




RESEARCH PAPER



Synthesis, antitumor activity, and molecular docking study of 2-cyclopentyloxyanisole derivatives: mechanistic study of enzyme inhibition

Walaa M. El-Husseiny^a, Magda A.-A. El-Sayed^{a,b} , Adel S. El-Azab^c , Nawaf A. AlSaif^c, Mohammed M. Alanazi^c and Alaa A.-M. Abdel-Aziz^c 

^aDepartment of Pharmaceutical Organic Chemistry, Faculty of Pharmacy, Mansoura University, Mansoura, Egypt; ^bDepartment of Pharmaceutical Chemistry, Faculty of Pharmacy, Horus University, New Damietta, Egypt; ^cDepartment of Pharmaceutical Chemistry, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia

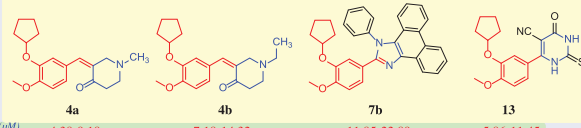
ABSTRACT

A series of 24 compounds was synthesised based on a 2-cyclopentyloxyanisole scaffold **3–14** and their *in vitro* antitumor activity was evaluated. Compounds **4a**, **4b**, **6b**, **7b**, **13**, and **14** had the most potent antitumor activity (IC_{50} range: 5.13–17.95 μM), compared to those of the reference drugs celecoxib, afatinib, and doxorubicin. The most active derivatives **4a**, **4b**, **7b**, and **13** were evaluated for their inhibitory activity against COX-2, PDE4B, and TNF- α . Compounds **4a** and **13** potently inhibited TNF- α (IC_{50} values: 2.01 and 6.72 μM , respectively) compared with celecoxib (IC_{50} =6.44 μM). Compounds **4b** and **13** potently inhibited COX-2 (IC_{50} values: 1.08 and 1.88 μM , respectively) comparable to that of celecoxib (IC_{50} =0.68 μM). Compounds **4a**, **7b**, and **13** inhibited PDE4B (IC_{50} values: 5.62, 5.65, and 3.98 μM , respectively) compared with the reference drug roflumilast (IC_{50} =1.55 μM). The molecular docking of compounds **4b** and **13** with the COX-2 and PDE4B binding pockets was studied.

HIGHLIGHTS

- Antitumor activity of new synthesized cyclopentyloxyanisole scaffold was evaluated.
- The powerful antitumor **4a**, **4b**, **6b**, **7b** & **13** were assessed as COX-2, PDE4B & TNF- α inhibitors.
- Compounds **4a**, **7b**, and **13** exhibited COX-2, PDE4B, and TNF- α inhibition.
- Compounds **4b** and **13** showed strong interactions at the COX-2 and PDE4B binding pockets.

GRAPHICAL ABSTRACT



	4a	4b	7b	13
Antitumor activity: IC_{50} (μM)	4.38-9.18	7.18-14.32	11.85-22.89	5.86-11.45
COX-2 inhibition: IC_{50} (μM)	3.34	1.08	24.02	1.88
PDE-4B inhibition: IC_{50} (μM)	5.62	11.62	5.65	3.98
TNF α inhibition: IC_{50} (μM)	2.01	17.67	13.94	6.72

ARTICLE HISTORY

Received 14 January 2020
Revised 24 February 2020
Accepted 2 March 2020



KEYWORDS

Synthesis; 2-cyclopentyloxyanisole scaffold; antitumor activity; enzyme inhibition assay; docking study

Introduction

Cancer, the uncontrolled growth of cells that invade adjacent healthy tissues, is the most fatal disease in the world¹. Therefore, the design and synthesis of new molecules with promising and potential antitumor activity is of great importance^{1–10}. The clinical use of drug combinations has led to various side effects, whereas the use of single molecules that target multiple molecular mechanisms is the currently preferred therapeutic strategy and is under investigation by medicinal chemists^{11–13}.

Cyclooxygenase-2 isoenzyme (COX-2) inhibitors, such as celecoxib (**A**; Figure 1), have been reported to have antitumor activities^{8,14,15}. The COX-2 isoenzyme is overexpressed in numerous human cancers, such as breast, lung, hepatocellular, gastric, ovarian, prostate, and colon cancers^{8,14–16}. There are two anticancer mechanisms associated with COX-2 inhibition: the first, termed the COX-2-dependent anticancer mechanism, is selective inhibition with the restoration of normal apoptosis; the second is the COX-2-independent mechanism, which occurs through the induction

CONTACT Alaa A.-M. Abdel-Aziz  almoenes@ksu.edu.sa  Department of Pharmaceutical Chemistry, College of Pharmacy, P.O. Box 2457, King Saud University, Riyadh 11451, Saudi Arabia

© 2020 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

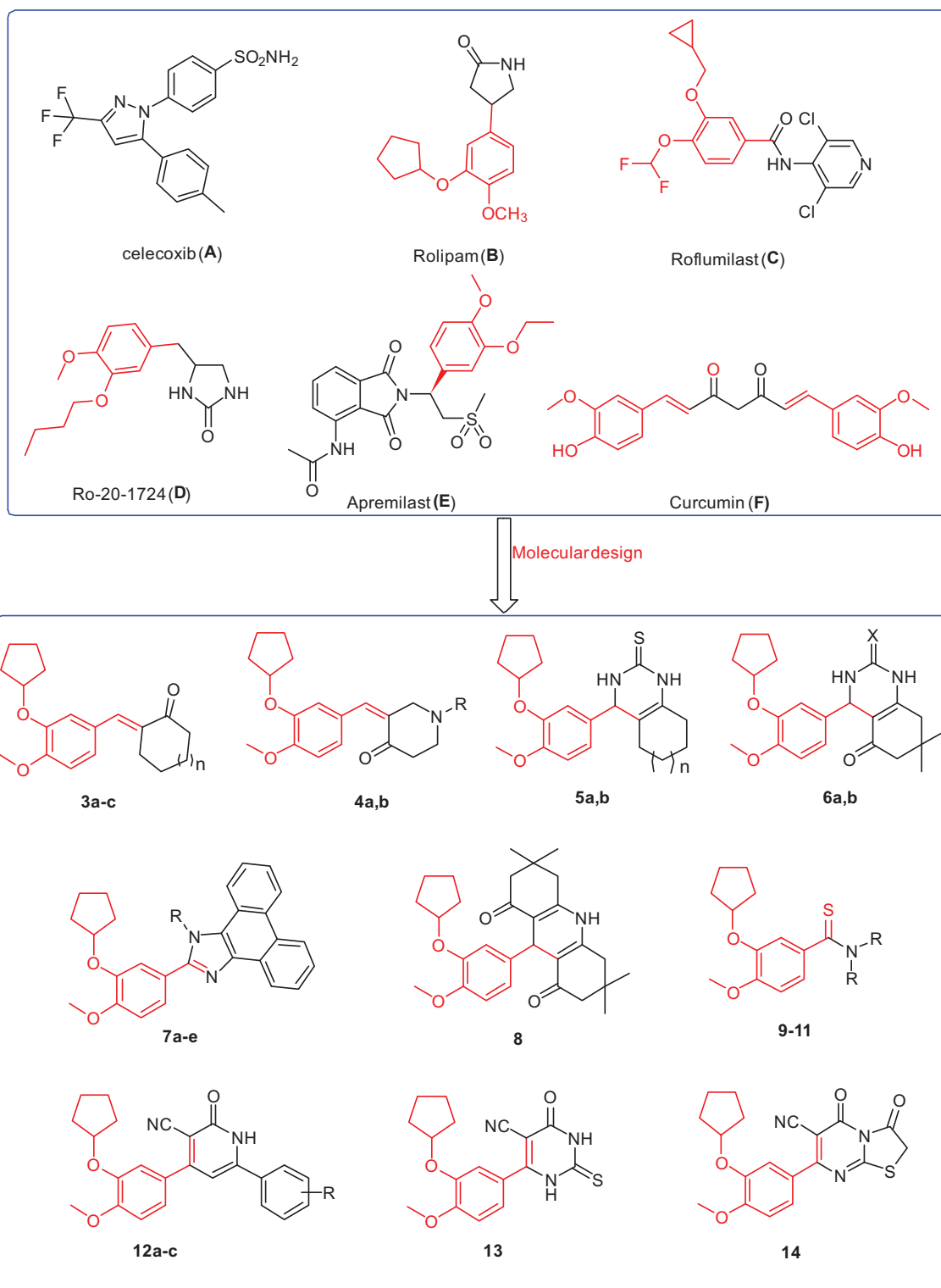


Figure 1. The structures of the reported antitumor agents (A–F) with COX-2 or PDE4 and the designed compounds 3–14.

of apoptosis or inhibition of cell proliferation¹⁷. These results indicated that COX-2 enzyme inhibition was an interesting molecular target for the treatment of cancer^{8,14–17}. In addition, phosphodiesterase isoenzyme 4 (PDE4) is responsible for inactivation and hydrolysis of 3',5'-cyclic adenosine monophosphate (cAMP) and subdivided into four subtypes, PDE4A to PDE4D^{18–20}. The secondary messenger cAMP is important for various cellular processes such as proliferation, growth, migration, differentiation, and apoptosis^{18–20}. These isoenzymes of cAMP-PDE expressed in several

cancer cells, such as colon cancer, melanoma, prostate cancer, myeloma, pancreatic cancer, B cell lymphoma, kidney cancer, and lung cancer^{18–27}. Recently, it was reported that PDE4 inhibitors possess antiproliferative effects, and inhibit the tumour cell growth of several types of cancers; thus, PDE4 inhibitors are a promising novel target for cancer therapy^{18–27}. Rolipam (**B**; Figure 1)^{18,22,23}, roflumilast (**C**; Figure 1)^{18,22,23}, Ro-20-1724 (**D**; Figure 1)²³, and apremilast (**E**; Figure 1)²³ are PDE4 inhibitors that reduced the growth of colon cancer cells through regulation of

the level of intracellular cAMP, leading to the induction of apoptosis. Roflumilast (**C**; Figure 1) was approved by FDA as a PDE4 inhibitor and used for the treatment of chronic obstructive pulmonary disease²⁶ and was successfully tested in lung cancer and B-cell lymphoma²⁵. In contrast, an increase in the level of intracellular cAMP by the inhibition of PDE4 isoenzymes leads to inhibition of the production of tumour necrosis factor- α (TNF- α)²⁸. TNF- α is a central mediator of inflammation, and thus provides a molecular link between chronic inflammation and the development of malignancies^{29–32}. In addition, TNF- α is overexpressed in various cancer cells such as liver cancer, kidney cancer, and gallbladder cancer and supports tumour growth and metastasis^{29–32}. The aforementioned results indicated that the inhibition of PDE4 enzyme activity^{18–27} and the suppression of the production of TNF- α ^{28–32} are an interesting target for the treatment of cancer.

Compounds containing 2-cyclopentyloxyanisole analogues are reported to be PDE4 inhibitors with anticancer activities, such as rolipram (**B**; Figure 1), roflumilast (**C**; Figure 1), and apremilast (**E**; Figure 1)^{18,22,23}. Meanwhile, compounds bearing chalcone structures constitute the main building block of several natural products with potential antitumor activity, such as curcumin (**F**; Figure 1)^{7,9,33}. It was reported that curcumin exerts antitumor activity against colon cancer through inhibition of the COX-2 isoenzyme³⁴. Recently, curcumin was shown to have *in vitro* anti-angiogenic effects and *in vivo* anticancer activity through the inhibition of PDE isoenzymes³⁵. Indeed, several compounds possessing heterocyclic core structures, such as quinazoline^{2–4}, quinoline^{9,10}, pyrimidine³⁶, pyridine⁹, imidazole⁶, have potential antitumor activity.

Based on the aforementioned data, and to continue our efforts to develop new molecules as effective antitumor agents, we have reported (i) the synthesis of new derivatives incorporating chalcone derivatives based on the 2-cyclopentyloxyanisole core structure; (ii) the preparation of 2-cyclopentyloxyanisole bearing heterocyclic moieties such as quinazoline, quinoline, pyridine, pyrimidine, and imidazole ring systems; (iii) the synthesis of 2-cyclopentyloxyanisole bearing thioamide moieties; (iv) a comparison of the effectiveness of heterocyclic derivatives versus the chalcone and thioamide derivatives; and (v) an evaluation of the *in vitro*

antitumor activity against different human cancers: liver cancer (HePG2 cell line), colon cancer (HCT-116 cell line), breast cancer (MCF-7 cell line), prostate cancer (PC3 cell line), and cervical cancer (HeLa cell line); (vi) a study of the structure–activity relationship (SAR) for the synthesised 2-cyclopentyloxyanisole structure with diverse substituent moieties regarding antitumor activities; (vii) an evaluation of the *in vitro* COX-2 and PDE4B, and TNF- α inhibitory abilities of the most promising compounds; and (viii) a molecular modelling study of the binding mode of the target molecules in the COX-2 and PDE 4 pockets.

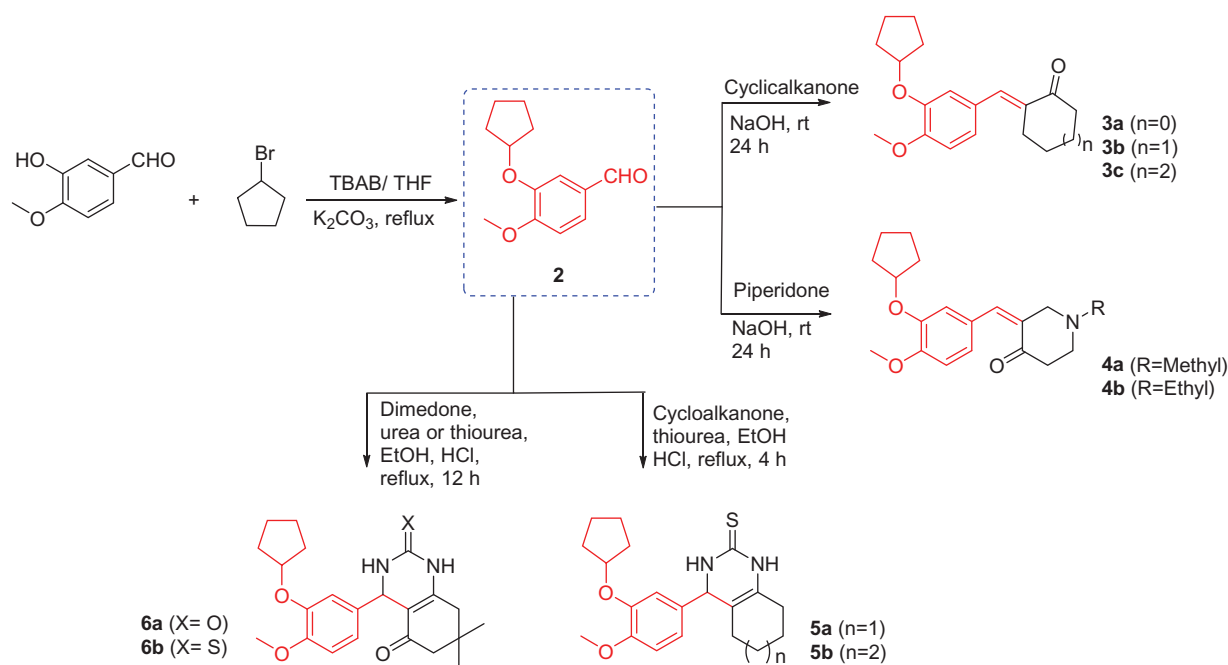
Experimental methods

Chemistry

Melting points were recorded by using a Fisher-Johns melting point apparatus and were uncorrected. ¹H NMR and ¹³C NMR spectra (500 MHz) were obtained in DMSO-*d*₆ and CHCl₃-*d* on a JOEL Nuclear Magnetic Resonance 500 spectrometer at Mansoura University, Faculty of Science, Egypt. Mass spectrometric analyses were performed by using a JEOL JMS-600H spectrometer at Mansoura University, Faculty of Science (Assiut, Egypt). The reaction times were determined by using a TLC technique on silica gel plates (60 F245, Merck, Kenilworth, NJ) and the spots were visualised by UV irradiation at 366 nm or 245 nm. The synthesis of 3-(cyclopentyloxy)-4-methoxybenzaldehyde (**2**) and 6-(3-(cyclopentyloxy)-4-methoxyphenyl)-4-oxo-2-thioxo-1,2,3,4-tetrahydropyrimidine-5-carbonitrile (**13**) are described elsewhere^{18,37,38}.

Synthesis of compounds 3a–c, 4a, and 4b

To a mixture of 3-(cyclopentyloxy)-4-methoxybenzaldehyde (**2**) (1.0 mmol, 0.22 g) and cyclic ketones (3.0 mmol) in ethanol (15 ml), NaOH (2.0 mmol, 0.08 g) was added whilst stirring at 0 °C. The reaction mixture was then stirred at room temperature for 24 h, poured on crushed ice, and the obtained solid was filtered, washed with water, and recrystallised from methanol (Scheme 1).



Scheme 1. Synthesis of the designed compounds 3–6.

2-(3-(Cyclopentylloxy)-4-methoxybenzylidene)cyclopentanone (3a)

Yield, 65%; melting point [MP] 252–254 °C. ¹H NMR spectrum (DMSO-d₆), δ, ppm: 1.53–1.56 (2H, m), 1.62–1.65 (4H, m), 1.70–1.74 (4H, m), 1.86–1.89 (2H, m), 2.89–2.91 (2H, m), 3.87 (3H, s), 4.74–4.77 (1H, m), 7.05–7.07 (1H, d, *J* = 8.0 Hz), 7.07–7.08 (1H, d, *J* = 8.0 Hz), 7.21 (1H, s), 7.74 (1H, s). IR spectrum, ν, cm⁻¹: 2957, 2872, 1703, 1620, 954, 642. C₁₈H₂₂O₃ MS: *m/z* 287 (M⁺+1), 286 (M⁺).

2-(3-(Cyclopentylloxy)-4-methoxybenzylidene)cyclohexanone (3b)

Yield, 60%; MP 245–247 °C. ¹H NMR spectrum (DMSO-d₆), δ, ppm: 1.60–1.68 (6H, m), 1.81–1.93 (8H, m), 2.91–2.93 (2H, m), 3.85 (3H, s), 4.75–4.79 (1H, m), 7.03–7.04 (1H, d, *J* = 8.1 Hz), 7.06–7.07 (1H, d, *J* = 8.0 Hz), 7.25 (1H, s), 7.77 (1H, s). IR spectrum, ν, cm⁻¹: 2953, 2870, 1705, 1621, 951, 638. C₁₉H₂₄O₃ MS: *m/z* 301 (M⁺+1), 300 (M⁺).

2-(3-(Cyclopentylloxy)-4-methoxybenzylidene)cycloheptanone (3c)

Yield, 63%; MP 250–252 °C. ¹H NMR spectrum (DMSO-d₆), δ, ppm: 1.50–1.60 (3H, m), 1.80–1.81 (2H, m), 1.82–1.85 (6H, m), 1.89–1.91 (5H, m), 2.68–2.71 (2H, m), 3.86 (3H, s), 4.73–4.75 (1H, m), 6.84 (1H, s), 6.86–6.88 (1H, d, *J* = 7.9 Hz), 6.89–6.90 (1H, d, *J* = 8.0 Hz), 7.44 (1H, s). IR spectrum, ν, cm⁻¹: 2950, 2871, 1710, 1616, 954, 639. C₂₀H₂₆O₃ MS: *m/z* 315 (M⁺+1), 314 (M⁺).

3-(3-(Cyclopentylloxy)-4-methoxybenzylidene)-1-methylpiperidin-4-one (4a)

Yield, 70%; MP 253–255 °C. ¹H NMR spectrum (DMSO-d₆), δ, ppm: 1.55–1.58 (2H, m), 1.64–1.73 (4H, m), 1.79–1.86 (4H, m), 2.15 (2H, s), 2.42 (3H, s), 2.91–2.95 (2H, m), 3.71 (3H, s), 4.74–4.78 (1H, q, *J* = 5.5 Hz), 6.66–6.74 (2H, m), 6.89–6.94 (2H, m). IR spectrum, ν, cm⁻¹: 2955, 2872, 1708, 1620, 956, 640. C₁₉H₂₅NO₃ MS: *m/z* 317 (M⁺+2), 316 (M⁺+1), 315 (M⁺).

3-(3-(Cyclopentylloxy)-4-methoxybenzylidene)-1-ethylpiperidin-4-one (4b)

Yield, 68%; MP 249–251 °C. ¹H NMR spectrum (DMSO-d₆), δ, ppm: 1.29–1.32 (3H, t, *J* = 4.5 Hz), 1.52–1.54 (2H, m), 1.62–1.68 (4H, m), 1.81–1.85 (4H, m), 2.43–2.45 (2H, m), 2.88–2.92 (2H, m), 2.93–2.95 (2H, m), 3.73 (3H, s), 4.73–4.79 (1H, m), 6.66–6.77 (2H, m), 6.86–6.95 (2H, m). IR spectrum, ν, cm⁻¹: 2954, 2870, 1708, 1624, 958, 644. C₂₀H₂₇NO₃ MS: *m/z* 331 (M⁺+2), 330 (M⁺+1), 329 (M⁺).

Synthesis of compounds 5a and 5b

To a solution of 3-(cyclopentylloxy)-4-methoxybenzaldehyde (**2**) (5 mmol, 1.1 g), thiourea (5 mmol, 380 mg), and cyclic ketones (7.5 mmol) in ethanol (25 ml), four drops of concentrated hydrochloric acid were added. The reaction mixture was heated under reflux for 4 h, and the solvent was evaporated under vacuum. The obtained solid was dissolved in H₂O and the solution was neutralised with ammonia solution. The precipitated solid was filtered, washed with water, and crystallised from ethanol (Scheme 1).

4-(3-(Cyclopentylloxy)-4-methoxyphenyl)-3,4,5,6,7,8-hexahydroquinazoline-2(1H)-thione (5a)

Yield, 55%; MP 199–201 °C. ¹H NMR spectrum (CHCl₃-d), δ, ppm: 0.80–0.86 (4H, m), 1.20–1.25 (4H, m), 1.83–1.89 (4H, m), 1.91–1.95

(4H, m), 3.83 (3H, s), 4.67 (1H, s), 4.78–4.93 (1H, m), 6.76 (1H, s), 6.80 (1H, s), 6.82 (1H, s), 6.83–6.86 (1H, d, *J* = 8.0 Hz), 7.13–7.16 (1H, d, *J* = 8.1 Hz). IR spectrum, ν, cm⁻¹: 3422, 3240, 2960, 2871, 1630, 1260. C₂₀H₂₆N₂O₂S MS: *m/z* 360 (M⁺+2), 359 (M⁺+1), 358 (M⁺).

4-(3-(Cyclopentylloxy)-4-methoxyphenyl)-1,3,4,5,6,7,8,9-octahydro-2H-cyclohepta[d]pyrimidine-2-thione (5b)

Yield, 52%; MP 205–207 °C. ¹H NMR spectrum (CHCl₃-d), δ, ppm: 0.83–0.88 (6H, m), 1.19–1.24 (2H, m), 1.25–1.29 (2H, m), 1.61–1.66 (4H, m), 1.82–1.93 (4H, m), 3.84 (3H, s), 4.67 (1H, s), 4.78–4.92 (1H, m), 6.76 (1H, s), 6.81 (1H, s), 6.84 (1H, s), 6.81–6.85 (1H, d, *J* = 7.9 Hz), 7.10–7.12 (1H, d, *J* = 8.0 Hz). IR spectrum, ν, cm⁻¹: 3426, 3243, 2963, 2873, 1632, 1262. C₂₁H₂₈N₂O₂S MS: *m/z* 374 (M⁺+2), 373 (M⁺+1), 372 (M⁺).

Synthesis of compounds 6a and 6b

To a solution of 3-(cyclopentylloxy)-4-methoxybenzaldehyde (**2**) (5 mmol, 1.1 g), urea or thiourea (5 mmol), and dimedone (7.5 mmol, 1.1 g) in ethanol (25 ml), four drops of concentrated hydrochloric acid were added. The reaction mixture was heated under reflux for 12 h and the solvent was evaporated under vacuum. The obtained solid was dissolved in H₂O and the solution was neutralised by using ammonia solution. The precipitated solid was filtered, washed with water, and re-crystallised from DMF (Scheme 1).

4-(3-(Cyclopentylloxy)-4-methoxyphenyl)-7,7-dimethyl-4,6,7,8-tetrahydroquinazoline-2,5(1H,3H)-dione (6a)

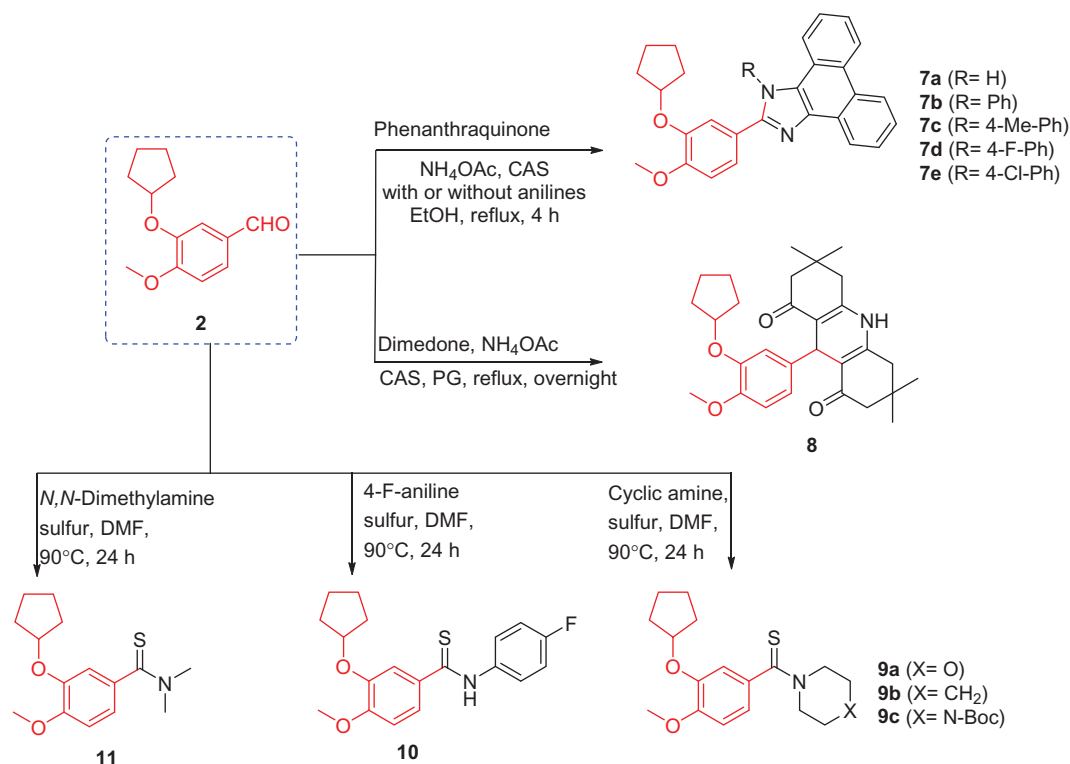
Yield, 80%; MP 230–232 °C. ¹H NMR spectrum (DMSO-d₆), δ, ppm: 0.99 (3H, s), 1.02 (3H, s), 1.05 (1H, s), 1.54–1.58 (4H, m), 1.78–1.89 (4H, m), 2.40–2.43 (3H, t, *J* = 6.5 Hz), 3.74 (3H, s), 4.67 (1H, s), 4.72–4.76 (1H, q, *J* = 3.5 Hz), 6.67 (1H, s), 6.68 (1H, s), 6.70 (1H, s), 6.73–6.74 (1H, d, *J* = 6.5 Hz), 6.75–6.76 (1H, d, *J* = 6.5 Hz). IR spectrum, ν, cm⁻¹: 3420, 3243, 2957, 2872, 1620, 1260. C₂₂H₂₈N₂O₄ MS: *m/z* 386 (M⁺+2), 385 (M⁺+1), 384 (M⁺).

4-(3-(Cyclopentylloxy)-4-methoxyphenyl)-7,7-dimethyl-2-thioxo-2,3,4,6,7,8-hexahydroquinazolin-5(1H)-one (6b)

Yield, 78%; MP 233–235 °C. ¹H NMR spectrum (DMSO-d₆), δ, ppm: 0.99 (3H, s), 1.01 (3H, s), 1.55 (2H, s), 1.77–1.82 (4H, m), 1.87–1.92 (4H, m), 2.44 (2H, s), 3.73 (3H, s), 4.68 (1H, s), 4.71–4.73 (1H, m), 6.66 (1H, s), 6.68 (1H, s), 6.69 (1H, s), 6.72–6.74 (1H, d, *J* = 7.5 Hz), 6.75–6.77 (1H, d, *J* = 6.5 Hz). IR spectrum, ν, cm⁻¹: 3425, 3245, 2960, 2870, 1623, 1264. C₂₂H₂₈N₂O₃S MS: *m/z* 402 (M⁺+2), 401 (M⁺+1), 400 (M⁺).

Synthesis of compound 7a

A mixture of 3-(cyclopentylloxy)-4-methoxybenzaldehyde (**2**) (5 mmol, 1.1 g), 9,10-phenanthraquinone (5 mmol, 1.04 g), ammonium acetate (15 mmol, 1.17 g), and CAS or iodine (5 mol%) in ethanol (25 ml) was heated under reflux for 4 h. The reaction mixture was cooled to room temperature, poured on crushed ice, and extracted with ethyl acetate. The extract was evaporated under vacuum to yield a precipitate, which was collected and re-crystallised from acetone (Scheme 2).



Scheme 2. Synthesis of the designed compounds 7–11.

2-(3-(Cyclopentyloxy)-4-methoxyphenyl)-1*H*-phenanthro[9,10-*d*]imidazole (7a)

Yield, 85%; MP 290–292 °C. ¹H NMR spectrum (DMSO-*d*₆), δ , ppm: 1.62 (2H, s), 1.79–1.82 (4H, m), 1.97 (2H, s), 3.84 (3H, s), 4.97 (1H, s), 7.17–7.18 (1H, d, J =8.0 Hz), 7.62 (2H, s), 7.72 (2H, s), 7.87 (2H, s), 8.55–8.56 (2H, d, J =6.5 Hz), 8.83–8.85 (2H, d, J =7.5 Hz). ¹³C NMR spectrum (DMSO-*d*₆), δ , ppm: 18.56, 23.68, 32.38, 55.67, 56.03, 79.95, 112.36, 112.85, 119.25, 121.88, 123.71, 125.02, 126.95, 127.07, 136.83, 147.14, 149.34, 150.92. IR spectrum, ν , cm⁻¹: 3422, 2964, 2864, 930, 615. C₂₇H₂₄N₂O₂ MS: m/z 409 (M⁺+1), 408 (M⁺).

Synthesis of compounds 7b–e

A mixture of 3-(cyclopentyloxy)-4-methoxybenzaldehyde (**2**) (5 mmol, 1.1 g), 9,10-phenanthraquinone (5 mmol, 1.04 g), ammonium acetate (15 mmol, 1.17 g), the appropriate aniline (5 mmol), and CAS or iodine (5 mol%) in ethanol (25 ml) was heated under reflux for 4 h. The formed precipitate was filtered, washed with ethanol, and crystallised from DMF (Scheme 2).

2-(3-(Cyclopentyloxy)-4-methoxyphenyl)-1-phenyl-1*H*-phenanthro[9,10-*d*]imidazole (7b)

Yield, 82%; MP 295–297 °C. ¹H NMR spectrum (DMSO-*d*₆), δ , ppm: 1.52 (2H, s), 1.60–1.65 (4H, m), 1.70–1.71 (2H, d, J =6.5 Hz), 3.74 (3H, s), 4.49 (1H, s), 6.96–6.98 (2H, d, J =8.0 Hz), 7.01–7.03 (1H, d, J =8.0 Hz), 7.29–7.31 (2H, d, J =7.5 Hz), 7.51–7.54 (1H, t, J =7.5 Hz), 7.62–7.77 (7H, m), 8.67–8.68 (1H, d, J =7.5 Hz), 8.85–8.87 (1H, d, J =8.5 Hz), 8.90–8.92 (1H, d, J =8.5 Hz). IR spectrum, ν , cm⁻¹: 2960, 2869, 932, 618. C₃₃H₂₈N₂O₂ MS: m/z 485 (M⁺+1), 484 (M⁺).

2-(3-(Cyclopentyloxy)-4-methoxyphenyl)-1-(4-methylphenyl)-1*H*-phenanthro[9,10-*d*]imidazole (7c)

Yield, 80%; MP 291–294 °C. ¹H NMR spectrum (DMSO-*d*₆), δ , ppm: 1.52 (4H, s), 1.64–1.68 (4H, m), 2.07 (3H, s), 3.75 (3H, s), 4.36 (1H, s), 6.89 (1H, s), 6.98–6.70 (1H, d, J =8.5 Hz), 7.12–7.14 (1H, d, J =8.0 Hz), 7.32–7.37 (2H, q, J =9.0 Hz), 7.50–7.54 (3H, q, J =7.5 Hz), 7.56–7.58 (2H, d, J =8.0 Hz), 7.65–7.68 (1H, t, J =7.5 Hz), 7.74–7.77 (1H, t, J =7.5 Hz), 8.66–8.67 (1H, d, J =7.5 Hz), 8.85–8.86 (1H, d, J =8.5 Hz), 8.90–8.92 (1H, d, J =9.0 Hz). IR spectrum, ν , cm⁻¹: 2968, 2877, 942, 632. C₃₄H₃₀N₂O₂ MS: m/z 499 (M⁺+1), 498 (M⁺).

2-(3-(Cyclopentyloxy)-4-methoxyphenyl)-1-(4-fluorophenyl)-1*H*-phenanthro[9,10-*d*]imidazole (7d)

Yield, 86%; MP 290–292 °C. ¹H NMR spectrum (DMSO-*d*₆), δ , ppm: 1.50–1.54 (2H, m), 1.60–1.64 (4H, m), 1.68–1.71 (2H, m), 3.86 (3H, s), 4.94–4.99 (1H, m), 6.85 (1H, s), 6.94–6.96 (1H, d, J =7.5 Hz), 7.10–7.11 (1H, d, J =7.5 Hz), 7.29–7.31 (2H, m), 7.40–7.49 (5H, m), 7.62–7.64 (1H, t, J =8.0 Hz), 7.71–7.74 (1H, t, J =8.0 Hz), 8.65–8.66 (1H, d, J =8.5 Hz), 8.81–8.83 (1H, d, J =9.0 Hz), 8.86–8.87 (1H, d, J =8.5 Hz). IR spectrum, ν , cm⁻¹: 2968, 2875, 940, 636. C₃₃H₂₇FN₂O₂ MS: m/z 505 (M⁺+3), 503 (M⁺+1), 502 (M⁺).

2-(3-(Cyclopentyloxy)-4-methoxyphenyl)-1-(4-chlorophenyl)-1*H*-phenanthro[9,10-*d*]imidazole (7e)

Yield, 84%; MP 294–296 °C. ¹H NMR spectrum (DMSO-*d*₆), δ , ppm: 1.52–1.56 (2H, m), 1.60–1.66 (4H, m), 1.69–1.73 (2H, m), 3.83 (3H, s), 4.91–4.95 (1H, m), 6.80 (1H, s), 6.94–6.96 (1H, d, J =8.0 Hz), 7.12–7.14 (1H, d, J =8.0 Hz), 7.25–7.29 (2H, m), 7.40–7.47 (5H, m), 7.61–7.63 (1H, t, J =7.5 Hz), 7.72–7.73 (1H, t, J =7.0 Hz), 8.65–8.67 (1H, d, J =8.50 Hz), 8.79–8.81 (1H, d, J =8.5 Hz), 8.84–8.86 (1H, d,

$J=9.0$ Hz). IR spectrum, ν , cm^{-1} : 2965, 2873, 942, 635. $\text{C}_{33}\text{H}_{27}\text{ClN}_2\text{O}_2$ MS: m/z 520 (M^++2), 519 (M^++1), 518 (M^+).

Synthesis of compound 8

To a solution of 3-(cyclopentyloxy)-4-methoxybenzaldehyde (**2**) (5 mmol, 1.1 g), dimedone (10 mmol, 1.47 g), and ammonium acetate (5 mmol, 0.39 g) in propylene glycol (20 ml), CAS or iodine (5 mol%) was added. The reaction mixture was heated under reflux overnight, cooled to room temperature, and poured on crushed ice. The obtained solid was filtered, washed with water, and re-crystallised from ethanol (Scheme 2).

9-(3-(Cyclopentyloxy)-4-methoxyphenyl)-3,3,6,6-tetramethyl-3,4,6,7,9,10-hexahydroacridine-1,8(2H,5H)-dione (8)

Yield, 77%; MP 286–287 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 0.86 (6H, s), 0.99 (6H, s), 1.53–1.55 (2H, m), 1.63–1.67 (4H, m), 1.69–1.71 (2H, m), 1.98–2.00 (2H, d, $J=6.5$ Hz), 2.14–2.15 (2H, d, $J=5.5$ Hz), 2.29–2.30 (2H, d, $J=5.5$ Hz), 2.41–2.43 (2H, d, $J=6.5$ Hz), 3.63 (3H, s), 4.55–4.58 (1H, q, $J=6.0$ Hz), 4.72 (1H, s), 6.60–6.62 (1H, d, $J=8.0$ Hz), 6.69–6.71 (2H, d, $J=6.0$ Hz), 9.26 (1H, s). ^{13}C NMR spectrum (DMSO- d_6), δ , ppm: 23.52, 26.39, 29.16, 31.89, 32.07, 32.28, 50.27, 55.38, 79.38, 111.44, 111.59, 115.02, 119.52, 139.68, 146.14, 147.58, 148.95, 149.07, 194.39. IR spectrum, ν , cm^{-1} : 3420, 2968, 2872, 1735, 1738. $\text{C}_{29}\text{H}_{37}\text{NO}_4$ MS: m/z 464 (M^++1), 463 (M^+).

Synthesis of compounds 9a–c, 10, and 11

A solution of 3-(cyclopentyloxy)-4-methoxybenzaldehyde (**2**) (5 mmol, 1.1 g), appropriate amine derivatives (25 mmol), and precipitated sulphur (12.5 mmol, 0.40 g) in DMF (15 ml) was heated at 90 °C for 24 h. The reaction was monitored by TLC and, after completion, was cooled to room temperature and poured on crushed ice. The formed precipitate was filtered, washed with water, and re-crystallised from methanol (Scheme 2).

(3-(Cyclopentyloxy)-4-methoxyphenyl)(morpholino)methanethione (9a)

Yield, 70%; MP 190–192 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.55–1.56 (2H, d, $J=2.5$ Hz), 1.67–1.70 (4H, m), 1.86–1.87 (2H, d, $J=4.0$ Hz), 3.58–3.59 (4H, d, $J=3.0$ Hz), 3.75 (3H, s), 4.27 (4H, s),

4.75–4.78 (1H, t, $J=5.5$ Hz), 6.83–6.85 (2H, t, $J=8.0$ Hz), 6.92–6.94 (1H, d, $J=8.0$ Hz). IR spectrum, ν , cm^{-1} : 2956, 2848, 1516, 1223, 1163, 925, 813, 631. $\text{C}_{17}\text{H}_{23}\text{NO}_3\text{S}$ MS: m/z 323 (M^++2), 322 (M^++1), 321 (M^+).

(3-(Cyclopentyloxy)-4-methoxyphenyl)(piperidin-1-yl)methanethione (9b)

Yield, 72%; MP 193–195 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.50–1.57 (4H, q, $J=6.5$ Hz), 1.66–1.69 (8H, t, $J=6.0$ Hz), 1.85–1.86 (2H, d, $J=4.0$ Hz), 3.52–3.53 (2H, d, $J=5.0$ Hz), 3.75 (3H, s), 4.22–4.23 (2H, d, $J=5.5$ Hz), 4.75–4.78 (1H, t, $J=6.0$ Hz), 6.78–6.80 (2H, d, $J=7.5$ Hz), 6.91–6.92 (1H, d, $J=8.5$ Hz). IR spectrum, ν , cm^{-1} : 2955, 2846, 1512, 1225, 1166, 920, 810, 630. $\text{C}_{18}\text{H}_{25}\text{NO}_2\text{S}$ MS: m/z 321 (M^++2), 320 (M^++1), 319 (M^+).

tert-Butyl 4-(3-(cyclopentyloxy)-4-methoxyphenylcarbonothioyl)piperazine-1-carboxylate (9c)

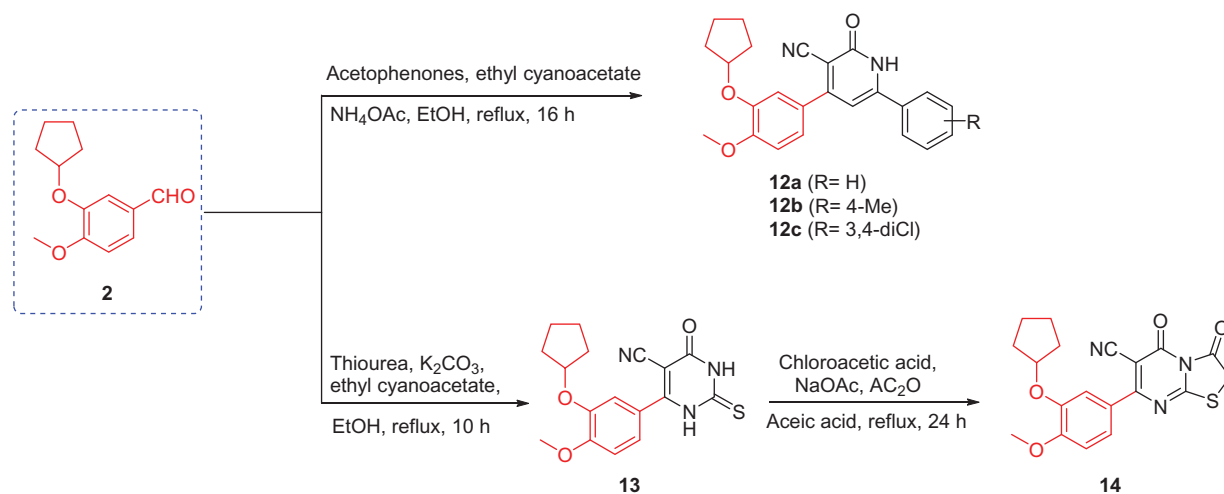
Yield, 68%; MP 191–193 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.39 (9H, s), 1.53–1.55 (2H, m), 1.68–1.70 (4H, t, $J=4.5$ Hz), 1.86–1.87 (2H, m, $J=4.5$ Hz), 3.30–3.33 (4H, m), 3.56–3.59 (4H, m), 3.75 (3H, s), 4.75–4.77 (1H, t, $J=5.5$ Hz), 6.84–6.86 (2H, t, $J=7.0$ Hz), 6.92–6.95 (1H, t, $J=8.0$ Hz). IR spectrum, ν , cm^{-1} : 2958, 2848, 1514, 1224, 1160, 929, 812, 633. $\text{C}_{22}\text{H}_{32}\text{N}_2\text{O}_4\text{S}$ MS: m/z 421 (M^++1), 420 (M^+).

3-(Cyclopentyloxy)-N-(4-fluorophenyl)-4-methoxybenzothioamide (10)

Yield, 75%; MP 194–196 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.55–1.58 (2H, d, $J=6.0$ Hz), 1.70–1.74 (4H, m), 1.91–1.94 (2H, d, $J=4.0$ Hz), 3.81 (3H, s), 4.83–4.85 (1H, t, $J=5.5$ Hz), 7.06–7.07 (1H, d, $J=7.5$ Hz), 7.19–7.23 (2H, m), 7.25–7.28 (2H, m), 7.34 (1H, s), 7.49–7.50 (1H, d, $J=7.5$ Hz), 8.48 (1H, s). IR spectrum, ν , cm^{-1} : 2951, 2848, 1510, 1225, 1162, 921, 814, 633. $\text{C}_{19}\text{H}_{20}\text{FNO}_2\text{S}$ MS: m/z 347 (M^++2), 346 (M^++1), 345 (M^+).

3-(Cyclopentyloxy)-4-methoxy-N,N-dimethylbenzothioamide (11)

Yield, 71%; MP 189–191 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.52–1.55 (2H, m), 1.68–1.70 (4H, t, $J=5.0$ Hz), 1.71–1.73 (2H, m), 3.16 (3H, s), 3.46 (3H, s), 3.75 (3H, s), 4.75–4.77 (1H, t, $J=5.5$ Hz), 6.84–6.87 (2H, m), 6.91–6.92 (1H, d, $J=8.5$ Hz). ^{13}C NMR spectrum



Scheme 3. Synthesis of the designed compounds 12–14.

(DMSO- d_6), δ , ppm: 23.53, 32.15, 43.09, 43.98, 55.57, 79.47, 111.30, 113.04, 118.95, 135.54, 145.96, 149.78, 198.94. IR spectrum, ν , cm^{-1} : 2956, 2851, 1514, 1229, 1159, 920, 814, 636. $\text{C}_{15}\text{H}_{21}\text{NO}_2\text{S}$ MS: m/z 280 (M^++1), 279 (M^+).

Synthesis of compounds 12a–c

A mixture of 3-(cyclopentyloxy)-4-methoxybenzaldehyde (**2**) (2 mmol, 0.44 g), the appropriate acetophenone derivatives (2 mmol), ethyl cyanoacetate (2 mmol, 0.23 g), and ammonium acetate (16 mmol, 1.24 g) in ethanol (10 ml) was heated under reflux for 16 h. The reaction mixture was cooled to room temperature, filtered, washed with ethanol, and re-crystallised from acetone (Scheme 3).

4-(3-(Cyclopentyloxy)-4-methoxyphenyl)-2-oxo-6-phenyl-1,2-dihydropyridine-3-carbonitrile (12a)

Yield, 88%; MP > 300 °C; ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.57–1.58 (2H, d, $J=6.0$ Hz), 1.71–1.76 (4H, m), 1.89–1.91 (2H, t, $J=11.5$ Hz), 3.82 (3H, s), 4.88–4.90 (1H, m), 6.77 (1H, s), 7.10–7.12 (1H, d, $J=10.0$ Hz), 7.30 (2H, s), 7.33 (1H, s), 7.51–7.56 (3H, m), 7.87–7.88 (2H, d, $J=5.0$ Hz). ^{13}C NMR spectrum (DMSO- d_6), δ , ppm: 23.62, 32.27, 55.71, 79.71, 112.04, 114.40, 116.94, 121.41, 127.78, 128.08, 128.94, 131.13, 146.82, 151.58. IR spectrum, ν , cm^{-1} : 3445, 2964, 2220, 1630, 1510, 1265, 810. $\text{C}_{24}\text{H}_{22}\text{N}_2\text{O}_3$ MS: m/z 387 (M^++1), 386 (M^+).

4-(3-(cyclopentyloxy)-4-methoxyphenyl)-2-oxo-6-(*p*-tolyl)-1,2-dihydropyridine-3-carbonitrile (12b)

Yield, 84%; MP > 300 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.56–1.57 (2H, t, $J=4.0$ Hz), 1.70–1.76 (4H, m), 1.88–1.92 (2H, m), 2.36 (3H, s), 3.81 (3H, s), 4.86–4.89 (1H, m), 6.74 (1H, s), 7.10–7.12 (1H, d, $J=8.5$ Hz), 7.28–7.30 (2H, q, $J=4.0$ Hz), 7.31 (1H, s), 7.33–7.34 (2H, d, $J=8.0$ Hz), 7.77–7.79 (2H, d, $J=7.0$ Hz). ^{13}C NMR spectrum (DMSO- d_6), δ , ppm: 20.92, 23.59, 32.24, 55.69, 79.69, 112.02, 114.39, 116.98, 121.34, 127.63, 128.14, 129.49, 141.27, 146.77, 151.52. IR spectrum, ν , cm^{-1} : 3447, 2959, 2216, 1629, 1514, 1263, 807; $\text{C}_{25}\text{H}_{24}\text{N}_2\text{O}_3$ MS: m/z 401 (M^++1), 400 (M^+).

4-(3-(Cyclopentyloxy)-4-methoxyphenyl)-6-(3,4-dichlorophenyl)-2-oxo-1,2-dihydropyridine-3-carbonitrile (12c)

Yield, 81%; MP > 300 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.52–1.57 (2H, m), 1.69–1.76 (4H, m), 1.89–1.94 (2H, m), 3.82 (3H, s), 4.86–4.89 (1H, m), 7.11–7.13 (2H, d, $J=8.0$ Hz), 7.30–7.31 (2H, d, $J=2.5$ Hz), 7.31–7.32 (1H, d, $J=2.5$ Hz), 7.33–7.34 (1H, d, $J=2.5$ Hz), 7.79–7.81 (2H, d, $J=9.0$ Hz). IR spectrum, ν , cm^{-1} : 3443, 2964, 2222, 1635, 1508, 1268, 808. $\text{C}_{24}\text{H}_{20}\text{Cl}_2\text{N}_2\text{O}_3$ MS: m/z 456 (M^++2), 454 (M^+).

Synthesis of compound 14

A mixture of compound **13** (1 mmol, 0.34 g), chloroacetic acid (1 mmol, 0.10 g), anhydrous sodium acetate (4 mmol, 0.33 g) in acetic anhydride (2 ml), and glacial acetic acid (10 ml) was heated under reflux for 24 h. The reaction mixture was cooled to room temperature and poured into crushed ice. The obtained solid was filtered, washed with water, and crystallised from methanol (Scheme 3).

7-(3-(Cyclopentyloxy)-4-methoxyphenyl)-3,5-dioxo-2,3-dihydro-5H-thiazolo[3,2-*a*]pyrimidine-6-carbonitrile (14)

Yield, 55%; MP 265–267 °C. ^1H NMR spectrum (DMSO- d_6), δ , ppm: 1.50–1.53 (2H, m), 1.69–1.72 (4H, m), 1.88–1.90 (2H, m), 3.79 (3H, s), 4.21 (2H, s), 4.76–4.79 (1H, m), 7.01 (1H, s), 7.35 (1H, s), 7.38 (1H, s). IR spectrum, ν , cm^{-1} : 2962, 2229, 1655, 16,450, 1217, 986. $\text{C}_{19}\text{H}_{17}\text{N}_3\text{O}_4\text{S}$ MS: m/z 385 (M^++2), 383 (M^+).

Biological evaluation

In vitro antitumor activity evaluation assay

The antitumor activity was performed by using the tetrazolium salt 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2*H*-tetrazolium bromide (MTT) assay in accordance with an established method³⁹.

In vitro COX-2 inhibition assay

The colorimetric COX-2 inhibition assay was performed in accordance with the manufacturer's instructions (Kit 560101, Cayman Chemical, Ann Arbor, MI)^{40–42}.

In vitro TNF- α inhibition assay

The concentration of TNF- α was measured by human-specific sandwich enzyme-linked immunosorbent assay (ELISA) in accordance with the manufacturer's instructions (no. 589201, Cayman Chemical, Ann Arbor, MI)^{43,44}.

Docking methodology

The molecular docking technique was performed by using MOE 2008.10, from the Chemical Computing Group Inc.⁴⁵ in accordance with previously established methods^{18,40–42}.

Results and discussion

Chemistry

The synthetic strategies used to obtain the target compounds are presented in Schemes 1–3. The O-alkylation of isovanillin (**1**) with bromocyclopentane was successively conducted in the presence of K_2CO_3 and a phase transfer catalyst tetrabutylammonium bromide (TBAB) in THF to obtain the key intermediate 3-cyclopentyloxy-4-methoxybenzaldehyde (**2**) that provided the core structure of phosphodiesterase-4 inhibitors³⁷. Tetrabutylammonium bromide successively exhibited the character of phase transfer catalyst in an environmentally friendly procedure under mild conditions³⁷.

Synthesis of compounds 3–6

First, the cyclocondensation of 3-cyclopentyloxy-4-methoxybenzaldehyde (**2**)³⁷ with cyclic ketones in the ethanolic solution of sodium hydroxide afforded chalcones **3a–c** and **4a,b** in good yields (Scheme 1). In addition, the one-pot cyclocondensation reaction of **2** with the cyclic ketone (cyclohexanone/cycloheptanone/dimedone) and urea or thiourea in ethanol containing few drops of concentrated hydrochloric acid yielded the quinazoline derivatives **5a,b** and **6a,b**⁴⁶, as shown in Scheme 1.

Synthesis of compounds 7–11

The synthesis of imidazole via multicomponent reactions (MCRs) was achieved through the cyclocondensation of 1,2-diketone, an aldehyde, and ammonium acetate using a catalytic amount of ceric ammonium sulphate (CAS) or molecular iodine^{47,48} (Scheme 2). Thus, a one-pot synthesis achieved phenanthroimidazole derivatives **7a–e** in good yield via the cyclocondensation of 9,10-phenanthraquinone, 3-cyclopentyloxy-4-methoxybenzaldehyde (**2**), and ammonium acetate in the presence of 5% mole of iodine or CAS. Furthermore, acridinedione **8** was prepared by a one-pot, three-component cyclocondensation reaction of 3-cyclopentyloxy-4-methoxybenzaldehyde (**2**), 1,3-dicarbonyl compound (dimedone), and ammonium acetate in the presence of a catalytic amount of 5% CAS using polyethylene glycol (PEG) as a solvent⁴⁹. Thioamides **9a–c**, **10**, and **11** were synthesised⁵⁰ by the reaction of elemental sulphur (S₈), 3-cyclopentyloxy-4-methoxybenzaldehyde (**2**), and secondary amines, such as piperidine, morpholine, N-Boc-piperazine, and dimethylamine, or primary amines, such as 4-fluoroaniline in dimethylformamide (DMF), under heating condition.

Synthesis of compounds 12–14

The MCRs of 3-cyclopentyloxy-4-methoxybenzaldehyde (**2**), ethyl cyanoacetate, an appropriate acetophenone, and ammonium acetate in EtOH at reflux temperature gave pyridine-3-carbonitrile derivatives **12a–c** in good yield. In contrast, the reaction of 3-cyclopentyloxy-4-methoxybenzaldehyde (**2**) with ethyl cyanoacetate and thiourea in an ethanolic solution of K₂CO₃ afforded 6-(3-(cyclopentyloxy)-4-methoxyphenyl)-4-oxo-2-thioxo-1,2,3,4-tetrahydropyrimidine-5-carbonitrile (**13**)^{18,38}. Compound **13** was cyclised

with chloroacetic acid in the presence of acetic anhydride and anhydrous sodium acetate in glacial acetic acid to yield thiazolo[3,2-*a*]pyrimidine-3,5-dione derivative **14**⁵¹ (Scheme 3).

Biological evaluation

Antitumor evaluation using the MTT assay

Compounds **3a–c**, **4a,b**, **5a,b**, **6a,b**, **7a–e**, **8**, **9a–c**, **10**, **11**, **12a–c**, **13**, and **14** were screened for their *in vitro* antitumor activity by using the standard 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay against five human cancers: HePG2, HCT-116, MCF-7, PC3, and HeLa cell lines³⁹. The antitumor activities of the synthesised compounds **3–14** and the reference drugs, celecoxib, afatinib, and doxorubicin, are shown in Table 1^{8–10}. Compounds **3a–c**, incorporating the cycloalkanone core, possessed strong to weak antitumor activity against some of the investigated cell lines (IC₅₀ ≅ 19.34–95.96 μM). Interestingly, the replacement of the cycloalkanone moieties, such as in compounds **3a–c**, with a piperidin-4-one fragment, such as compound **4a,b**, resulted in a sharp increase in antitumor activity (IC₅₀ ≅ 4.38–14.32 μM) against all of the investigated five cell lines, compared with the reference drug, celecoxib (IC₅₀ ≅ 25.6–36.08 μM), afatinib (IC₅₀ values of 5.4–11.4 μM), and doxorubicin (IC₅₀ ≅ 4.17–8.87 μM).

Moreover, the introduction of quinazoline-2-thione or pyrimidine-2-thione moieties, instead of a piperidin-4-one moiety, as in compounds **5a,b**, resulted in a sharp decrease in antitumor activity against all the investigated five cancer cell lines, with IC₅₀ values in the range 46.29–92.37 μM. In contrast, the replacement of the quinazoline-2-thione fragment, as in compound **5a**, with quinazoline-2,5-dione and 2-thioxo-quinazolin-5-one fragments at the same position, such as compounds **6a** and **6b**, resulted in a sharp increase in antitumor activity against all the investigated cancer cell lines, for HePG2 (IC₅₀ values of 18.53 and 16.05 μM, respectively), HCT-116 (IC₅₀ values of 30.49 and 25.41 μM, respectively), MCF-7 (IC₅₀ values of 28.62 and 10.27 μM, respectively), PC3 (IC₅₀ values of 27.44 and 17.95 μM, respectively), and HeLa (IC₅₀ values of 19.12 and 13.49 μM, respectively), compared with celecoxib (IC₅₀ values of 25.6, 29.54, 31.28, 30.69, and 36.08 μM, respectively), afatinib (IC₅₀ values of 5.4, 11.4, 7.1, 7.7, and 6.2 μM, respectively), and doxorubicin (IC₅₀ values of 4.50, 5.23, 4.17, 8.87, and 5.57 μM, respectively).

Moreover, weak antitumor activity against some of the tested cancer cell lines was exhibited by some polycyclic derivatives incorporating imidazole and quinoline ring systems, such as compounds **7a** and **7c** (IC₅₀ ≅ 53.18–90.34 μM), whereas compounds **7e** and **8** showed moderate antitumor activity against some selected cancer cell lines (IC₅₀ ≅ 29.8–46.97 μM). Unexpectedly, derivative **7b** showed a sharp increase in antitumor activity compared with the structural analogues **7a**, **c**, **d**, and **8**, with IC₅₀ values of 13.68, 19.67, 11.85, 22.89, and 17.18 μM against HePG2, HCT-116, MCF-7, PC3, and HeLa cancer cell lines, respectively.

In contrast, the introduction of thioamide fragments in the 2-cyclopentyloxyanisole scaffold resulted in variable antitumor activity against the tested cancer cell lines; for example, compounds **9a–c** showed strong to moderate antitumor activity (IC₅₀ ≅ 24.85–48.93 μM) in comparison with thioamide **10** (IC₅₀ ≅ 47.32–79.12 μM) and **11** (IC₅₀ ≅ 88.63–96.79 μM). Furthermore, replacement of the thioamide moiety with a pyridine fragment, such as in compounds **12a–c**, retained the antitumor activity against all cancer cell lines, as indicated by their IC₅₀ values in the range 38.14–83.42 μM. In contrast, the 2-cyclopentyloxyanisole scaffold bearing the pyrimidine ring system, such as

Table 1. *In vitro* antitumor activity of the designed compounds, celecoxib, afatinib, and doxorubicin against human tumour cells.

Compound no.	IC ₅₀ (μM) ^a				
	HePG2	HCT-116	MCF-7	PC3	HeLa
3a	95.96 ± 5.2	>100	56.14 ± 2.6	51.43 ± 3.0	59.12 ± 3.8
3b	53.87 ± 3.7	80.56 ± 3.9	23.81 ± 1.5	19.34 ± 1.8	26.11 ± 1.9
3c	86.90 ± 4.5	93.46 ± 5.1	>100	>100	81.65 ± 4.7
4a	6.04 ± 0.5	4.38 ± 0.4	5.13 ± 0.3	9.18 ± 0.8	7.24 ± 0.7
4b	10.96 ± 1.1	9.48 ± 0.8	7.18 ± 0.8	14.32 ± 1.2	8.56 ± 0.9
5a	73.41 ± 3.7	66.48 ± 3.8	92.37 ± 5.2	78.95 ± 4.1	84.26 ± 4.6
5b	59.08 ± 3.5	61.13 ± 3.6	81.20 ± 4.3	55.17 ± 3.1	46.29 ± 3.0
6a	18.53 ± 1.7	30.49 ± 1.8	28.62 ± 1.6	27.44 ± 2.1	19.12 ± 1.7
6b	16.05 ± 1.4	25.41 ± 1.7	10.27 ± 1.1	17.95 ± 1.6	13.49 ± 1.4
7a	78.21 ± 4.4	90.34 ± 4.9	89.79 ± 4.3	>100	77.64 ± 4.6
7b	13.68 ± 1.2	19.67 ± 1.4	11.85 ± 1.3	22.89 ± 1.9	17.18 ± 1.5
7c	57.08 ± 3.9	81.19 ± 4.2	65.32 ± 3.4	68.06 ± 3.5	53.18 ± 3.7
7e	29.89 ± 2.1	44.82 ± 2.3	42.41 ± 2.2	46.97 ± 2.7	38.05 ± 2.5
8	41.82 ± 3.0	70.52 ± 3.5	60.48 ± 2.8	55.82 ± 3.2	43.47 ± 2.9
9a	32.87 ± 2.3	48.13 ± 2.4	35.17 ± 1.9	29.23 ± 2.3	37.50 ± 2.5
9b	24.85 ± 1.9	39.07 ± 2.2	37.09 ± 2.0	31.50 ± 2.4	28.37 ± 2.3
19c	36.27 ± 2.5	52.87 ± 2.7	48.93 ± 2.3	33.39 ± 2.6	40.61 ± 2.8
10	49.86 ± 3.5	79.12 ± 3.8	64.10 ± 3.1	47.32 ± 2.9	52.50 ± 3.7
11	91.23 ± 4.8	96.79 ± 5.5	94.27 ± 4.7	88.63 ± 5.0	90.89 ± 4.9
12a	45.24 ± 3.4	76.05 ± 3.6	71.63 ± 3.9	79.83 ± 4.0	65.72 ± 4.1
12b	38.14 ± 2.8	67.74 ± 3.5	58.28 ± 2.7	61.45 ± 3.3	45.69 ± 3.2
12c	59.63 ± 4.0	83.42 ± 4.3	66.07 ± 3.7	73.48 ± 3.8	62.76 ± 3.9
13	8.71 ± 0.7	7.66 ± 0.6	6.93 ± 0.5	11.45 ± 1.1	5.86 ± 0.6
14	20.11 ± 1.8	34.93 ± 1.9	9.62 ± 0.9	15.31 ± 1.3	12.48 ± 1.2
Celecoxib	25.6 ± 2.3	29.54 ± 2.1	31.28 ± 2.5	30.69 ± 2.7	36.08 ± 2.8
Afatinib	5.4 ± 0.25	11.4 ± 1.26	7.1 ± 0.49	7.7 ± 0.57	6.2 ± 0.67
DOX	4.50 ± 0.2	5.23 ± 0.3	4.17 ± 0.2	8.87 ± 0.6	5.57 ± 0.4

DOX: doxorubicin.

^aIC₅₀, compound concentration required to inhibit tumour cell proliferation by 50% (mean ± SD, *n* = 3). IC₅₀, (μM): 1–10 (very strong), 11–25 (strong), 26–50 (moderate), 51–100 (weak), and above 100 (non-cytotoxic). Compound **7d** had an IC₅₀ of >100 μM.

compounds **13** and **14**, exhibited strong antitumor activities against the cancer cell lines tested ($IC_{50} \cong 5.86\text{--}20.11 \mu\text{M}$). In brief, the compounds **4a**, **4b**, **7b**, and **13** exhibited the strongest antitumor activities among the designed compounds against the HeG2, HCT-116, MCF-7, PC3, and HeLa cancer cell lines ($IC_{50} \cong 4.38\text{--}22.89 \mu\text{M}$).

Structure–activity relationship of antitumor activity

According to the aforementioned antitumor activity, the SARs for the designed compounds indicated the following. (i) *N*-Methylpiperidin-4-one derivative **4a** and *N*-ethylpiperidin-4-one derivative **4b** exhibited higher antitumor activity ($IC_{50} \cong 4.38\text{--}14.32 \mu\text{M}$) than the corresponding cycloalkanones **3a–c** ($IC_{50} \cong 19.34$ to $>100 \mu\text{M}$). It was clear that the derivative with *N*-methylpiperidin-4-one **4a** had greater antitumor activity against all tested cancer cell lines ($IC_{50} \cong 4.38\text{--}9.18 \mu\text{M}$) than the *N*-ethylpiperidin-4-one derivative **4b** ($IC_{50} \cong 7.18\text{--}14.32 \mu\text{M}$). (ii) Similarly, cyclohexanone derivative **3b** exhibited greater antitumor activity against MCF-7 ($IC_{50}=23.81 \mu\text{M}$), PC3 ($IC_{50}=19.34 \mu\text{M}$), and HeLa ($IC_{50}=26.11 \mu\text{M}$) cancer cells than cyclopentanone derivative **3a** ($IC_{50} \cong 51.43$ to $>100 \mu\text{M}$), and cycloheptanone derivative **3c** ($IC_{50} \cong 81.65$ to $>100 \mu\text{M}$). (iii) Compounds incorporating a quinoxaline fragment, such as quinoxaline-2,5(1H,3H)-dione derivative **6a** ($IC_{50} \cong 18.53\text{--}30.49 \mu\text{M}$) and 2-thioxoquinazolin-5(1H)-one derivative **6b** ($IC_{50} \cong 10.27\text{--}25.41 \mu\text{M}$) showed higher antitumor activity than the corresponding derivatives quinazoline-2(1H)-thione **5a**, and pyrimidine-2-thione **5b** ($IC_{50} \cong 46.29\text{--}92.37 \mu\text{M}$). (iv) The 2-cyclopentyloxyanisole scaffold bearing the bulky polycyclic 1H-phenanthro[9,10-d]imidazoles **7a,c,d,e** ($IC_{50} \cong 29.89$ to $>100 \mu\text{M}$), and acridine-1,8(2H,5H)-dione **8** ($IC_{50} \cong 41.82\text{--}70.52 \mu\text{M}$) showed lower antitumor activity than the corresponding 2-cyclopentyloxyanisole scaffold bearing quinazoline moiety **6a,b** ($IC_{50} \cong 10.27\text{--}30.49 \mu\text{M}$). Interestingly, the derivative **7b** with the phenyl ring at position 1 of 1H-phenanthro[9,10-d]imidazole core structure ($IC_{50} \cong 11.85\text{--}22.89 \mu\text{M}$) showed a sharp increase in antitumor activity in comparison with derivatives **7a,c,d,e** and had approximately similar activity with compound **6b** ($IC_{50} \cong 10.27\text{--}25.41 \mu\text{M}$). (v) The antitumor activities of the 2-cyclopentyloxyanisole scaffold bearing a methanethione fragment, such as *N*-(4-fluorophenyl)benzothioamide derivative **10** ($IC_{50} \cong 47.32\text{--}79.12 \mu\text{M}$) and *N,N*-dimethylbenzothioamide derivative **11** ($IC_{50} \cong 88.63\text{--}96.79 \mu\text{M}$), were less potent than derivatives that contained morpholinomethanethione derivative **9a** ($IC_{50} \cong 29.23\text{--}48.13 \mu\text{M}$), piperidin-1-ylmethanethione derivative **9b** ($IC_{50} \cong 24.85\text{--}39.07 \mu\text{M}$), and *tert*-butyl piperazine-1-carboxylate derivative **9c** ($IC_{50} \cong 33.39\text{--}52.87 \mu\text{M}$). (vi) The pyrimidine derivatives, 4-oxo-2-thioxo-1,2,3,4-tetrahydropyrimidine-5-carbonitrile derivative **13** and 3,5-dioxo-2,3-dihydro-5H-thiazolo[3,2-*a*]pyrimidine-6-carbonitrile derivative **14**, had potent antitumor activities ($IC_{50} \cong 5.86\text{--}20.11 \mu\text{M}$) compared with that of the pyridine derivatives, 6-aryl-2-oxo-1,2-dihydropyridine-3-carbonitriles **12a–c**, which have moderate to weak antitumor activity ($IC_{50} \cong 38.14\text{--}83.42 \mu\text{M}$) against all tested cancer cells. Briefly, the structure–activity correlation of antitumor activity revealed that compounds **4a**, **4b**, **6b**, **7b**, **13**, and **14** were the most active compounds, whereas compound **7d** was the only derivative that had no antitumor activity against any of the tested cancer cell lines.

COX-2 inhibition assay

Several compounds that possess COX-2 inhibition activity have shown potent antitumor activities that may be attributable to the role of the COX-2 enzyme in cell proliferation^{8,14–17}. Accordingly,

Table 2. *In vitro* inhibitory effects of COX-2, PDE-4B, and TNF- α of the antitumor compounds **4a**, **4b**, **7b**, and **13**.^a

Compound no.	IC_{50} (μM) ^a		
	COX-2 inhibition	PDE-4B inhibition	TNF α inhibition
4a	3.34	5.62	2.012
4b	1.08	11.62	17.67
7b	24.02	5.65	13.94
13	1.88	3.98	6.72
Roflumilast	–	1.55	–
Celecoxib	0.68	–	6.44

^a IC_{50} value is the compound concentration required to produce 50% inhibition.

the four compounds (**4a**, **4b**, **7b**, and **13**) that exhibited the greatest antitumor activity, as well as celecoxib (used as the reference drug) were subjected to colorimetric COX-2 inhibition assays by using a COX-2 assay kit (catalogue no. 560101, Cayman Chemicals Inc., Ann Arbor, MI). The measured IC_{50} (μM) values are shown in Table 2, and are expressed as the means of three acquired determinations^{40–42}. The IC_{50} values of celecoxib for COX-2 inhibition are found to be $0.68 \mu\text{M}$. It is clear that compounds **4b** and **13** were found to be the most active inhibitors of COX-2, with IC_{50} values of 1.08 and $1.88 \mu\text{M}$, respectively, whereas compound **4a** exhibited lower COX-2 inhibitory effect with an IC_{50} value of $3.34 \mu\text{M}$. In contrast, compound **7b** showed a very low inhibitory effect, with an IC_{50} value for COX-2 inhibition of $24.02 \mu\text{M}$. Briefly, a small heterocyclic substituent on the 2-cyclopentyloxyanisole core, such as the piperidine ring in compounds **4a** and **4b** and the pyrimidine ring in compound **13**, exhibited higher COX-2 inhibition in comparison with the polycyclic 1H-phenanthro[9,10-d]imidazole in compound **7b**. The reduced inhibitory effect of compound **7b** on COX-2 may be attributed to the bulkiness of the polycyclic system, which interferes with the COX-2 binding interactions.

PDE-4B enzyme assay

Compounds that inhibit PDE4 were recently shown to possess effective antitumor activities owing to the overexpression of PDE4 in cancer and its role in cell proliferation and tumour cell growth^{18–27}. The compounds that were the most active antitumor agents, such as compounds **4a**, **4b**, **7b**, and **13**, were subjected to a PDE4B inhibition assay using roflumilast as a reference drug; the IC_{50} values are presented in Table 2. Compound **13** showed the highest inhibition against PDE4B, with an IC_{50} value of $3.98 \mu\text{M}$ comparable to that of the reference drug roflumilast ($IC_{50}=1.55 \mu\text{M}$), whereas compounds **4a** and **7b** were found have moderate activity, with IC_{50} values of 5.62 and $5.65 \mu\text{M}$, respectively. Compound **4b** possessed the lowest activity against PDE4B, with an IC_{50} value of $11.62 \mu\text{M}$. From the structural study of the tested derivatives, including **4a**, **4b**, **7b**, and **13**, we concluded that the 2-cyclopentyloxyanisole scaffold bearing a cyanopyrimidine fragment, such as compound **13**, increased the PDE4B inhibitory activity in comparison with other heterocyclic derivatives.

TNF- α inhibition assay

TNF- α has been reported as a target for cancer treatment; presently, TNF antagonists are under clinical investigation in phase I and II trials as single agents for cancer therapy^{29–32}. Accordingly, compounds **4a**, **4b**, **7b**, and **13**, which are the most active antitumor agents, were subjected to the TNF- α inhibition assay using celecoxib as a reference drug⁴³; the IC_{50} values are presented in Table 2. Compound **4a** possessed potent TNF- α inhibitory effect,

with an IC_{50} value of 2.01 μM , comparable with the reference drug celecoxib ($IC_{50}=6.44 \mu M$), whereas compound **13** was found to be an effective inhibitor, with an IC_{50} value of 6.72 μM , similar to the TNF- α inhibitory effect of the reference drug celecoxib ($IC_{50}=6.44 \mu M$). In contrast, compounds **4b** and **7b** were the least active derivatives, with IC_{50} values of 17.67 and 13.94 μM , respectively.

Molecular modelling analysis

Molecular modelling and docking analysis is an important technique used to establish the theoretical interaction between the bioactive molecules and the target enzyme and receptor to understand their binding mode^{52,53}. Therefore, a molecular docking analysis was performed by using MOE 2008.10 software and viewer utility (Chemical Computing Group Inc., Montreal, Canada) in accordance with the standard MOE procedure⁴⁵.

Docking with the COX-2 isoenzyme

The molecular interaction of the most active compounds, **4b** and **13**, with the COX-2 isoenzyme was studied by molecular docking. The crystal structure of the COX-2 isoenzyme interacting with its inhibitor SC-558 was obtained from the RSC Protein Data Bank (PDB code: 1CX2)⁵⁴. The putative binding site of the COX-2 isoenzyme (Figure 2), which is responsible for the hydrogen bonds and

hydrophobic interactions with its inhibitors, consists of key amino acid residues, such as Arg510, Gln192, Arg120, Tyr355, His90, Val523, Ser353, and Ile517. The docking procedure was validated by including the bound inhibitor SC-558 for a one-ligand run docking calculation.

The bound ligand SC-558 exhibited two types of hydrogen bonds, classical and non-classical hydrogen bonds. Four classical hydrogen bonding interactions were observed with Arg513, His90, Arg120, and Tyr355. In addition, three non-classical hydrogen bonds connected the amino acids Tyr385, Phe518, and Ala516, and the benzenesulfonamide and 4-bromophenyl fragments of SC-558 through CH-O and CH-Br interactions (Figure 2, upper panel).

Interestingly, compounds **4b** and **13**, which were the most active COX-2 inhibitors, were placed in the same binding site of the inhibitor SC-558 (Figure 2). Compound **4b**, which has nearly similar COX-2 inhibition activity as celecoxib, accommodated an orientation within the COX-2 binding site (Figure 2, left lower panel), in which the *N*-ethylpiperidine-4-one fragment was located towards the secondary pocket of the COX-2 isoenzyme and interacted with the amino acid residues of Arg513, His90, Leu352, and Gln192. In general, when compound **4b** was docked into the enzyme pocket, nine hydrogen bonds were formed with the surrounding amino acids lining the pocket. One of these interactions was a classical hydrogen bond between the carbonyl (C=O) group of the *N*-ethylpiperidine-4-one fragment and the OH group of the Tyr355 residue (3.06 Å). Moreover, eight non-bonding interactions,

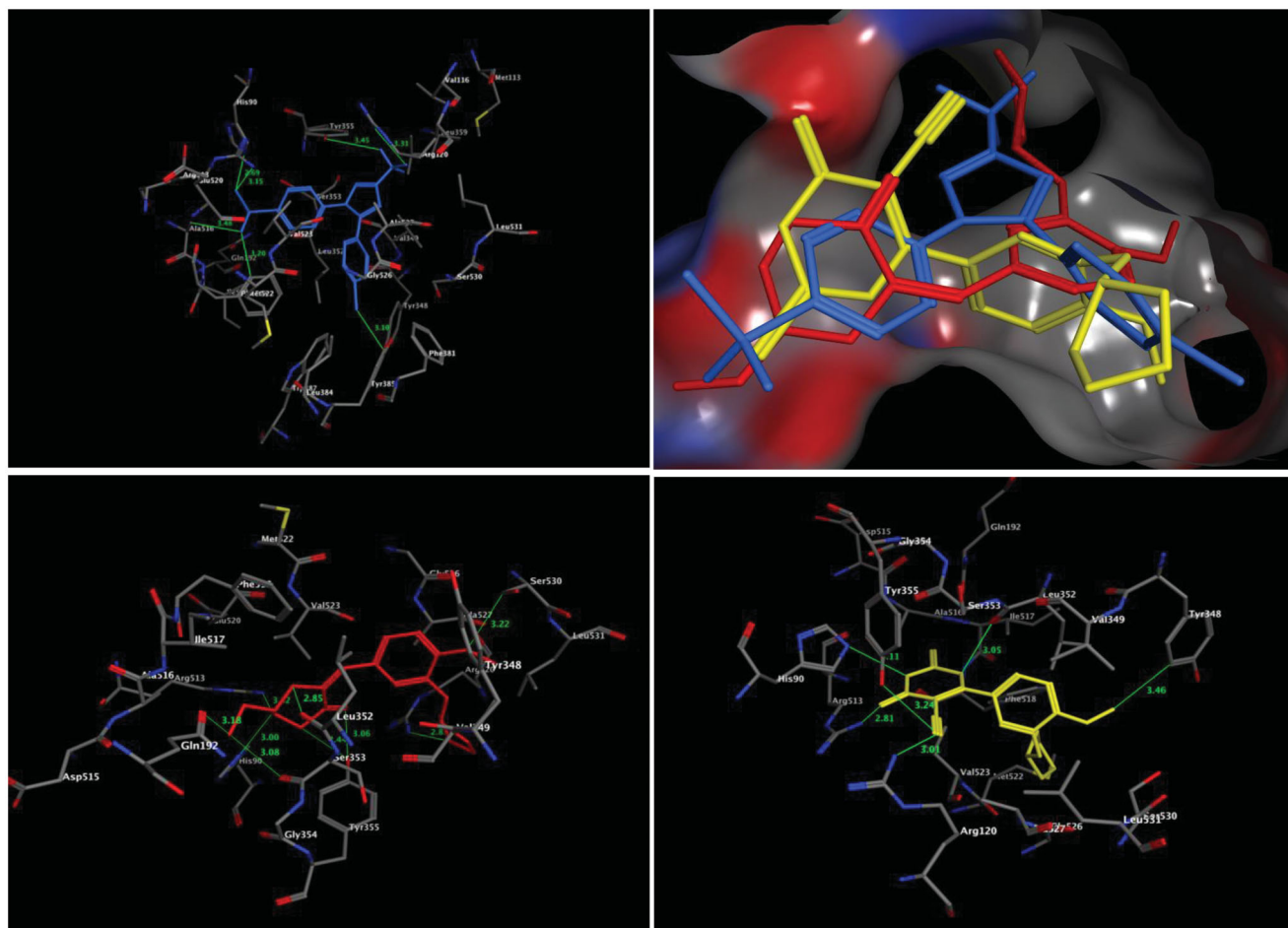


Figure 2. Three-dimensional (3D) orientation of the docked ligand SC-558 (upper left panel); docked compounds **4b** (lower left panel), and **13** (lower right panel) in the active pocket of the COX-2 enzyme (H bond interactions are shown as green lines). Upper right panel showed the alignment of SC-558, **4b**, and **13** in the active pocket of the COX-2 enzyme.

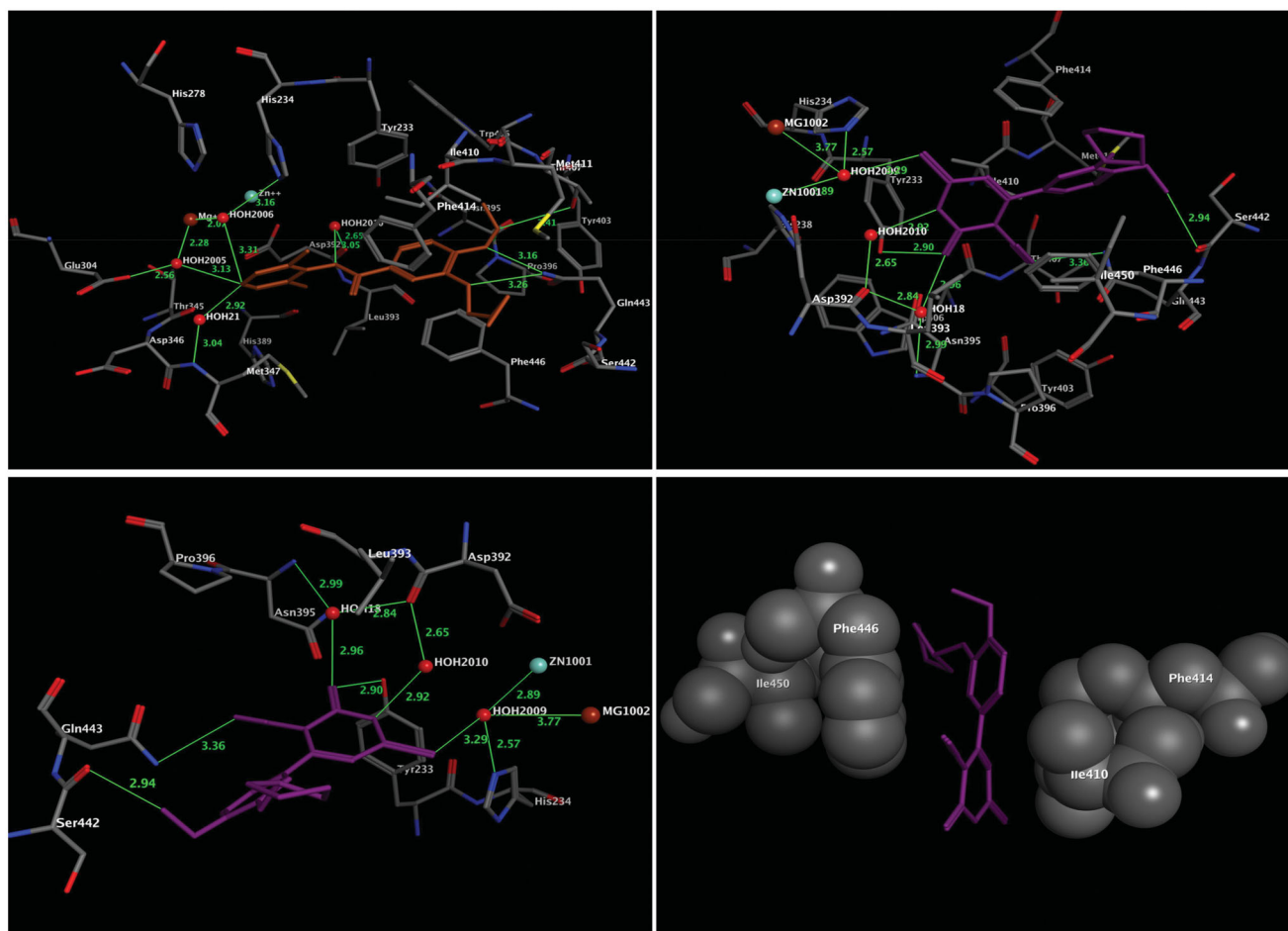


Figure 3. Three-dimensional (3D) orientation of the docked roflumilast (upper left panel); docked compound **13** (upper right panel), in the active pocket of the PDE4B enzyme (H bond interactions are shown as green lines). Lower left panel showed near picture of compound **13** in the active pocket of the PDE4B enzyme. Lower right panel showed the hydrophobic interactions of compound **13** in the active pocket of the PDE4B enzyme.

namely non-classical hydrogen bonds were formed, among the two bonds of the OH of the Tyr355 residue, and the C=O of the Leu352 residue with the CH₂ of the piperidine-4-one moiety (3.44 Å, and 2.85 Å, respectively), and among two more bonds among the C=O fragments of the Gln192 and Ser353 residues and the CH₃ moiety of *N*-ethylpiperidine-4-one (3.18 Å and 3.08 Å, respectively). The amino acid residues Arg513 and His90 formed additional two bonds between their HN groups and the CH₂ of the piperidine-4-one ring (3.52 Å and 3.00 Å, respectively). Finally, the amino acid residues Arg120 and Ser530 formed two non-classical hydrogen bonds with the cyclopentyl and methoxyl moieties of the anisole core structure (NH-CH₂, 2.87 Å; and CH₂-OCH₃, 3.22 Å, respectively). The overall outcome of the molecular docking of compound **4b**, with respect to non-classical hydrogen bonds, showed that compound **4b** had more hydrophobic interactions with the protein than the bound ligand SC-558.

The molecular docking analysis of compound **13** showed that the 4-oxo-2-thioxo-1,2,3,4-tetrahydropyrimidine-5-carbonitrile moiety was the main fragment responsible for COX-2 activity, which interacted with the surrounding amino acid residues of the active pocket of the COX-2 isoenzyme, such as Arg513, His90, Tyr348, Tyr355, and Arg120 (Figure 2, right lower panel). Four classical and one non-classical hydrogen bonding interactions were formed between the abovementioned amino acid residues and compound **13**. The nitrile group (CN) of compound **13** formed two classical hydrogen bonds with Arg120 (3.01 Å) and Tyr355 (3.24 Å), whereas the 4-oxo-tetrahydropyrimidine ring system interacted

with amino acid residues Arg513 and His90 through two classical hydrogen bonds (2.81 Å and 3.11 Å, respectively). The final interaction was the hydrophobic interaction between Tyr348 and the methoxyl moiety of anisole through a CH₂-π bond, with a non-bonding distance of 3.46 Å.

Docking with the PDE4B enzyme

The binding mode of the most active compound, **13**, within the PDE4B enzyme was analysed by using molecular docking. The crystal structure of the PDE4B enzyme bound with its inhibitor roflumilast was obtained from the RSC Protein Data Bank (PDB code: 1XMU)⁵⁵. The binding site of the PDE4B enzyme (Figure 3), which is responsible for the formation of coordination bonds, hydrogen bonds, and hydrophobic interactions with its inhibitor roflumilast, has three main sites for interaction: the solvent-filled metal coordination pocket, including both zinc and magnesium; the conserved residue Gln443; and the hydrophobic pocket. The amino acid residues Phe414, Ile410, Phe446, and Ile450 were the key residues that formed the tunnel, and were responsible for the accommodation of the hydrophobic interaction with the bound inhibitor, roflumilast. The molecular docking procedure was validated by performing a one-ligand run docking calculation for the bound inhibitor roflumilast. The results of the docking calculation of compound **13** are presented in Figure 3 (upper right panel). From the docking results, it was clear that the 2-cyclopentylloxaniso scaffold and the pyrimidine ring adapted for hydrophobic

recognition at the binding cavity lining with the amino acid residues Phe414, Ile410, Phe446, and Ile450 (Figure 3, lower right panel), similar to the bound inhibitor roflumilast (Figure 3, upper left panel). In contrast, the methoxyl group of the 2-cyclopentyllox-yanisole scaffold formed a non-classical hydrogen bond with Ser442 (2.94 Å), whereas the conserved residue Gln443 interacted with the pyrimidine ring system through the nitrile moiety by the formation of hydrogen bond with a distance of 3.36 Å (Figure 3, lower left panel). Moreover, the pyrimidine ring projected towards the metal-coordinating site filled with water molecules. Accordingly, the thione (C=S) moiety of the pyrimidine ring is coordinated with Zn and Mg ions, mediated by HOH2009, and formed a hydrogen bond with the amino acid residue His234. Meanwhile, the carbonyl oxygen (C=O) of the pyrimidine formed one hydrogen bond with Tyr233 (2.90 Å) and another two hydrogen bonds with the amino acid residues Asn395 and Asp392, mediated by HOH18. Finally, the internal NH group of pyrimidine ring was adapted to form a hydrogen bond with Asp392 mediated by HOH18.

Briefly, in comparison of compound **13** with the bound inhibitor roflumilast, both compounds accommodated approximately similar interactions at the hydrophobic clamp site (Phe414, Ile410, Phe446, and Ile450) and the metal coordination site.

Conclusions

A series of compounds incorporating 2-cyclopentyllox-yanisole scaffold bearing a variety of ring systems—cycloalkanones **3a–c** and **4a–b**, quinazolines **5a–b** and **6a–b**, fused imidazoles **7a–e**, fused quinoline **8**, thioamides **9a–c**, **10**, and **11**, pyridines **12a–c**, and pyrimidines **13** and **14** was synthesised. These compounds were evaluated for their *in vitro* antitumor activity in five human cancer cell lines: HePG2, HCT-116, MCF-7, PC3, and HeLa. The antitumor activity of compounds **4a**, **4b**, **6b**, **7b**, **13**, and **14** indicated that these derivatives were the most potent antitumor agents among the tested compounds, with IC₅₀ values of 5.13–17.95 μM in the tested cancer cell lines. The antitumor results of the synthesised compounds were comparable with the reference drug celecoxib (IC₅₀ values of 25.6–36.08 μM), afatinib (IC₅₀ values of 5.4–11.4 μM), and doxorubicin (IC₅₀ values of 4.17–8.87 μM). In addition, the compounds that were most active as antitumor agents, **4a**, **4b**, **7b**, and **13**, were assayed for their ability to inhibit COX-2, PDE4B, and TNF-α. The results indicated that compounds **4b** and **13** exhibited effective COX-2 inhibitory activity, with IC₅₀ values of 1.08 and 1.88 μM, respectively, which were comparable with celecoxib (IC₅₀=6.44 μM). In addition, compounds **4a** and **13** inhibited the PDE4B enzyme, with an IC₅₀ value of 5.62 and 3.98 μM, respectively, which was comparable with roflumilast (IC₅₀=1.55 μM), whereas these compounds had potent TNF-α inhibitory effect, with IC₅₀ values of 2.01 and 6.72 μM, respectively, which were comparable with the reference drug celecoxib (IC₅₀=6.44 μM). Compounds **4b** and **13** were docked into the COX-2 and PDE4B binding sites and exhibited similar binding characteristics to that of bound inhibitor SC-558 for the COX-2 enzyme and the bound inhibitor roflumilast for the PDE4B enzyme.

Acknowledgements

The authors thank the Deanship of Scientific Research and RSSU at the King Saud University for their technical support.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

The authors express their appreciation to the Deanship of Scientific Research at King Saud University for funding the work through the research group project No. RGP-163.

ORCID

Magda A.-A. El-Sayed  <http://orcid.org/0000-0001-9599-9248>

Adel S. El-Azab  <http://orcid.org/0000-0001-7197-1515>

Alaa A.-M. Abdel-Aziz  <http://orcid.org/0000-0002-3362-9337>

References

- (a) Avendaño C, Menéndez J. Medicinal chemistry of anti-cancer agents. Amsterdam: Elsevier; 2008. (b) Varmus H. The new era in cancer research. Science 2006;312:1162–5. (c) Eckhardt S. Recent progress in the development of anti-cancer agents. Curr Med Chem Anticancer Agents 2002;2: 419–39.
- (a) El-Azab AS, Al-Omar MA, Abdel-Aziz AAM, et al. Design, synthesis and biological evaluation of novel quinazoline derivatives as potential antitumor agents: molecular docking study. Eur J Med Chem 2010;45:4188–98. (b) Al-Suwaidan IA, Alanazi AM, Abdel-Aziz AAM, et al. Design, synthesis and biological evaluation of 2-mercapto-3-phenethylquinazoline bearing anilide fragments as potential antitumor agents: molecular docking study. Bioorg Med Chem Lett 2013;23: 3935–41. (c) Alanazi AM, Al-Suwaidan IA, Abdel-Aziz AAM, et al. Design, synthesis and biological evaluation of some novel substituted 2-mercapto-3-phenethylquinazolines as antitumor agents. Med Chem Res 2013;22:5566–77.
- (a) Alanazi AM, Abdel-Aziz AAM, Al-Suwaidan IA, et al. Design, synthesis and biological evaluation of some novel substituted quinazolines as antitumor agents. Eur J Med Chem 2014;79:446–54. (b) Alanazi AM, Abdel-Aziz AAM, Shower TZ, et al. Synthesis, antitumor and antimicrobial activity of some new 6-methyl-3-phenyl-4(3H)-quinazolinone analogues: in silico studies. J Enzyme Inhib Med Chem 2016; 31:721–35. (c) El-Azab AS, Al-Dhfyhan A, Abdel-Aziz AAM, et al. Synthesis, anticancer and apoptosis-inducing activities of quinazoline–isatin conjugates: epidermal growth factor receptor-tyrosine kinase assay and molecular docking studies. J Enzyme Inhib Med Chem 2017;32:935–44.
- (a) Mohamed MA, Ayyad RR, Shower TZ, et al. Synthesis and antitumor evaluation of trimethoxyanilides based on 4(3H)-quinazolinone scaffolds. Eur J Med Chem 2016;112: 106–13. (b) Al-Suwaidan IA, Abdel-Aziz AAM, Shower TZ, et al. Synthesis, antitumor activity and molecular docking study of some novel 3-benzyl-4(3H)quinazolinone analogues. J Enzyme Inhib Med Chem 2016;31:78–89. (c) El-Azab AS, Abdel-Aziz AAM, Ghabbour HA, et al. Synthesis, in vitro anti-tumour activity, and molecular docking study of novel 2-substituted mercapto-3-(3,4,5-trimethoxybenzyl)-4(3H)-quinazolinone analogues. J Enzyme Inhib Med Chem 2017;32: 1229–39.
- (a) Abdel-Aziz AAM. Novel and versatile methodology for synthesis of cyclic imides and evaluation of their cytotoxic,

- DNA binding, apoptotic inducing activities and molecular modeling study. *Eur J Med Chem* 2007;42:614–26.(b) El-Azab AS, Alanazi AM, Abdel-Aziz NI, et al. Synthesis, molecular modeling study, preliminary antibacterial, and antitumor evaluation of N-substituted naphthalimides and their structural analogues. *Med Chem Res* 2013;22:2360–75.
6. (a) El-Deeb IM, Bayoumi SM, El-Sherbeny MA, et al. Synthesis and antitumor evaluation of novel cyclic arylsulfonyleureas: ADME-T and pharmacophore prediction. *Eur J Med Chem* 2010;45:2516–30.(b) Abdel-Aziz AAM, El-Azab AS, El-Subbagh HI, et al. Design, synthesis, single-crystal and preliminary antitumor activity of novel arenesulfonylimidazolidin-2-ones. *Bioorg Med Chem Lett* 2012;22:2008–14.(c) Alanazi AM, El-Azab AS, Al-Swaidan IA, et al. Synthesis, single-crystal, in vitro antitumor evaluation and molecular docking of 3-substituted 5,5-diphenylimidazolidine-2,4-dione derivatives. *Med Chem Res* 2013;22:6129–42.
 7. (a) Al-Suwaitan IA, Abdel-Aziz NI, El-Azab AS, et al. Antitumor evaluation and molecular docking study of substituted 2-benzylidenebutane-1,3-dione, 2-hydrizonobutane-1,3-dione and trifluoromethyl-1H-pyrazole analogues. *J Enzyme Inhib Med Chem* 2015;30:679–87.(b) El-Sherbeny MA, Abdel-Aziz AAM, Ahmed MA. Synthesis and antitumor evaluation of novel diarylsulfonylurea derivatives: molecular modeling applications. *Eur J Med Chem* 2010;45:689–97.
 8. (a) El-Husseiny WM, El-Sayed MAA, Abdel-Aziz NI, et al. Structural alterations based on naproxen scaffold: synthesis, evaluation of antitumor activity and COX-2 inhibition, and molecular docking. *Eur J Med Chem* 2018;158:134–43.(b) El-Azab AS, Abdel-Aziz AAM, Abou-Zeid LA, et al. Synthesis, antitumour activities and molecular docking of thiocarboxylic acid ester-based NSAID scaffolds: COX-2 inhibition and mechanistic studies. *J Enzyme Inhib Med Chem* 2018;33:989–98.
 9. El-Husseiny WM, El-Sayed MA, Abdel-Aziz NI, et al. Synthesis, antitumour and antioxidant activities of novel α,β -unsaturated ketones and related heterocyclic analogues: EGFR inhibition and molecular modelling study. *J Enzyme Inhib Med Chem* 2018;33:507–18.
 10. El-Sayed MA, El-Husseiny WM, Abdel-Aziz NI, et al. Synthesis and biological evaluation of 2-styrylquinolines as antitumour agents and EGFR kinase inhibitors: molecular docking study. *J Enzyme Inhib Med Chem* 2018;33:199–209.
 11. (a) Abdel-Aziz AAM, El-Azab AS, Alanazi AM, et al. Synthesis and potential antitumor activity of 7-(4-substituted piperazin-1-yl)-4-oxoquinolines based on ciprofloxacin and norfloxacin scaffolds: in silico studies. *J Enzyme Inhib Med Chem* 2016;31:796–809.(b) Abdel-Aziz AAM, Asiri YA, Al-Agamy MH. Design, synthesis and antibacterial activity of fluoroquinolones containing bulky arenesulfonyl fragment: 2D-QSAR and docking study. *Eur J Med Chem* 2011;46:5487–97.
 12. Bayomi SM, El-Kashef HA, El-Ashmawy MB, et al. Synthesis and biological evaluation of new curcumin analogues as antioxidant and antitumor agents: molecular modeling study. *Eur J Med Chem* 2015;101:584–94.
 13. (a) Stanković T, Dinic J, Podolski-Renić A, et al. Dual inhibitors as a new challenge for cancer multidrug resistance treatment. *Curr Med Chem* 2019;26:6074–106.(b) Raghavendra NM, Pingili D, Kadasi S, et al. Dual or multi-targeting inhibitors: the next generation anticancer agents. *Eur J Med Chem* 2018;143:1277–300.
 14. (a) Dai ZJ, Ma XB, Kang HF, et al. Antitumor activity of the selective cyclooxygenase-2 inhibitor, celecoxib, on breast cancer in vitro and in vivo. *Cancer Cell Int* 2012;12:53.(b) Ghosh N, Chaki R, Mandal V, et al. COX-2 as a target for cancer chemotherapy. *Pharmacol Rep* 2010;62:233–44.
 15. (a) Blanke C. Role of COX-2 inhibitors in cancer therapy. *Cancer Invest* 2004;22:271–82.(b) Basu GD, Pathangey LB, Tinder TL, et al. Mechanisms underlying the growth inhibitory effects of the cyclo-oxygenase-2 inhibitor celecoxib in human breast cancer cells. *Breast Cancer Res* 2005;7:422–35.
 16. (a) Vosooghi M, Amini M. The discovery and development of cyclooxygenase-2 inhibitors as potential anticancer therapies. *Expert Opin Drug Discov* 2014;9:255–67.(b) Claria J. Cyclooxygenase-2 biology. *Curr Pharm Des* 2003;9:2177–90.
 17. Gurpinar E, Grizzle WE, Piazza GA. COX-independent mechanisms of cancer chemoprevention by anti-inflammatory drugs. *Front Oncol* 2013;3:181–18.
 18. (a) Hirsh L, Dantes A, Suh BS, et al. Phosphodiesterase inhibitors as anti-cancer drugs. *Biochem Pharmacol* 2004;68:981–8.(b) Almatary AM, Elmorsy MA, El Husseiny WM, et al. Design, synthesis, and molecular modeling of heterocyclic bioisostere as potent PDE4 inhibitors. *Arch Pharm (Weinheim)* 2018;351:e1700403.(c) Sandeep G, Bhasker S, Ranganath YS. Phosphodiesterase as a novel target in cancer chemotherapy. *Int J Pharmacol* 2009;7:1.
 19. (a) Drees M, Zimmermann R, Eisenbrand G. 3',5'-Cyclic nucleotide phosphodiesterase in tumor cells as potential target for tumor growth inhibition. *Cancer Res* 1993;53:3058–61.(b) Savai R, Pullamsetti SS, Banat GA, et al. Targeting cancer with phosphodiesterase inhibitors. *Expert Opin Investig Drugs* 2010;19:117–31.
 20. (a) Pullamsetti SS, Banat GA, Schmall A, et al. Phosphodiesterase-4 promotes proliferation and angiogenesis of lung cancer by crosstalk with HIF. *Oncogene* 2013;32:1121–34.(b) Selige J, Hatzelmann A, Dunkern TJ. The differential impact of PDE4 subtypes in human lung fibroblasts on cytokine-induced proliferation and myofibroblast conversion. *Cell Physiol* 2011;226:1970–80.
 21. (a) Domvri K, Zarogoulidis K, Ziogas N, et al. Potential synergistic effect of phosphodiesterase inhibitors with chemotherapy in lung cancer. *J Cancer* 2017;8:3648–56.(b) Yeo CD, Kim YA, Lee HY, et al. Roflumilast treatment inhibits lung carcinogenesis in benzo(a)pyrene-induced murine lung cancer model. *Eur J Pharmacol* 2017;812:189–95.
 22. (a) Kim DU, Kwak B, Kim SW. Phosphodiesterase 4B is an effective therapeutic target in colorectal cancer. *Biochem Biophys Res Commun* 2019;508:825–31.(b) Tsunoda T, Ota T, Fujimoto T, et al. Inhibition of phosphodiesterase-4 (PDE4) activity triggers luminal apoptosis and AKT dephosphorylation in a 3-D colonic-crypt model. *Mol Cancer* 2012;11:46.
 23. (a) Murata K, Sudo T, Kameyama M, et al. Cyclic AMP specific phosphodiesterase activity and colon cancer cell motility. *Clin Exp Metastasis* 2000;18:599–604.(b) Nishi K, Luo H, Ishikura S, et al. Apremilast induces apoptosis of human colorectal cancer cells with mutant KRAS. *Anticancer Res* 2017;37:3833–9.
 24. (a) Cooney JD, Aguiar RC. Phosphodiesterase 4 inhibitors have wide-ranging activity in B-cell malignancies. *Blood* 2016;128:2886–90.(b) Kelly K, Mejia A, Suhasini AN, et al. Safety and pharmacodynamics of the PDE4 inhibitor roflumilast in advanced B-cell malignancies. *Clin Cancer Res* 2017;23:1186–92.

25. Kelly K, Mejia A, Suhasini AN, et al. Safety and pharmacodynamics of the PDE4 inhibitor roflumilast in advanced B-cell malignancies. *Clin Cancer Res* 2017;23:1186–92.
26. (a) Parikh N, Chakraborti AK. Phosphodiesterase 4 (PDE4) inhibitors in the treatment of COPD: promising drug candidates and future directions. *Curr Med Chem* 2016;23:129–41.(b) Contreras S, Milara J, Morcillo E, et al. Selective inhibition of phosphodiesterases 4A, B, C and D isoforms in chronic respiratory diseases: current and future evidences. *Curr Pharm Des* 2017;23:2073–83.
27. Mouratidis PX, Colston KW, Bartlett JB, et al. Antiproliferative effects of CC-8062 and CC-8075 in pancreatic cancer cells. *Pancreas* 2009;38:78–84.
28. Balasubramanian G, Narayanan S, Andiappan L, et al. In vivo effective dibenzo[b,d]furan-1-yl-thiazoles as novel PDE-4 inhibitors. *Bioorg Med Chem* 2016;24:5702–16.
29. (a) Brenner D, Blaser H, Mak TW. Regulation of tumour necrosis factor signalling: live or let die. *Nat Rev Immunol* 2015;15:362–74.(b) Lebec H, Ponce R, Preston BD, et al. Tumor necrosis factor, tumor necrosis factor inhibition, and cancer risk. *Curr Med Res Opin* 2015;31:557–74.
30. (a) Chuang MJ, Sun KH, Tang SJ, et al. Tumor-derived tumor necrosis factor- α promotes progression and epithelial-mesenchymal transition in renal cell carcinoma cells. *Cancer Sci* 2008;99:905–13.(b) Liu XL, Li FQ, Liu XL, et al. TNF- α , HGF and macrophage in peritumoural liver tissue relate to major risk factors of HCC recurrence. *Hepatogastroenterology* 2013;60:1121–6.
31. (a) Mikami S, Mizuno R, Kosaka T, et al. Expression of TNF- α and CD44 is implicated in poor prognosis, cancer cell invasion, metastasis and resistance to the sunitinib treatment in clear cell renal cell carcinomas. *Int J Cancer* 2015;136:1504–14.(b) Zhu G, Du Q, Wang X, et al. TNF- α promotes gallbladder cancer cell growth and invasion through autocrine mechanisms. *Int J Mol Med* 2014;33:1431–40.
32. (a) Yu M, Zhou X, Niu L, et al. Targeting transmembrane TNF- α suppresses breast cancer growth. *Cancer Res* 2013;73:4061–74.(b) Zidi I, Mestiri S, Bartegi A, et al. TNF- α and its inhibitors in cancer. *Med Oncol* 2010;27:185–98.
33. (a) Katsori AM, Hadjipavlou-Litina D. Chalcones in cancer: understanding their role in terms of QSAR. *Curr Med Chem* 2009;16:1062–81.(b) Karthikeyan C, Moorthy N, Ramasamy S, et al. Advances in chalcones with anticancer activities. *Recent Pat Anticancer Drug Discov* 2014;10:97–115.
34. (a) Goel A, Boland CR, Chauhan DP. Specific inhibition of cyclooxygenase-2 (COX-2) expression by dietary curcumin in HT-29 human colon cancer cells. *Cancer Lett* 2001;172:111–8.(b) Lev-Ari S, Strier D, Kazanov L, Madar-Shapiro L, et al. Celecoxib and curcumin synergistically inhibit the growth of colorectal cancer cells. *Clin Cancer Res* 2005;11:6738–44.(c) Lee SH, Lee GH, Park SY, et al. Apoptotic effects of curcumin via the regulation of COX-2/ VASP signaling molecules in MCF-7 breast cancer cells. *Cancer Prev Res* 2012;17:19–26.
35. (a) Abusnina A, Keravis T, Zhou Q, et al. Tumour growth inhibition and anti-angiogenic effects using curcumin correspond to combined PDE2 and PDE4 inhibition. *Thromb Haemost* 2015;113:319–22.(b) Abusnina A, Keravis T, Yougbaré I, et al. Anti-proliferative effect of curcumin on melanoma cells is mediated by PDE1A inhibition that regulates the epigenetic integrator UHRF1. *Mol Nutr Food Res* 2011;55:1677–89.(c) Yi YX, Gaurav A, Akowuah GA. Docking studies of curcumin and analogues with various phosphodiesterase 4 subtypes. *Curr Drug Discov Technol* 2018.
36. Ahmed NM, Youns M, Soltan MK, et al. Design, synthesis, molecular modelling, and biological evaluation of novel substituted pyrimidine derivatives as potential anticancer agents for hepatocellular carcinoma. *J Enzyme Inhib Med Chem* 2019;34:1110–20.
37. Youssef KM, El-Sherbeny MA, El-Shafie FS, et al. Synthesis of curcumin analogues as potential antioxidant, cancer chemopreventive agents. *Arch Pharm (Weinheim)* 2004;337:42–54.
38. Kambe S, Saito K, Kishi H, et al. A one-step synthesis of 4-Oxo-2-thioxopyrimidine derivatives by the ternary condensation of ethyl cyanoacetate, aldehydes, and thiourea. *Synthesis* 1979;4:287.
39. (a) Denizot F, Lang R. Rapid colorimetric assay for cell growth and survival. *J Immunol Methods* 1986;89:271–7.(b) Vega-Avila E, Pugsley MK. An overview of colorimetric assay methods used to assess survival or proliferation of mammalian cells. *West Pharmacol Soc* 2011;54:10–4.(c) Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods* 1983;65:55–63.
40. (a) Abdel-Aziz AAM, Angeli A, El-Azab AS, et al. Synthesis and anti-inflammatory activity of sulfonamides and carboxylates incorporating trimellitimidates: dual cyclooxygenase/carbonic anhydrase inhibitory actions. *Bioorg Chem* 2019;84:260–8.(b) Uddin MJ, Rao PP, Knaus EE. Design and synthesis of acyclic triaryl (Z)-olefins: a novel class of cyclooxygenase-2 (COX-2) inhibitors. *Bioorg Med Chem* 2004;12:5929–40.(c) El-Sayed MA, Abdel-Aziz NI, Abdel-Aziz AAM, et al. Design, synthesis, and biological evaluation of substituted hydrazone and pyrazole derivatives as selective COX-2 inhibitors: molecular docking study. *Bioorg Med Chem* 2011;19:3416–24.(d) El-Sayed MA, Abdel-Aziz NI, Abdel-Aziz AAM, et al. Synthesis, biological evaluation and molecular modeling study of pyrazole and pyrazoline derivatives as selective COX-2 inhibitors and anti-inflammatory agents. Part 2. *Bioorg Med Chem* 2012;20:3306–16.
41. (a) Abdel-Sayed MA, Bayomi SM, El-Sherbeny MA, et al. Synthesis, anti-inflammatory, analgesic, COX-1/2 inhibition activities and molecular docking study of pyrazoline derivatives. *Bioorg Med Chem* 2016;24:2032–42.(b) Abdel-Aziz AAM, El-Azab AS, Abou-Zeid LA, et al. Synthesis, anti-inflammatory, analgesic and COX-1/2 inhibition activities of anilides based on 5,5-diphenylimidazolidine-2,4-dione scaffold: molecular docking studies. *Eur J Med Chem* 2016;115:121–31.
42. (a) Abdel-Aziz AAM, Abou-Zeid LA, ElTahir KEH, et al. Synthesis, anti-inflammatory, analgesic, COX-1/2 inhibitory activities and molecular docking studies of substituted 2-mercapto-4(3H)-quinazolinones. *Eur J Med Chem* 2016;121:410–21.(b) Abdel-Aziz AAM, Abou-Zeid LA, ElTahir KE, et al. Design, synthesis of 2,3-disubstituted 4(3H)-quinazolinone derivatives as anti-inflammatory and analgesic agents: COX-1/2 inhibitory activities and molecular docking studies. *Bioorg Med Chem* 2016;24:3818–28.
43. Funakoshi-Tago M, Shimizu T, Tago K, et al. Celecoxib potently inhibits TNF α -induced nuclear translocation and activation of NF- κ B. *Biochem Pharmacol* 2008;76:662–71.
44. Gupta M, Kaur G. Aqueous extract from the *Withania somnifera* leaves as a potential anti-neuroinflammatory agent: a mechanistic study. *J Neuroinflammation* 2016;13:193–209.

45. Molecular Operating Environment (MOE 2008.10) of Chemical Computing Group. Inc. Canada. Available from: <http://www.chemcomp.com>. 2008.
46. El-Zahar MI, Abd El-Karim SS, Haiba ME. Synthesis and cytotoxic evaluation of some novel 6- (benzofuran-2-yl)-4-(4-fluorophenyl) pyridines. *World J Chem* 2008;4:182–194.
47. (a) Davidson D, Weiss M, Jelling M. The action of ammonia on benzil. *J Org Chem* 1937;2:319–27.(b) Behmadi H, Roshani M, Saadati MS. Synthesis of phenanthrimidazole from 9,10-phenanthraquinone and aldehydes by molecular iodine as catalyst. *Chin Chem Lett* 2009;20:5–8.
48. Arshia P, Azim A, Shaikh KA. Ceric ammonium nitrate catalyzed efficient one-pot synthesis of 2, 4, 5-triarylimidazoles. *Res J Pharm Biol Chem Sci* 2010;1:943–951.
49. Neelam PP, Rajesh HV, Hitesh SP. Ceric Ammonium Nitrate (CAN)-Catalyzed Multicomponent Reactions: An efficient catalyst for green organic synthesis. *Synthetic Commun* 2015;45:2399–2425.
50. (a) Daniel LP, Carsten B. Recent advances in the Willgerodt-Kindler reaction. *Chem Soc Rev* 2013;42:7870–7880. (b) Aghapoor K, Mohsenazadeh F, Khanalizadeh G, et al. The Willgerodt-Kindler reaction in water: high chemoselectivity of benzaldehydes over acetophenones. *Monatshefte Chem Chem Month* 2007;138:61.
51. (a) El-Zahar MI, Abd El-Karim SS, Haiba ME, et al. Synthesis, antitumor activity and molecular docking study of novel benzofuran-2-yl pyrazole pyrimidine derivatives. *Acta Pol Pharm Drug Res* 2011;68:357–73.(b) Hussain SM, El-Reedy AM, Hassan Rezk AM, et al. Reactions with 2-mercaptopyrimidines. Synthesis of some new thiazolo[3,2-a]- and triazolo[4,3-a]pyrimidines. *J Heterocycl Chem* 1987;24:1605–10.
52. (a) Al-Suwaidan IA, Alanazi AM, El-Azab AS, et al. Molecular design, synthesis and biological evaluation of cyclic imides bearing benzenesulfonamide fragment as potential COX-2 inhibitors. Part 2. *Bioorg Med Chem Lett* 2013;23:2601–5.(b) Alanazi AM, El-Azab AS, Al-Suwaidan IA, et al. Structure-based design of phthalimide derivatives as potential cyclooxygenase-2 (COX-2) inhibitors: anti-inflammatory and analgesic activities. *Eur J Med Chem* 2015;92:115–23.(c) Abdel-Aziz AAM, El Tahir KEH, Asiri YA. Synthesis, anti-inflammatory activity and COX-1/COX-2 inhibition of novel substituted cyclic imides. Part 1: molecular docking study. *Eur J Med Chem* 2011;46:1648–55.(d) Abdel-Aziz AA, El-Azab AS, AlSaif NA, et al. Synthesis, anti-inflammatory, cytotoxic, and COX-1/2 inhibitory activities of cyclic imides bearing 3-benzenesulfonamide, oxime, and β -phenylalanine scaffolds: a molecular docking study. *J Enzyme Inhib Med Chem* 2020;35:610–21.
53. (a) El-Gamal MI, Bayomi SM, El-Ashry SM, et al. Synthesis and anti-inflammatory activity of novel (substituted)benzylidene acetone oxime ether derivatives: molecular modeling study. *Eur J Med Chem* 2010;45:1403–14.(b) Goda FE, Abdel-Aziz AAM, Ghoneim HA. Synthesis and biological evaluation of novel 6-nitro-5-substituted aminoquinolines as local anesthetic and anti-arrhythmic agents: molecular modeling study. *Bioorg Med Chem* 2005;13:3175–83.(c) Goda FE, Abdel-Aziz AAM, Attef OA. Synthesis, antimicrobial activity and conformational analysis of novel substituted pyridines: BF₃-promoted reaction of hydrazine with 2-alkoxy pyridines. *Bioorg Med Chem* 2004;12:1845–52.(d) El-Azab AS, Mary YS, Panicker CY, et al. DFT and experimental (FT-IR and FT-Raman) investigation of vibrational spectroscopy and molecular docking studies of 2-(4-oxo-3-phenethyl-3,4-dihydroquinazolin-2-ylthio)-N-(3,4,5-trimethoxyphenyl) acetamide. *J Mol Struct* 2016;1113:133–45.(e) El-Azab AS, Abdel-Aziz AA, Ahmed HEA, et al. Exploring structure–activity relationship of S-substituted 2-mercaptoquinazolin-4(3H)-one including 4-ethylbenzenesulfonamides as human carbonic anhydrase inhibitors. *J Enzyme Inhib Med Chem* 2020;35:598–609.
54. Kurumbail RG, Stevens AM, Gierse JK, et al. Structural basis for selective inhibition of cyclooxygenase-2 by anti-inflammatory agents. *Nature* 1996;384:644–8.
55. Card GL, England BP, Suzuki Y, et al. Structural basis for the activity of drugs that inhibit phosphodiesterases. *Structure* 2004;12:2233–47.