

MATERIALS SCIENCE

Formation of 2D and 3D multi-tori mesostructures via crystallization-driven self-assembly

Gerald Guerin^{1,2*}, Menandro Cruz^{2,3}, Qing Yu²

The fabrication of three-dimensional (3D) objects by polymer self-assembly in solution is extremely challenging. Here, multi-tori mesostructures were obtained from the crystallization-driven self-assembly of a coil-crystalline block copolymer (BCP) in mixed solvents. The formation of these structures follows a multistep process. First, the BCP self-assembles into amorphous micrometer-large vesicles. Then, the BCP confined in these mesosized vesicles crystallizes. This second step leads to the formation of objects with shapes ranging from closed 3D multi-tori spherical shells to 2D toroid mesh monolayers, depending on the solvent mixture composition. This approach demonstrates how topological constraints induced by the specific interactions between coil-crystalline BCP and solvents can be used to prepare mesostructures of complex morphologies.

INTRODUCTION

In recent years, the study of the crystallization-driven self-assembly (CDSA) of coil-crystalline block copolymers (BCPs) in solution has become an important branch of polymer self-assembly. The living-like properties of CDSA demonstrated by Wang *et al.* (1) offer seemingly endless possibilities to generate structures of controlled size and morphology, opening the doors to the potential development of a multitude of devices (2–5). The main morphology reported was one-dimensional (1D), i.e., elongated homomicelles (6–10), as well as multi-block (11–13), patchy (14–17), and asymmetric comicelles (18, 19). As the manipulation of 1D core-crystalline micelles became more common, sophisticated objects such as 2D (20–22), hybrid (23, 24), and mesostructures (25, 26) started to be presented.

The fabrication of isolated 3D mesoscale polymeric objects via bottom-up self-assembly, however, remains a daunting task. There are no compelling strategies that lead to the formation of 3D functional materials. The very few attempts to prepare these structures used preformed core-crystalline micelles as building blocks (27, 28). This approach, however, failed to produce 3D mesostructures that persist in the solid state.

In the present work, we describe the successful preparation of 3D mesostructures as large as pollen grains (>10 μm). This unique type of structure was obtained by the CDSA of coil-crystalline BCPs in a mixture of solvents that are poor for both the coil and crystalline blocks. More specifically, we studied the self-assembly of polyferrocenyldimethylsilane-*block*-polystyrene (PFS₂₆-*b*-PS₃₀₆, with $M^{\text{PS}} \approx 30,000$) in an acetone/decane mixture containing 25 volume % of acetone, a composition where PFS₂₆-*b*-PS₃₀₆ microphase separation is anticipated (29).

We show that the presence of PFS₅₅-*b*-PI₅₀₀ (PI = polyisoprene) crystallites at an early stage of the self-assembly is essential for forming these micrometer large 3D multi-tori structures (Fig. 1 and fig. S1). Only perforated films could be observed when no PFS-*b*-PI seed micelles were present in the solution. We also show that the

addition of an aliquot of a third solvent into the acetone/decane mixture strongly affects the morphology of final structure, either by greatly enhancing the sphericity of the multi-tori mesostructures or by inducing the formation of toroid mesh monolayers. Above all, the 3D multi-tori mesostructures have proven to be very stable at room temperature once transferred onto a substrate, preserving their morphology indefinitely, a prerequisite for the fabrication of 3D functional materials.

RESULTS

Preparation of multi-tori by self-assembly

The samples were prepared by quickly injecting a solution of PFS₂₆-*b*-PS₃₀₆ in tetrahydrofuran (THF; 8 μl , $c = 10 \text{ mg ml}^{-1}$) to 1 ml of a 25/75 (v/v%) acetone/decane mixture preheated at 35°C, and to which 20 μl of PFS₅₅-*b*-PI₅₀₀ rod-like micelles in decane ($c = 0.2 \text{ mg ml}^{-1}$) had been added (Fig. 1A). The solution was annealed for 1 hour and then cooled to room temperature. Within 1 hour after the heating was stopped, an aliquot of the solution was deposited on a transmission electron microscopy (TEM) grid at room temperature. The sample was finally imaged by scanning electron microscopy (SEM) and high-resolution TEM (HRTEM).

The protocol described above leads to the formation of a mixture of isolated toroids and open spherical multi-tori shells of ca. 20 μm diameter with a broad size distribution (Fig. 1, B to E). These shells look like broken or collapsed hollow spheres, consisting of toroids glued together. The toroids that compose the multi-tori appear to be relatively narrowly distributed in size (Fig. 1E), with an inner diameter close to 400 nm and an outer diameter of ca. 1 μm (because very few toroids lay flat on the TEM grid, the dimensions indicated are only semiquantitative).

It is worth noting that the multi-tori structures are unstable and collapse in on themselves when annealed at room temperature for a few extra hours in solution (fig. S2), but they preserve their morphology indefinitely once taken out of the solution (by depositing them on a TEM grid, for example). This allowed us to reimage the same TEM grid by SEM and HRTEM.

Intuitively, one would believe that the genesis of these massive objects is hierarchical and genuinely straightforward: First, PFS₂₆-*b*-PS₃₀₆ toroids form by CDSA, and then, they are glued together by the remaining PFS₂₆-*b*-PS₃₀₆ unimer in a sequence similar to that

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¹Shanghai Key Laboratory of Advanced Polymeric Materials, Key Laboratory for Ultrafine Materials of Ministry of Education, School of Materials Science and Engineering, East China University of Science and Technology, Shanghai 200237, China. ²Department of Chemistry, University of Toronto, 80 St. George Street, Toronto, ON M5S 3H6, Canada. ³Melville Laboratory for Polymer Synthesis, Department of Chemistry, University of Cambridge, Lensfield Road, Cambridge, UK. *Corresponding author. Email: gguerin@ecust.edu.cn

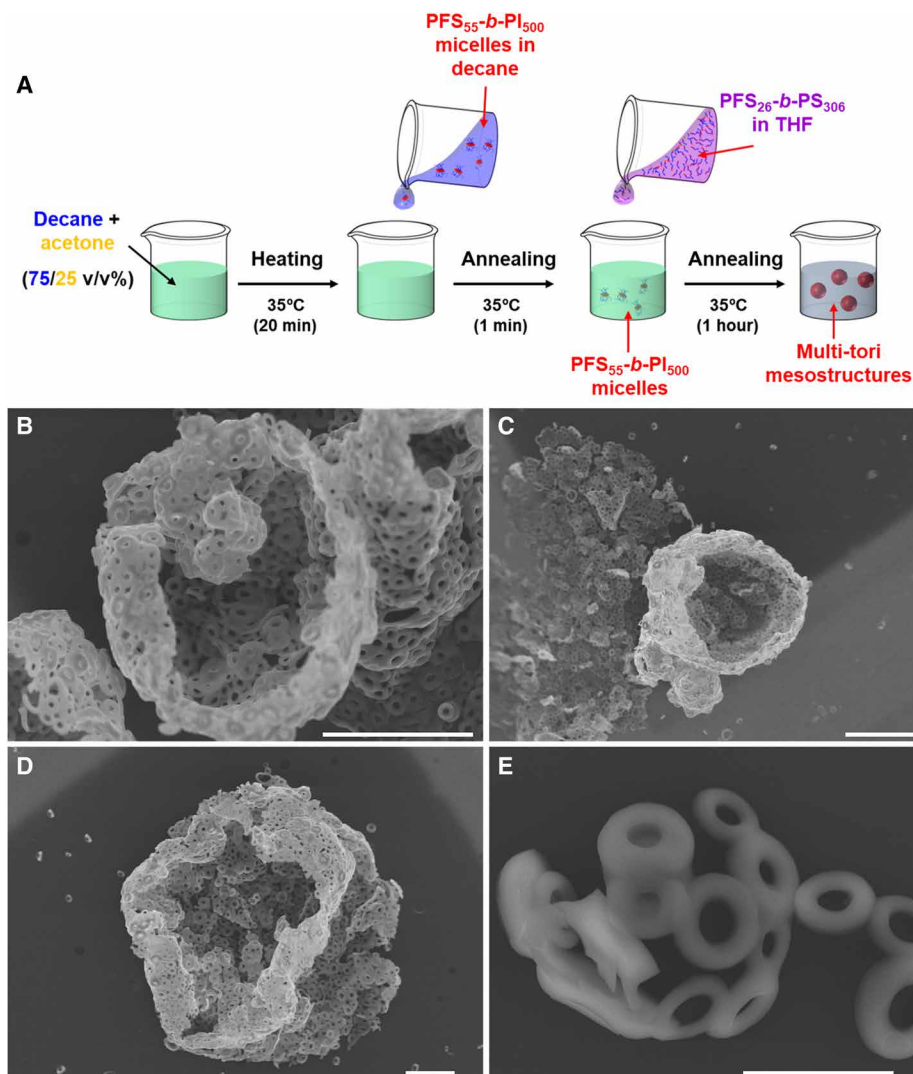


Fig. 1. Examples of 3D multi-tori mesostructures. (A) Protocol followed to prepare the multi-tori mesostructures. (B to E) Representative SEM micrographs of spherical hollow mesostructures consisting of a network of toroids. The structures were obtained by the self-assembly of PFS₂₆-b-PS₃₀₆ in a mixture of 1.02 ml of acetone/decane (25/75, v/v%), containing 20 μ l of PFS₅₅-b-PI₅₀₀ core-crystalline elongated micelles ($c = 0.2 \text{ mg ml}^{-1}$), and preheated at 35°C. An aliquot of PFS₂₆-b-PS₃₀₆ dissolved in THF (8 μ l, $c = 10 \text{ mg ml}^{-1}$) was then injected, and the solution was annealed for 1 hour at 35°C and cooled to room temperature; TEM grids were prepared within 1 hour after heating. Scale bars, 10 μ m (B to D) and 2 μ m (E).

described by Cai *et al.* (30). These authors prepared sponge-like nanoporous superstructures using PFS₄₈-b-[PMVS(OH)₂]₃₂₆ BCP to glue already prepared PI₁₀₄₀-b-P2VP₂₀₀ toroids.

We, thus, studied the sample by SEM at very short annealing times, i.e., 30 s and 5 min (Fig. 2 and fig. S3), hoping to capture images of isolated toroids, the expected building block and first step toward the formation of the 3D multi-tori structures. When we looked at the sample annealed for only 30 s (Fig. 2, A to C, and fig. S3, A to C), we were astonished to observe several large broken spherical shells of the size of the massive multi-tori structures and with a broad size distribution ranging from ca. 5 μ m to more than 20 μ m. The larger structures had a bumpy surface and appeared to have broken and flattened onto the TEM grid (Fig. 2, A and B), while the smaller structures were more raspberry-like, with bulges, or drupelets (to use a morphological analogy to raspberries), resembling the kippah micelles described by Azzam and Eisenberg (31)

(Fig. 2C). In some cases, a hole located on the top of the drupelets is visible (Fig. 2C). After 5 min of annealing (Fig. 2, D to F, and fig. S3, D to F), the flattened structures are still observable, but they mainly consist of an assembly of a very large number of drupelets. The holes in the drupelets also appear to become wider as pointed out in Fig. 2F.

To further understand how these large vesicular mesostructures transformed into multi-tori, we performed a kinetic experiment and followed the evolution of these structures by TEM (Fig. 3). We prepared six vials containing a mixture of acetone/decane (25/75, v/v%) and the same amount of PFS₅₅-b-PI₅₀₀ rod-like micelles. To each vial preheated at 35°C, we added a solution of PFS₂₆-b-PS₃₀₆ in THF and annealed the solutions for different times. We took aliquots of the samples at 0.5, 5, 10, 20, 30, and 60 min and deposited them on TEM grids. For this experiment, we were particularly interested in studying the morphological evolution of the drupelets constituting

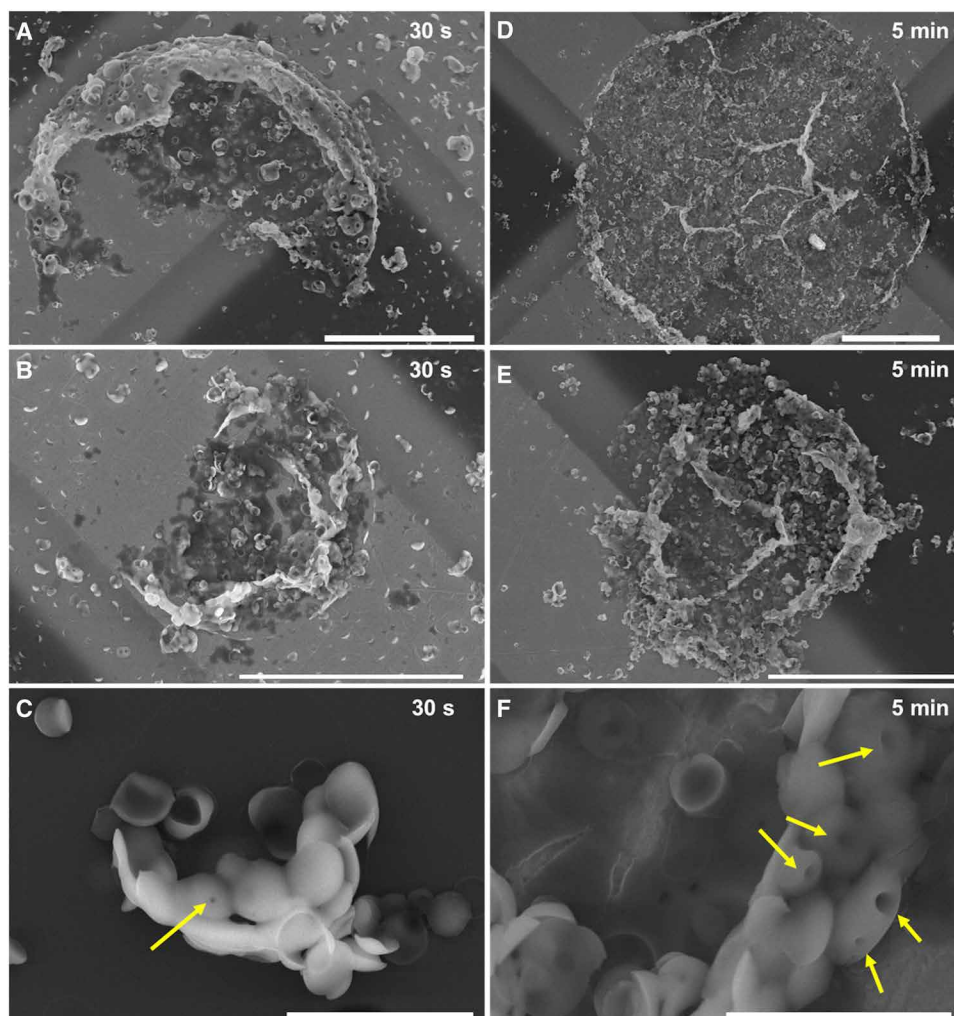


Fig. 2. SEM images of the 3D multi-tori mesostructures at short annealing times. Twenty microliters of a decane solution of PFS₅₅-*b*-PI₅₀₀ seeds ($c = 0.2 \text{ mg ml}^{-1}$) was added to 1 ml of acetone/decane mixture. The solution was then left to equilibrate at 35°C, and an aliquot of PFS₂₆-*b*-PS₃₀₆ dissolved in THF ($8 \mu\text{l}$, $c = 10 \text{ mg ml}^{-1}$) was injected. The solution was then annealed at 35°C. Samples were taken out from the solution and deposited on a TEM grid 30 s (**A to C**) and 5 min (**D to F**) after injection of PFS₂₆-*b*-PS₃₀₆ unimer. The yellow arrows in (C) and (F) point at holes forming in the drupelets. Scale bars, 20 μm (A, B, D, and E) and 2 μm (C and F).

the mesostructures. HRTEM image of the sample annealed for 30 s (Fig. 3A and fig. S4, A and B) shows that, at this very short annealing time, the structures are made of what appears to be supramolecular assemblies of small spherical micelles.

After 5 min of annealing (Fig. 3B and fig. S4, C and D), the structures are still composed of small spherical micelles, but one can find, very occasionally, some curved elongated micelles as indicated by the arrows in Fig. 3B. When the sample was annealed for 10 min (Fig. 3, C and D, and fig. S4, E and F), the number as well as the length of the elongated micelles increased. These 1D micelles appear to be concentric and are present in every structure, along with a large amount of spherical micelles. At this stage, however, the concentric elongated micelles do not look like toroids. After 20 min of annealing (Fig. 3E and fig. S5, A and B), the vast majority of the spherical micelles have disappeared and have been replaced by concentric elongated micelles. Last, the toroids become clearly observable after 30 min of annealing (Fig. 3F and fig. S5, C and D). No obvious variation of the structures could be seen between 30 and 60 min of annealing (Fig. 3, F to H, and fig. S5, E and F).

Elongated micelles are typical of core-crystalline micelles obtained by CDSA of PFS BCPs. They were first reported by Massey *et al.* (32), who also demonstrated the role of PFS crystallization in the formation of these rod-like structures (33). Since then, a plethora of studies have confirmed the crucial role played by the crystallization of the PFS block in the formation of these 1D structures (1, 34–37). We thus expected the concentric elongated micelles that assembled into toroids to have a crystalline core. To test this hypothesis, we studied a sample by x-ray diffraction (XRD). This experiment proved to be very challenging because we needed to prepare ca. 10 mg of product, while our usual experiments were performed with less than 0.1 mg. We first confirmed by SEM (fig. S6A) that the sample contained toroids, and we studied it by XRD. The XRD pattern is shown in fig. S6B. In addition to the very broad peaks centered at $2\theta \approx 10^\circ$ and $2\theta \approx 19^\circ$ due to the amorphous halo of PS, we found a slightly broad peak centered at $2\theta \approx 13.7^\circ$, corresponding to a d -spacing of 6.45 Å, typical of PFS core-crystalline micelles (34–37).

To ascertain the role of the PFS₅₅-*b*-PI₅₀₀ seeds in the formation of the multi-tori structures, we injected a solution of PFS₂₆-*b*-PS₃₀₆

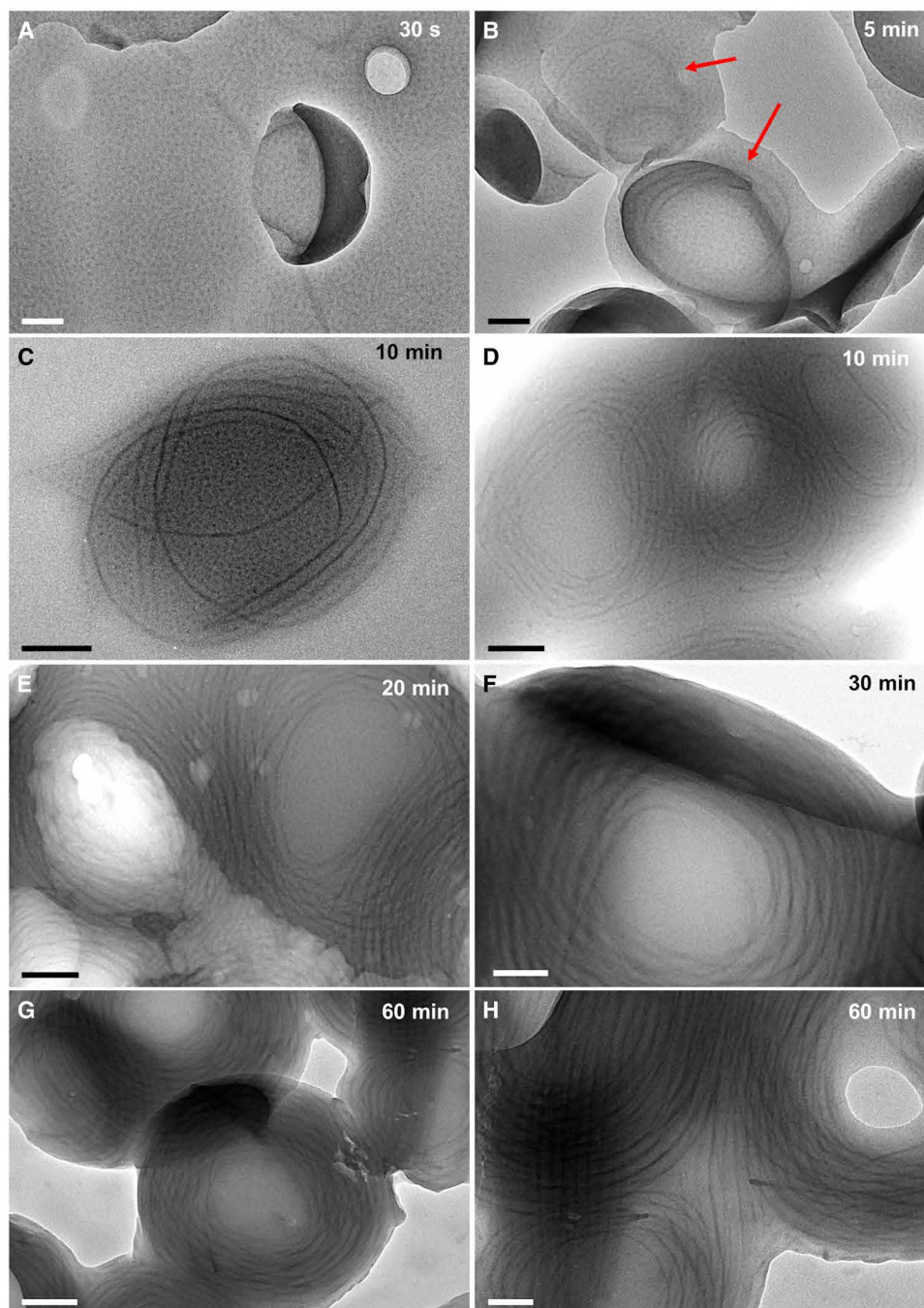


Fig. 3. Kinetic study of the formation of the toroids constituting the multi-tori mesostructures. Six vials were prepared. In each vial, 20 μl of a decane solution of PFS₅₅-*b*-PI₅₀₀ seeds ($c = 0.2 \text{ mg ml}^{-1}$) was added to 1 ml of acetone/decane mixture. The solution was then left to equilibrate at 35°C, and an aliquot of PFS₂₆-*b*-PS₃₀₆ dissolved in THF (8 μl , $c = 10 \text{ mg ml}^{-1}$) was injected. The solution was then annealed at 35°C. Samples were taken out from the solution and deposited on a TEM grid 30 s (A), 5 min (B), 10 min (C and D), 20 min (E), 30 min (F), and 1 hour (G and H) after injection of PFS₂₆-*b*-PS₃₀₆ unimer. The red arrows in (B) point at the elongated micelles that started to appear in the drupelets after 5 min of annealing. Scale bars, 100 nm (A, B, F, and H) and 200 nm (C to E and G).

unimer in a decane/acetone mixture preheated at 35°C that did not contain any PFS₅₅-*b*-PI₅₀₀ seeds. In the absence of PFS₅₅-*b*-PI₅₀₀ micelles, we could not observe any multi-tori objects on the TEM grid, but only ill-defined perforated films (fig. S7A). A closer look at these structures by HRTEM (fig. S7, B to F) showed that they contain the spherical-like micelles that we observed at the early stage of the kinetics experiment (Fig. 3, A and B) along with a negligible number

of elongated micelles (fig. S7F). This result confirmed that the crystallization of the PFS block is the driving force for the formation of the elongated micelles observed inside the toroids.

In a second control experiment, we postponed the addition of the PFS₅₅-*b*-PI₅₀₀ seeds: We first injected the solution of PFS₂₆-*b*-PS₃₀₆ unimer in the acetone/decane mixture, and we annealed the solution at 35°C for 1 hour before to add the PFS₅₅-*b*-PI₅₀₀ seed solution. Just

after the addition of the seeds, we let the solution cool to room temperature, and after one extra hour of aging at room temperature, we studied the structures by SEM (fig. S8A) and HRTEM (fig. S8, B to F). Here again, we could only observe thick and smooth films that contained a large amount of spherical objects similar to those present at the early stage of the multi-tori formation (Fig. 3, A to C, and fig. S3) along with a relatively small number of straight elongated micelles. The occurrence of the elongated micelles was, however, more frequent than in the absence of PFS₅₅-*b*-PI₅₀₀ seeds.

From these two control experiments, we infer that the PFS₂₆-*b*-PS₃₀₆ vesicular mesostructure is a transient morphology. It gradually evolves with time to decrease the surface area between PFS₂₆-*b*-PS₃₀₆ and the poor solvent mixture, leading to the formation of a thick film. The film is then too thick to confine micelle growth, and the usual rectilinear elongated micelles seeded by the late addition of the PFS₅₅-*b*-PI₅₀₀ seeds are observed.

Additional information can also be obtained from SEM images of the edges of broken mesostructures (Fig. 4, A and B). When the structures fracture (either in solution or when deposited on the

TEM grid), the crack appears to mostly propagate through the toroids and not between them. A high-magnification SEM image (Fig. 4B and fig. S9) of a fractured toroid shows the presence of the curved micelles in the toroid as well as what appears to be some micelles that are still bridging the two sections of the broken toroid. This result indicates that the connection between toroids is at least as strong as the toroids themselves. Another surprising finding is presented in Fig. 4D, where one clearly observes that the toroids are highly asymmetric, with a thickness of ca. 300 nm. They are flat on one side and inflated on the other, seemingly preserving the overall shape and orientation of the drupelets, as depicted in Fig. 4 (E and F).

The results presented in Figs. 2 to 4 disprove our intuitive rationale and show that the formation of these 3D mesostructures follows a unique pathway (Fig. 4, E and F). The first step consists of the supramolecular self-assembly of PFS₂₆-*b*-PS₃₀₆ micelles with an amorphous core. The structures formed are micrometer large raspberry-like vesicles (Fig. 4E) with a broad size distribution. In the second step, the elongated core-crystalline PFS₅₅-*b*-PI₅₀₀ micelles initiate

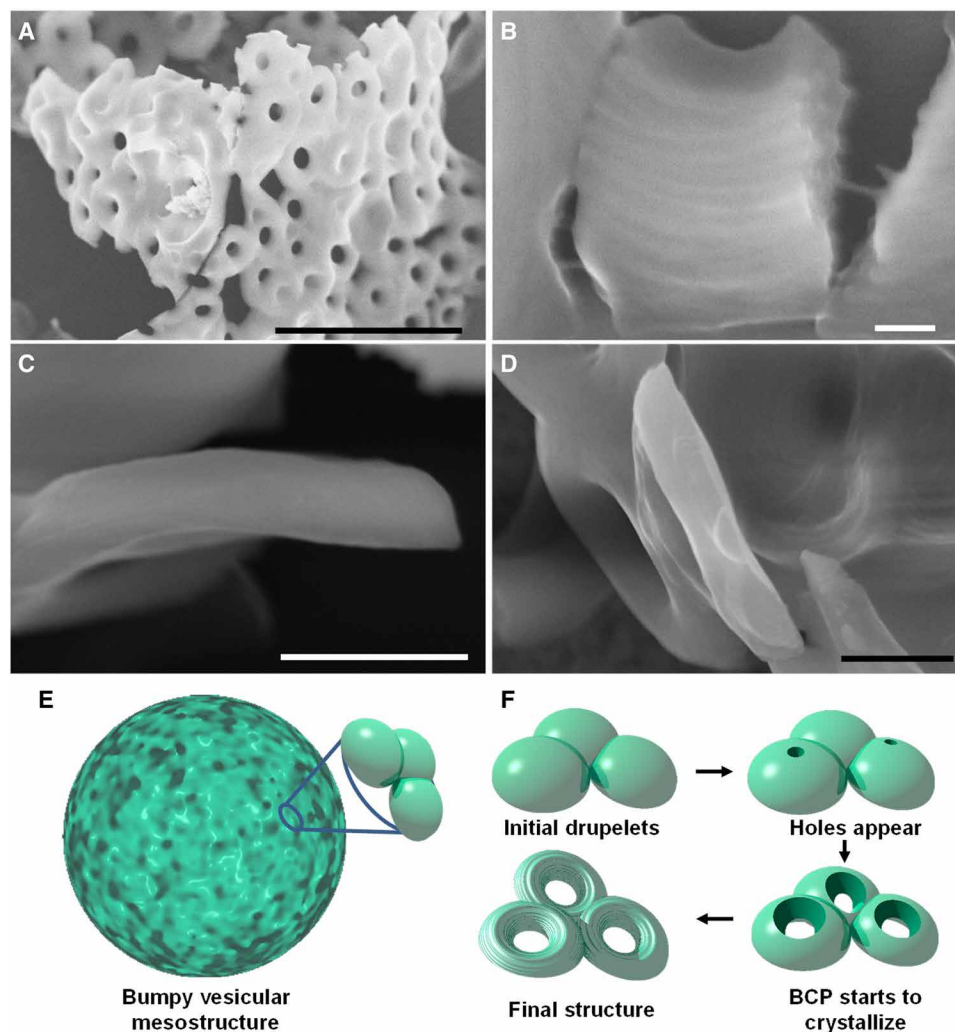


Fig. 4. Understanding the formation of the 3D multi-tori mesostructures. SEM micrographs of (A) the edge of a broken 3D multi-tori mesostructured, (B) a broken toroid, and (C and D) the side view of toroids of 3D multi-tori mesostructures. (E) Schematic representation of a micrometer large bumpy shell. (F) Schematic representation of the formation of the toroids. Scale bars, 5 μ m (A and C), 100 nm (B), and 500 nm (D).

the crystallization of PFS₂₆-*b*-PS₃₀₆ located in the drupelets (Fig. 4F), resulting in a sphere-to-cylinder morphology transition.

The sphere-to-cylinder transition of PFS micelles in polar solvents has been reported on several occasions (37–40), but it was unexpected in our system. In the present case, the morphology transition takes place in a confined environment, and we believe that the formation of the toroids is dictated by the raspberry-like topography of the thin vesicular mesostructures: As the PFS₂₆-*b*-PS₃₀₆ elongated micelles extend, they follow the path with the smallest radius of curvature, growing parallel to the drupelet equatorial plane. As a consequence, the PFS₂₆-*b*-PS₃₀₆ micelles grow in concentric rings and form toroids, as observed in Fig. 3 (E and F) and fig. S5 and as schematically represented in Fig. 4F.

The formation of these multi-tori shells is extremely sensitive to the ratio of acetone/decane used, as shown in figs. S10 and S11. These structures were only present for mixture compositions varying from 24 to 26 volume % of acetone (fig. S10). From 20 to 23 volume % (fig. S11, A to C), a mixture of toroids and 10- μ m-long fibers (bundles of micelles) could be found on the TEM grids, while perforated films made of bundles of elongated micelles (fig. S11, D to F) were observed in the range of 27 to 30 volume % of acetone.

Cowie and McEwen (29) thoroughly studied the cosolvency effect of acetone/*n*-alkane binary mixtures on PS and showed that PS with a molecular weight (M_w) of 20 400 g mol⁻¹ (close to that of the PS block used in this study) dissolves in acetone/decane mixtures ranging from ca. 60 to 95 volume % of acetone near room temperature. The unique self-assembly behavior presented here, however, occurs in a narrow range of solvent mixtures that is outside of the cosolvency window for the PS block (29). It is thus still unclear whether the self-assembly is directly related to the demixing of

PFS₂₆-*b*-PS₃₀₆ in the specific solvent mixture or whether the presence of the PFS block affects the range of the cosolvency window.

While the fractured 3D mesostructures shown in Fig. 1 are clearly hollow, they are not as spherical as represented in Fig. 4E, a consequence, we believe, of the rapid crystallization of PFS₂₆-*b*-PS₃₀₆ in the drupelets, which would disrupt the roundness of the massive objects in solution. To slow down the crystallization and improve the sphericity of the mesostructures, we added 20 μ l of methyl ethyl ketone (MEK), a better solvent for the PFS block, to the acetone/decane mixture, keeping the amount of acetone and decane unchanged. We then followed the same procedure as described previously, adding PFS₅₅-*b*-PI₅₀₀ micelles and then PFS₂₆-*b*-PS₃₀₆ at 35°C.

As shown in Fig. 5, the spherical structures formed in the presence of MEK were much better defined than those prepared in acetone/decane mixtures alone. For example, some closed or nearly closed structures could be observed on the TEM grid (Fig. 5, A and B, and fig. S12), which was not the case in acetone/decane mixtures. There is also little doubt that the opened structures shown in Fig. 5 (C and D) were 3D multi-tori mesospheres that subsequently fractured (fig. S13). The structures remained, however, broadly disperse in size, with diameters ranging from ca. 15 to 40 μ m.

There have been several reports on the preparation of curved crystals by polymer crystallization at liquid/liquid interfaces. The most noticeable studies were reported by Li and co-workers (41–44), who prepared crystalsomes by crystallizing either polymers or BCPs at the interface of an oil-in-water miniemulsion. In the present case, such a mechanism would be relatively surprising because it requires a liquid/liquid phase separation in a solvent mixture (acetone/decane) that is normally miscible over all compositions and over a wide range of temperatures (29).

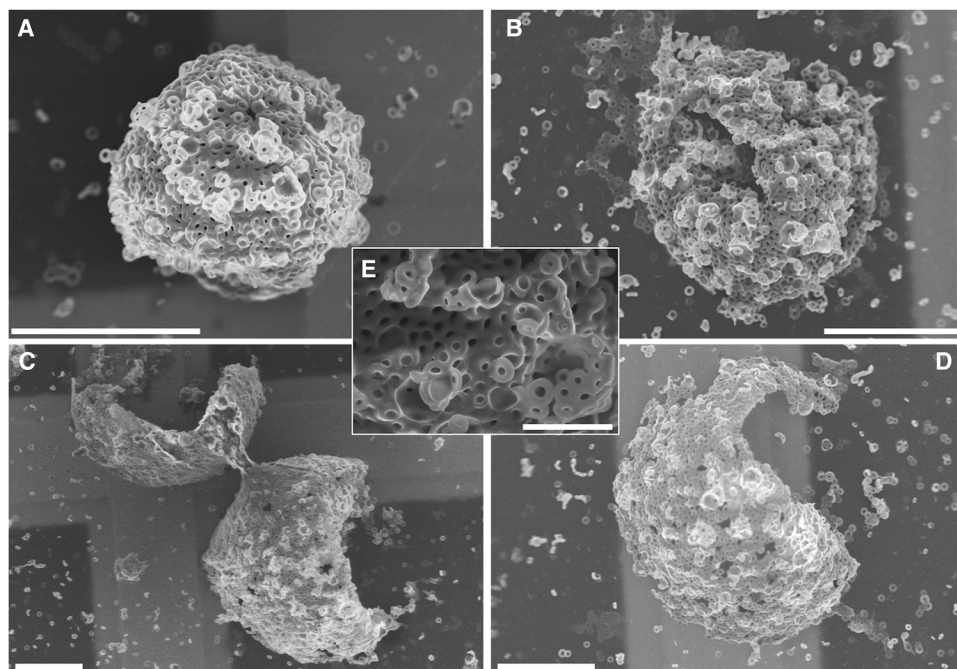


Fig. 5. Effect of an aliquot of MEK on the morphology of 3D multi-tori mesostructures. (A to E) Representative SEM micrographs of spherical hollow mesostructures consisting of a network of toroids. The mesostructures were prepared by adding 20 μ l of MEK to a mixture of 1.02 ml of acetone/decane (25/75, v/v%), containing 20 μ l of PFS₅₅-*b*-PI₅₀₀ core-crystalline elongated micelles ($c = 0.2$ mg ml⁻¹). The solution was preheated at 35°C, and an aliquot of PFS₂₆-*b*-PS₃₀₆ dissolved in THF (8 μ l, $c = 10$ mg ml⁻¹) was injected. The solution was then annealed for 1 hour at 35°C and cooled to room temperature. TEM grids were prepared within 1 hour after heating. Scale bars, 20 μ m (A to D) and 5 μ m (E).

To test whether the presence of PFS₂₆-*b*-PS₃₀₆ unimer could favor the formation of an emulsion-like suspension, we replaced MEK by a solvent that is miscible with acetone, but strongly immiscible with decane, to facilitate a possible microphase separation. Three solvents were tested: *N,N*-dimethylacetamide (DMAc) and *N,N'*-dimethylformamide (DMF), which are good solvents for PS,

and acetonitrile (ACN), a poor solvent for PS. At first, we used DMAc, a good solvent for PS. Figure 6 (A to C) and fig. S14 show the structures obtained in the presence of 10 μ l of DMAc. As anticipated, instead of the 3D mesostructures presented in Figs. 1 to 5, one observes very large objects of ill-defined shapes, with the long section ranging from 50 to 100 μ m (Fig. 6A). Although the structures

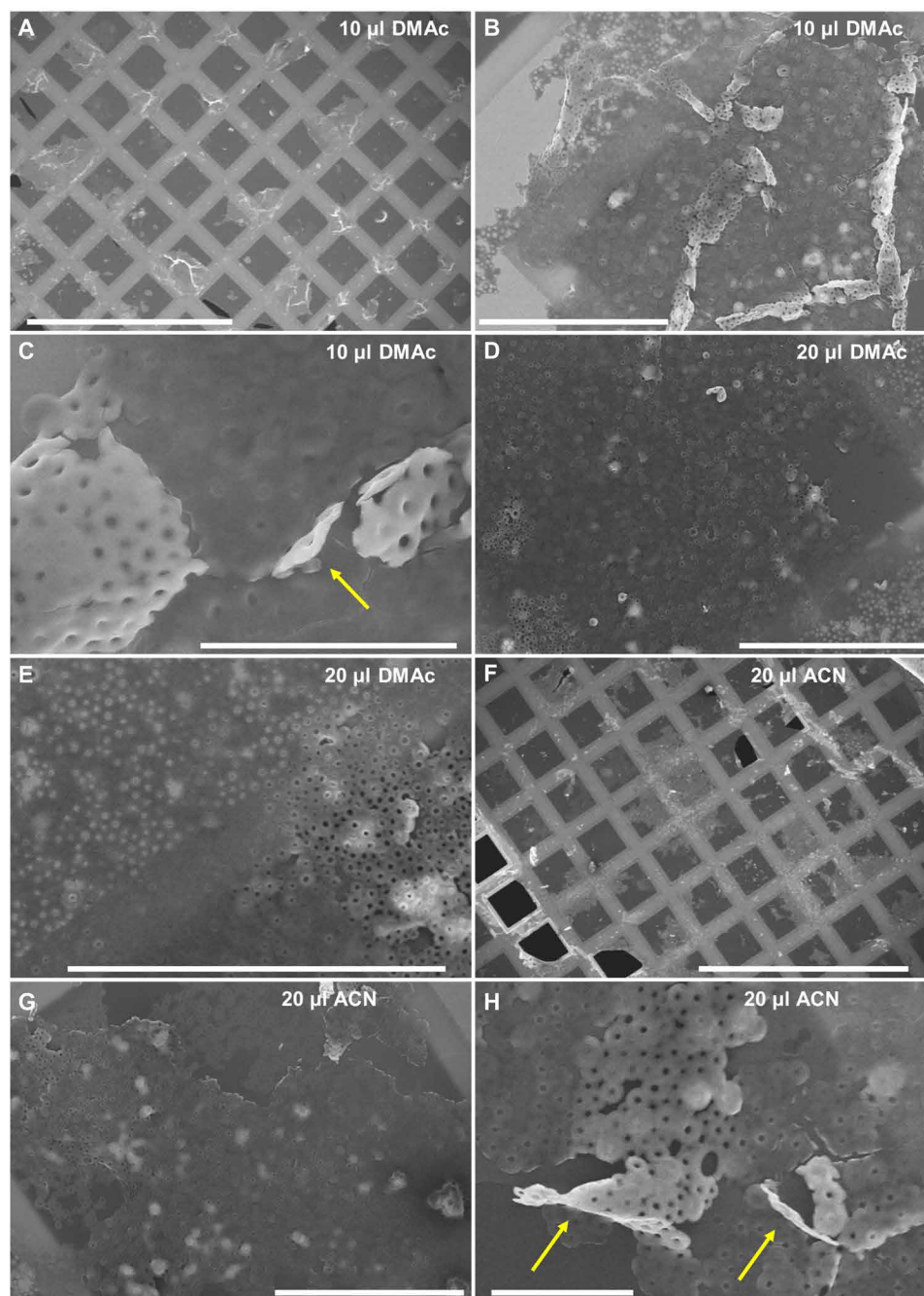


Fig. 6. Effect of an aliquot of DMAc and ACN on the morphology of multi-tori mesostructures. Representative SEM micrographs of mesostructures consisting of a network of toroids. The mesostructures were prepared by adding (A to C) 10 μ l of DMAc, (D and E) 20 μ l of DMAc, or (F to H) 20 μ l of ACN to a mixture of 1.02 ml of acetone/decane (25/75, v/v%), containing 20 μ l of PFS₅₅-*b*-PI₅₀₀ core-crystalline elongated micelles ($c = 0.2 \text{ mg ml}^{-1}$). The solution was preheated at 35°C, and an aliquot of PFS₂₆-*b*-PS₃₀₆ dissolved in THF (8 μ l, $c = 10 \text{ mg ml}^{-1}$) was injected. The solution was then annealed for 1 hour at 35°C and cooled to room temperature. The yellow arrows in (C) and (H) point at sections of the monolayer perpendicular to the TEM grid and protruding toward the reader. TEM grids were prepared within 1 hour after heating. Scale bars, 500 μ m (A and F), 40 μ m (B, D, E, and G), and 10 μ m (C and H).

deposited on the TEM grids have different morphologies, they all appear to be flat with ridges on their top, as one would expect from semirigid hollow objects collapsing onto a flat surface. A closer look at these structures (Fig. 6, B and C) confirms that they consist of a network of toroids. Similar structures could also be observed when 10 μl of DMF (fig. S15, A to C) was used, instead of DMAc. When ACN was used (fig. S15, D to F), SEM revealed the presence of broken sphere-like structures along with the flattened structures observed for DMAc and DMF. From these results, we conclude that the presence of a solvent immiscible with decane favored droplet coalescence to decrease the total surface area of the droplets.

To further increase droplet coalescence, we prepared solvent mixtures containing 20 μl of either DMAc (Fig. 6, D and E) or ACN (Fig. 6, F to H). This slight increase led to the formation of toroid mesh monolayers (Fig. 6, D to H, and fig. S16), as pointed out by the yellow arrows in Fig. 6 (C and H). These monolayers have a surface area of hundreds of square micrometers and a thickness corresponding to that of the toroids, i.e., around 300 nm, as evaluated from the images shown in Fig. 3 (C and D).

DISCUSSION

Here, we describe the preparation of 3D multi-tori structures via the CDSA of PFS₂₆-*b*-PS₃₀₆ in a 25/75 (v/v%) acetone/decane mixture, triggered by the presence of core-crystalline PFS₅₅-*b*-PI₅₀₀ elongated micelles. Both solvents are poor for each block. We show that these 3D structures are the result of a multistep process. In the first step, PFS₂₆-*b*-PS₃₀₆ unimer self-assembles into micrometer-large raspberry-like vesicles. In the absence of core-crystalline micelles, the films remain mostly amorphous, but thicken with time, to give large perforated films.

In the presence of PFS₅₅-*b*-PI₅₀₀ seed crystallites, the PFS₂₆-*b*-PS₃₀₆ BCP inside the thin raspberry-like vesicles appears to quickly undergo a sphere-to-cylinder transition, leading to the formation of core-crystalline elongated micelles. Because of this short nucleation time, the seeded growth of the PFS₂₆-*b*-PS₃₀₆ micelles is confined inside the wall of the vesicular mesostructures, and the elongated micelles are forced to grow concentrically along the equatorial plane of the droplets, resulting in the formation of toroids. This mechanism is particularly remarkable and differs strongly from those previously reported in the literature (45, 46).

The morphology of the multi-tori structures could also be tuned by adding an aliquot (10 or 20 μl) of a third solvent into the acetone/decane mixture. When MEK was used, the sphericity of the 3D objects was strongly increased, while 300-nm-thick toroid mesh monolayers could be obtained via the addition of a solvent immiscible with decane (DMAc, DMF, or ACN). These drastic changes in the final morphology of the multi-tori mesostructures induced by these subtle variations in solvent composition demonstrate the high potential of this approach to fabricate novel functional materials. It paves the way for new strategies to generate a plethora of intricate structures via bottom-up self-assembly.

MATERIALS AND METHODS

Materials

Decane (99+ %), THF (99+ %), acetone (99.9+ %), DMAc (99+ %), DMF (99+ %), and ACN (99.9+ %) were purchased from Sigma-Aldrich and used without further purification. PFS₅₅-*b*-PI₅₀₀ ($M_n = 48,200$, $D = 1.1$) is the same sample reported in (25). PFS₂₆-*b*-PS₃₀₆ ($M_n = 40,600$,

PDI = 1.06) was synthesized in the group of I. Manners by anionic polymerization following established procedures (47).

SEM and HRTEM

SEM images were taken using a high-resolution Hitachi S-5200 instrument at a voltage of 5 kV. Samples were prepared by placing one drop of solution on a formvar-carbon-coated grid. The sample was left on the grid for few minutes (up to 5 min), and then the edge of the droplet was touched with a filter paper to remove excess liquid, allowing the grid to dry. The samples are very stable in the dry state; thus, the same grid could also be used for HRTEM imaging several months after they were deposited on the TEM grids. The HRTEM images were taken with a JEOL JEM-2100 instrument at an accelerating voltage of 200 kV.

TEM

TEM imaging of the PFS₅₅-*b*-PI₅₀₀ core-crystalline micelles was performed on a Hitachi HT7700 TEM operating at an accelerating voltage of 80 kV. The TEM images were analyzed using the image processing program ImageJ.

Powder XRD experiments

Powder XRD experiments were performed on a Rigaku Ultima IV diffractometer at room temperature using Cu K α radiation ($\lambda = 1.5406 \text{ \AA}$). The diffraction pattern was collected with an angular resolution of 0.02° per step and a scan speed of 1° per min.

Measurement of PFS₅₅-*b*-PI₅₀₀ micelle length by TEM

To trigger the CDSA of the PFS₂₆-*b*-PS₃₀₆, we used a decane solution of core-crystalline micelles PFS₅₅-*b*-PI₅₀₀ with an extremely broad length distribution. More than 700 micelles were measured by hand using the software ImageJ (National Institutes of Health, USA). The number and weight average length of the micelles, L_n and L_w , respectively, were calculated as

$$L_n = \frac{\sum_{i=1}^N N_i L_i}{\sum_{i=1}^N N_i} \quad L_w = \frac{\sum_{i=1}^N N_i L_i^2}{\sum_{i=1}^N N_i L_i}$$

where N_i is the number of micelles of length L_i , and N is the total number of micelles examined. We found $L_n = 530 \text{ nm}$, $L_w = 1210 \text{ nm}$, and $L_w/L_n = 2.3$. The seed micelle solution contains a large majority of relatively short micelles (ca. 300 nm long), along with few micrometer-long micelles. A representative TEM image of these PFS₅₅-*b*-PI₅₀₀ micelles, as well as their length distribution, is shown in fig. S17.

Preparation of 3D multi-tori mesostructures by self-assembly

The samples were prepared by injecting a solution of PFS₂₆-*b*-PS₃₀₆ in THF (8 μl , $c = 10 \text{ mg ml}^{-1}$) to 1 ml of a 25/75 (v/v%) acetone/decane mixture preheated at 35°C, and to which 20 μl of PFS₅₅-*b*-PI₅₀₀ rod-like micelles in decane ($c = 0.2 \text{ mg ml}^{-1}$) had been added. The solution was annealed for 1 hour and then cooled to room temperature. Within 1 hour after the heating was stopped, an aliquot of the solution was deposited on a TEM grid. The sample was finally imaged by SEM and HRTEM.

Preparation of mesostructures (i) in the absence of PFS₅₅-*b*-PI₅₀₀ seeds and (ii) with a delayed addition of PFS₅₅-*b*-PI₅₀₀ seeds

We annealed a PFS₂₆-*b*-PS₃₀₆ unimer solution in acetone/decane for 1 hour at 35°C, in the absence of PFS₅₅-*b*-PI₅₀₀ micelles. We then took out the solution to let it cool in air and split it in two vials (S₁ and S₂; 0.33 ml each). S₁ was aged for one additional hour at room temperature, while we added an aliquot (7 μl in decane) of a solution of PFS₅₅-*b*-PI₅₀₀ micelles to S₂, just after it was taken out of the bath, and also let it age for one additional hour at room temperature. The samples were then examined by SEM and HRTEM.

An aliquot of the solution was deposited on a TEM grid held with an anti-capillary tweezer. The solution was left on the grid for 1 or 2 min, and excess solvent was removed by touching the side of the grid with a Kimwipe. The overall procedure took only few minutes, and we did not expect any preferential solvent evaporation that would affect the solvent ratio during this procedure. In addition, SEM imaging was performed on uncoated samples, which means that substantial height differences throughout the samples could lead to large differences in contrast due to charging effects.

Kinetic study of the formation of the 3D multi-tori mesostructures

The samples were prepared by injecting a solution of PFS₂₆-*b*-PS₃₀₆ in THF (8 μl, $c = 10 \text{ mg ml}^{-1}$) to six vials, each containing 1 ml of a 25/75 (v/v%) acetone/decane mixture preheated at 35°C, and to which 20 μl of PFS₅₅-*b*-PI₅₀₀ rod-like micelles in decane ($c = 0.2 \text{ mg ml}^{-1}$) had been added. At a given annealing time (namely, 0.5, 5, 10, 20, 30, and 60 min), an aliquot of the sample was taken out of the solution and deposited on the TEM grid. The excess of liquid was quickly removed with a filter paper. The two first samples (annealed for 30 s and 5 min) were imaged by SEM and HRTEM, and the four other samples were studied by HRTEM only.

Preparation of the sample for the XRD experiment

We injected 100 μl of a highly concentrated solution of PFS₂₆-*b*-PS₃₀₆ in THF ($c = 110 \text{ mg ml}^{-1}$) to 6 ml of a 25/75 (v/v%) acetone/decane mixture preheated at 35°C and containing 150 μl of PFS₅₅-*b*-PI₅₀₀ seed crystallites dispersed in decane ($c = 1 \text{ mg ml}^{-1}$). As soon as PFS₂₆-*b*-PS₃₀₆ dissolved in THF was added, the solution became cloudy and a small amount precipitated out, most likely due to the high concentration of the unimer in THF. We thus transferred the cloudy solution in a second vial and annealed this vial for 1 hour at 35°C. The solution was then cooled to room temperature. After the sample was aged overnight, it fully precipitated out of the solution, settling down at the bottom of the vial. We then removed the solvent, verified by SEM that this specific protocol led to the formation of toroids, and performed XRD on the resulting powder. SEM imaging was performed with a Hitachi S-4800 instrument at a voltage of 3 kV. The sample was prepared by placing the powder onto a conducting tape.

Preparation of 3D multi-tori mesostructures in the presence of an aliquot of MEK

The mesostructures were prepared by adding 20 μl of MEK to a mixture of 1.02 ml of acetone/decane (25/75, v/v%), containing 20 μl of PFS₅₅-*b*-PI₅₀₀ core-crystalline elongated micelles ($c = 0.2 \text{ mg ml}^{-1}$). The solution was preheated at 35°C, and an aliquot of PFS₂₆-*b*-PS₃₀₆ dissolved in THF (8 μl, $c = 10 \text{ mg ml}^{-1}$) was injected. The solution

was then annealed for 1 hour at 35°C and cooled to room temperature. TEM grids were prepared within 1 hour after heating.

Preparation of 2D toroid mesh monolayers in the presence of an aliquot of DMAc, DMF, or ACN

The mesostructures were prepared by adding 10 μl of DMAc to a mixture of 1.02 ml of acetone/decane (25/75, v/v%), containing 20 μl of PFS₅₅-*b*-PI₅₀₀ core-crystalline elongated micelles ($c = 0.2 \text{ mg ml}^{-1}$). The solution was preheated at 35°C, and an aliquot of PFS₂₆-*b*-PS₃₀₆ dissolved in THF (8 μl, $c = 10 \text{ mg ml}^{-1}$) was injected. The solution was then annealed for 1 hour at 35°C and cooled to room temperature. TEM grids were prepared within 1 hour after heating.

SUPPLEMENTARY MATERIALS

Supplementary material for this article is available at <http://advances.sciencemag.org/cgi/content/full/6/16/eaaz7301/DC1>

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