

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

TAXONOMY, CLASSIFICATION AND NOMENCLATURE OF VIRUSES

Claude M. Fauquet, The Scripps Research Institute, Division of Plant Biology, La Jolla, California, USA

Copyright © 1999 Academic Press



History of Virus Classification and Virus Nomenclature

Humans feel the need to classify natural entities and the viruses are no exception. As in other biological systems, virus classification is an approximate and imperfect exercise. Like any other type of classification, it is a totally artificial and human-driven activity without any natural base. However, science requires workable descriptions of living systems and their constituent parts, and, when achieved properly, classifications are extremely useful for showing similar characteristics and properties across populations. Unfortunately for virus taxonomy no fossil record exists and so evolutionary relationships are very speculative, meaning that only a logical and precise virus classification can provide indications of the evolution of viruses. Appropriately chosen classification criteria are also informative in the case of newly discovered viruses. In theory, nomenclature and classification are totally independent, but for viruses both issues are often considered at the same time. As a result, taxonomic names for the viruses have always been the subject of passionate discussions and the taxonomic status of viruses is a sensitive and critical issue.

Virus classification is a relatively new exercise, as the first evidence for existence of a virus was only presented at the end of the nineteenth century by Beijerinck in 1898. It was not until 1927 that Johnson, a plant virologist, drew attention to the need for a system of virus nomenclature and classification. First efforts to classify viruses utilized a range of ecological and biological criteria, including pathogenic properties in the case of human and animal viruses, and symptoms for plant viruses. For example, viruses sharing the pathogenic properties causing hepatitis (e.g. hepatitis A virus, hepatitis B virus, yellow fever virus, and Rift Valley fever virus) were grouped together as 'the hepatitis viruses'. Virology developed substantially in the 1930s and early classifications for the viruses reflected these advances. In 1939, Holmes published a classification of plant viruses dependent on host reactions and differential host species, using a binomial-trinomial nomenclature based on the name of the infected plant; however, only 89 viruses were

described and classified in this way. With the development of electron microscopy and biochemical studies in the 1950s, the first virus groupings based on common virion properties emerged: like the Herpesvirus group described by Andrewes in 1954, the Myxovirus group by Andrewes et al, in 1955, and the Poxvirus group by Fenner and Burnet in 1957. During this period there was also an explosion of newly discovered viruses; in response, several individuals and committees independently proposed virus classification systems but none was widely adopted by the scientific community. It became obvious that only an international association of virologists could propose a comprehensive and universally acceptable system of virus classification.

At the 1966 International Congress for Microbiology held in Moscow, the International Committee on Nomenclature of Viruses (ICNV) was established by an international group of 43 virologists. An international organization was set up with the aim of developing a taxonomy and nomenclature system for all viruses that would be recognized worldwide. The name of the ICNV was changed in 1974 to the more appropriate International Committee on Taxonomy of Viruses (ICTV), which remains active today. ICTV, the unique official committee of the Virology Division, is now considered the international official body for all matters related to taxonomy and nomenclature of viruses.

Since the founding of the ICTV, all virologists have agreed that the hundreds of viruses isolated from different organisms should be classified together in a unique system, but separate from other microorganisms such as fungi, bacteria and mycoplasma. However, there was much controversy on the way to do it. Lwoff, Horne and Tournier argued for the adoption of a system classifying viruses into subphyla, classes, orders, suborders and families. Descending hierarchical divisions would have been based on nucleic acid type (DNA or RNA), strandedness (single or double), presence or absence of an envelope, capsid symmetry, and so on. The hierarchy of this system has never been recognized by the ICTV; nevertheless, the types of criteria used became the basis of the universal taxonomy system now in place, and all ICTV reports have used this scheme. Until 1990, no hierarchical

classification level higher than the family was used, however, the system has recently begun to move in this direction. A first order, Mononegavirales, was accepted in 1990, and another two, Caudovirales and Nidovirales, were adopted in 1996. In its nonlinnean structure, the scheme is quite different from that used in the taxonomy of bacteria and other organisms. Nevertheless, the usefulness of the scheme is being demonstrated by its wide application. It has replaced all competing classification schemes for all viruses and no one would now dispute with the ICTV the international mandate to name and classify viruses.

Since its establishment, a total of seven virus taxonomic reports (also known by the names of the ICTV Presidents acting as Editors in Chief of the reports) have been published by the ICTV: Wildy in 1971; Fenner in 1976; Matthews in 1979; Matthews in 1982; Francki et al in 1991; Murphy et al in 1995; and van Regenmortel et al in 1999. At the first meeting in Mexico City in 1970, two families with a corresponding two genera and 24 floating genera were adopted to begin the grouping of the vertebrate, invertebrate and bacterial viruses. In addition, 16 plant virus groups were designated, as reported by Matthews in 1983. The fifth ICTV report, edited by Francki et al in 1991, described one order, 40 families, nine subfamilies, 102 genera, two floating genera and two subgenera for vertebrate, invertebrate, bacterial and fungal viruses, and 32 groups and seven subgroups for plant viruses. While most virologists shifted to placing viruses in families and genera, plant virologists retained the term 'groups' until 1993. It was only in 1995, as described in the sixth ICTV report, that the ICTV proposed a uniform system for all viruses, with two orders, 50 families, nine subfamilies, 126 genera, 23 floating genera and four subgenera encompassing 2644 assigned viruses. Most recently, at the 28th meeting of the ICTV in March 1998 in San Diego, California, the Universal Virus Classification was adopted; this comprises three orders, 56 families, nine subfamilies, 203 genera, 30 floating genera and a total of 3954 species, strains and/ or serotypes of species and tentative species. It is a general trend that the number of described taxa and the number of species of viruses is increasing steadily, easily explained by the increasing complexity of the virus classification and by the amount of data available to demarcate viruses.

With precise and complete descriptions available for a large number of virus families, this classification now constitutes a valuable source of information for new 'unknown' viruses. Therefore, the ICTV classification is not only a taxonomic exercise for virus evolutionists but also a valuable diagnostic tool and educational system for virologists, teachers, medical doctors and epidemiologists.

How does the ICTV operate?

The ICTV is a committee of the Virology Division, which is in turn part of the International Union of Microbiological Societies. The ICTV is a nonprofitmaking organization composed of prominent virologists representing countries from throughout the world who work to designate virus names and taxa through a democratic process. The ICTV operates through a number of committees, subcommittees and study groups consisting of more than 492 eminent virologists with expertise in viruses infecting humans, animals, insects, protozoa, archaea, bacteria, mycoplasma, fungi, algae, yeasts and plants. Taxonomic proposals are initiated and formulated by individuals or by the study groups. These proposals are revised and accepted by the corresponding subcommittees and presented for executive committee approval. All decisions are then ratified at a plenary session (or also now by postal vote) held at each Virology Congress where all members of ICTV and more than 50 representatives of national microbiological societies are represented. At present, there are 47 study groups working in concert with six subcommittees - namely, the vertebrate, invertebrate, plant, bacteria, fungus and virus data subcommittees. The ICTV does not impose any taxonomic terms or taxa but ensures that all propositions are compatible with ICTV rules for homogeneity and consistency. The ICTV regularly publishes reports describing all existing virus taxa with a list of classified viruses as well as descriptions of virus families and genera. An Internet web site, where the most important information relative to virus taxonomy is made available, is updated regularly. The sixth report was published by Murphy et al (1995) and the seventh by van Regenmortel et al (1999).

The increasing number of virus species and virus strains being identified, together with the explosion of data on many descriptive aspects of viruses and viral diseases, and particularly sequence data, has led the ICTV to launch an international virus database project. This project, termed ICTVdB, is scheduled to be fully operational and accessible to the scientific community around the year 2000. The ICTVdB, in addition to the taxonomic descriptions of all the taxa, will comprise all the information available about each virus species, and later each virus strain, for all the descriptors necessary to identify and recognize all viruses.

A Universal System for Virus Classification

There are currently two systems in use for classifying organisms: the linnean and the adansonian systems.

The former is the monothetic hierarchical classification applied by Linnaeus to plants and animals, while the adansonian is a polythetic hierarchical system initially proposed by Adanson in 1763. In 1984 Maurin and collaborators suggested applying the linnean classification system to the viruses. Although convenient to use, this system has shortcomings when applied to the classification of viruses. Firstly, it is difficult to appreciate the validity of a particular criteria. For example, it may not be appropriate to use the number of genomic components as a hierarchical criteria. Secondly, there are no obvious reasons for prioritizing criteria, and in consequence it is difficult to rank all the available criteria. For instance, is the nature of the genome (DNA/RNA) more important than the sense of the coding sequence of the genome or the shape of the virus particles?

The adansonian system considers all available criteria at once and makes several classifications. taking the criteria into consideration successively. The criteria leading to the same classifications are considered as correlated and are therefore not discriminatory. Subsequently, a subset of criteria are considered, and the process is repeated until all criteria can be ranked to provide the best discrimination of the species. This system has not been used frequently in the past owing to its labor-intensive nature, but this situation has changed as a result of the power and availability of today's computer technology. Furthermore, qualitative and quantitative data can be simultaneously considered when generating such a classification. In the case of viruses, it was determined by Harrison and collaborators in 1971 that at least 60 characters could be used for a complete virus description (Table 1). Thus, the limiting factor for applying the adansonian system is now not its labor-intensive nature but the lack of data for many of the viruses.

In addition, the increasing number of viral nucleic acid sequences being reported, in combination with the appropriate computer software, allows the comparison of viruses to generate different phylogenetic trees, according to the gene or set of genes used, as for example proposed by Koonin in 1991, Dolja and Koonin in 1991 and Dolja et al in 1991. However, to date, none of them has satisfactorily provided a clear classification of all viruses. A multidimensional classification, taking into account all the criteria necessary to describe viruses, would probably be the most appropriate way of representing a virus classification, but again the shortcomings of data for some viruses would prevent the use of this system in the foreseeable future.

For almost 25 years, the ICTV has been classifying viruses essentially at the family and genus levels using a nonsystematic polythetic approach. Viruses were

clustered first in genera and then in families. A subset of characters, including physicochemical, structural, genomic and biological criteria, is then used to compare and group viruses. This subset of characters may change from one family to another, according to the availability of the data and the importance of a particular character for a particular family. It is obvious that there is no homogeneity in this respect throughout the virus classification and that virologists weigh the criteria differently in this subjective process, leading to the generation of a nonhomogeneous classification. Nevertheless, over time we can see stability of the current ICTV classification at the genus and family level. When sequence, genomic organization and replicative cycle data are subsequently used for taxonomic purposes, they usually confirm the actual classification. It is also obvious that hierarchical classifications above the family level will encounter conflicts between phenotypic and genotypic criteria and that virologists will have to consider the entire classification process in order to progress in this direction.

Currently, and for practical reasons only, virus classification is structured according to the presentation indicated in Tables 2 and 3. This 'Order of Presentation of the Viruses' does not reflect any hierarchical or phylogenetic classification but only a convenient order of presentation of the virus taxa. Since a taxonomic structure above the level of family (with the exception of the orders Mononegavirales, Caudovirales and Nidovirales) has not been developed extensively, any listing must be arbitrary. The order of presentation of virus families and genera follows four criteria: (1) the nature of the viral nucleic acid; (2) the strandedness of the nucleic acid; (3) the use of a reverse transcription process (DNA or RNA); and (4) the positive or negative sense of gene coding on the encapsidated genome. These four criteria give rise to six clusters comprising the 86 families and floating genera of viruses. In the past, two other criteria were also taken in account: the presence or absence of a lipid envelope and the segmentation of the genome as mono-, bi-, tri-, tetra- or multipartite. However, it has become clear that the presence of an envelope was entirely related to the nature of the host and that families could comprise genera having viruses with segmented or nonsegmented genomes, but sharing all other properties, including genome organization and sequence homology. These criteria have been therefore abandoned.

The Virus Species Concept and its Application

In 1991 the ICTV accepted the concept that viruses

Table 1 Virus family descriptors used in virus taxonomy

I Virion properties

A Morphology properties of virions

- 1 Size
- 2 Shape
- 3 Presence or absence of an envelope and peplomers
- 4 Capsomeric symmetry and structure

B Physical properties of virions

- 1 Molecular mass
- 2 Buoyant density
- 3 Sedimentation coefficient
- 4 pH stability
- 5 Thermal stability
- 6 Cation (Mg2+, Mn2+) stability
- 7 Solvent stability
- 8 Detergent stability
- 9 Radiation stability

C Properties of genome

- 1 Type of nucleic acid DNA or RNA
- 2 Strandedness: single-stranded or double-stranded
- 3 Linear or circular
- 4 Sense: positive, negative or ambisense
- 5 Number of segments
- 6 Size of genome or genome segments
- 7 Presence or absence and type of 5' terminal cap
- 8 Presence or absence of 5' terminal covalently linked polypeptide
- 9 Presence or absence of 3' terminal poly(A) tract (or other specific tract)
- 10 Nucleotide sequence comparisons

D Properties of proteins

- 1 Number
- 2 Size
- 3 Functional activities (especially virion transcriptase, virion reverse transcriptase, virion hemagglutinin, virion neuraminidase, virion fusion protein)
- 4 Amino acid sequence comparisons

E Lipids

- 1 Presence or absence
- 2 Nature

F Carbohydrates

- 1 Presence or absence
- 2 Nature

Il Genome organization and replication

- 1 Genome organization
- 2 Strategy of replication of nucleic acid
- 3 Characteristics of transcription
- 4 Characteristics of translation and post-translational processing
- 5 Site of accumulation of virion proteins, site of assembly, site of maturation and release
- 6 Cytopathology, inclusion body formation

III Antigenic properties

- 1 Serological relationships
- 2 Mapping epitopes

IV Biological properties

- 1 Host range, natural and experimental
- 2 Pathogenicity, association with disease
- 3 Tissue tropisms, pathology, histopathology

Table 1 Continued

- 4 Mode of transmission in nature
- 5 Vector relationships
- 6 Geographic distribution

Adapted from ICTV guidelines for family descriptions.

exist as species, in a similar manner to other organisms, and adopted a definition for a virus species proposed by van Regenmortel in 1990: 'A virus species is a polythetic class of viruses that constitutes a replicating lineage and occupies a particular ecological niche.' This simple definition and the position taken by the ICTV has already had, and will continue to have, a profound effect on virus classification. Effectively, in the sixth ICTV report virus names were indicated in 'List of species' but they were in fact a 'List of virus names' with undefined taxonomic status. In the seventh ICTV report, according to the polythetic nature of the species definition, a 'List of species-demarcating criteria' is provided for each genus, indicating how virus species can be identified in this particular genus. Viruses are then differentiated in species and tentative species according to this list of criteria and the availability of information to demarcate the species.

First, it is intended to define for each genus the criteria demarcating a virus species, and, second, to compare these criteria from one genus to the next, searching for homogeneity throughout the virus classification. Naturally this list of criteria should follow the polythetic nature of the species definition and more than one criteria should be used to determine a new species. It is obvious that most of the criteria in the list of demarcating criteria are shared amongst the different genera, within and across families; namely, host range, serological relationships, vector transmission type, tissue tropism, genome rearrangement and sequence homology (Table 4). However, if the types of criteria are similar, the levels of demarcation clearly differ from one family to another. This may reflect differences in appreciation from one family to another but also the differential ranking of a particular criterion in different families. The huge differences (up to 30%) in sequences among nucleoproteins of species of lentiviruses does not have the same biological significance as small differences in capsid protein sequences (1-10%) of species of potyviruses, and therefore universal levels of sequence identity for similar genes may not exist for viruses! The levels of demarcation may even change from one gene to another within the same family. Homogenization of the application of the species definition concept throughout the virus definition will be the next challenge of ICTV for the eighth report to be published by 2002. This, in turn, will contribute to homogeneity of the genus and family demarcation criteria (Table 4) and will permit creation of new families or merging of existing families. However, it is important to note that the nature of the demarcating criteria at the genus level will probably not change as these have passed the test of time. Despite the fact that they were mostly established using biochemical and structural criteria, they remained valid when correlated with genome organization and sequence data.

The Universal Virus Classification

The present universal system of virus taxonomy is set arbitrarily at hierarchical levels of order, family (in some cases subfamily), genus and species. Lower hierarchical levels, such as subspecies, strain, serotype, variant, pathotype and isolate are established by international specialty groups and/or by culture collections, but not by the ICTV. However some of them may be indicated in the ICTV report for information or because in the past these names were listed as 'viruses' in previous reports.

Virus species

The species taxon is always regarded as the most important taxonomic level in classification but it has proved to be the most difficult to apply to the viruses. ICTV definition of a virus species was long considered to be 'a concept that will normally be represented by a cluster of strains from a variety of sources, or a population of strains from a particular source, which have in common a set or pattern of correlating stable properties that separates the cluster from other clusters of strains' as stated by Matthews in 1982 and by Francki et al in 1991. This was a general definition, which was in fact not very useful for practically delineating species in a particular family. Furthermore, this definition directly addressed the definition of a virus strain, which had never been attempted in the history of virus taxonomy. In 1991, the ICTV Executive Committee accepted a definition proposed by van Regenmortel in 1990 (see above). This definition states: 'A virus species is a polythetic class of viruses that constitutes a replicating lineage

e viruses
the
ō
presentation
ŏ
Order
N
aple

The dabM viruses Caudovirales Myovindes Tit-like viruses" Siphovindas Siphovindas Siphovindas Formit with viruses and security of the viruses of the viruses and se	Order	Family	Subfamily	Genus	Type species	Host
A viruses Myoundse "T4-like viruses" "P2-like viruses" "P2-like viruses" "P1-like viruses" "P1-like viruses" "P1-like viruses" "P2-like viruses" "P3-like viruses" "P4-like viruses" "P3-like viruses" "P4-like viruses" "P3-like viruses" "P4-like viruses" "P5-like	The DNA viruses					
Myoviridae T14-like viruses" "P1-like viruses" "Sp01-like viruses" "SP01-like viruses" "SP01-like viruses" "AH-like viruses" "T1-like viruses" "C2-like viruses" "P22-like viruses" "P23-like viruses" "P24-like viruses" "P24-like viruses" "P25-like viruse	The dsDNA viruses					
"T4-like viruses" "P1-like viruses" "P2-like viruses" "Mu-like viruses" "A-like viruses" "A-like viruses" "T6-like viruses" "T6-like viruses" "P22-like viruses" "P23-like viruses" "P24-like viruses" "P24-like viruses" "P24-like viruses" "P25-like viruses" "P25-like viruses" "P26-like viruses" "P26	Caudovirales	Myoviridae				
"P1-like viruses" "P2-like viruses" "Mu-like viruses" "SP01-like viruses" "A-like viruses" "T1-like viruses" "C2-like viruses" "P22-like viruses" "Pasmavirus "Pasmavirus "SNDV-like viruses" "SNDV-like viruses" "SNDV-like viruses" "SNDV-like viruses" "SNDV-like viruses" "SNDV-like viruses"				"T4-like viruses"	Enterobacteria phage T4	Bacteria
"P2-like viruses" "Mu-like viruses" "SP01-like viruses" "#Hite viruses" "#Hite viruses" "T1-like viruses" "#2-like viruses" "#22-like viruses" "#23-like viruses" "#24-like viruses" "#24-like viruses" "#24-like viruses" "#25-like viruses" "#2				"P1-like viruses"	Enterobacteria phage P1	Bacteria
"Mu-like viruses" "SP01-like viruses" "P11-like viruses" "T1-like viruses" "T2-like viruses" "C2-like viruses" "P22-like viruses" "P23-like viruses" "P23-like viruses" "P24-like viruses" "P24-like viruses" "P24-like viruses" "P25-like viruses" "P25-like viruses" "P26-like viruses" "P26-like viruses" "P26-like viruses" "P27-like viruse				"P2-like viruses"	Enterobacteria phage P2	Bacteria
"SP01-like viruses" "#Hike viruses" "#Hike viruses" "T1-like viruses" "T5-like viruses" "#M1-like viruses" "#M1-like viruses" "#M1-like viruses" "##################################				"Mu-like viruses"	Enterobacteria phage Mu	Bacteria
" " " " " " " " " " " " "				"SP01-like viruses"	Bacillus phage SP01	Bacteria
"Y-like viruses" "T5-like viruses" "T5-like viruses" "C2-like viruses" "C2-like viruses" "P22-like viruses" "P23-like viruses" "P24-like viruses" "P24-like viruses" "P25-like viruses" "P25-like viruses" "P25-like viruses" "P26-like viruses" "P27-like viruses" "P26-like viruses" "P27-like viruses" "P26-like viruses" "P27-like viruses"				" ## AH-like viruses"	Halobacterium virus <i>ϕH</i>	Archaea
"A-like viruses" "T1-like viruses" "T5-like viruses" "C2-like viruses" "C2-like viruses" "P22-like viruses" "A29-like viruses" "A17-like viruses" "A29-like viruses" "Andivirus "Andivirus "Andivirus "Andipoxvirus "Anipoxvirus "Anipoxvirus "Anipoxvirus "Anipoxvirus "Anipoxvirus		Siphoviridae				
"T1-like viruses" "T5-like viruses" "C2-like viruses" "C2-like viruses" "P22-like viruses" "Pasmavirus "Plasmavirus "Subtrivirus "Subtrivir				">-like viruses"	Enterobacteria phage >>	Bacteria
"T5-like viruses" "L5-like viruses" "C2-like viruses" "P22-like viruses" "Paramovirus "SNDV-like viruses"				"T1-like viruses"	Enterobacteria phage T1	Bacteria
"L5-like viruses" "C2-like viruses" "P22-like viruses" "P22-like viruses" "P22-like viruses" "P22-like viruses" "P23-like viruses" "A24-like viruses" "A25-like viruses" "A26-like viruses" "A26-like viruses" "A36 "A36 Chordopoxvirinae Chordopoxvirinae Orthopoxvirus Avipoxvirus Avipoxvirus Capripoxvirus Capripoxvirus Capripoxvirus				"T5-like viruses"	Enterobacteria phage T5	Bacteria
"C2-like viruses" "#M1-like viruses" "P22-like viruses" "P22-like viruses" " P22-like viruses" " Pasmavirus " Parapoxvirus " Parapoxvirus				"L5-like viruses"	Mycobacterium phage L5	Bacteria
" " " " " " " " " " " " " " " " " " "				"c2-like viruses"	Lactococcus phage c2	Bacteria
### T7-like viruses" ##################################				" ψ M1-like viruses"	Methanobacterium virus ψM1	Archaea
"T7-like viruses" "P22-like viruses" "P22-like viruses" fae		Podoviridae				
"P22-like viruses" "#429-like viruses" Jae Corticovirus Jae Corticovirus Idae Lipothrixvirus Rudivirus Rudivirus Chordopoxvirinae Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Avipoxvirus Capripoxvirus Capripoxvirus				"T7-like viruses"	Enterobacteria phage T7	Bacteria
"#429-like viruses" Iae Corticovirus Iae Corticovirus Idae Lipothrixvirus Rudivirus Rudivirus Rudivirus Parapoxvirus Chordopoxvirinae Chordopoxvirins Avipoxvirus Capripoxvirus Capripoxvirus Capripoxvirus				"P22-like viruses"	Enterobacteria phage P22	Bacteria
Tectivirus lae Corticovirus lae Plasmavirus lidae Lipothrixvirus Rudivirus Rudivirus Rudivirus Parapoxvirus Avipoxvirus Parapoxvirus Parapoxvirus Avipoxvirus Capripoxvirus Capripoxvirus				"¢29-like viruses"	Bacillus phage 429	Bacteria
lae Corticovirus lae Plasmavirus idae Lipothrixvirus lae Rudivirus lae Fusellovirus "SNDV-like viruses" Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus		Tectiviridae		Tectivirus	Enterobacteria phage PRD1	Bacteria
idae Lipothrixvirus Plasmavirus Rudivirus Rudivirus Rudivirus Rudivirus Fusellovirus "SNDV-like viruses" Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus		Corticoviridae	!	Corticovirus	Alteromonas phage PM2	Bacteria
idae Lipothrixvirus Rudivirus Iae Fusellovirus "SNDV-like viruses" Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus		Plasmaviridae		Plasmavirus	Acholeplasma phage L2	Mycoplasma
Hudivirus Fusellovirus "SNDV-like viruses" Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus Capripoxvirus		Lipothrixviridae		Lipothrixvirus	Thermoproteus virus 1	Archaea
"SNDV-like viruses" Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus		Rudiviridae		Rudivirus	Sulfolobus virus SIRV1	Archaea
"SNDV-like viruses" Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus		Fuselloviridae		Fusellovirus	Sulfolobus virus SSV1	Archaea
Chordopoxvirinae Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus	į			"SNDV-like viruses"	Sulfolobus virus SNDV	Archaea
Orthopoxvirus Parapoxvirus Avipoxvirus Capripoxvirus		Poxviridae				
Sr Sn			Chordopoxvirinae			
s sr				Orthopoxvirus	Vaccinia virus	Vertebrates
SI				Parapoxvirus	Orf virus	Vertebrates
				Avipoxvirus	Fowlpox virus	Vertebrates
				Capripoxvirus	Sheeppox virus	Vertebrates

pen
ă
ၓ
N
0
욮

Table 2 Continued					
Order	Family	Subfamily	Genus	Type species	Host
			orani produceo 1	Moomo vieno	Vortohratoe
			Leponboxinas	Mykonia viius	ע פו ופתו מופס
			Suipoxvirus	Swinepox virus	Vertebrates
			Molluscipoxvirus	Molluscum contagiosum virus	Vertebrates
			Yatapoxvirus	Yaba monkey tumor virus	Vertebrates
		Entomopoxvirinae			
			Entomopoxvirus A	Melolontha melolontha entomopoxvirus	Invertebrates
			Entomopoxvirus B	Amsacta moorei entomopoxvirus	Invertebrates
			Entomopoxvirus C	Chironomus luridus entomopoxvirus	Invertebrates
	Asfarviridae		Asfivirus	African swine fever virus	Vertebrates ^b
	Iridoviridae				
			Iridovirus	Chilo iridescent virus	Invertebrates
			Chloriridovirus	Mosquito iridescent virus	Invertebrates
			Ranavirus	Frog virus 3	Vertebrates
			Lymphocystivirus	Flounder virus	Vertebrates
	Phycodnaviridae				
			Chlorovirus	Paramecium bursaria Chlorella virus 1	Algae
			Prasinovirus	Micromonas pusilla virus SP1	Algae
			Prymnesiovirus	Chrysochromulina brevifilum virus	Algae
			Phaeovirus	Ectocarpus siliculosus virus 1	Algae
	Baculoviridae				
			Nucleopolyhedrovirus	Autographa californica	Invertebrates
			Granulovirus	riaciscoporprisarios Cydia pomonella granulovirus	Invertebrates
	Herpesviridae				
		Alphaherpesvirinae			
			Simplexvirus	Human herpesvirus 1	Vertebrates
			Varicellovirus	Human herpesvirus 3	Vertebrates
			"Marek's disease-like	Marek's disease virus	Vertebrates
			"II TV-like vinsee"	Infactions larypootrachaitie vinus	Vertebrates
		Betaherpesvirinae			
			Cytomegalovirus	Human herpesvirus 5	Vertebrates
			Muromegalovirus	Mouse cytomegalovirus 1	Vertebrates
			Roseolovirus	Human herpesvirus 6	Vertebrates

ed
ntin
රි
Ŋ
훒
Œ

Order	Family	Subfamily	Genus	Type species	Host
		Gammaherpesvirinae			
			Lymphocryptovirus	Human herpesvirus 4	Vertebrates
			Rhadinovirus	Ateline herpesvirus 2	Vertebrates
			"Ictalurid herpes-like viruses"	Ictalurid herpesvirus 1	Vertebrates
	Adenoviridae				
			Mastadenovirus	Human adenovirus 2	Vertebrates
			Aviadenovirus	Fowl adenovirus 1	Vertebrates
			Rhizidiovirus	Rhizidiomyces virus	Fungi
	Polyomaviridae				
		:	Polyomavirus	Murine polyomavirus	Vertebrates
	Papillomaviridae				
			Papillomavirus	Cottontail rabbit papillomavirus	Vertebrates
	Polydnaviridae				
			Ichnovirus	Campoletis sonorensis virus	Invertebrates
			Bracovirus	Cotesia melanoscela virus	Invertebrates
	Ascoviridae		Ascovirus	Spodoptera frugiperda ascovirus	Invertebrates
The ssDNA viruses					
	Inoviridae			;	
			Inovirus	Coliphage fd	Bacteria
			Plectrovirus	Acholoplasma phage L51	Mycoplasma
	Microviridae				
			Microvirus	Coliphage <i>♦X174</i>	Bacteria
			Spiromicrovirus	Spiroplasma phage 4	Spiroplasma
			Bdellomicrovirus	Bdellovibrio phage MAC1	Bacteria
			Chlamydiamicrovirus	Chlamydia phage 1	Bacteria
	Geminiviridae				
			Mastrevirus	Maize streak virus	Plants
			Curtovirus	Beet curly top virus	Plants
			Begomovirus	Bean golden mosaic virus	Plants
	Circoviridae		Circovirus	Chicken anemia virus	Vertebrates
			Nanovirus	Subterranean clover stunt virus	Plants

Table 2 Continued

Oppor	,	O. Money.		***	11.
Older	raimiy	Subramily	Genus	lype species	Host
	Parvoviridae				
		Parvovirinae			
			Parvovirus	Mice minute virus	Vertebrates
			Erythrovirus	B19 virus	Vertebrates
			Dependovirus	Adeno-associated virus 2	Vertebrates
		Densovirinae			
			Densovirus	Junonia coenia densovirus	Invertebrates
			Iteravirus	Bombyx mori densovirus	Invertebrates
			Brevidensovirus	Aedes aegypti densovirus	Invertebrates
The DNA and RNA reverse transcribing viruses	se transcribing viruses				
	Hepadnaviridae				
			Orthohepadnavirus	Hepatitis B virus	Vertebrates
			Avihepadnavirus	Duck hepatitis B virus	Vertebrates
	Caulimoviridae				
			Caulimovirus	Cauliflower mosaic virus	Plants
			"PVCV-like viruses"	Petunia vein-clearing virus	Plants
			"SbCMV-like viruses"	Soybean chlorotic mottle virus	Plants
			"CsVMV-like viruses"	Cassava vein mosaic virus	Plants
			Badnavirus	Commelina yellow mottle virus	Plants
	:		"RTBV-like viruses"	Rice tungro bacilliform virus	Plants
	Pseudoviridae				in the latest and the
			Pseudovirus	Saccharomyces cerevisiae Ty1 virus	Yeast, Plants
			Hemivirus	Drosophila melanogaster copia virus	Yeast, Invertebrates
	Metaviridae				
			Metavirus	Saccharomyces cerevisiae Ty3 virus	Yeast, Plants, Invertebrates
			Errantivirus	Drosophila melanogaster gypsy virus	Invertebrates
	Retroviridae				;
			Alpharetrovirus	Avian leukosis virus	Vertebrates
			Betaretrovirus	Mason-Pfizer monkey virus	Vertebrates
			Gammaretrovirus	Mouse mammary tumor virus	Vertebrates
			Deltaretrovirus	Bovine leukemia virus	Vertebrates
			Epsilonretrovirus	Walleye dermal sarcoma virus	Vertebrates
			Lentivirus	Human immunodeficiency virus 1	Vertebrates
			Spumavirus	Human spumavirus	Vertebrates

9
3
₽
⊆
0
Contir
Q
0
Table 2

Order	Family	Subfamily	Genus	Type species	Host
The RNA viruses					
The dsRNA viruses					
	Cystoviridae		Cystovirus	Pseudomonas phage φ6	Bacteria
	Reoviridae		111		
			Orthoreovirus	Reovirus 3	Vertebrates
			Orbivirus	Bluetongue virus 1	Vertebrates
			Rotavirus	Simian rotavirus SA11	Vertebrates
			Coltivirus	Colorado tick fever virus	Vertebrates
			Aquareovirus	Golden shiner virus	Vertebrates
			Cypovirus	Bombyx mori cypovirus 1	Invertebrates
			Fijivirus	Fiji disease virus	Plants
			Phytoreovirus	Wound tumor virus	Plants
			Oryzavirus	Rice ragged stunt virus	Plants
!	Birnaviridae				
			Aquabimavirus	Infectious pancreatic necrosis virus	Vertebrates
			Avibirnavirus	Infectious bursal disease virus	Vertebrates
			Entomobirnavirus	Drosophila X virus	Invertebrates
	Totiviridae				
			Totivirus	Saccharomyces cerevisiae virus L-A	Fungi
			Giardiavirus	Giardia lamblia virus	Protozoa
			Leishmaniavirus	Leishmania RNA virus 1-1	Protozoa
	Partitiviridae				
			Partitivirus	Gaeumannomyces graminis virus 019/6-A	Fungi
			Chrysovirus	Penicillium chrysogenum virus	
			Alphacryptovirus	White clover cryptic virus 1	Plants
			Betacryptovirus	White clover cryptic virus 2	Plants
	Hypoviridae		Hypovirus	Cryphonectria hypovirus 1-EP713	Fungi
			Varicosavirus	Lettuce big-vein virus	Plants
The negative-stranded ssRNA viruses	ssRNA viruses				
	Bomaviridae				
			Bornavirus	Borna disease virus	Vertebrates

7
ā
=
\simeq
_
#2
ō
べ
u
-
α
-
ž
ž

Order	Family	Subfamily	Genus	Type species	Host
	Filoviridae				
			"Ebola-like viruses" Zaïre	Ebola virus	Vertebrates
			"Marburg-like viruses"	Marburg virus	Vertebrates
	Paramyxoviridae				
		Paramyxovirinae			
			Respirovirus	Human parainfluenza virus 1	Vertebrates
			Morbillivirus	Measles virus	Vertebrates
			Rubulavirus	Mumps virus	Vertebrates
		Pneumovirinae			
			Pneumovirus	Human respiratory syncytial virus	Vertebrates
	Dhohdo: dido		Metapneumovirus	Turkey rhinotracheitis virus	Vertebrates
	niabuoviildae				
			Vesiculovirus	Vesicular stomatitis Indiana virus	Vertebrates
			Lyssavirus	Rabies virus	Vertebrates
			Ephemerovirus	Bovine ephemeral fever virus	Vertebrates
			Novirhabdovirus	Infectious hematopoletic necrosis virus	Vertebrates
			Cytorhabdovirus	Lettuce necrotic yellows virus	Plants
			Nucleorhabdovirus	Potato yellow dwarf virus	Plants
	Orthomyxoviridae				
			Influenzavirus A	Influenza A virus	Vertebrates
			Influenzavirus B	Influenza B virus	Vertebrates
			Influenzavirus C	Influenza C virus	Vertebrates
		:	Thogotovirus	Thogoto virus	Vertebrates
	Bunyaviridae				
			Bunyavirus	Bunyamwera virus	Vertebrates
			Hantavirus	Hantaan virus	Vertebrates
			Nairovirus	Nairobi sheep disease virus	Vertebrates
			Phlebovirus	Sandfly fever Sicilian virus	Vertebrates
			Tospovirus	Tomato spotted wilt virus	Plants
			Tenuívirus	Rice stripe virus	Plants
			Ophiovirus	Citrus psorosis virus	Plants
	Arenaviridae		Arenavirus	Lymphocytic choriomeningitis virus	Vertebrates
			Deltavirus	Hepatitis delta virus	Vertebrates

þe	
ij	
ပွဲ	
N	
5	
a	

Order	Subfamily		F	
	Sublamily	Genus	lype species	Host
The positive-stranded ssRNA viruses				
L.eviviridae				
		Levivirus	Enterobacteria phage MS2	Bacteria
		Allolevivirus	Enterobacteria phage Qβ	Bacteria
Narnaviridae				
		Namavirus	Saccharomyces cerevisiae 20S namavirus	Yeast
		Mitovirus	Cryphonectria parasitica NB631 virus	Yeast
Picomaviridae	*			
		Enterovirus	Poliovirus 1	Vertebrates
		Rhinovirus	Human rhinovirus 1A	Vertebrates
		Hepatovirus	Hepatitis A virus	Vertebrates
		Cardiovirus	Encephalomyocarditis virus	Vertebrates
		Aphthovirus	Foot-and-mouth disease virus O	Vertebrates
		Parechovirus	Human echovirus 22	Vertebrates
		"Cricket paralysis-like viruses"	Cricket paralysis virus	Invertebrates
Sequiviridae				
		Sequivirus	Parsnip yellow fleck virus	Plants
	, chart	Waïkavirus	Rice tungro spherical virus	Plants
Comoviridae				
		Comovirus	Cowpea mosaic virus	Plants
		Fabavirus	Broad bean wilt virus 1	Plants
		Nepovirus	Tobacco ringspot virus	Plants
Potyviridae				
		Potyvirus	Potato virus Y	Plants
		Rymovirus	Ryegrass mosaic virus	Plants
		Macluravirus	Maclura mosaic virus	Plants
		Ipomovirus	Sweet potato mild mottle virus	Plants
		Bymovirus	Barley yellow mosaic virus	Plants
		Tritimovirus	Wheat streak mosaic virus	Plants
Caliciviridae				
		Vesivirus	Swine vesicular exanthema virus	Vertebrates
		Lagovirus	Rabbit hemorrhagic disease virus	Vertebrates

Continued	
N	
Table	

Order	Family	Subfamily	Genus	Type species	Host
			"Norwalk-like viruses"	Norwalk virus	Vertebrates
			"Sapporo-like viruses"	Sapporo virus	Vertebrates
			"Hepatitis E-like viruses"	Hepatitis E virus	Vertebrates
	Astroviridae		Astrovirus	Human astrovirus 1	Vertebrates
	Nodaviridae				
			Alphanodavirus	Nodamura virus	Invertebrates
			Betanodavirus	Striped jack nervous necrosis virus	Vertebrates
	Tetraviridae				
			Betatetravirus	Nudaurelia capensis β virus	Invertebrates
			Omegatetravirus	Nudaurelia capensis ω virus	Invertebrates
			Sobemovirus	Southern bean mosaic virus	Plants
			Marafivirus	Maize rayado fino virus	Plants
	Luteoviridae				
			Luteovirus	Barley yellow dwarf virus – MAV	Plants
			Polerovirus	Potato leafroll virus	Plants
			Enamovirus	Pea enation mosaic virus 1	Plants
			Umbravirus	Carrot mottle virus	Plants
	Tombusviridae				
			Avenavirus	Oat chlorotic stunt virus	Plants
			Aureusvirus	Pothos latent virus	Plants
			Carmovirus	Carnation mottle virus	Plants
			Dianthovirus	Camation ringspot virus	Plants
			Machlomovirus	Maize chlorotic mottle virus	Piants
			Necrovirus	Tobacco necrosis virus	Plants
			Panicovirus	Panicum mosaic virus	Plants
			Tombusvirus	Tomato bushy stunt virus	Plants
Nidovirales					
	Coloriaviidae				
			Coronavirus	Avian infectious bronchitis virus	Vertebrates
	Arterivindae		Torovirus	Berne virus	Vertebrates
	O. C.		Arterivirus	Equine arteritis virus	Vertebrates

per	
ontin	
Ŏ N	
ble 2	
酉	

Order	Family	Subfamily	Genus	Type species	Host
	Flaviviridae				
			Flavivirus	Yellow fever virus	Vertebrates
			Pestivirus	Bovine diarrhea virus	Vertebrates
			Hepacivirus	Hepatitis C virus	Vertebrates
	Togaviridae				
			Alphavirus	Sindbis virus	Vertebrates
			Rubivirus	Rubella virus	Vertebrates
			Tobamovirus	Tobacco mosaic virus	Plants
			Tobravirus	Tobacco rattle virus	Plants
			Hordeivirus	Barley stripe mosaic virus	Plants
			Furovirus	Soil-borne wheat mosaic virus	Plants
			Pomovirus	Potato mop-top virus	Plants
			Pecluvirus	Peanut clump virus	Plants
			Benyvirus	Beet necrotic yellow vein virus	Plants
	Bromoviridae				
			Alfamovirus	Alfalfa mosaic virus	Plants
			Bromovirus	Brome mosaic virus	Plants
			Cucumovirus	Cucumber mosaic virus	Plants
			llarvirus	Tobacco streak virus	Plants
			Oleavirus	Olive latent virus 2	Plants
			Ourniavirus	Ourmia melon virus	Plants
			Idaeovirus	Rasberry bushy dwarf virus	Plants
	Closteroviridae				
			Closterovirus	Beet yellows virus	Plants
			Crinivirus	Lettuce infectious yellows virus	Plants
			Capillovirus	Apple stem grooving virus	Plants
			Trichovirus	Apple chlorotic leaf spot virus	Plants
			Vitivirus	Grapevine virus A	Plants
			Tymovirus	Turnip yellow mosaic virus	Plants
			Carlavirus	Camation latent virus	Plants

Table 2 Continued

Order	Family	Subfamily	Genus	Type species	Host
			Potexvirus	Potato virus X	Plants
			Allexivirus	Shallot virus X	Plants
			Foveavirus	Apple stem pitting virus	Plants
	Barnaviridae		Barnavirus	Mushroom bacilliform virus	Fungi
Unassigned viruses The subviral agents:	Unassigned viruses The subviral agents: viroids, satellites and agents	ents of spongiform encephalopathies (prions)	lopathies (prions)		
Subviral agent	Family	Genus	Type species	Host	st
Viroids					
	Pospiviroidae				
		Pospiviroid	Potato spindle tuber viroid	viroid Plants	nts
		Hostuviroid	Hop stunt viroid	Plants	nts
		Cocadviroid	Coconut cadang-cadang viroid	lang viroid Plants	nts
		Apscaviroid	Apple scar skin viroid	7 Plants	nts
		Coleviroid	Coleus blumei viroid 1	1 Plants	nts
	Avsunviroidae				
		Avsunviroid	Avocado sunblotch viroid	iroid Plants	nts
		Pelamoviroid	Peach latent mosaic virus	virus Plants	ints
Satellites				Plants	nts
				inve	invertebrates
				Fungi	igi
Prions				Ved	Vertebrates
				Fungi	igr

^a Quotes are used to denote taxon names that are not approved ICTV international names, and are thus temporary until formal names are approved.

^b Vertebrate arthropod-borne viruses are listed according to their vertebrate hosts.

Table 3 Orders, families and floating genera of viruses according to the seventh ICTV report (1999)

								Number of	Number of species		
Criteria	Order	Family	Floating genus	Morphology	Genome configuration	Genome size (kb)	Virus host	Species	Strains/ serotypes	Tentative	Total
dsDNA	Caudovirales	Myoviridae		Tailed phage	1 linear	336	Bacteria, archaea	15	23	117	155
		Siphoviridae		Tailed phage	1 linear	53	Bacteria, archaea	7	0	137	144
		Podoviridae		Tailed phage	1 linear	40	Bacteria, archaea	œ	5	99	98
		Tectiviridae		Isometric	1 linear	16	Bacteria	4	0	88	45
		Corticoviridae		Isometric	1 circular supercoiled	10	Bacteria	-	0	α	က
		Plasmaviridae		Pleomorphic	1 circular	12	Mycoplasma	-	0	7	ထ
		Lipothrixviridae		Rod	1 linear	16	Archaea	Ŋ	0	0	8
		Rudiviridae		Rod	1 linear	33-36	Archaea	ત	0		က
		Fuselloviridae		Lemon-shape	1 circular supercoiled	15	Archaea	-	0	0	-
			"SNDV-like viruses"	Droplet-shape	1 circular	20	Archaea	-	0	0	-
		Poxviridae		Ovioid	1 linear	130–375	Vertebrate, invertebrate	62	œ	23	93
		Asfarviridae		Isometric	1 circular	170-190	Vertebrate	-	0	0	-
		Iridoviridae		Isometric	1 linear	160-400	Vertebrate, invertebrate	17	4	ო	24
		Phycodnaviridae		Isometric	1 linear	250-350	Algae	27	0	38	65
		Baculoviridae		Bacilliform	1 circular supercoiled	90–230	Invertebrate	17	ဖ	7	30
		Herpesviridae		Isometric	1 linear	120-220	Vertebrate	26	0	65	121
		Adenoviridae		Isometric	1 linear	32-48	Vertebrate	56	102	32	163
			Rhizidiovirus	Isometric	1 linear	27	Fungus	-	0	0	-
		Polyomaviridae		Isometric	1 circular	ა	Vertebrate	12	4	0	16
		Papillomaviridae		Isometric	1 circular	6.8-8.4	Vertebrate	7	0	88	92
		Polydnaviridae		Rod, fusiform	1 circular supercoiled	2–28	Invertebrate	29	0	0	29
		Ascoviridae		Ovoid and bacilliform	1 circular	100-180	Invertebrate	ო	0	-	4
								330	159	628	117

Table 3 Continued

								Number (Number of species		1
Criteria	Order	Family	Floating genus	Morphology	Genome configuration	Genome size (kb)	Virus host	Species	Strains/ serotypes	Tentative	Tota!
ssDNA		Inoviridae		Rod	1 circular	7–20	Bacteria, mycoplasma	36	7	ഹ	84
		Microviridae		Isometric	1 circular	9	Bacteria, spiroplasma	7	0	88	40
		Geminivirus		Isometric	1 or 2 circular	မှ	Plant	94	81	10	106
		Circoviridae		Isometric	1 circular	1.7-2.3	Vertebrate	ო	0	,	4
			Nanovirus	Isometric	6-9 circular	6-9	Plant	4	0	-	2
		Parvoviridae		Isometric	1 - strand	8	Vertebrate, invertebrate	88	0	16	54
:								182	0	99	257
ssDNA RT		Hepadnaviridae		Isometric	1 circular strand	က	Vertebrate	ς.	0	8	7
dsDNA RT		Caulimoviridae		Isometric, bacilliform	1 circular	ας.	Plant	56	0	&	8
ssRNA RT		Pseudoviridae		Ovoid	1 linear	5-8	Yeast, plant	15	0	0	5
ssRNA RT		Metaviridae		Isometric	1 linear	4-10	Yeast, fungus, Invertebrate	18	0	-	19
ssRNA RT		Retroviridae		Spherical	dimer 1 + segment	7–10	Vertebrate	29	4	α.	105
								123	4	5	180
dsRNA		Cystoviridae		Isometric	3 segments	17	Bacteria	-	0	0	-
		Reoviridae		Isometric	10–12 segments	19-62	Vertebrate, invertebrate, plant	8	256	36	357
		Birnaviridae		Isometric	2 segments	9	Vertebrate, invertebrate	4	21	-	56
		Totiviridae		Isometric	1 segment	2-7	Fungus, protozoa	8	0	2	23
		Partitiviridae		Isometric	2 segments	3-10	Fungus, plant	90	0	15	45
		Hypoviridae		Pleomorphic	1 segment	9-13	Fungus	ဗ	0	8	r.
			Varicosavirus	Rod	2 segments	14	Plant	-	0	က	4
14		;						119	277	60	461
negative ssRNA	negative <i>mononegavirales</i> ssRNA	Bomavirdae		Spherical	1 – segment	6	Vertebrate	-	0	-	N
		Filoviridae		Bacilliform	1 - segment	13	Vertebrate	ເດ	19	0	24

Table 3 Continued

								Number	Number of species		
:		:	;		Genome	Genome	,		Strains/		
Criteria	Order	Family	Floating genus	Morphology	configuration	size (kb)	Virus host	Species	serotypes	Tentative	Total
		Paramyxoviridae		Helical	1 - segment	15-16	Vertebrate	31	ιO	8	38
		Rhabdoviridae		Bacilliform	1 - segment	10-13	Vertebrate, plant	37	0	142	179
		Orthomyxoviridae		Helical	8 - segments	13-14	Vertebrate	S	, -	0	9
		Bunyaviridae		Spherical	3 – segments	12-23	Vertebrate, plant	69	236	99	395
			Tenuivirus	Filaments	4 -?segments	s 15–19	Plant	9	0	Ŋ	Ξ
			Ophiovirus	Filaments	3 - segments	12	Plant	က	0	0	က
		Arenaviridae		Spherical	2 - segments	=	Vertebrate	19	27	8	48
			Deltavirus	Spherical	1 circular – strand	1.7	Vertebrate	-	0	0	-
								201	288	218	707
Positive ssRNA		Leviviridae		Isometric	1 + segment	4	Bacteria	4	18	35	22
		Namaviridae		Ribonucleic complex	1 + segment	2.5	Yeast	က	0	0	က
		Picornaviridae		Isometric	1 + segment	7-8.5	Vertebrate	16	105	137	258
			"CrPV-like viruses"	Isometric	1 + segment	9–10	Invertebrate	rS	0	0	Ŋ
		Sequiviridae		Isometric	1 + segment	9-12	Plant	Ŋ	0	0	2
		Comoviridae		Isometric	2 + segments	9-16	Plant	20	0	တ	29
		Potyviridae		Rod	1 or 2 +	8-12	Plant	106	0	92	198
		Caliciviridae		Isometric	1 + segment	00	Vertehrate	Œ	40	α	74
			"HEV-like viruses" Isometric	3" Isometric	1 + segment	7	Vertebrate	-	. 0	0	-
		Astroviridae		Isometric	1 + segment	7-8	Vertebrate	9	5	0	19
		Nodaviridae		Isometric	2 + segments	ro 2	Vertebrate, invertebrate	4	0	0	4
		Tetravirldae		Isometric	1 + segment	2	Invertebrate	တ	0	0	6
			Sobemovirus	Isometric	1 + segment	4	Plant	Ξ	0	က	14
			Marafivirus	Isometric	1 + segment	6-7	Plant	ო	0	0	က
		Luteoviridae		Isometric	1 or 2 + segment	6-9	Plant	œ	0	Ξ	6
			Umbravirus	No particles	1 + segment	4	Plant	7	0	15	22
		Tombusviridae		Isometric	1 or 2 +	4-5.5	Plant	38	0	=	49
					segment				į	1	

Table 3 Continued

								Number	Number of species		
Criteria	Order	Family	Floating genus	Morphology	Genome configuration	Genome size (kb)	Virus host	Species	Strains/ serotypes	Tentative	Total
	Nidovirales	Coronaviridae		Pleomorphic	1 + segment	28-33	Vertebrate	16	r.	-	22
		Arteriviridae		Spherical	1 + segment	13-16	Vertebrate	4	0	0	4
		Flaviviridae		Isometric	1 + segment	10-12	Vertebrate	22	47	9	110
		Togaviridae		Isometric	1 + segment	10-13	Vertebrate	23	9	0	59
SSRNA			Tobamovirus	Rod	1 + segment	9	Plant	16	0	က	6
Positive sense			Tobravirus	Rod	2 + segments	11-6	Plant	ဇ	0	0	က
			Hordeivirus	Rod	3 + segments	01	Plant	4	0	0	4
			Furovirus	Rod	2 + segments	<u>1</u>	Plant	-	0	4	2
			Pomovirus	Rod	3 + segments	12	Plant	4	0	0	4
			Pecluvirus	Rod	2 + segments	10	Plant	Ø	0	0	Ø
			Benyvirus	Rod	4 (or 5) + segments	14-16	Plant	2	0	0	N
		Bromoviridae		Isometric, bacilliform	3 + segments	6-8	Plant	58	0	0	88
			Ourmiavirus	Bacilliform	3 + segments	1	Plant	က	0	0	က
			Idaeovirus		3 + segments	80	Plant	-		0	-
		Closteroviridae		Rod	1 or 2 +	15-19	Plant	18	0	16	श्च
					segments						
			Capillovirus	Rod	1 + segment	7	Plant	ო	0	-	4
			Trichovirus	Rod	1 + segment	7.5	Plant	က	0	-	4
			Vitivirus	Rod	1 + segment	7.5	Plant	4	0	-	ß
			Tymovirus	isometric	1 + segment	9	Plant	20	0	-	21
			Carlavirus	Rod	1 + segment	2-8	Plant	31	0	53	90
			Potexvirus	Rod	1 + segment	9	Plant	56	0	18	4
			Allexivirus	Rod	1 + segment	တ	Plant	9	0	က	6
			Foveavirus	Rod	1 + segment	6-8	Plant	2	0	-	က
		Barnaviridae		Bacilliform	1 + segment	4	Fungus	-	0	-	8
								565	234	403	1202
		Unassigned viruses		1	1	ı	ΑI	30	0	0	30
								1550	101	1393	3954
		Viroids		1	1	ŀ	Plant	27	0	œ	32
		Satellites		1	1	1	Plant	33	0	9	33

Table 4 List of criteria demarcating different virus taxa

I Order

Common properties between several families including:

Biochemical composition

Virus replication strategy

Particle structure (to some extent)

General genome organization

II Family

Common properties between several genera including:

Biochemical composition

Virus replication strategy

Nature of the particle structure

Genome organization

III Genus

Common properties within a genus including:

Virus replication strategy

Genome size, organization and/or number of segments

Sequence homologies (hybridization properties)

Vector transmission

IV Species

Common properties within a species including:

Genome rearrangement

Sequence homologies (hybridization properties)

Serological relationships

Vector transmission

Host range

Pathogenicity

Tissue tropism

Geographical distribution

and occupies a particular ecological niche.' The major advantage in this definition is that it can accommodate the inherent variability of viruses and is not dependent on the existence of a unique characteristic. Members of a polythetic class are defined by more than one property and no single property is absolutely essential and necessary. Thus in each family it might be possible to determine the set of properties of the class 'species' and to check if the family members are species of this family or if they belong to a lower taxonomic level. The ICTV is currently conducting this exercise throughout all virus families. This exercise should ultimately result in an excellent evaluation of a precise definition of each virus species in the entire classification.

Several practical matters are related to the definition of a virus species with the goal of improving the usefulness of virus classification. These include: (1) homogeneity of the different taxa; (2) diagnosis-related matters; (3) virus collections; (4) evolution studies; (5) biotechnology; (6) sequence database

projects; (7) virus database projects; and now (8) intellectual property rights.

Virus families and genera

There is no formal definition for a genus, but it is commonly considered as: 'a population of virus species that share common characteristics and are different from other populations of species'. Although this definition is somewhat elusive, this level of classification seems enduring and useful; some genera have been moved from one family to another over the years, but the composition and description of the genera has remained stable. The characters defining a genus differ from one family to another and there is a tendency to create genera with fewer differences between them. Upon examination, there is more and more evidence that the members of a genus have a common evolutionary origin. The use of subgenera has been abandoned in current virus classification.

Notwithstanding the creation of the ICTV, plant

virologists continued to classify plant viruses in 'groups', refusing to place them in genera and families. However, owing to obvious similarities, plant reoviruses and rhabdoviruses had been integrated into the families Reoviridae and Rhabdoviridae (Table 2). This position was mostly due to plant virologists' refusal to accept binomial nomenclature. Since this form of nomenclature was withdrawn from the ICTV classification rules in 1995, they subsequently accepted the placing of plant viruses into species, genera and families as shown in the sixth ICTV report. However, there are still 30 of so-called 'floating genera' that do not pertain to any family. This is mostly due to the fact that plant virologists prefer to accumulate data on virus species and genera before clustering appropriate genera in families. It is remarkable that this attitude has also been adopted by other virologists as a convenient way of classifying viruses, without having to move genera out of families when it becomes apparent that they are part of a distinct family. For example, the members of the floating genus 'cricket paralysis-like viruses' share enough properties with picornaviruses to be included in the family *Picornaviridae*; however, they also possess properties that would justify their classification in a separate family. Only new data or new viruses will permit a definitive position, therefore for the time being it remains a floating genus. Similarly the same strategy is used to create a floating genus 'ictalurid herpes-like viruses', within the family Herpesviridae, although in this case it is a floating genus within the family because of uncertainty as to whether the members of this genus should be classified in one of the existing subfamilies or to a new subfamily.

Virus orders

As mentioned above, the higher hierarchy levels for virus classification are extremely difficult to establish. Despite several propositions in the past, only three have been accepted: Caudovirales, Mononegavirales and Nidovirales. The first virus order, Mononegavirales, was established in 1990 and comprises the nonsegmented single-stranded RNA negative-sense viruses, namely the families Bornaviridae, Filoviridae, Paramyxoviridae and Rhabdoviridae. This decision was taken because of the great similarity of many criteria between these families, including their replication strategy. A second order, Caudovirales, contains all the families of double-stranded DNA phages possessing a tail, including the families Myoviridae, Podoviridae and Siphoviridae. A third order, Nidovirales, comprising the families Coronaviridae and Arteriviridae, was accepted in 1996 because of the impossibility of grouping together these two taxonomic entities, which share many properties and yet are so different, as a single family. Many members of the ICTV advocate the creation of many more orders, but it has been decided to proceed cautiously to avoid creation of short-lived orders. The creation of formal taxa higher than the orders, for example, kingdoms, classes and subclasses, has not been considered by the ICTV.

Virus Taxa Descriptions

Virus classification continues to evolve with the technologies available for describing viruses. The first wave of descriptions, those before 1940, mostly took into account the visual symptoms of the diseases caused by viruses, along with their modes of transmission. A second wave, between 1940 and 1970, brought together an enormous amount of information from studies of virion morphology (electron microscopy, structural data), biology (serology and virus properties) and physicochemical properties of viruses (nature and size of genome, number and size of viral proteins). Since 1970, the third wave of virus descriptions has included genome and replicative information as well as molecular relationships with virus hosts. There is a correlative modification of the list of virus descriptors and Table 1 lists the family and genera descriptors which are used in the current ICTV report. Figures 1-5 are diagrammatic representations of families and genera of viruses infecting vertebrates, invertebrates, plants, fungi, yeasts, protozoa and bacteria. The most recent wave of information used to classify viruses is naturally nucleotide and amino acid sequences. It is becoming more and more prevalent in virus taxonomy, as exemplified by the presence of a significant number of 'phylogenetic trees' in the seventh ICTV report, and by the huge number of scientific publications on this topic. Some scientists promote the concept of 'quantitative taxonomy' aimed at demonstrating that virus sequences contain all the coding information required for all the biological properties of the viruses. This is in complete agreement with the polythetic concept of the virus species definition, as demonstrated for example by Padidam et al in 1995, van Regenmortel et al in 1997, Hyppia et al in 1998, and Aleman et al in 1999.

The impact of descriptions on virus classification has been particularly influenced by electron microscopy and of the negative staining technique for virions. This technique had an immediate influence on diagnostics and classification of viruses. With negative staining, viruses could be identified from poorly purified preparations of all tissue types, and informa-

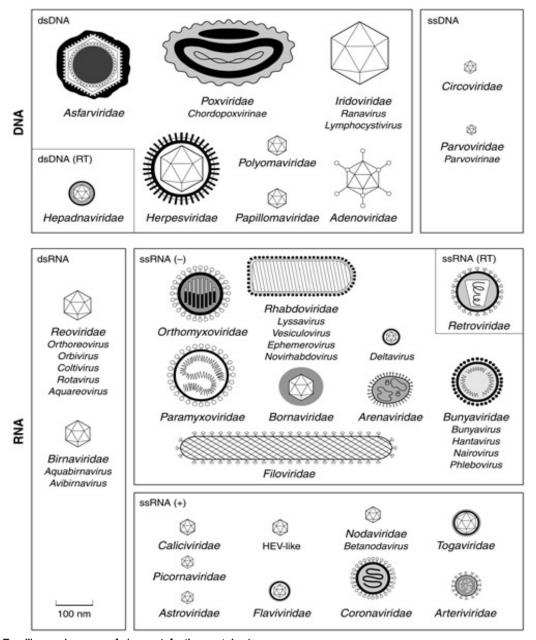


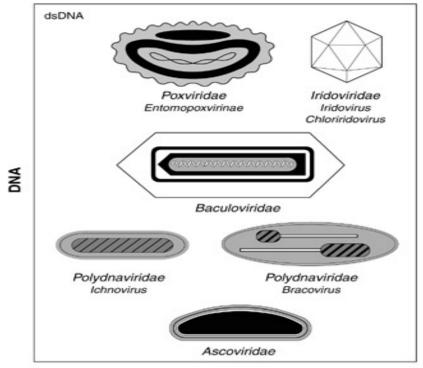
Figure 1 Families and genera of viruses infecting vertebrates.

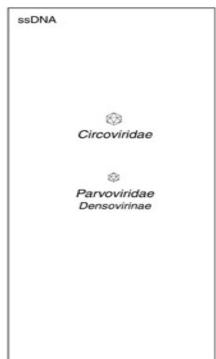
tion about size, shape, structure and symmetry could be quickly provided. As a result, virology progressed simultaneously for all viruses infecting animals, insects, plants and bacteria. Thin sections of infected tissues brought a new dimension to virus classification by providing information about virion morphogenesis and cytopathogenic effects. These techniques, in conjunction with the determination of the nature of the genome, provided a major source of information for the system of virus classification established in the 1980s, as shown by the large number of viruses listed in the fifth ICTV report in 1989.

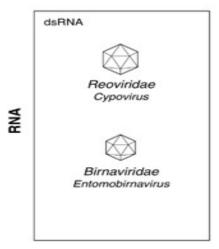
In many instances the properties of viruses belong-

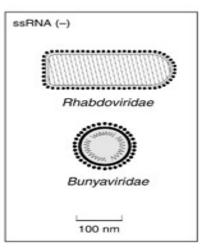
ing to the same genus are correlated. Thus, the classification of a few of them will likely be sufficient to allow the classification of a new virus into an established genus. For example, a plant virus with filamentous particles of 700-850 nm and transmitted by aphids is likely to be a member of the genus Potyvirus. Establishment of new genera in the future will require more information. Most of the properties listed in Table 3 will have to be precisely analyzed to warrant the formation of a new genus.

Table 3 lists 45 different categories of properties but each category includes many items. Lists of virus descriptors usually comprise 1000-2000 descriptors.









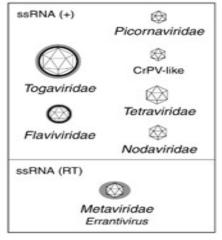


Figure 2 Families and genera of viruses infecting invertebrates.

The establishment of a universal list of virus descriptors is under way and should be adopted by ICTV around 2000 with the establishment of the ICTVdB. It will contain a common set of descriptors for all viruses and subsets for specific viruses in relation to their specific hosts (human, animal, insect, plant and bacterial).

A Uniform Nomenclature of Viral Taxa

When a genus is approved by ICTV, a type species is designated. However, none of these type species have received a new international name and only English

names are used. Latinized binomial names for virus names have been supported by animal and human virologists of ICTV for many years, but have never been implemented. This suggestion was in fact withdrawn from ICTV nomenclature rules in 1990 and consequently such names as Herpesvirus varicella or Polyomavirus hominis should not be used. For several years, plant virologists have adopted a different nomenclature, using the vernacular name of a virus but replacing the word 'virus' by the genus name; for example, Cucumber mosaic cucumovirus and Tobacco mosaic tobamovirus. Though this usage is favored by many scientists, and examples of such practice can

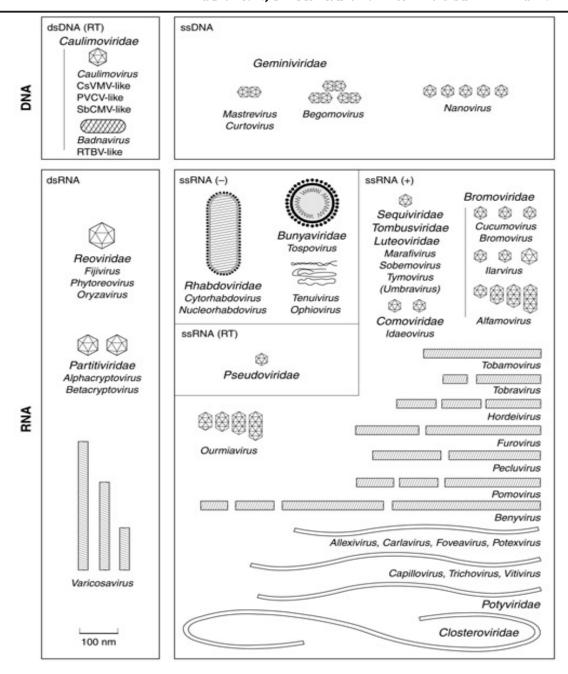


Figure 3 Families and genera of viruses infecting plants.

be found for human, animal and insect viruses (e.g. Human rhinovirus, Canine calicivirus, Acheta densovirus...), it has not been universally adopted by the ICTV.

The ICTV has set rules for virus nomenclature and orthography of taxonomic names that are regularly revisited and improved. The last word of international virus species names is 'virus', the international genus names universally end in '...virus', the international subfamily names end in '...virinae', the international family names end with '...viridae', and

the international order names are ending in '...virales'. In formal taxonomic usage, the virus order, family, subfamily, genus and species names are all printed in italics (or underlined) and the first letter is capitalized. For all taxa except the species names, new names are created de novo following ICTV guidelines, but in the case of virus names English vernacular form is used. In formal usage, the name of the taxon precedes the name of the taxonomic unit; for example, 'the family Picornaviridae' or 'the genus Rhinovirus'. In informal vernacular usage, order,

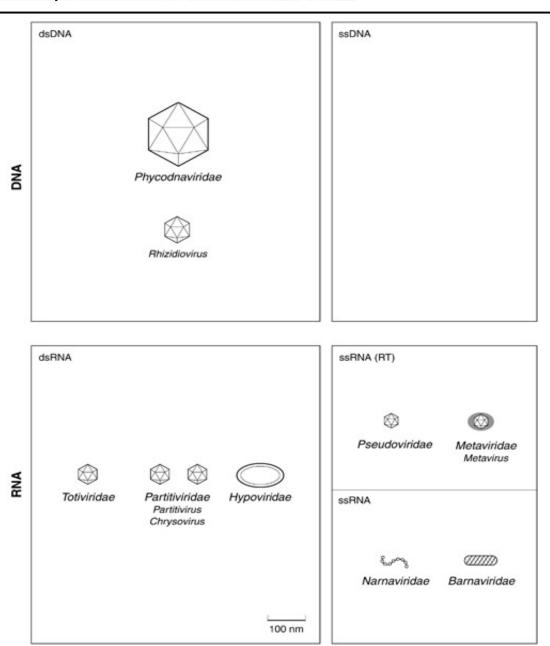


Figure 4 Families of viruses infecting algae, fungi, yeasts and protozoa.

family, subfamily, genus and species names are written in lower case Roman script; they are not capitalized nor italicized (or underlined). Additionally, in informal usage, the name of the taxon should not include the formal suffix, and it should follow the term for the taxonomic unit; for example, 'the mononegavirales order', 'the adenovirus family', 'the avihepadnavirus genus' or 'the tobacco mosaic virus' species. Virus names are often abbreviated for convenient reasons, but ICTV has not set up guidelines to generate such abbreviations. The ICTV reports list abbreviations most commonly used by

specialists and the ICTV reports help virologists to identify duplicates of abbreviations in order to decrease the number of such duplicates. In 1988 plant virologists initiated the publication of such lists and have indicated guidelines for the creation of new virus names and new abbreviations. These guidelines were last published in 1991 by Fauquet and Martelli and will be updated again in 1999.

To avoid ambiguous virus identifications, it has been recommended to journal editors that published papers follow ICTV guidelines for proper virus identification and nomenclature, and that viruses should be cited

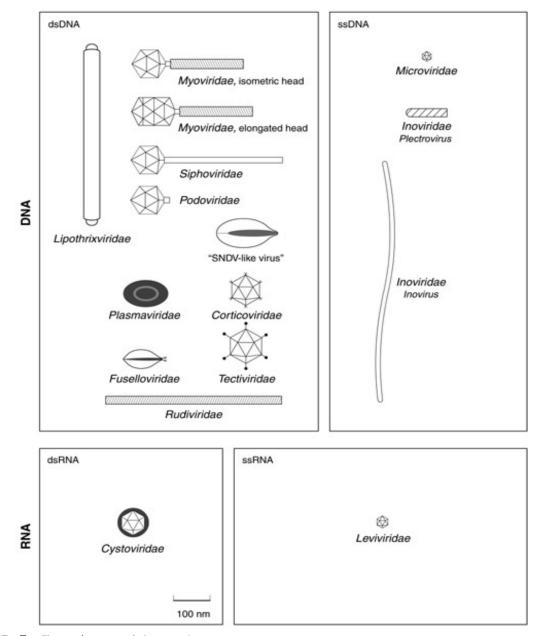


Figure 5 Families and genera of viruses infecting bacteria.

with their full taxonomic terminology when they are first mentioned in an article. For example:

- Order Caudovirales, family Podoviridae, genus 'T7-like viruses', species Enterobacteria phage T7.
- Order Mononegavirales, family Paramyxoviridae, subfamily Paramyxovirinae, genus Rubulavirus, species Mumps virus.
- Order Nidovirales, family Coronaviridae, genus Coronavirus, species Avian infectious bronchitis virus.

- Family Iridoviridae, genus Iridovirus, species Chilo iridescent virus.
- Family Picornaviridae, genus Enterovirus, species Poliovirus, serotype Human poliovirus 1.
- Genus Tobamovirus, species Tobacco mosaic virus.

See also: Phage taxonomy and classification; Virus structure: Atomic structure, Principles of virus structure.

Further Reading

Fauquet CM and Martelli GP (1995) Up-dated ICTV list of names and abbreviations of viruses, viroids and satellites infecting plants. *Arch. Virol.* 140: 393.

Francki RIB, Milne RG and Hatta T (1985) Atlas of Plant Viruses. Boca Raton, CRC Press.

Francki RIB, Fauquet CM, Knudson DL and Brown F (1991) Classification and Nomenclature of Viruses. Fifth Report of the International Committee on Taxonomy of Viruses. Vienna: Springer.

Lwoff A, Horne R and Tournier P (1962) A system of viruses. Cold Spring Harb. Symp. Quant. Biol.

Matthews REF (1983) The History of Viral Taxonomy. A

Critical Appraisal of Viral Taxonomy, pp 1-35. Boca Raton: CRC Press.

Murphy FA, Fauquet CM, Bishop DHL et al (eds) (1995) Virus Taxonomy. Sixth Report of the International Committee on Taxonomy of Viruses. Vienna: Springer

van Regenmortel MHV, Bishop DHL, Fauquet CM et a. (1997) Guidelines to the demarcation of virus species Arch. Virol. 142: 1505.

van Regenmortel MHV, Fauquet CM, Bishop DHL et a. (eds) (1999) Virus Taxonomy. Seventh Report of the International Committee on Taxonomy of Viruses. New York: Academic Press.

TENUIVIRUSES



Bryce W. Falk, Department of Plant Pathology, University of California, Davis, California, USA Copyright © 1999 Academic Press

History

The viruses in the genus Tenuivirus (tenuiviruses) have been recognized as important plant pathogens since the early 1900s. However, in 1977 it was first recognized that rice stripe virus (RSV)-infected rice (Oryza sativa L.) plants contained unusual finestranded particles. These appeared as circular or branched filaments and were distinctly different from particles associated with other plant viruses. Subsequent work showed that plants infected by several other viruses contained similar particles. This and the somewhat unique biological properties shared by these viruses led to the early grouping together of RSV and other plant viruses, including maize stripe virus (MSpV) and rice hoja blanca virus (RHBV). Since 1995, RSV and related viruses have been grouped together by the International Committee on Taxonomy of Viruses (ICTV) as members of the genus Tenuivirus (tenuiviruses).

Taxonomy and Classification

There are currently six species within the genus *Tenuivirus*, and at least five tentative species (**Table** 1). The definitive tenuiviruses are: rice stripe virus (RSV; the type species of the genus); MSpV; RHBV; rice grassy stunt virus (RGSV); *Echinochloa* hoja blanca virus (EHBV) and *Urochloa* hoja blanca virus (UHBV). All tenuiviruses exhibit similar properties, and in recent years molecular biological analyses have shown that they are quite distinct from other

currently recognized plant viruses. The genus *Tenuivirus* is not placed within a formally recognized virus family; however, tenuiviruses appear in some ways to be more closely related to viruses in the genus *Phlebovirus* of the family *Bunyaviridae*, than they are to other plant viruses.

Biology, Host Range and Vector Transmission

The plant host ranges of all tenuiviruses are limited to monocotyledenous plant species within the family Poaceae, including many plants which are important food crops (i.e. rice (Oryza sativa L.) and maize (Zea mays L.)). The symptoms induced in infected plants are generally similar for the different tenuiviruses and includes general leaf striping, a distinct white coloring of the leaf stripes and stunting (Fig. 1).

Tenuiviruses generally are not mechanically transmissible to their plant hosts; however, mechanical transmission has been reported in limited instances but only under specific conditions. All tenuiviruses are transmitted to plants by specific planthoppers (Homoptera: Delphacidae, see Table 1 and Fig. 2). Transmission of a given tenuivirus is specific and may be limited to planthoppers of a single species. Compared to other vectors of plant viruses (i.e. aphids or whiteflies), delphacid planthoppers are not generally thought of as common vectors; however, they are perfectly adapted to be vectors of tenuiviruses. Delphacid planthoppers which vector tenuiviruses