these in vivo data with in vitro experiments, we isolated four types of hepatic

cells. The predominant cells in the liver are hepatocvtes (Hc), which numerically represent about 60% of the total liver cell population. Hc were isolated according to Berry and Friend (1969) and Seglen (1976), partially modified by Braunwald et al. (1991). The Kupffer cells (Kc), liver endothelial cells (Ec) and Ito cells (Ic) make up the sinusoidal wall. The Kc, which are macrophages in the liver and have immunological functions, are found inside the lumen of the hepatic sinusoid at strategic positions; the Ec form a barrier which separates the perisinusoidal space from the sinusoid and possesses characteristic fenestrae which act as filtration barriers and also provide a means of communication between the sinusoid and the space of Disse; the Ic, for fat

Ec endothelial cell. =

- Hc = hepatocyte.
- Ito cell. Ic =
- i.p. = intraperitoneally.
- Kupffer cell. Kc =
- mAb monoclonal antibody.

MHV mouse hepatitis virus. =

m.o.i. multiplicity of infection. PFU plaque-forming unit. =

S spike. =

wild type. wt



Fig. 1. Mortality (A) of mice infected with MHV3 and kinetics of virus growth in the liver (B).

Groups of 40 BALB/c mice were inoculated i.p. with 100 PFU of MHV3 wt (\*), MHV3 51.6 ( $\Box$ ) or MHV3 Cl12 ( $\Delta$ ). The animals were observed and the mortality recorded. At subsequent intervals, groups of 5 mice were sacrificed, the livers removed and the virus titrated. The virus titres reported as log<sub>10</sub> PFU/g of tissue are the average of 5 different determinations.

storage, are usually located in the sinusoidal recesses within the space of Disse and are known to play a role in storing vitamin A and maintaining the sinusoidal vascular structure. The technique for isolation of Kc and Ec has already been described (Steffan *et al.*, 1986), and Ic were prepared according to a previously described procedure (Chen *et al.*, 1989).

Virus multiplication in cultured hepatic cells is showed in figure 2. In all cell cultures, MHV3 wt, the most virulent, was capable of replicating faster than the other isolates, consistently reaching higher titres. MHV3 51.6, of medium virulence, replicated in the cultures, except in Ec where basal titres were found throughout the experiment. The less virulent MHV3 Cl12 was capable of replicating, reaching low titres only in Ic and showing basal titres in the other cells throughout the experiment.

In this work, we compared the *in vivo* and *in vitro* virulence of two MHV3 mutants which escaped neutralization by one of two different anti-mAb. The procedure that we used for isolating the mutants led to the obtaining of a virus with a mutation in a determinant of the S glycoprotein, which is limited enough to enable replication in the L929 cell line used to propagate the virus. The data obtained indicate that the modified *in vivo* virulence observed for two MHV3 mutants could reflect an impairment of the replication in hepatic cells. Indeed, only basic viral titres were detected in some cultured hepatic cells. The basic titres found throughout the



Fig. 2. Kinetics of MHV3 multiplication in cultured hepatic cells.

Cultures of Kupffer cells (A), endothelial liver cells (B), hepatocytes (C) or Ito cells (D) were infected with 0.1 m.o.i. of MHV3 wt (\*), MHV3 51.6 ( $\Box$ ) or MHV3 Cl12 ( $\Delta$ ). Supernatants were collected at different times after infection and the virus titrated. The virus titres, reported as  $\log_{10}$  PFU/ml, is the average of 5 different experiments.

experiments, in Ec infected with MHV3 51.6 and in Kc, Ec and Hc infected with MHV3 Cl12, indicate that, although these cells are permissive for the virus, its replication is very limited, though sufficient to maintain virus titre for a long period of time.

The first hepatic cells in contact with the blood are Kc and Ec. After virus multiplication and lysis of these cells, the virus reaches the parenchymal cells, where its multiplication leads to focal necrosis that can be responsible for physiological dysfunction associated with fatal hepatitis. It should be borne in mind that impairment in adsorption of a given virus to strategically placed cells, such as Ec or Kc, may be a limiting factor in viral multiplication. This has important implications for the outcome of infection.

An alteration in the immune functions of BALB/c mice consecutive to the infection, as well as the selection of revertant viruses possessing a wild phenotype, could account for the observed results. However, the latter hypothesis has been excluded, since viruses isolated from the livers of mice infected by each of the mutants were still resistant to neutralization by relevant mAb. Our results are consistent with the notion that our MV3 escape mutants display at least one single mutation in a determinant of the S glycoprotein. These mutations could be responsible for impairment in recognition of a cellular receptor by the virus in hepatic cells (Martin et al., 1990), thus hindering its multiplication in the liver and delaying the evolution of the disease.

Key-words: Hepatitis, MHV3, Replication, Permissivity; Hepatic cells, Escape mutants, Virulence.

#### Acknowledgements

This work was subsidized in part by a grant from the Direction des Recherches, Etudes et Techniques  $n^{\circ}$  90.34.111.00.470.75.01, and in part by the scientific collaboration between the Institut National de la Santé et de la Recherche Médicale (INSERM), France, and the Conselho Nacional de Pesquisa (CNPq), Brazil.

## Corrélation entre le pouvoir pathogène de deux mutants d'échappement du virus de l'hépatite murine 3 et la permissivité des cellules hépatiques en culture primaire

Deux mutants d'échappement à la neutralisation du MHV3 (MHV 51.6 et MHV Cl12) montrent, en comparaison au MHV3 sauvage (wt), un ralentissement de leur multiplication chez des souris sensibles BALB/c et une atténuation du pouvoir pathogène provoquant un retard considérable de la mortalité. Le MHV3 wt se réplique à des titres élevés dans toutes les cellules hépatiques en culture. Le MHV3 51.6, de virulence intermédiaire, se multiplie dans toutes les cellules hépatiques excepté dans les cellules endothéliales. Le mutant qui tue le plus tardivement les souris (MHV Cl12) se multiplie uniquement dans les cellules de Ito, les autres étant non permissives. Ces résultats montrent une corrélation directe entre la virulence du MHV3 et son aptitude à se répliquer de manière efficace dans les cellules hépatiques in vitro.

*Mots-clés*: Hépatite, MHV3, Réplication, Permissivité; Cellules hépatiques, Mutants d'échappement, Virulence.

#### References

- Berry, M.N. & Friend, D.S. (1969), High-yield preparation of isolated rat liver parenchymal cells. J. Cell. Biol., 43, 506-520.
- Braunwald, J., Nonnenmacher, H., Pereira, C.A. & Kirn, A. (1991), Increased susceptibility to mouse hepatitis type 3 (MHV3) infection induced by a hypercholesterolemic diet with increased adsorption of MHV3 to primary hepatocytes cultures. *Res. Virol.*, 142, 5-15.
- Chen, W., Gendrault, J.L., Steffan, A.M., Jeandidier, E. & Kirn, A. (1989), Isolation, culture and main characteristics of mouse fat-storing cells: interaction with viruses. *Hepatology*, 9, 352-362.
- Daziel, R.G., Lampert, P.W., Talbot, P.J. & Buchmeier, M.J. (1986), Site-specific alteration of murine hepatitis virus type 4 peplomer glycoprotein E2 results in reduced virulence. J. Virol., 59, 463-471.
- Fazekas de St Groth, S. & Scheidegger, D. (1980), Production of monoclonal antibodies: strategy and tactics. J. Immunol. Methods, 35, 1-21.
- Fleming, J.O., Trousdale, M.D., El-zaatari, F., Stohlman, S.A. & Weiner, L.P. (1986), Pathogenicity of antigenic variants of murine coronavirus JHM selected with monoclonal antibodies. J. Virol., 58, 869-875.
- Lai, M.C.C., Brayton, P.R., Armen, R.C., Pugh, C. & Stohlman, S.A. (1981), Mouse hepatitis virus A59:

mRNA structure and genetic localization of the sequence divergence from hepatotropic strain MHV3. J. Virol., 39, 823-834.

- Lucchiari, M.A., Martin, J.P., Modolell, M. & Pereira, C.A. (1991), Acquired immunity of A/J mice to mouse hepatitis virus 3 infection: dependence on interferon gamma synthesis and macrophage sensitivity to interferon gamma. J. Gen. Virol., 72, 1317-1322.
- Martin, J.P., Koehren, F., Rannou, J.J. & Kirn, A. (1988), Temperature-sensitive mutants of mouse hepatitis virus type 3 (MHV3): isolation, biochemical and genetic characterization. Arch. Virol., 100, 147-160.
- Martin, J.P., Chen, W., Obert, G. & Koehren, F. (1990), Characterization of attenuated mutants of MHV3: importance of the E2 protein in organ tropism and

infection of isolated liver cells (Eds. D. Cavanagh & T.D.K. Brown, Plenum Press). Adv. Exp. Med. Biol., 276, 403-410.

- Seglen, P.O. (1976), Preparation of isolated rat liver cells. Meth. Cell. Biol., 13, 29-81.
- Steffan, A.M., Pereira, C.A. & Kirn, A. (1986), Role of the sinusoidal cells in the course of the hepatitis induced by mouse hepatitis virus type 3 (MHV3) in mice, *in* "Cells of the hepatic sinusoid, vol. I" (A. Kirn, D.L. Knook and E. Wisse) (pp. 377-378). The Kupffer Cell Foundation, Rijswijk.
- Wege, H., Winter, J. & Meyermann, R. (1988), The peplomer protein E2 of coronavirus JHM as a determinant of neurovirulence: definition of critical epitopes by variant analysis. J. Gen. Virol., 69, 87-98.