# Oncologist<sup>®</sup>

# Metastatic Thymoma Harboring a Deleterious *BRCA2* Mutation Derives Durable Clinical Benefit from Olaparib

DANIEL R. PRINCIPE ,<sup>a</sup> SUNEEL D. KAMATH,<sup>b</sup> HIDAYATULLAH G. MUNSHI,<sup>b</sup> NISHA A. MOHINDRA<sup>b</sup>

<sup>a</sup>Medical Scientist Training Program, University of Illinois College of Medicine, Chicago, Illinois, USA; <sup>b</sup>Feinberg School of Medicine, Northwestern University, Chicago, Illinois, USA

Disclosures of potential conflicts of interest may be found at the end of this article.

### Abstract \_

Thymomas comprise a group of rare epithelial neoplasms of the anterior mediastinum. Whereas localized disease carries a favorable prognosis, the majority of patients with metastatic thymomas experience progression or recurrence over a 10-year period. Although targeted therapies have become standard of care in many malignancies, no clinically actionable mutations have consistently been identified in metastatic thymomas. Here, we describe a patient with an aggressive thymoma complicated by extensive pleural metastases. Over a 16-year period, she progressed on multiple treatment regimens. To identify additional treatment options, tissue from a pleural metastasis was sent for next-generation sequencing, revealing mutations in *BRCA2*, tyrosine kinase 2, and SET domain containing 2. Based on supporting evidence for poly (ADP-ribose) polymerase (PARP) inhibition in other BRCA-mutated tumors, the patient was started on the PARP inhibitor olaparib. She derived significant clinical benefit from treatment, with imaging showing overall stabilization of her disease. Here, we review the genotyping results of her tumor and discuss the functional and clinical significance of the mutations in her cancer as well as implications for managing patients with advanced BRCA-mutant thymomas. *The Oncologist* 2020;25:301–305

### **KEY POINTS**.

- Targeted therapy has yet to enter the standard clinical management of metastatic thymomas.
- Patients with BRCA2-mutant thymomas may benefit from poly (ADP-ribose) polymerase inhibition.

### PATIENT STORY \_

A 60-year-old woman with a history of autoimmune hepatitis was diagnosed with both thymoma (World Health Organization type B) and papillary thyroid cancer in September 2003. At this time, she underwent resection of the thymoma as well as thyroidectomy followed by iodine-131 therapy for her thyroid cancer. Subsequent positron emission tomography imaging revealed mediastinal lymphadenopathy that was treated with external beam radiation. Two years after diagnosis, imaging identified a solitary left pleural lesion pathologically confirmed to be metastatic thymoma. When the lesion began enlarging after another 2 years, she was treated with sandostatin LAR and prednisone. Over the next 10 years, she was treated with multiple regimens, including a combination of carboplatin and etoposide, single-agent gemcitabine, CAP (cyclophosphamide, doxorubicin, and cisplatin), single-agent pemetrexed, capecitabine, and single-agent everolimus. She was also treated with the anti-insulin-like growth factor 1 receptor antibody cixutumumab on a clinical trial. Although cixutumumab halted significant progression for nearly 2 years,

in each case she eventually progressed on treatment and ultimately developed extensive, bulky pleural metastases (Fig. 1).

To identify additional treatment options, a sample of her tumor was sent for FoundationOne next-generation sequencing (NGS), which identified mutations in BRCA2, tyrosine kinase 2 (TYK2), and SET domain containing 2 (SETD2). Although neither TYK2 nor SETD2 mutations are clinically actionable, cancers harboring inactivation mutations in BRCA1 or BRCA2 can respond to poly (ADP-ribose) polymerase (PARP) inhibitors [1]. Thus, the patient was started on olaparib and has remained on treatment for 14 months with overall stable disease. The patient had some reduction in tumor size (approximate of 20% reduction in the sum of the longest diameter for all target lesions); however, this did not meet RECIST criteria for a partial response (Fig. 1). Because of toxicity of nausea and fatigue, the patient has remained on dose-reduced olaparib and has since been able to wean down her pain regimen for management of pleural-based cancer pain.

Correspondence: Nisha A. Mohindra, M.D., Northwestern University Feinberg School of Medicine, 676 N St Clair St., Suite 850, Chicago, Illinois 60611, USA. Telephone: 312-695-2351; e-mail: nisha.mohindra@nm.org Received May 22, 2019; accepted for publication September 24, 2019; published Online First on November 1, 2019. http://dx.doi.org/10.1634/theoncologist.2019-0393

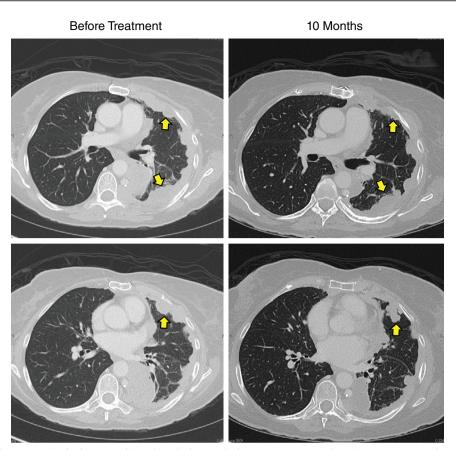


Figure 1. Computed tomography (CT) image of the chest before and after treatment. Baseline chest CT imaging showing extensive metastatic pleural-based nodules on the left (arrows) from metastatic thymoma, and overall stable disease after 10 months on olaparib.

### **MOLECULAR TUMOR BOARD**

Precision medicine has become standard of care in several cancers. Tyrosine kinase inhibitors targeting the BCR-ABL fusion protein have revolutionized the treatment of chronic myeloid leukemia [2], and many therapies targeting molecular alterations have made an impact in breast cancer [3], non-small cell lung cancer, and others. Comprehensive genomic profiling is also changing the treatment paradigm toward molecularly targeted therapies, regardless of histology. For instance, a trial of 1,144 patients with any solid tumor evaluated in a large phase I program showed that patients harboring distinct molecular aberrations treated with a matched targeted therapy had significant improvements in overall response rates, time to treatment failure, and overall survival [4]. Although a patient with thymic carcinoma overexpressing mutant KIT responded to imatinib [5], such approaches have yet to enter the management of metastatic thymomas. Although thymic epithelial tumors frequently harbor mutations in GTF2I, HRAS, NRAS, and TP53, none of these mutations are clinically actionable at this time [6]. Similarly, although thymic epithelial tumors can be clustered into four genetic subtypes (38% GTF2I mutant, 33% Tcell enriched, 21% chromosome unstable, 8% chromosome stable), these groupings have yet to influence clinical practice [7].

## GENOTYPING RESULTS AND INTERPRETATION OF MOLECULAR RESULTS

The FoundationOne Heme (Foundation Medicine, Cambridge, MA) assay is recommended by the manufacturer for

thymomas, and it uses whole-genome shotgun library construction and hybrid-capture NGS to provide a focused interrogation of 406 genes and 31 introns, 265 RNAs associated with gene fusions, and information regarding the tumor mutational burden and microsatellite instability status. Briefly, massively parallel sequencing is done using 50-200 ng of DNA from a formalin-fixed paraffin-embedded tissue specimen. Hybrid-capture-selected libraries are sequenced to high uniform depth. FoundationOne sequences patient tissues to a median depth of approximately 500× unique coverage for DNA and RNA to an average of  $\sim$ 6.9 million unique pairs (targeting >500× coverage by non-polymerase chain reaction duplicate read pairs, with >99% of exons at coverage >100×) using the Illumina HiSeq 2500 platform (Illumina, San Diego, CA, https://www.illumina. com). The panel identifies base substitutions, insertions and deletions, copy number alterations, and rearrangements [8]. In our patient, the results revealed three distinct mutations: *TYK2*<sup>V15A</sup>, *SETD2*<sup>T1652fs\*14</sup>, and *BRCA2*<sup>K1800fs\*16</sup>.

Although each of these genes has been investigated in other cancers, none has a clearly defined role in thymic cancers. For instance, *TYK2* encodes for a nonreceptor JAK family tyrosine kinase with contradictory roles in inflammation. Although TYK2 is involved in interleukin (IL)-12 and interferon signaling, it also acts downstream of IL-10, and *Tyk2* knockout mice are unable to fully generate or respond to IL-10 [9, 10]. In cancer, the effects of TYK2 are similarly dichotomous. Although TKY2 is seemingly required for oncogenic fibroblast growth factor and epidermal growth factor signaling, TYK2 inhibition also appears to impede anticancer CD8 responses [11]. Interestingly, in



Table 1. Frequency of BRCA	mutations in breast	, ovarian, and	thymic cancers	in genomic databases
		, ,	,	8

Data set	Sample size	BRCA1 mutation	BRCA2 mutation	Combined
Breast Cancer (MSK)	1,918	1.30%	3.18%	4.38%
Breast Cancer (METABRIC)	2,506	1.68%	1.80%	3.43%
Breast Cancer (TCGA)	1,066	2.53%	2.72%	5.07%
Ovarian Cancer (TCGA)	585	3.08%	2.56%	5.64%
Thymoma (TCGA)	124	0%	1.61%	1.61%

Abbreviations: METABRIC, Molecular Taxonomy of Breast Cancer International Consortium; MSK, Memorial Sloan Kettering; TCGA, The Cancer Genome Atlas.

murine models of allografted thymoma cells, genetic deletion of *TYK2* accelerated tumor development largely because of defects in CD8-mediated cytotoxicity [12]. Although interesting, the *TYK2*<sup>V15A</sup> mutation has not been described in meta-static thymoma and its clinical significance is unknown.

SETD2 is a histone methyltransferase specific for lysine-36 of histone H3, and it is generally considered a tumor suppressor gene [13]. SETD2 is frequently altered in other malignancies, including approximately 30% of pediatric high-grade gliomas and 15% of clear cell renal cell carcinomas [14, 15]. In breast cancer, SETD2 is often downregulated, and lower SETD2 mRNA levels have been associated with both local and distant recurrences and poor survival [16]. Similarly, in B-cell acute lymphoblastic leukemia, loss of SETD2 is associated with chemotherapy resistance through loss of DNA damage recognition and impaired apoptotic signaling [17]. Although the specific SETD2<sup>T1652fs\*14</sup> frame-shift mutation observed in our patient has not been described, it occurs in the coding region for the SET domain (AA 1561-1667) and is likely analogous to the oncogenic SETD2<sup>R1625C</sup> mutation [18]. However, the clinical significance of this mutation is unclear at this time.

Finally, our patient's tumor harbored a BRCA2K1800fs\*16 frame-shift mutation with presumptive loss of function. After learning this, our patient was referred to a hospital geneticist. A blood sample was sent for *BRCA2* sequencing ( $\pm$ 20 base pairs of adjacent intronic sequence) through Invitae, and this mutation was found to be germline, although the patient had no personal or family history of BRCA-associated cancers. Whereas the role of BRCA2 in thymic cancers is largely unexplored, familial BRCA1 and BRCA2 mutations have been well described in breast and ovarian cancers. Although familial BRCA mutations drive only a relatively small percentage of these cancers, women with an inherited BRCA1 mutation have a lifetime risk of 65%-80% of developing breast cancer and 37%-62% for ovarian cancer. Similarly, women carrying familial BRCA2 mutations have a lifetime risk of 45%-85% for breast cancer and 11%-23% for ovarian cancer [19]. BRCA1 and BRCA2 are established tumor suppressor genes with distinct but important roles in DNA damage response and repair pathways [20]. Given the strong evidence supporting the use of PARP inhibitors in BRCA-mutant ovarian cancers [1], our patient was started on olaparib.

# Functional and Clinical Significance of the Specific Mutation in Thymic Cancer

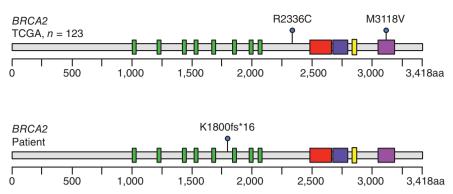
Little is known regarding the role of BRCA mutations in thymic cancers. To date, there has only been one reported case of a *BRCA1*-mutant metastatic thymoma, although this patient also developed thyroid and ovarian cancers during her lifetime

[21]. A similar report described a family with frequent thymomas, potentially attributed to a germline translocation of the *BRCA2*-associated DNA repair protein RAD51B [22]. Another family was reported to have high rates of BRCA-linked breast/ovarian cancers and thymomas, although there was insufficient evidence to suggest a familial cancer syndrome caused by a single unifying mutation [23]. Of note, one series of 72 patients with thymoma or thymic carcinoma identified 1 patient with an ataxia-telangiectasia mutated (*ATM*) gene mutation, which can affect DNA repair mechanisms similar to BRCA mutations [24]. Interestingly, deletion of ATM *in vivo* is associated with spontaneous thymoma formation, whereas restoration of ATM expression reverses this phenotype [25].

Despite these observations, the frequency of BRCA mutations in thymomas is unknown. We therefore evaluated the established The Cancer Genome Atlas (TCGA) genomic database for mutations in either BRCA1 or BRCA2 using cBioportal [26, 27]. Of 123 thymoma patients, none had a BRCA1 mutation, and two (1.61%) had mutations in BRCA2 (Table 1). Of the observed BRCA2 mutations, one consisted of a single amino acid substitution of unknown significance (M3118V) at the oligonucleotide/ oligosaccharide-binding domain 3 (AA: 3052 - 3190). The second mutation was another substitution (R2336C) analogous to the oncogenic R2336H/P mutations [28, 29]. Our patient displayed a frame-shift mutation at the 1800 position (Fig. 2). While this particular mutation has not been characterized, several similar BRCA2 frame-shift mutations have been identified in breast and ovarian cancers, many leading to missing or nonfunctional proteins [30]. However, given the small sample size of the TCGA cohort, the overall rate of oncogenic BRCA mutations in patients with thymomas remains unclear. Given the efficacy of olaparib in our patient, PARP inhibition in BRCA-mutated advanced thymomas warrants further investigation.

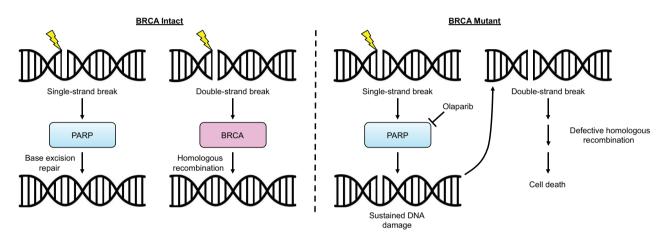
## POTENTIAL STRATEGIES TO TARGET THE PATHWAY AND IMPLICATIONS FOR CLINICAL PRACTICE

The mechanistic link between BRCA and PARP inhibition is well established. In brief, PARP is critical for single-strand break repair (SSBR) [31]. As such, PARP inhibitors such as olaparib prevent SSBR, leading to double strand breaks (DSBs). As *BRCA1* and *BRCA2* play key roles in DSB repair, namely, through homologous recombination [32], the use of olaparib in BRCA-mutant cancers leads to the accumulation of irreparable DSBs, ultimately causing cell death (Fig. 3). As discussed, this approach has shown significant efficacy in BRCA-mutant ovarian and breast cancers. Olaparib became the first Food and Drug Administration (FDA)-approved PARP inhibitor for patients with ovarian cancer with germline BRCA mutations in 2014. This was based on results



**Figure 2.** Known *BRCA2* mutations in patients with thymomas. Using the TCGA genomic dataset, we evaluated the known mutations of *BRCA1* and *BRCA2* in 123 patients with thymomas. Two patients harbored point mutations leading to single amino acid substitutions either at R2236C or M3118V at the oligonucleotide/oligosaccharide-binding, domain 3. In contrast, our patient had a frameshift mutation at the K1800 position, presumptively causing loss of function.

Abbreviation: TCGA, The Cancer Genome Atlas.



**Figure 3.** Schema describing the efficacy of PARP inhibition in BRCA-deficient tumor cells. Under physiologic conditions, DNA damage causing single-strand breaks (SSBs) leads to the rapid synthesis and recruitment of PARP. This promotes base excision repair, leading to repair of SSBs. In the case of double-strand breaks (DSBs), BRCA proteins interact with a variety of additional factors to promote repair via homologous recombination. In the setting of BRCA deficiency, the repair of DSBs is significantly impaired. Therefore, when PARP is inhibited with olaparib, cells lack the ability to repair SSBs, leading to the accumulation of DSBs. With no means of repairing these DSBs, cells eventually undergo programmed cell death. Abbreviation: PARP, poly (ADP-ribose) polymerase.

from Study 19, a randomized, placebo-controlled trial showing a statistically significant improvement in both progression-free and overall survival [33]. Three PARP inhibitors, olaparib, rucaparib, and niraparib, have now been FDA approved as maintenance therapy after platinum-based chemotherapy in patients with ovarian cancer with either germline or somatic BRCA mutations or homologous recombination deficiency [34–36]. Olaparib was also FDA approved in 2018 for germline BRCA-mutated breast cancer based on the randomized, phase III OlympiAD trial showing an improved response rate (59.9% vs. 28.8%) and progression-free survival (7.0 vs. 4.2 months) compared with standard therapy [37].

Additionally, emerging evidence suggests there are additional mutations that may be indicative of homologous recombination deficiencies. These include alterations in *ATM*, *BARD1*, *BRIP1*, *CHEK2*, *FAAP20*, *FAN1*, *FANCE*, *FANCM*, *PALB2*, *POLQ*, *RAD51B*, *RAD51C*, and *RAD51D* [38–40]. Several of these mutations, namely, *ATM* and *PALB2*, have correlated with responsiveness to PARP inhibition in other malignancies including metastatic prostate cancer [41]. Given these results, and those seen in our patient, PARP inhibition warrants further investigation in other

malignancies with germline or somatic BRCA mutations and potentially other mutations in DNA repair pathways.

### **PATIENT UPDATE**

After 14 months on treatment, the patient reports that her pain is under control without new chest/abdominal pain or other related symptoms. The first 3 months on olaparib were complicated by nausea and fatigue, but after a dose adjustment, the patient is tolerating treatment well. Her most recent imaging demonstrates overall stable disease with no new pleural-based nodules.

GLOSSARY OF GENOMIC TERMS AND NOMENCLATURE DSBs: double-strand breaks NGS: next-generation sequencing SSBs: single strand breaks SSBR: single-strand break repair

#### ACKNOWLEDGMENTS

We thank our patient for giving us the permission to include her clinical information in this report, and we wish



her well in her continued recovery. This work was supported by NIH F30CA236031 to D.R.P.

**AUTHOR CONTRIBUTIONS** 

- Conception/design: Daniel R. Principe, Suneel D. Kamath, Hidayatullah G. Munshi, Nisha A. Mohindra
- Provision of study material or patients: Daniel R. Principe, Suneel D. Kamath, Hidayatullah G. Munshi, Nisha A. Mohindra
- Collection and/or assembly of data: Daniel R. Principe, Suneel D. Kamath, Hidayatullah G. Munshi, Nisha A. Mohindra

#### **References** \_

**1.** Konecny GE, Kristeleit RS. PARP inhibitors for BRCA1/2-mutated and sporadic ovarian cancer: Current practice and future directions. Br J Cancer 2016;115:1157–1173.

**2.** An X, Tiwari AK, Sun Y et al. BCR-ABL tyrosine kinase inhibitors in the treatment of Philadelphia chromosome positive chronic myeloid leukemia: A review. Leuk Res 2010;34:1255–1268.

**3.** Maximiano S, Magalhaes P, Guerreiro MP et al. Trastuzumab in the treatment of breast cancer. BioDrugs 2016;30:75–86.

**4.** Tsimberidou AM, Iskander NG, Hong DS et al. Personalized medicine in a phase I clinical trials program: The MD Anderson Cancer Center initiative. Clin Cancer Res 2012;18:6373–6383.

**5.** Strobel P, Hartmann M, Jakob A et al. Thymic carcinoma with overexpression of mutated KIT and the response to imatinib. N Engl J Med 2004;350:2625–2626.

**6.** Radovich M, Pickering CR, Felau I et al. The integrated genomic landscape of thymic epithelial tumors. Cancer Cell 2018;33:244–258 e210.

**7.** Lee HS, Jang HJ, Shah R et al. Genomic analysis of thymic epithelial tumors identifies novel subtypes associated with distinct clinical features. Clin Cancer Res 2017;23:4855–4864.

**8.** He J, Abdel-Wahab O, Nahas MK et al. Integrated genomic DNA/RNA profiling of hematologic malignancies in the clinical setting. Blood 2016;127:3004–3014.

**9.** Shaw MH, Freeman GJ, Scott MF et al. Tyk2 negatively regulates adaptive Th1 immunity by mediating IL-10 signaling and promoting IFN-gamma-dependent IL-10 reactivation. J Immunol 2006;176:7263–7271.

**10.** Hashiguchi T, Oyamada A, Sakuraba K et al. Tyk2-dependent bystander activation of conventional and nonconventional Th1 cell subsets contributes to innate host defense against *Listeria monocytogenes* infection. J Immunol 2014;192: 4739–4747.

**11.** Ubel C, Mousset S, Trufa D et al. Establishing the role of tyrosine kinase 2 in cancer. Oncoimmunology 2013;2:e22840.

**12.** Simma O, Zebedin E, Neugebauer N et al. Identification of an indispensable role for tyrosine kinase 2 in CTL-mediated tumor surveillance. Cancer Res 2009;69:203–211.

**13.** Li J, Duns G, Westers H et al. SETD2: An epigenetic modifier with tumor suppressor functionality. Oncotarget 2016;7:50719–50734.

**14.** Cancer Genome Atlas Research Network. Comprehensive molecular characterization of clear cell renal cell carcinoma. Nature 2013;499: 43–49.

www.TheOncologist.com

**15.** Fontebasso AM, Liu XY, Sturm D et al. Chromatin remodeling defects in pediatric and young adult glioblastoma: A tale of a variant histone 3 tail. Brain Pathol 2013;23:210–216.

**16.** Al Sarakbi W, Sasi W, Jiang WG et al. The mRNA expression of SETD2 in human breast cancer: Correlation with clinico-pathological parameters. BMC Cancer 2009;9:290.

**17.** Mar BG, Chu SH, Kahn JD et al. SETD2 alterations impair DNA damage recognition and lead to resistance to chemotherapy in leukemia. Blood 2017;130:2631–2641.

**18.** Hacker KE, Fahey CC, Shinsky SA et al. Structure/function analysis of recurrent mutations in SETD2 protein reveals a critical and conserved role for a SET domain residue in maintaining protein stability and histone H3 Lys-36 trimethylation. J Biol Chem 2016;291: 21283–21295.

**19.** Balmana J, Diez O, Rubio IT et al. BRCA in breast cancer: ESMO clinical practice guidelines. Ann Oncol 2011;22(suppl 6):vi31–vi34.

**20.** Roy R, Chun J, Powell SN. BRCA1 and BRCA2: Different roles in a common pathway of genome protection. Nat Rev Cancer 2011;12: 68–78.

**21.** Yi EJ, Park JH, Lee HW et al. BRCA1 gene mutation in thymic malignant melanoma. Ann Thorac Surg 2013;96:677–680.

**22.** Nicodeme F, Geffroy S, Conti M et al. Familial occurrence of thymoma and autoimmune diseases with the constitutional translocation t(14; 20)(q24.1;p12.3). Genes Chromosomes Cancer 2005;44:154–160.

**23.** Zhang X, Wang T, Wang W et al. Does familial breast cancer and thymoma suggest a cancer syndrome? A family perspective. Gene 2015;573: 333–337.

**24.** Enkner F, Pichlhofer B, Zaharie AT et al. Molecular profiling of thymoma and thymic carcinoma: Genetic differences and potential novel therapeutic targets. Pathol Oncol Res 2017;23: 551–564.

**25.** Di Siena S, Campolo F, Gimmelli R et al. Atm reactivation reverses ataxia telangiectasia phenotypes in vivo. Cell Death Dis 2018;9:314.

**26.** Cerami E, Gao J, Dogrusoz U et al. The cBio cancer genomics portal: An open platform for exploring multidimensional cancer genomics data. Cancer Discov 2012;2:401–404.

**27.** Gao J, Aksoy BA, Dogrusoz U et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 2013;6:pl1.

**28.** Claes K, Poppe B, Coene I et al. BRCA1 and BRCA2 germline mutation spectrum and

Data analysis and interpretation: Daniel R. Principe, Suneel D. Kamath, Hidayatullah G. Munshi, Nisha A. Mohindra

Manuscript writing: Daniel R. Principe, Suneel D. Kamath, Hidayatullah G. Munshi, Nisha A. Mohindra

Final approval of manuscript: Daniel R. Principe, Suneel D. Kamath, Hidayatullah G. Munshi, Nisha A. Mohindra

**DISCLOSURES** The authors indicated no financial relationships.

frequencies in Belgian breast/ovarian cancer families. Br J Cancer 2004;90:1244–1251.

**29.** Laitman Y, Simeonov M, Herskovitz L et al. Recurrent germline mutations in BRCA1 and BRCA2 genes in high risk families in Israel. Breast Cancer Res Treat 2012;133:1153–1157.

**30.** Petrucelli N, Daly MB, Feldman GL. Hereditary breast and ovarian cancer due to mutations in BRCA1 and BRCA2. Genet Med 2010;12: 245–259.

**31.** Fisher AE, Hochegger H, Takeda S et al. Poly (adp-ribose) polymerase 1 accelerates singlestrand break repair in concert with poly(adpribose) glycohydrolase. Mol Cell Biol 2007;27: 5597–5605.

**32.** Liu Y, West SC. Distinct functions of BRCA1 and BRCA2 in double-strand break repair. Breast Cancer Res 2002;4:9–13.

**33.** Ledermann J, Harter P, Gourley C et al. Olaparib maintenance therapy in platinumsensitive relapsed ovarian cancer. N Engl J Med 2012;366:1382–1392.

**34.** Moore K, Colombo N, Scambia G et al. Maintenance olaparib in patients with newly diagnosed advanced ovarian cancer. N Engl J Med 2018;379:2495–2505.

**35.** Coleman RL, Oza AM, Lorusso D et al. Rucaparib maintenance treatment for recurrent ovarian carcinoma after response to platinum therapy (ARIEL3): A randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2017; 390:1949–1961.

**36.** Mirza MR, Monk BJ, Herrstedt J et al. Niraparib maintenance therapy in platinum-sensitive, recurrent ovarian cancer. N Engl J Med 2016;375:2154–2164.

**37.** Robson M, Im SA, Senkus E et al. Olaparib for metastatic breast cancer in patients with a germline BRCA mutation. N Engl J Med 2017; 377:523–533.

**38.** Hoppe MM, Sundar R, Tan DSP et al. Biomarkers for homologous recombination deficiency in cancer. J Natl Cancer Inst 2018; 110:704–713.

**39.** Riaz N, Blecua P, Lim RS et al. Pan-cancer analysis of bi-allelic alterations in homologous recombination DNA repair genes. Nat Commun 2017;8:857.

**40.** Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. Nature 2011;474:609–615.

**41.** Mateo J, Carreira S, Sandhu S et al. DNArepair defects and olaparib in metastatic prostate cancer. N Engl J Med 2015;373:1697–1708.