



## Review article

## Childhood diarrhoeal diseases in developing countries

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## ABSTRACT

Diarrhoeal diseases collectively constitute a serious public health challenge globally, especially as the leading cause of death in children (after respiratory diseases). Childhood diarrhoea affecting children under the age of five accounts for approximately 63% of the global burden. Accurate and timely detection of the aetiology of these diseases is very crucial; but conventional methods, apart from being laborious and time-consuming, often fail to identify difficult-to-culture pathogens. The aetiological agent of an average of up to 40% of cases of diarrhoea cannot be identified. This review gives an overview of the recent trends in the epidemiology and treatment of diarrhoea and aims at highlighting the potentials of metagenomics technique as a diagnostic method for enteric infections.

## 1. Introduction

World health organization defines diarrhoea as the passage of three or more loose or liquid stools per day (or more frequent passage than is normal for the individual (WHO, 2007). Worldwide, diarrhoeal diseases are reported as the leading cause of mortality among children aged five years and below (UNICEF, 2018). In some parts of the world, they account for higher mortality rates than all other causes combined (Petri et al., 2008; UNICEF, 2018). Childhood diarrhoea affecting children five years old and below accounts for approximately 63% of the global diarrhoea burden (Walker et al., 2012; Zhang et al., 2016), and is the second significant cause of infant mortality in developing nations (Platts-Mills et al., 2015; Kotloff, 2017) where poor sanitation and insufficient potable water supply are key factors (Chakravarty et al., 2017; Squire and Ryan, 2017). In Africa, Asia, and South America, diarrhoea accounts for one in eight deaths among children younger than 5 years per annum (Keddy et al., 2016; Kotloff, 2017) and an estimated 16% of child deaths in Nigeria annually (Charveva et al., 2015). In Ogun State, South-West Nigeria, diarrhoea is one of the three diseases (the others being typhoid fever and cholera) which together are the second most prevalent water-related disease (Omole et al., 2015).

Acute diarrhoea, the passage of stools with abnormal consistency and frequency in a day (e.g. more than three times) which lasts for less than two weeks, is a syndrome that is frequently not subject to differential diagnosis in medical practice (Petri et al., 2008; Zhang et al., 2016).

Bacterial diarrhoeal cases are the most prevalent of all diarrhoeal cases around the globe (Zhang et al., 2018). Commonly reported enteric bacterial diarrhoeal diseases and the causative agents are botulism (*Clostridium botulinum*), Campylobacter gastroenteritis (*Campylobacter jejuni*), cholera (*Vibrio cholerae*), *Escherichia coli* gastroenteritis, Salmonellosis (various *Salmonella* serovars), Shigellosis (*Shigella spp.*), and Staphylococcal food poisoning (*Staphylococcus aureus* enterotoxins) (Humphries and Linscott, 2015; Tarr et al., 2018). Children below five years of age have the most at risk from foodborne pathogens, including Shiga toxin-producing *Escherichia coli* O157, *Campylobacter*, *Shigella*, *Yersinia*, *Salmonella*, and *Cryptosporidium* (Liu et al., 2014a, b; Kotloff, 2017). In the past few decades, the awareness in handwashing has tremendously reduced the burden of diarrhoea caused by enteric bacteria and protozoans, yet, it has less impact on diarrhoea caused by viruses (Tagbo et al., 2019).

The mouth is the typical portal of entry for gastrointestinal pathogens, which are ingested alongside contaminated food and water (Rodulfo et al., 2012). Also, they are acquired via contact with diarrhoeic animals and their contaminated environments or with the faecal matter of a diarrhoeic person (Humphries and Linscott, 2015; Squire and Ryan, 2017). While in their gastrointestinal habitat, these pathogens, through a variety of pathological mechanisms by which they could be typed, trigger the over secretion of fluid in the lumen of the small intestine associated with electrolyte imbalance, and eventual diarrhoea (Humphries and Linscott, 2015; Crawford et al., 2017). In spite of the huge burden of

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diarrhoea, it is preventable with modern science and public health intervention (Centres for Disease Control and Prevention, 2016a, b). In some cases, diarrhoea may be self-limiting. In severe infections, however, antibiotics may be prescribed to prevent possible death. The resulting risk, however, is antibiotic resistance, which is an important public health threat towards the treatment of diarrhoea (Centres for Disease Control and Prevention, 2013). The continuous surveillance of antimicrobial resistance is an epidemiological strategy at tracking new and emerging resistances to some of the last-line antibiotics.

Accurate diagnosis of diarrhoeal pathogens is necessary for surveillance, prevention, and control of diarrhoea (Ranjbar et al., 2014; Tarr et al., 2018). Traditional, phenotypic tests such as Gram staining, bacteriological culture and recording of colonial characteristics, and biochemical tests form the mainstay of laboratory diagnosis in less developed countries (Liu et al., 2014a, b). However, such tests take longer turnaround time to identify slow-growing bacteria, resulting in delayed treatment of patients (Khan and Jahan, 2017; Maciel and Leite, 2018). In many other cases, the results of these tests, even when considered in concert, are false negatives (Miller et al., 2013).

Conventional epidemiological typing methods which include biotyping, antibiogram, and serotyping are quite useful in describing temporal epidemiological studies (MacCannell, 2013). However, aetiological agents of approximately 40% of gastroenteritis cases go undetected by these methods, complicating diagnosis and treatment (Finkbeiner et al., 2008; Khan and Jahan, 2017).

Molecular approaches have increasingly brought to light significant viral, parasitic, and bacterial enteric pathogens and also their virulent traits (Zhou et al., 2016). Most molecular techniques employ polymerase chain reaction (PCR) to detect deoxyribonucleic acid (DNA) in a sample (Chang et al., 2013). One of the tools in molecular epidemiology is microbial typing. Microbial typing helps to diagnose the aetiology and the route of transmission of infection, identify virulent and resistant strains and evaluate the impact of control measures of infectious diseases (Ranjbar et al., 2014). Next-generation sequencing (NGS), a technique which quickly and extensively sequences a mixed population of DNA or ribonucleic acid (RNA) genomes have enhanced the study of infectious disease epidemiology (Platts-Mills et al., 2013). Metagenomics, which is the technique that directly sequences and analyses the total nucleic acids isolated from a sample, without culture, has a promising prospect in the field of infectious disease diagnosis (Decuyper et al., 2016).

Globally, researchers are already utilizing metagenomics in the aetiology and antimicrobial resistance surveillance of diarrhoea (Table 1). Conversely, most of the studies on diarrhoea aetiology in Sub-Saharan

Africa has focused more on the prevalence, antimicrobial profile, and risk factors (Godana and Mengistie, 2013; Akingbade et al., 2013; Chiyangi et al., 2017). A few studies have reported the molecular characterisation of enteric pathogens isolated from diarrhoea patients (Ifeanyi et al., 2014; Tian et al., 2016). This review aims at highlighting recent trends in the epidemiology and treatment of enteric infections and gives an overview of the recent molecular methods of diagnosis. In furtherance, the potentials of metagenomics as a diagnostic method for diarrhoea infections will be highlighted.

## 2. Epidemiology of diarrhoea

### 2.1. Severity and trend of diarrhoea among children

The four major infectious diseases that cause death in children under the age of 5 are pneumonia, diarrhoeal diseases, malaria and measles (Figure 1). However, 25% and 31% of the overall diarrhoeal burden has been attributed to diarrhoea among children below the age of 5 in Africa and Asia respectively (Walker et al., 2012), especially Sub-Saharan Africa which has the highest death rate (Figure 2). Despite the global decline in the diarrhoea mortality rate from 11% in 2010 to 9% in 2015, it is still the second cause of mortality among children under five years of age (Liu et al., 2012; Liu et al., 2016a, b).

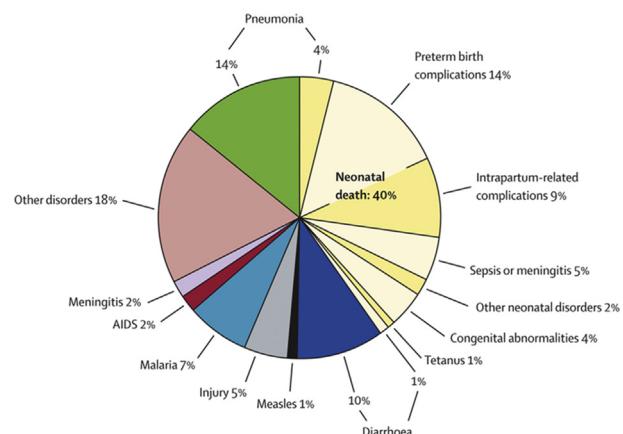
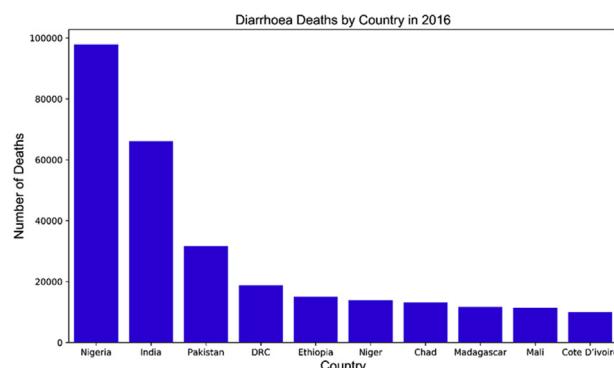


Figure 1. Causes of child deaths worldwide; Source: Liu et al. (2012)..

Table 1. Application of metagenomics in diarrhoea diagnosis.

Country	References	Metagenomic method	Age (year)	Sample size (n)	Type of organisms detected
Australia	Finkbeiner et al. (2008)	Micro-mass sequencing	0–5	12	Viruses
Japan	Nakamura et al. (2008)	454 Roche	34	1	Bacteria
Bangladesh	Smits et al. (2012)	454 Roche	All	105	Viruses
Germany	Loman et al., 2013	Deep Amplicon sequencing	n/a	45	Bacteria
Bangladesh	Monira et al. (2013)	454 parallel sequencing	2–3	9	Bacteria
Australia	Holtz et al. (2014)	454 Titanium platform	0–5	8	Viruses
Nicaragua	Becker-Dreps et al. (2015)	16S rRNA amplicon sequencing	0–5	66	Bacteria
Coted'Ivoire	Schneeberger et al. (2016)	Shotgun	All	4	Multi-pathogens
U.S. A	Youmans et al. (2015)	16S rRNA	Adults	99	Bacteria and viruses
China	Sun et al. (2016)	Deep amplicon sequencing	0–5	7	Viruses
U.S. A	Zhou et al. (2016)	Shotgun/WGS	0–18	22	Multi-pathogens
Israel	Braun et al. (2017)	16S rRNA amplicon sequencing	All	196	Bacteria
Denmark	Joensen et al. (2017)	Deep amplicon sequencing	0–95	58	Bacteria
Bangladesh	Kieser et al. (2018)	16S rRNA amplicon sequencing	0–2	71	Bacteria
Ethiopia	Aiemjoy et al. (2019)	RT-PCR/Illumina MiSeq	0–5	269	Viruses
China	Liu et al. (2019)	16S rDNA amplicon sequencing	0–1	20	Bacteria
Cameroun	Yinda et al. (2019)	Deep amplicon sequencing	0–89	221	Viruses



**Figure 2.** Data showing the high rate of diarrhoea morbidity among children below 5 years in Nigeria and some Asian Countries. Source: Computed by the Researcher with data from <https://data.unicef.org/topic/child-health/diarrhoeal-disease/> Accessed 17<sup>th</sup> February 2019.

## 2.2. Types of diarrhoea

### i. Acute watery diarrhoea

Acute watery diarrhoea often presents with sudden development of an unusual frequent stooling of mostly fluid. Other signs are vomiting, fever, nausea and abdominal pain (Dipasquale et al., 2018). In the gastrointestinal tract, absorption of over 90% of the physiologic net fluid takes place in the proximal small intestine. The pathogenic mechanism that leads to diarrhoea occurs when enteric pathogens attack the proximal small intestine (Thiagarajah et al., 2015). Acute watery diarrhoea is often caused by enterotoxin-secreting bacteria such as enterotoxigenic *Escherichia coli* (ETEC), and *Vibrio cholerae*, which cause fluid loss without cellular injury (Willey et al., 2013). Viruses such as rotaviruses and caliciviruses that damage the intestinal epithelium also cause fluid loss. Besides, they have more tendency to cause fever, vomiting and watery stools without blood and mucus (Tagbo et al., 2019). Usually, cases of watery diarrhoea run an acute but brief (1–3 days) self-limiting duration.

### ii. Dysentery

As defined by World Health Organization, dysentery is bloody diarrhoea, i.e. any diarrhoeal episode in which the loose or watery stools contain visible red blood (UNICEF and WHO, 2009). Dysentery is most often caused by *Shigella* species (bacillary dysentery) or *Entamoeba histolytica* (amoebic dysentery). Dysentery starts with the sudden onset of repeated stooling. However, unlike acute watery diarrhoea, stools are often smaller in quantity and are characterized by blood and pus. Thus, it is also referred to as acute bloody diarrhoea. Dysentery usually presents with fever, tenesmus, abdominal pain, and cramps; vomiting occurs less often (Wang et al., 2019). Inflammation of the colon (the part of the large intestine that extends from the cecum to the rectum) due to infection by one of a number of enteric pathogens leads to dysentery. The main cause of dysentery in children is the Shigellae (Tickell et al., 2017). *Campylobacter jejuni* and enteroinvasive *E. coli* or salmonellae of many serotypes being relatively less frequent causes. *Entamoeba histolytica* seldom causes dysentery in young children (Khan and Jahan, 2017; Delfino Vubil et al., 2018). Dysentery usually requires antimicrobial therapy (Williams and Berkley, 2018).

### iii. Persistent Diarrhoea

Persistent diarrhoea is acute as well as prolonged (at least 14 days) rather than brief also referred to as chronic diarrhoea (Centres for Disease Control and Prevention, 2016a, b). The case may begin with the passage of frequent watery or bloody stool but last for a long period thus causing loss of weight. It accounts for less than ten percent of all

diarrhoea but is responsible for 30 to 50 percent of death caused by diarrhoea (Black, 1993). Persistent diarrhoea has different causes which are either infectious or non-infectious. The infectious causes include intestinal parasites (Cryptosporidium, Cyclospora, *E. histolytica*, Giardia, Microsporidia), bacteria (*Aeromonas*, *Campylobacter*, *C. difficile*, *E. coli*, *Plesiomonas*, *Salmonella*, *Shigella*), and viruses (norovirus, rotavirus). While the non-infectious causes include altered immune function, disorders of the pancreas, medications (antibiotics), heritable metabolic disorders (enzyme deficiency), intolerance to some food products (gluten, lactose), intestinal disorders, disorders of the thyroid, and reduced blood flow to the intestines (Centres for Disease Control and Prevention, 2016a, b; Holtman et al., 2016; Spitz et al., 2016; El-Chammas et al., 2017). Major organisms that are responsible for persistent diarrhoea are enteroaggregative *E. coli*, *Shigella*, and *Cryptosporidium* (DuPont, 2016). The pathogenesis of persistent diarrhoea could be multifactorial and fundamentally based on continuous damage to the mucosal linings of the intestines due to several infections with different pathogens (Giannattasio et al., 2016). Malnutrition also increases the likelihood of death in children with persistent diarrhoea. Evidence-based studies have established a strong relationship between chronic diarrhoea and HIV-positive patients in developing countries (Agholi et al., 2013; Gebremedhin et al., 2013; Kumurya and Gwarzo, 2013; Rostami et al., 2014).

## 2.3. Aetiology of diarrhoeal diseases

The aetiology of diarrhoea may be infectious or non-infectious. In non-infectious diarrhoea cases, factors such as food intolerances (lactose and gluten), intestinal complications (irritable bowel syndrome, ulcerative colitis, Crohn's disease, and celiac disease), and reactions to drugs (Humphries and Linscott, 2015).

Before the late 1960s, less than 20% of enteric infection symptoms could be linked to a specific aetiological agent by any known diagnostic method (Ahmad et al., 2010). Diarrhoeal diseases are caused by several viral, bacterial, and protozoan species (Platts-Mills et al., 2015; Zhang et al., 2016). Co-infections by a spectrum of enteric pathogens is the norm in diarrhoeal diseases (Serrano and Millán, 2014; Becker et al., 2015). Rotaviruses and diarrhoeagenic *E. coli* (DEC) are the most reported enteropathogens globally, with the DEC being particularly important in resource low countries (Onanuga et al., 2014).

Evidence based studies from Sudan (Adam et al., 2018), Burkina Faso (Bonkoungou et al.; 2013; Ouedraogo et al., 2016), China (Zhang et al., 2016), Nigeria (Enitan et al., 2019), and other endemic regions reveals that a significant amount of diarrhoea episodes in children are caused by enteric viruses. Rotaviruses, Noroviruses, Adenoviruses, Bocaviruses and Caliciviruses have been implicated in childhood diarrhoea (Aktas et al., 2019). However, recent reports show rotaviruses as the major cause of fatal cases among children below 5 years old (Gatinu et al., 2016; Nnukwu et al., 2017; Giri et al., 2019). Moreover, group A rotavirus, in particular, is the prominent aetiological agent that is responsible for infantile gastroenteritis globally, causing an estimated 20% of diarrhoea-related deaths in children below the age of five. Low-income countries and those which have no running RVA vaccination programmes are particularly affected by the rotavirus group A (RVA) diarrhoea (Gatinu et al., 2016; Zhang et al., 2016; Crawford et al., 2017).

Diarrhoeagenic *E. coli* (DEC) has been grouped into six pathotypes based on their pathogenic processes and clinical features. These subtypes are enteropathogenic *E. coli* (EPEC), enterohemorrhagic *E. coli* (EHEC), enterotoxigenic *E. coli* (ETEC), enteroinvasive *E. coli* (EIEC), enteropathogenic *E. coli* (EAEC), and diffusely adherent *E. coli* (DAEC). DEC pathotypes have also been further categorised based on their virulence mechanisms (Yu et al., 2015; Zhang et al., 2016; Thakur et al., 2018). Also, there are important regional variations in the prevalence of the different DEC pathotypes. DEC pathotypes from different locations around the globe are genetically diverse (Yu et al., 2018).

In outbreak situations, *Vibrio cholerae*, *Escherichia coli* and *Clostridium difficile* have been reported as the causative agents of diarrhoea in Bangkok (Dalsgaard et al., 1999), China (Scheutz et al., 2011; Escher et al., 2014), Yemen (Camacho et al., 2018; Weill et al., 2019), Iran (Tajeddin et al., 2019) and Nigeria (Eko et al., 1994; Lawoyin et al., 1999; Usman et al., 2005; Elimian et al., 2019). The role of viruses as agents of diarrhoea outbreaks among children cannot be overemphasized. Astroviruses in Bangkok (Sirinavin et al., 2006), Noroviruses in China (Rogers et al., 2019), and Rotaviruses in India and Botswana (Barman et al., 2006; Erick, 2020) have been implicated in childhood diarrhoea outbreaks.

The parasitic diarrhoeal diseases of public health importance are Amebiasis (*Entamoeba histolytica*), Cryptosporidiosis (*Cryptosporidium spp*) and Giardiasis (*Giardia lamblia*). The two predominant causes that have been reported are *Cryptosporidium* and *Giardia*, of which *Cryptosporidium* is the more medically important across the world (Kotloff et al., 2013; Squire and Ryan, 2017; Troeger et al., 2017). However, studies from China (Yu et al., 2019), Ghana (Nkrumah and Nguah, 2011), Nigeria (Ighobgoja and Ikeh, 1997; Akingbade et al., 2013), Kenya (Mbae et al., 2013) and Buzigi and Uganda, 2015 have detected other enteric parasites such as *G. duodenalis*, *E. bieneusi*, *Schistosoma mansoni* and *C. cayetanensis* among diarrhoeic children, though at very low proportion when compared to bacteria and viruses.

#### 2.4. Mode of transmission of diarrhoea

Diarrhoea-causing pathogens are usually spread via the faecal-oral route through the consumption of food or fluid infected with faecal matter, or direct contact with infected stools (Pires et al., 2015; Kirk et al., 2017).

#### 2.5. Risk factors for diarrhoea in children

##### i. Duration of breastfeeding

Breastfeeding has a huge impact on the health status of children. A significant correlation exists between breastfeeding and diarrhoea episodes (Godana and Mengistie, 2013; Siregar et al., 2018). Sub-optimal breastfeeding increases the risk of developing diarrhoea because breast milk can confer proper functioning of the gut immune system in infants, both for a short- and long-term duration (Bartick et al., 2017; Ogbo et al., 2018). Also, breast milk decreases a newborn's risk of contracting gut diseases because it is made up of antibodies, immunoglobulin A (IgA), which confer protection against pathogenic bacteria and harmonize the activity of white blood cells (Willey et al., 2013; Hennet and Borsig, 2016).

In Africa, the rate of breastfeeding in Nigeria is lower than that in Uganda, Ethiopia, and Tanzania (Ogbo et al., 2017). This would mean that there might be a higher risk of infants presenting with diarrhoea in Nigeria than in the other countries of comparison (Figure 2). Infants should be given breast milk within the first hour after birth and breastfeeding should be exclusively practised during the first six months before feeding with supplementary foods (WHO, 2002). The initiation of supplementary food before the end of the first six months increases the risk of contamination, especially in the less developed countries where potable water and basic sanitation is lacking (Desmennu et al., 2017).

##### ii. The Source of drinking water

Consumption of contaminated water is a viable means of transmission of diarrhoea-causing pathogens. Contamination may occur at the water source, during storage by unhygienic packaging, or during mealtimes through contact with unwashed hands or exposure (Pires et al., 2015; Wasihun et al., 2018). The availability of potable water may not necessarily be taken for granted in every part of the world; but the challenge of insufficient or outright lack of potable water is worse in less developed countries, thus predisposing their populations to a higher burden of

diarrhoeal diseases (Omole et al., 2015). Unhygienic handling of drinking water is also an attributable factor for diarrhoea in children (Oloruntoba et al., 2014; Chakravarty et al., 2017).

##### iii. Hygiene and Sanitation

Non-adherence of mothers/caregivers to the practice of handwashing predisposes their children/wards to diarrhoea (Chakravarty et al., 2017). Infant feeding bottles, which can be easily contaminated with faecal bacteria may, in turn, contaminate milk held in them, and milk encourages bacteria growth if not consumed immediately (Oloruntoba et al., 2014; Joshua et al., 2016; Wasihun et al., 2018). Also, unhygienic sewage disposal and the use of crude toilet facilities may predispose children to diarrhoea (Kapwata et al., 2018).

##### iv. Age

Globally, the prevalence of diarrhoea has been reported to be higher in children than in adults, especially among children below five years (Walker et al., 2013; Charyeva et al., 2015; Zhang et al., 2016). Diarrhoea incidence among children has been reported to be highest in the age group below 24 months and declines with an increase in age (Walker et al., 2012; Akinnibosun and Nwafor, 2015).

##### v. The Level of Maternal Educational

The impact of the educational level of mothers in the health status of their children is important because it is directly related to their awareness levels. Mothers without formal education are more likely to have children who will suffer diarrhoeal diseases when compared to educated mothers (Desmennu et al., 2017). Moreover, because access to formal education is limited in rural areas where more males than females go to school, there is a higher rate of uneducated mothers in such locations thus increasing the risk of children having diarrhoea in rural environments (Jolaiya Tolu et al., 2016).

Apart from the factors listed above, vitamin A deficiency (Elalfy et al., 2014; Stevens et al., 2015), zinc deficiency (Walker et al., 2013; Troeger et al., 2020), childhood wasting (Mokomane et al., 2018; Troeger et al., 2018), low use of ORS (Charyeva et al., 2015), mother's handwashing (Abuzerr et al., 2019; Alemayehu et al., 2020), childhood stunting (Larsen et al., 2017) and low rotavirus vaccine coverage (Troeger et al., 2020) have been indicated as risk for childhood diarrhoea in developing countries.

### 3. Approaches to diarrhoeal diseases diagnosis

The polymicrobial nature of diarrhoea aetiology requires the medical microbiologist to harness the differential diagnosis towards establishing the causative agent of the disease (Panchalingam et al., 2012; Humphries and Linscott, 2015). Accurate detection of the etiologic agent is a very important step for diarrhoeal disease surveillance and control activities (Tarr et al., 2018).

#### 3.1. Conventional diagnostics

The traditional approach to the diagnosis of infectious diarrhoea includes Gram staining and microscopic examination, plate culture (and records of morphological characteristics), toxin assay, antigen-antibody assay, and biochemical testing (Ahmad et al., 2010). They are still being routinely used in clinical laboratories to detect and identify enteric pathogens (Liu et al., 2011). They have good sensitivity and specificity (Platts-Mills et al., 2013), but are quite laborious (Loman et al., 2013), because they must include a battery of methods to detect a spectrum of potential viruses, bacteria, and parasites (Chang et al., 2013). Also, time of identification takes longer, a factor which may threaten the chance of

proper antibiotic and supportive therapy and ultimately of survival for a patient in a critical condition.

Laboratory diagnosis that is culture-based often produce low yield for enteropathogens and this may hamper antibiotic therapy (Forbes et al., 2017). Although stool microscopy for parasite detection is broadly used, it is, however, insensitive, time-consuming, and requires equipment and training (Panchalingam et al., 2012; DuPont, 2016). Moreover, bacteria referred to as 'viable but not culturable' (VBNC), as well as, 'difficult to culture' (DTC), are often missed out of the identification process. It has been estimated that only about 1% of bacteria are culturable (Allan, 2014).

### 3.2. Molecular diagnosis of diarrhoeal diseases

Molecular diagnostics, which generally utilise the amplification of DNA or RNA, are increasingly relevant in infectious disease diagnosis. Molecular epidemiology uses the genetic sequence of microorganisms and their hosts to describe the disease patterns as well as gain insight into possible gene function and origin by describing the distribution of genes or gene according to person, place, and time characteristics. Microbial DNA can be detected in all types of samples. The two approaches to molecular detection of microbes are the culture-based and culture-independent methods.

Culture-dependent molecular methods require that microbes are grown in culture media in the laboratory under optimum growth requirements before DNA is extracted for further molecular processing. On the other hand, culture-independent molecular methods do not require microbes to be cultivated on culture plates for DNA isolation. Rather, DNA is extracted directly from samples (Gosalbes et al., 2012; Loman et al., 2013).

### 3.3. Polymerase chain reaction

Polymerase chain reaction (PCR) is by far the most widely used method for nucleic acid amplification. Among several biomarkers for pathogen detection, nucleic acids are the ultimate. Molecular targets for many enteropathogens are known. Some researchers have utilised multiplex PCR (Liu et al., 2011; Onanuga et al., 2014), array singleplex PCR (Liu et al., 2014a, b; Platts-Mills et al., 2013), and quantitative PCR (DuPont, 2016) in diarrhoeal research.

### 3.4. Metagenomics

Metagenomics, which is the culture-independent genomic analysis of microbial communities, has emerged as an influential new research tool in microbiology over the last twenty years (Allan, 2014). There are basically two approaches, the deep amplicon sequencing (DAS) or metagenome shotgun sequencing (MSS) (Miller et al., 2013). In diarrhoea diagnosis, the Bacterial DAS strategy typically entails the use of universal primers such as 16S rRNA (Miller et al., 2013; Decuyper et al., 2016). On the other hand, shotgun sequencing utilizes the process of randomly breaking up DNA sequences into many small pieces and then rearranging them by targeting regions of overlap, thus generating sequencing data that are naturally immune to primer bias (Hao and Chen, 2012). The shotgun approach has a wider coverage in terms of application in microbial community studies.

Other molecular biology techniques deal with the extraction of genomic DNA from an individual organism or cell from a pure isolate, whereas, metagenomics focuses on the direct investigation of total genomic DNA from clinical specimens. Rich phylogenetic information can be harnessed from amplicons of either or both of the two variant regions V3 and/or V6 of 16S ribosomal DNA (rDNA) of the bacteria in samples when pyro-sequenced (Miller et al., 2013).

Unlike the DAS, shotgun metagenomic sequencing approach can potentially detect all of the microbes present in a sample, despite their kingdom of origin, by sequencing all the nucleic acids extracted from a

specimen (Zhou et al., 2016). Therefore, its application in the analysis of the diversity and the metabolic potential of microbial communities is indispensable, especially as only a few microorganisms in nature are culturable (Padmanabhan et al., 2013; Loman et al., 2013). Shotgun metagenomics has the potential to detect completely novel genes (Fredericks, 2013). Apart from the detection of enteric pathogens, shotgun metagenomics has other applications in diarrhoea epidemiology. It can be used to investigate the phylogenetic diversity of microbial genes or gene products, antimicrobial resistance genes, and virulence genes (Hu et al., 2013; Allan, 2014; Zhou et al., 2016; Braun et al., 2017).

Recent reports from around the world suggest the potential of metagenomics as a useful tool in diarrhoea diagnosis (Becker-Dreps et al., 2015; Sun et al., 2016; Kieser et al., 2018; Aiemjoy et al., 2019). Bender and Dien Bard (2018), predict a rising frequency in the application of metagenomic techniques in paediatric medicine. Enteric pathogens, including viruses, bacteria, and protozoans have been identified using different metagenomics methods such as micro-mass sequencing, pyro-sequencing, amplicon sequencing, shotgun sequencing, and whole-genome sequencing (Table 1). Despite the many advantages of metagenomics in pathogen detection, the major drawback is the enormous data that are often generated which require in-depth bioinformatics software application.

### 3.5. Bioinformatics

A very crucial part of metagenomics technique is the application of bioinformatics tools, especially for data analysis. The application of gene sequencing in epidemiology brings about huge data (Padmanabhan et al., 2013). Bioinformatics is the discipline that focuses on the development of ways to use computer software's to characterise molecular components of living things. In the past few decades, molecular epidemiology focused on strain typing for outbreak and surveillance investigation. The ability to analyse genetic sequences from microbes has transformed the molecular epidemiology of infectious diseases. Therefore, modern molecular epidemiology relies on new phylogenetic methods that enable the analysis of genetic data to estimate epidemiologic parameters and link epidemic processes to pathogen evolution (Foxman and Goldberg, 2010; Gosalbes et al., 2012).

Recent studies reveal the significance of bioinformatics tools in the diagnosis of clinically challenging cases such as asymptomatic clinical presentations (Bodian et al., 2017; Pathak et al., 2019). Besides, there are pieces of evidence showing that bioinformatics is a very reliable epidemiological tool during disease and antimicrobial resistance surveillance (Bessoff et al., 2013; Gardner and Hall, 2013; Kaiser et al., 2016; Yodmeeklin et al., 2017). Also, with bioinformatics applications, scientists are already discovering new drug targets for the treatment of diarrhoeal diseases (Allan, 2014; Ugboko et al., 2019).

## 4. Treatment and management of diarrhoeal diseases

The common treatment options for diarrhoeal diseases are fluid replacement and antimicrobial therapy. Fluid replacement therapy also referred to as oral rehydration therapy (ORT) is especially necessary for young children (Iannotti et al., 2015; Bruzzese et al., 2018). In Nigeria, cereal-based oral therapies and home-made fluids have proven to be effective in diarrhoea management (Peter and Umar, 2018).

In some instances, diarrhoea cases may be self-limiting. However, during severe cases of infection (persistent diarrhoea or dysentery), antimicrobial agents are required (Breurec et al., 2016). The first line antimicrobial agents for childhood diarrhoea therapy are co-trimoxazole and metronidazole which could be administered empirically. Others include penicillin, erythromycin, amoxicillin, ampicillin, cefuroxime, ceftriaxone, tetracycline, chloramphenicol, and ampicillin/cloxacillin, azithromycin, ciprofloxacin, and rifaximin (Udoh and Meremikwu, 2017; Bruzzese et al., 2018). Howteerakul et al. (2004), reported co-trimoxazole, norfloxacin, colistin sulfate,

and nalidixic acid as the frequently recommended antibiotics for childhood therapy in Thailand. A study in Nigeria showed that metronidazole is the most prescribed antibiotic, followed by co-trimoxazole and gentamycin (Udoh and Meremikwu, 2017). Parenteral treatment with ceftriaxone or ciprofloxacin is recommended for severe diarrhoea cases (Bruzze et al., 2018). Childhood diarrhoea caused by *Shigella* is responsible for most cases of mortality in non-bloody diarrhoea (Liu et al., 2016a, b) and morbidity in moderate diarrhoea (Anderson et al., 2019), among children under five years in developing countries.

However, the emergence/re-emergence of antibiotic-resistant strains of enteric pathogens is becoming a huge threat (Willey et al., 2013). Worldwide, there are reports on multidrug-resistant strains of enteropathogens isolated from stools of children under five years (Elsherif et al., 2016). For instance, multidrug resistant *Shigella* spp has been reported in Ethiopia (Gebreegziabher et al., 2018), Mozambique (Vubil et al., 2018), and Nigeria (Ajayi et al., 2019).

Other treatment options include immunotherapy which is an alternative treatment option (Thu et al., 2017; Nagata et al., 2018; Zhao et al., 2019), fortified nutrition such as iron fortification and zinc replacement therapy (Paganini et al., 2016; Van Der Kam et al., 2016; Wessells et al., 2018), lactose-free diet (Iannotti et al., 2015), probiotics, though is limited awareness in Nigeria (Ajanya et al., 2018; Mokomane et al., 2018; Efunshile et al., 2019), and faecal microbiota transplantation which has been used mostly in *Clostridium difficile*-associated diarrhoea (Austin et al., 2014; Colman and Rubin, 2014; Barnes and Park, 2017). Oral zinc therapy could help reduce the severity and duration of diarrhoea in children above six months but may not have significant effect in children under six months of age (Lazzerini and Wanzira, 2016). Besides, Zinc absorption during diarrhoea may be reduced and vomiting may be a possible side effect (Ogunlesi et al., 2017; Somji et al., 2019). In Bangladesh, a combination of ORS, zinc and vitamin A supplementation was very effective at reducing death in children caused by diarrhoea (Billah et al., 2019).

#### 4.1. Prevention and management of diarrhoeal infections

An adequate supply of potable water, sanitation, and vaccination are the main means of preventing diarrhoeal infections (Desmennu et al., 2017).

##### a) Improved Water Supply and Sanitation

Acute diarrhoeal diseases can be prevented with a variety of measures focused on limiting the spread of organisms within the community and from person to person. Diarrhoeal diseases spread by the faecal-oral route. Therefore, handwashing is considered a key barrier to the transmission of enteric pathogens (Wolf et al., 2018). Studies from the developing world and from the U.S. and Australia childcare settings estimated 42%–47% reduction in the risk of diarrhoeal diseases by handwashing with soap (Bennett et al., 2014). Therefore, effective handwashing with soap should be practised by parents and caregivers before food preparation, before feeding their children and after leaving the toilet (Oloruntoba et al., 2014; Centres for Disease Control and Prevention, 2016a, b).

Globally, over 1 billion people lack access to improved drinking water supplies (Willey et al., 2013) and over 2.4 billion live without adequate sanitation (Darvesh et al., 2017). Strategies to improve the microbial quality of drinking water can be applied at the source or in the household. Water source strategy includes protected wells, boreholes and public tap stands (Chakravarty et al., 2017). Household strategies include improved water storage or approaches for treating water, such as chlorination, solar disinfection, filtration, or combined flocculation and disinfection (Darvesh et al., 2017).

##### b) Vaccines

Immunization has reduced the burden of diarrhoea in children. However, there are limited vaccines which protect against few and specific pathogens. There are two enteric infectious agents for which vaccines have been licensed for use at the present time, they include rotavirus and *V. cholerae* O1 (Preidis et al., 2011).

#### 4.2. Rotavirus vaccine

Rotavirus vaccination has been very effective in reducing the hospitalization and death rate caused by diarrhoea in Africa. Studies by Das et al. (2013), showed commendable reduction in rates of death (74%) and hospitalization (47–57%) caused by rotavirus in childhood diarrhoea. There are two types: a pentavalent rotavirus vaccine recommended for routine use in infants with three doses given at two, three, and six months respectively; and a monovalent rotavirus vaccine recommended as an alternative of two doses given at two and four months (Shah et al., 2017; Weldegebriel et al., 2018; Mwenda et al., 2018). Rotarix and RotaTeq manufactured in Belgium and New Jersey respectively have been effective in the prevention of diarrhoea (Mokomane et al., 2018; Soares-Weiser et al., 2019). Rotavac and BRV-PV produced in India have also proved to be promising. BRV-PV, is a heat-stable, live, oral bovine rotavirus pentavalent vaccine (given at 6, 10, and 14 weeks), which has undergone trial in Niger with a 66.7% efficacy against infantile diarrhoea caused by rotavirus (Isanaka et al., 2017; Seck et al., 2017).

#### 4.3. The cholera vaccine

The two oral cholera vaccines that are available for use and pre-qualified by the WHO are the oral, whole-cell, killed *V. cholerae* O1 vaccine supplemented with the B subunit of cholera toxin and the O139 vaccine without supplemented B with names as Dukoral, Shanchol, Euvichol, and mORC-Vax (Riddle et al., 2018; Boeckmann et al., 2019). These vaccines are given in two or three doses depending on age and produce protective efficacy against the moderate and severe disease of between 60%–85%, for up to two to three years after vaccination. They are less effective in children below two years and produce a shorter duration of protection in these individuals (Leung et al., 2012). A field trial of the oral cholera vaccine was done in Mozambique, where two doses of Dukoral showed 78% protective efficacy against disease in a cholera outbreak.

#### 4.4. New enteric vaccines

Recent trends show evidence of novel enteric vaccines. These include ETEC, *Shigella*, and norovirus vaccines. In the past two decades, research has been ongoing in the area of vaccine development using several methods which include live attenuated, killed whole-cell, and subunit techniques. Some of the new vaccines are in the preclinical and clinical stage of development (Bourgeois et al., 2016; Mani et al., 2016; Tennant et al., 2016; Mokomane et al., 2018).

#### 5. Conclusion

Diarrhoeal diseases remain an existential threat to global public health, especially to the children of ages five and below in low- and middle-income nations of the world, where access and the cost of quality healthcare remain below par and beyond affordable respectively. Treatment of childhood diarrhoeal diseases should be done within shorter turnaround times than ever before, given the need to reduce mortality due to them, especially during epidemic outbreaks. Conventional culture techniques, time-tested as they are, remain limited in offering rapid diagnosis, especially as a fair proportion of diarrhoeal pathogens are difficult-to-culture and yet-to-be-identified. Metagenomics and bioinformatics, in particular, offer great potentials in achieving a rapid diagnosis and epidemiological surveillance of diarrhoeal diseases,

which in turn will lead to the outcome of rapid management, treatment, prevention, and control of diarrhoeal diseases.

## Declarations

### Author contribution statement

Harriet U. Ugboko; Obinna C. Nwinyi; Solomon U. Oranusi: Conceived and designed the experiments; Performed the experiments; Analyzed and interpreted the data; John O. Oyewale: Contributed reagents, materials, analysis tools or data; Harriet U. Ugboko: Wrote the paper.

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### Competing interest statement

The authors declare no conflict of interest

### Additional information

No additional information is available for this paper.

## References

- Abuzerr, S., Nasseri, S., Yunesian, M., Hadi, M., Zinszer, K., Mahvi, A.H., Mohammed, S.H., 2020. Water, sanitation, and hygiene risk factors of acute diarrhea among children under five years in the Gaza Strip. *J. Water, Sanit. Hyg. Dev.* 10 (1), 111–123.
- Adam, M.A., Wang, J., Enan, K.A., Shen, H., Wang, H., El Hussein, A.R., et al., 2018. Molecular survey of viral and bacterial causes of childhood diarrhea in Khartoum state, Sudan. *Front. Microbiol.* 9, 112.
- Agholi, M., Hatam, G.R., Motazedian, M.H., 2013. HIV/AIDS-associated opportunistic protozoal diarrhea. *AIDS Res. Hum. Retrovir.* 29 (1), 35–41.
- Ahmad, N., Drew, W.L., Plorde, J.J., 2010. Sherris Medical Microbiology, fifth ed. McGraw-Hill Companies, USA, pp. 929–931.
- Aiemjoy, K., Altan, E., Aragie, S., Fry, D.M., Phan, T.G., Deng, X., et al., 2019. Viral species richness and composition in young children with loose or watery stool in Ethiopia. *BMC Infect. Dis.* 19 (1), 53.
- Ajanya, B.U., Attah, F., Mahmud, M.E., Owolabi, B.I., Adetoro, R.O., Adeniyi, K.A., Oyibosman, K.A., 2018. Therapeutic potency of probiotics in the treatment of gastrointestinal parasites. *J. Publ. Health Dent.* 1 (2), 22–30.
- Ajayi, O.I., Ojo, D.A., Akindutu, P.A., Akintokun, A.K., Akinrototye, K.P., 2019. Prevalence and antibiotic resistance profiles of serotypes of *Shigella* species isolated from community children in Odeda local government, Ogun state. *J. Environ. Treat. Tech.* 7 (3), 270–281.
- Akingbade, O.A., Akinjinmi, A.A., Ezechukwu, U.S., Okerentugba, P.O., Okonko, I.O., 2013. Prevalence of intestinal parasites among children with diarrhoea in Abeokuta, Ogun State, Nigeria. *Researcher* 5 (9), 66–73.
- Akinnibosun, F.I., Nwafor, F.C., 2015. Prevalence of diarrhoea and antibiotic susceptibility test in children below 5 years at University of Benin Teaching Hospital, Nigeria. *Int. Res. J. Publ. Environ. Health* 2 (4), 49–55.
- Aktas, O., Aydin, H., Timurkan, M.O., 2019. A molecular study on the prevalence and coinfections of Rotavirus, Norovirus, Astrovirus and Adenovirus in children with gastroenteritis. *Minerva Pediatr.* 71 (5), 431–437.
- Alemayehu, B., Ayele, B.T., Kloos, H., Ambelu, A., 2020. Individual and community-level risk factors in under-five children diarrhea among agro-ecological zones in southwestern Ethiopia. *Int. J. Hyg. Environ. Health* 224, 113447.
- Allan, E., 2014. Metagenomics: unrestricted access to microbial communities. *Virulence* 5 (3), 397–398.
- Anderson IV, J.D., Bagamian, K.H., Muhib, F., Amaya, M.P., Laytner, L.A., Wierzbka, T., Rheingans, R., 2019. Burden of enterotoxigenic *Escherichia coli* and shigella non-fatal diarrhoeal infections in 79 low-income and lower middle-income countries: a modelling analysis. *Lancet Global Health* 7 (3), e321–e330.
- Austin, M., Mellow, M., Tierney, W.M., 2014. Fecal microbiota transplantation in the treatment of *Clostridium difficile* infections. *Am. J. Med.* 127 (6), 479–483.
- Barman, P., Ghosh, S., Samajdar, S., Mitra, U., Dutta, P., Bhattacharya, S.K., et al., 2006. RT-PCR based diagnosis revealed importance of human group B rotavirus infection in childhood diarrhoea. *J. Clin. Virol.* 36 (3), 222–227.
- Barnes, D., Park, K.T., 2017. Donor considerations in fecal microbiota transplantation. *Curr. Gastroenterol. Rep.* 19 (3), 10.
- Bartick, M.C., Jegier, B.J., Green, B.D., Schwarz, E.B., Reinhold, A.G., Stuebe, A.M., 2017. Disparities in breastfeeding: impact on maternal and child health outcomes and costs. *J. Pediatr.* 181, 49–55.
- Becker, S.L., Chatigre, J.K., Gohou, J.P., Coulibaly, J.T., Leuppi, R., Polman, K., et al., 2015. Combined stool-based multiplex PCR and microscopy for enhanced pathogen detection in patients with persistent diarrhoea and asymptomatic controls from Côte d'Ivoire. *Clin. Microbiol. Infect.* 21 (6), 591–e1.
- Becker-Dreps, S., Allali, I., Monteagudo, A., Vilchez, S., Hudgens, M.G., Rogawski, E.T., et al., 2015. Gut microbiome composition in young Nicaraguan children during diarrhoea episodes and recovery. *Am. J. Trop. Med. Hyg.* 93 (6), 1187–1193.
- Bender, J.M., Dien Bard, J., 2018. Metagenomics in pediatrics: using a shotgun approach to diagnose infections. *Curr. Opin. Pediatr.* 30 (1), 125–130.
- Bennett, J.E., Dolin, R., Blaser, M.J., 2014. Principles and Practice of Infectious Diseases, eighth ed., 1. Elsevier Health Sciences. Canada.
- Bessoff, K., Sateriale, A., Lee, K.K., Huston, C.D., 2013. Drug repurposing screen reveals FDA-approved inhibitors of human HMG-CoA reductase and isoprenoid synthesis that block *Cryptosporidium parvum* growth. *Antimicrob. Agents Chemother.* 57 (4), 1804–1814.
- Billah, S.M., Raihana, S., Ali, N.B., Iqbal, A., Rahman, M.M., Khan, A.N.S., et al., 2019. Bangladesh: a success case in combating childhood diarrhoea. *J. Global Health* 9 (2).
- Black, R.E., 1993. Persistent diarrhea in children in developing countries. *Pediatr. Infect. Dis. J.* 12, 751–761.
- Bodian, D.L., Vilboux, T., Hourigan, S.K., Jenevein, C.L., Mani, H., Kent, K.C., et al., 2017. Genomic analysis of an infant with intractable diarrhea and dilated cardiomyopathy. *Mol. Case Stud.* 3 (6), a002055.
- Boeckmann, M., Roux, T., Robinson, M., Areal, A., Durusu, D., Wernecke, B., et al., 2019. Climate change and control of diarrhoeal diseases in South Africa: priorities for action Connections between temperature and diarrhoeal disease. *SAMJ: S. Afr. Med. J.* 109 (6), 359–361.
- Bonkoungou, I.J.O., Haukka, K., Österblad, M., Hakanen, A.J., Traoré, A.S., Barro, N., Siiton, A., 2013. Bacterial and viral etiology of childhood diarrhea in Ouagadougou, Burkina Faso. *BMC Pediatr.* 13 (1), 36.
- Bourgeois, A.L., Wierzbka, T.F., Walker, R.I., 2016. Status of vaccine research and development for enterotoxigenic *Escherichia coli*. *Vaccine* 34 (26), 2880–2886.
- Braun, T., Di Segni, A., BenShoshan, M., Asaf, R., Squires, J.E., Barhom, S.F., et al., 2017. Fecal microbial characterization of hospitalized patients with suspected infectious diarrhea shows significant dysbiosis. *Sci. Rep.* 7 (1), 1088.
- Breurec, S., Vanel, N., Bata, P., Chartier, L., Farra, A., Favenne, L., et al., 2016. Etiology and epidemiology of diarrhea in hospitalized children from low income country: a matched case-control study in Central African Republic. *PLoS Neglected Trop. Dis.* 10 (1).
- Bruzzone, E., Giannattasio, A., Guarino, A., 2018. Antibiotic treatment of acute gastroenteritis in children. *F1000Research* 7, 193–196.
- Buziggi, E., Uganda, K., 2015. Prevalence of intestinal parasites, and its association with severe acute malnutrition related diarrhea. *J. Biol. Agric. Healthc.* 5 (2).
- Camacho, A., Bouhenia, M., Alyusfi, R., Alkohlani, A., Naji, M.A.M., de Radigès, X., et al., 2018. Cholera epidemic in Yemen, 2016–18: an analysis of surveillance data. *Lancet Global Health* 6 (6), e680–e690.
- Centres for Disease Control and Prevention, 2013. Antibiotic Resistance Threats in the United States. Atlanta, GA, 2013, pp. 36–37, p. 7.
- Centres for Disease Control and Prevention, 2016a. Water, Sanitation, and Environmentally-Related Hygiene. [https://www.cdc.gov/healthywater/hygiene/disease/chronic\\_diarrhea.html](https://www.cdc.gov/healthywater/hygiene/disease/chronic_diarrhea.html). (Accessed 30 January 2020).
- Centres for Disease Control and Prevention, 2016b. National Centre for Emerging and Zoonotic Infectious Diseases (NCEZID). Division of Foodborne, Waterborne, and Environmental Diseases. <https://www.cdc.gov/ncezid/dfwed/edeb/index.html>. (Accessed 19 July 2017).
- Chakravarty, I., Bhattacharya, A., Das, S.K., 2017. Water, sanitation and hygiene: the unfinished agenda in the world health organization south-east Asia region. *WHO S. East Asia J. Public Health* 6 (2), 22.
- Chang, S.S., Hsieh, W.H., Liu, T.S., Lee, S.H., Wang, C.H., Chou, H.C., et al., 2013. Multiplex PCR system for rapid detection of pathogens in patients with presumed sepsis—a systemic review and meta-analysis. *PLoS One* 8 (5), e62323.
- Charyeva, Z., Cannon, M., Oguntunde, O., Garba, A.M., Sambisa, W., Bassi, A.P., et al., 2015. Reducing the burden of diarrhea among children under five years old: lessons learned from oral rehydration therapy corner program implementation in Northern Nigeria. *J. Health Popul. Nutr.* 34 (1), 4.
- Chiyanji, H., Muma, J.B., Malama, S., Manyahi, J., Abade, A., Kwenda, G., Matee, M.I., 2017. Identification and antimicrobial resistance patterns of bacterial enteropathogens from children aged 0–59 months at the University Teaching Hospital, Lusaka, Zambia: a prospective cross-sectional study. *BMC Infect. Dis.* 17 (1), 117.
- Colman, R.J., Rubin, D.T., 2014. Fecal microbiota transplantation as therapy for inflammatory bowel disease: a systematic review and meta-analysis. *J. Crohn's Colitis* 8 (12), 1569–1581.
- Crawford, S.E., Ramani, S., Tate, J.E., Parashar, U.D., Svensson, L., Hagbom, M., et al., 2017. Rotavirus infection. *Nat. Rev. Dis. Prim.* 3, 17083.
- Dalsgaard, A., Forslund, A., Bodhidatta, L., Serichantalergs, O., Pitartangsi, C., Pang, L., et al., 1999. A high proportion of *Vibrio cholerae* strains isolated from children with diarrhoea in Bangkok, Thailand are multiple antibiotic resistant and belong to heterogeneous non-O1, non-O139 O-serotypes. *Epidemiol. Infect.* 122 (2), 217–226.
- Darvesh, N., Das, J.K., Vaivada, T., Gaffey, M.F., Rasanathan, K., Bhutta, Z.A., 2017. Water, sanitation and hygiene interventions for acute childhood diarrhea: a systematic review to provide estimates for the Lives Saved Tool. *BMC Publ. Health* 17 (4), 776.
- Das, J.K., Tripathi, A., Ali, A., Hassan, A., Dojosoeady, C., Bhutta, Z.A., 2013. Vaccines for the prevention of diarrhea due to cholera, shigella, ETEC and rotavirus. *BMC Publ. Health* 13 (3), S11.
- Decuyper, S., Meehan, C.J., Van Puyvelde, S., De Block, T., Maltha, J., Palpouguini, L., et al., 2016. Diagnosis of bacterial bloodstream infections: a 16S metagenomics approach. *PLoS Neglected Trop. Dis.* 10 (2), e0004470.

- Delfino Vubil, S.A., Quintò, L., Ballesté-Delpierre, C., Nhampossa, T., Kotloff, K., Levine, M.M., et al., 2018. Clinical features, risk factors, and impact of antibiotic treatment of diarrhoea caused by *Shigella* in children less than 5 years in Manhiça District, rural Mozambique. *Infect. Drug Resist.* 11, 2095.
- Desmennu, A.T., Oluwasanu, M.M., John-Akinola, Y.O., Oladunni, O., Adebowale, S.A., 2017. Maternal education and diarrhoea among children aged 0-24 months in Nigeria. *Afr. J. Reprod. Health* 21 (3), 27–36.
- Dipasquale, V., Corica, D., Gramaglia, S.M., Valenti, S., Romano, C., 2018. Gastrointestinal symptoms in children: primary care and specialist interface. *Int. J. Clin. Pract.* 72 (6), e13093.
- DuPont, H.L., 2016. Persistent diarrhoea: a clinical review. *Jama* 315 (24), 2712–2723.
- Efunshile, A.M., Ezeanosike, O., Nwangwu, C.C., König, B., Jokelainen, P., Robertson, L.J., 2019. Apparent overuse of antibiotics in the management of watery diarrhoea in children in Abakaliki, Nigeria. *BMC Infect. Dis.* 19 (1), 275.
- Eko, F.O., Udo, S.M., Antia-Obong, O.E., 1994. Epidemiology and spectrum of vibrio diarrheas in the lower cross river basin of Nigeria. *Cent. Eur. J. Publ. Health* 2 (1), 37–41.
- Elalfy, M.S., Elagoua, I.A., Ibrahim, F.A., AbdElmessieh, S.K., Gadallah, M., 2014. Intracranial haemorrhage is linked to late onset vitamin K deficiency in infants aged 2–24 weeks. *Acta Paediatr.* 103 (6), e273–e276.
- El-Chammas, K., Williams, S.E., Miranda, A., 2017. Disaccharidase deficiencies in children with chronic abdominal pain. *J. Parenter. Enteral Nutr.* 41 (3), 463–469.
- Elimian, K.O., Musah, A., Mezue, S., Oyebanji, O., Yennan, S., Jinadu, A., et al., 2019. Descriptive epidemiology of cholera outbreak in Nigeria, January–November, 2018: implications for the global roadmap strategy. *BMC Publ. Health* 19 (1), 1264.
- Elsherif, R.H., Ismail, D.K., El-Kholby, Y.S., Gohar, N.M., Elnagdy, S.M., Elkraly, O.A., 2016. Integron-mediated multidrug resistance in extended-spectrum β-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* isolated from fecal specimens in Egypt. *J. Egypt. Publ. Health Assoc.* 91 (2), 73–79.
- Enitan, S.S., Ihongbe, J.C., Ochei, J.O., Oluremi, A.S., Ajulibe, G.E., 2019. Detection of rotavirus and adenovirus Co-infection among apparently healthy school aged children in Ilisan-remo community of Ogun state, Nigeria. *Asian J. Pediatr. Res.* 1–12.
- Erick, P., 2020. Botswana: country report on children's environmental health. *Rev. Environ. Health*.
- Escher, M., Scavia, G., Morabito, S., Tozzoli, R., Maugliani, A., Cantoni, S., et al., 2014. A severe foodborne outbreak of diarrhoea linked to a canteen in Italy caused by enteroinvasive *Escherichia coli*, an uncommon agent. *Epidemiol. Infect.* 142 (12), 2559–2566.
- Finkbeiner, S.R., Allred, A.F., Tarr, P.I., Klein, E.J., Kirkwood, C.D., Wang, D., 2008. Metagenomic analysis of human diarrhoea: viral detection and discovery. *PLoS Pathog.* 4 (2), e1000011.
- Forbes, J.D., Knox, N.C., Ronholm, J., Pagotto, F., Reimer, A., 2017. Metagenomics: the next culture-independent game changer. *Front. Microbiol.* 8, 1069.
- Foxman, B., Goldberg, D., 2010. Why the human microbiome project should motivate epidemiologists to learn ecology. *Epidemiology* (Cambridge, Mass.) 21 (6), 757.
- Fredricks, D.N., 2013. The Human Microbiota: How Microbial Communities Affect Health and Disease. John Wiley & Sons.
- Gardner, S.N., Hall, B.G., 2013. When whole-genome alignments just won't work: kSNP v2 software for alignment-free SNP discovery and phylogenetics of hundreds of microbial genomes. *PLoS One* 8 (12), e81760.
- Gatinu, B.W., Kiulia, N.M., Nyachieo, A., Macharia, W., Nyangao, J., Irimu, G., 2016. Clinical features associated with group A rotavirus in children presenting with acute diarrhoea at Kenyatta national hospital, Nairobi, Kenya. *J. Virol. Emerg. Dis.* 2 (1).
- Gebreeziabher, G., Asrat, D., Hagos, T., 2018. Isolation and antimicrobial susceptibility profile of *Shigella* and *Salmonella* species from children with acute diarrhoea in Mekelle Hospital and Semen Health Center, Ethiopia. *Ethiop. J. Health Sci.* 28 (2), 197–206.
- Gebremedhin, A., Gebremariam, S., Haile, F., Weldearegawi, B., Decotelli, C., 2013. Predictors of mortality among HIV infected children on anti-retroviral therapy in Mekelle Hospital, Northern Ethiopia: a retrospective cohort study. *BMC Publ. Health* 13 (1), 1047.
- Giannattasio, A., Guarino, A., Vecchio, A.L., 2016. Management of children with prolonged diarrhoea. *F1000Research* 5.
- Giri, S., Nair, N.P., Mathew, A., Manohar, B., Simon, A., Singh, T., et al., 2019. Rotavirus gastroenteritis in Indian children < 5 years hospitalized for diarrhoea, 2012 to 2016. *BMC Publ. Health* 19 (1), 69.
- Godana, W., Mengistie, B., 2013. Determinants of acute diarrhoea among children under five years of age in Derashe District, Southern Ethiopia. *Rural Rem. Health* 13 (3).
- Gosalbes, M.J., Abellán, J.J., Durban, A., Pérez-Cobas, A.E., Latorre, A., Moya, A., 2012. Metagenomics of human microbiome: beyond 16S rDNA. *Clin. Microbiol. Infect.* 18, 47–49.
- Hao, X., Chen, T., 2012. OTU analysis using metagenomic shotgun sequencing data. *PloS One* 7 (11), e49785.
- Hennet, T., Borsig, L., 2016. Breastfed at Tiffany's. *Trends Biochem. Sci.* 41 (6), 508–518.
- Holtzman, G.A., Kranenberg, J.J., Blunker, M.H., Ott, A., Lisman-van Leeuwen, Y., Berger, M.Y., 2016. *Dientamoeba fragilis* colonization is not associated with gastrointestinal symptoms in children at primary care level. *Fam. Pract. cmw111.*
- Holtz, L.R., Cao, S., Zhao, G., Bauer, I.K., Denno, D.M., Klein, E.J., et al., 2014. Geographic variation in the eukaryotic virome of human diarrhea. *Virology* 468, 556–564.
- Howteerakul, N., Higginbotham, N., Dibley, M.J., 2004. Antimicrobial Use in Children under Five Years with Diarrhea in a central Region Province, Thailand.
- Hu, Y., Yang, X., Qin, J., Lu, N., Cheng, G., Wu, N., et al., 2013. Metagenome-wide analysis of antibiotic resistance genes in a large cohort of human gut microbiota. *Nat. Commun.* 4, 2151.
- Humphries, R.M., Linscott, A.J., 2015. Laboratory diagnosis of bacterial gastroenteritis. *Clin. Microbiol. Rev.* 28 (1), 3–31.
- Iannotti, L.L., Trehan, I., Clitheroe, K.L., Manary, M.J., 2015. Diagnosis and treatment of severely malnourished children with diarrhoea. *J. Paediatr. Child Health* 51 (4), 387–395.
- Ifeanyi, C.I.C., Bassey, B.E., Ikeneche, N.F., Al-Gallas, N., 2014. Molecular characterization and antibiotic resistance of *Salmonella* in children with acute gastroenteritis in Abuja, Nigeria. *J. Infect. Dev. Ctries.* 8 (6), 712–719.
- Ighogboja, I.S., Ike, E.I., 1997. Parasitic agents in childhood diarrhoea and malnutrition. *W. Afr. J. Med.* 16 (1), 36–39.
- Isanaka, S., Guindo, O., Langendorf, C., Matar Seck, A., Plikaytis, B.D., Sayinzoga-Makombe, N., et al., 2017. Efficacy of a low-cost, heat-stable oral rotavirus vaccine in Niger. *N. Engl. J. Med.* 376 (12), 1121–1130.
- Joensen, K.G., Engsbro, A.Ø., Lukjancenko, O., Kaas, R.S., Lund, O., Westh, H., Aarestrup, F.M., 2017. Evaluating next-generation sequencing for direct clinical diagnostics in diarrhoeal disease. *Eur. J. Clin. Microbiol. Infect. Dis.* 36 (7), 1325–1338.
- Jolaiya Tolu, F., Smith Stella, I., Coker Akitoye, O., 2016. Knowledge and assessment of parents on diarrhoea and its management in Lagos, Nigeria. *Int. J. Health Sci. Res.* 6 (7), 138–143.
- Joshua, I.A., Biji, B.D., Gobir, A.A., Aliyu, A.A., Onyemochi, A., Nmadu, A.G., et al., 2016. Social characteristics and risk factors for diseases among internally displaced persons: a study of stefano's foundation camp in Jos, Nigeria. *Arch. Med. Surg.* 1 (2), 42.
- Kaiser, P., Regoes, R.R., Hardt, W.D., 2016. Population dynamics analysis of ciprofloxacin-resistant *S. Typhimurium* cells in a mouse model for *Salmonella* diarrhoea. In: *Bacterial Persistence*. Humana Press, New York, NY, pp. 189–203.
- Kapwata, T., Mathee, A., le Roux, W., Wright, C., 2018. Diarrhoeal disease in relation to possible household risk factors in South African Villages. *Int. J. Environ. Res. Publ. Health* 15 (8), 1665.
- Keddy, K.H., Smith, A.M., Page, N.A., 2016. GEMS extend understanding of childhood diarrhoea. *Lancet* 388 (10051), 1252–1254.
- Khan, N.T., Jahan, N., 2017. Prevalence of *E. Histolytica* associated dysentery in children in satellite town, quetta. *Epidemiology (Sunnyvale)* 7 (290), 2161–1165.
- Kieser, S., Sarker, S.A., Sakwinska, O., Foata, F., Sultana, S., Khan, Z., et al., 2018. Bangladeshi children with acute diarrhoea show faecal microbiomes with increased *Streptococcus* abundance, irrespective of diarrhoea aetiology. *Environ. Microbiol.* 20 (6), 2256–2269.
- Kirk, M.D., Angulo, F.J., Havelaar, A.H., Black, R.E., 2017. Diarrhoeal disease in children due to contaminated food. *Bull. World Health Organ.* 95 (3), 233.
- Kotloff, K.L., Nataro, J.P., Blackwelder, W.C., Nasrin, D., Farag, T.H., Panchalingam, S., et al., 2013. Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. *Lancet* 382 (9888), 209–222.
- Kotloff, K.L., 2017. The burden and aetiology of diarrheal illness in developing countries. *Pediatr. Clin.* 64 (4), 799–814.
- Kumurya, A.S., Gwarzo, M.Y., 2013. Cryptosporidiosis in HIV infected patients with diarrhoea in Kano state, North-western Nigeria. *J. AIDS HIV Res.* 5 (8), 301–305.
- Larsen, D.A., Grisham, T., Slawsky, E., Narine, L., 2017. An individual-level meta-analysis assessing the impact of community-level sanitation access on child stunting, anaemia, and diarrhea: evidence from DHS and MICS surveys. *PLoS Neglected Trop. Dis.* 11 (6).
- Lawoyin, T.O., Ogunbodede, N.A., Olumide, E.A.A., Onadeko, M.O., 1999. Outbreak of cholera in Ibadan, Nigeria. *Eur. J. Epidemiol.* 15 (4), 365–368.
- Lazzerini, M., Wanzira, H., 2016. Oral zinc for treating diarrhoea in children. *Cochrane Database Syst. Rev.* 12 (12). CD005436.
- Leung, D.T., Rahman, M.A., Mohasin, M., Patel, S.M., Aktar, A., Khanam, F., et al., 2012. Memory B cell and other immune responses in children receiving two doses of an oral killed cholera vaccine compared to responses following natural cholera infection in Bangladesh. *Clin. Vaccine Immunol.* 19 (5), 690–698.
- Liu, H., Guo, M., Jiang, Y., Cao, Y., Qian, Q., He, X., et al., 2019. Diagnosing and tracing the pathogens of infantile infectious diarrhea by amplicon sequencing. *Gut Pathog.* 11 (1), 12.
- Liu, H., Shen, Y., Yin, J., Yuan, Z., Jiang, Y., Xu, Y., et al., 2014a. Prevalence and genetic characterization of *Cryptosporidium*, *Enterocytozoon*, *Giardia* and *Cyclospora* in diarrhoeal outpatients in China. *BMC Infect. Dis.* 14 (1), 25.
- Liu, J., Kabir, F., Manne, J., Lertsethakarn, P., Begum, S., Gratz, J., et al., 2014b. Development and assessment of molecular diagnostic tests for 15 enteropathogens causing childhood diarrhoeal multicentre study. *Lancet Infect. Dis.* 14 (8), 716–724.
- Liu, J., Kibiki, G., Maro, V., Maro, A., Kumburu, H., Swai, N., et al., 2011. Multiplex reverse transcription PCR Luminex assay for detection and quantitation of viral agents of gastroenteritis. *J. Clin. Virol.* 50 (4), 308–313.
- Liu, J., Platts-Mills, J.A., Juma, J., Kabir, F., Nkeze, J., Okoi, C., et al., 2016a. Use of quantitative molecular diagnostic methods to identify causes of diarrhoea in children: a reanalysis of the GEMS case-control study. *Lancet* 388 (10051), 1291–1301.
- Liu, L., Johnson, H.L., Cousins, S., Perin, J., Scott, S., Lawn, J.E., et al., 2012. Global, regional, and national causes of child mortality: an updated systematic analysis for 2010 with time trends since 2000. *Lancet* 379 (9832), 2151–2161.
- Liu, L., Ozza, S., Hogan, D., Chu, Y., Perin, J., Zhu, J., et al., 2016b. Global, regional, and national causes of under-5 mortality in 2000–15: an updated systematic analysis with implications for the Sustainable Development Goals. *Lancet* 388 (10063), 3027–3035.
- Loman, N.J., Constantinidou, C., Christner, M., Rohde, H., Chan, J.Z.M., Quick, J., et al., 2013. A culture-independent sequence-based metagenomics approach to the investigation of an outbreak of Shiga-toxicogenic *Escherichia coli* O104: H4. *Jama* 309 (14), 1502–1510.
- MacCannell, D., 2013. Bacterial strain typing. *Clin. Lab. Med.* 33 (3), 629–650.

- Maciel, I.A., Leite, J.P.G., 2018. Use of quantitative molecular diagnostic methods to assess the aetiology, burden, and clinical characteristics of diarrhoea in children in low-resource settings: a reanalysis of the MAL-ED cohort study. *Lancet Global Health* 6, e1309–e1318.
- Mani, S., Wierzbka, T., Walker, R.I., 2016. Status of vaccine research and development for Shigella. *Vaccine* 34 (26), 2887–2894.
- Mbae, C.K., Nokes, D.J., Mulinge, E., Nyambura, J., Waruru, A., Kariuki, S., 2013. Intestinal parasitic infections in children presenting with diarrhoea in outpatient and inpatient settings in an informal settlement of Nairobi, Kenya. *BMC Infect. Dis.* 13 (1), 243.
- Miller, R.R., Montoya, V., Gardy, J.L., Patrick, D.M., Tang, P., 2013. Metagenomics for pathogen detection in public health. *Genome Med.* 5 (9), 81.
- Mokomane, M., Kasvosve, I., Melo, E.D., Pernica, J.M., Goldfarb, D.M., 2018. The global problem of childhood diarrhoeal diseases: emerging strategies in prevention and management. *Therapeut. Adv. Infect. Dis.* 5 (1), 29–43.
- Monira, S., Nakamura, S., Gotoh, K., Izutsu, K., Watanabe, H., Alam, N.H., et al., 2013. Metagenomic profile of gut microbiota in children during cholera and recovery. *Gut Pathog.* 5 (1), 1.
- Mwenda, J.M., Parashar, U.D., Cohen, A.L., Tate, J.E., 2018. Impact of rotavirus vaccines in Sub-Saharan African countries. *Vaccine* 36 (47), 7119–7123.
- Nagata, Y., Yamamoto, T., Kadowaki, M., 2018. Long-term effect of combined therapy of oral immunotherapy and Japanese traditional medicine kakkonto on food allergy mice. *J. Allergy Clin. Immunol.* 141 (2), AB242.
- Nakamura, S., Maeda, N., Miron, I.M., Yoh, M., Izutsu, K., Kataoka, C., et al., 2008. Metagenomic diagnosis of bacterial infections. *Emerg. Infect. Dis.* 14 (11), 1784.
- Nkrumah, B., Ngwuah, S.B., 2011. Giardia lamblia: a major parasitic cause of childhood diarrhoea in patients attending a district hospital in Ghana. *Parasites Vectors* 4 (1), 163.
- Nnukwu, S.E., Utsalo, S.J., Oyero, O.G., Ntemgwia, M., Ayukerbong, J.A., 2017. Point-of-care diagnosis and risk factors of infantile, rotavirus-associated diarrhoea in Calabar, Nigeria. *Afr. J. Lab. Med.* 6 (1), 1–5.
- Ogbo, F.A., Eastwood, J., Page, A., Efe-Aluta, O., Anago-Amanze, C., Kadiri, E.A., et al., 2017. The impact of sociodemographic and health-service factors on breast-feeding in sub-Saharan African countries with high diarrhoea mortality. *Publ. Health Nutr.* 20 (17), 3109–3119.
- Ogbo, F.A., Nguyen, H., Naz, S., Agho, K.E., Page, A., 2018. The association between infant and young child feeding practices and diarrhoea in Tanzanian children. *Trop. Med. Health* 46 (1), 2.
- Ogunlesi, T.A., Olowonyo, M.T., Runsewe-Abiodun, T.I., 2017. Pre-hospital use of oral rehydration therapy and zinc and the risk of dehydration in childhood diarrhoea. *J. Adv. Med. Med. Res.* 1–8.
- Oloruntoba, E.O., Folarin, T.B., Ayede, A.I., 2014. Hygiene and sanitation risk factors of diarrhoeal disease among under-five children in Ibadan, Nigeria. *Afr. Health Sci.* 14 (4), 1001–1011.
- Omole, D.O., Emenike, P.C., Tenebe, I.T., Akinde, A.O., Badejo, A.A., 2015. An assessment of water related diseases in a Nigerian community. *Res. J. Appl. Sci. Eng. Technol.* 10 (7), 776–781.
- Onanuga, A., Igbenegbu, O., Lamikanra, A., 2014. A study of the prevalence of diarrhoeagenic *Escherichia coli* in children from Gwagwalada, Federal Capital Territory, Nigeria. *Pan Afr. Med. J.* 17, 146.
- Ouedraogo, N., Kaplon, J., Bonkoungou, I.J.O., Traore, A.S., Pothier, P., Barro, N., Ambert-Balay, K., 2016. Prevalence and genetic diversity of enteric viruses in children with diarrhea in Ouagadougou, Burkina Faso. *PLoS One* 11 (4).
- Padmanabhan, R., Mishra, A.K., Raoult, D., Fournier, P.E., 2013. Genomics and metagenomics in medical microbiology. *J. Microbiol. Methods* 95 (3), 415–424.
- Paganini, D., Uyoga, M.A., Zimmermann, M.B., 2016. Iron fortification of foods for infants and children in low-income countries: effects on the gut microbiome, gut inflammation, and diarrhoea. *Nutrients* 8 (8), 494.
- Panchalingam, S., Antonio, M., Hossain, A., Mandomando, I., Ochieng, B., Oundo, J., et al., 2012. Diagnostic microbiologic methods in the GEMS-1 case/control study. *Clin. Infect. Dis.* 55 (suppl\_4), S294–S302.
- Pathak, S.J., Mueller, J.L., Okamoto, K., Das, B., Hertecant, J., Greenhalgh, L., et al., 2019. EPCAM mutation update: variants associated with congenital tufting enteropathy and Lynch syndrome. *Hum. Mutat.* 40 (2), 142–161.
- Peter, A.K., Umar, U., 2018. Combating diarrhoea in Nigeria: the way forward. *J. Microbiol. Exp.* 6 (4), 191–197.
- Petri Jr, W.A., Miller, M., Binder, H.J., Levine, M.M., Dillingham, R., Guerrant, R.L., 2008. Enteric infections, diarrhoea, and their impact on function and development. *J. Clin. Invest.* 118 (4), 1277.
- Pires, S.M., Fischer-Walker, C.L., Lanata, C.F., Devleesschauwer, B., Hall, A.J., Kirk, M.D., et al., 2015. Aetiology-specific estimates of the global and regional incidence and mortality of diarrhoeal diseases commonly transmitted through food. *PLoS One* 10 (12), e0142927.
- Platts-Mills, J.A., Babji, S., Bodhidatta, L., Gratz, J., Haque, R., Hatt, A., et al., 2015. Pathogen-specific burdens of community diarrhoea in developing countries: a multisite birth cohort study (MAL-ED). *Lancet Global Health* 3 (9), e564–e575.
- Platts-Mills, J.A., Liu, J., Houpt, E.R., 2013. New concepts in diagnostics for infectious diarrhoea. *Mucosal Immunol.* 6 (5), 876.
- Preidis, G.A., Hill, C., Guerrant, R.L., Ramakrishna, B.S., Tannock, G.W., Versalovic, J., 2011. Probiotics, enteric and diarrheal diseases, and global health. *Gastroenterology* 140 (1), 8–14.
- Ranjbar, R., Karami, A., Farshad, S., Giannanco, G., Mammina, C., 2014. Typing methods used in the molecular epidemiology of microbial pathogens: a how-to guide. *New Microbiol.* 37 (1), 1–15.
- Riddle, M.S., Chen, W.H., Kirkwood, C.D., MacLennan, C.A., 2018. Update on vaccines for enteric pathogens. *Clin. Microbiol. Infect.* 24 (10), 1039–1045.
- Rodulfo, H., Donato, M.D., Luiggi, J., Michelli, E., Millán, A., Michelli, M., 2012. Molecular characterization of *Salmonella* strains in individuals with acute diarrhoea syndrome in the State of Sucre, Venezuela. *Rev. Soc. Bras. Med. Trop.* 45 (3), 329–333.
- Rogers, B.R., Holmes, C.W., Hull, M., Westmoreland, D., Celma, C., Beard, S., et al., 2019. Persistent norovirus outbreaks in a hospital setting—The role of environmental contamination. *J. Infect.* 79 (3), 277–287.
- Rostami, M.N., Nikmanesh, B., Haghchi-Ashtiani, M.T., Monajemzadeh, M., Douraghi, M., Ghavavand, Z., Kashi, L., 2014. Isospora belli associated recurrent diarrhea in a child with AIDS. *J. Parasit. Dis.* 38 (4), 444–446.
- Scheutz, F., Nielsen, E.M., Frimodt-Møller, J., Boisen, N., Morabito, S., Tozzoli, R., et al., 2011. Characteristics of the enteroaggregative *Shiga* toxin/verotoxin-producing *Escherichia coli* O104: H4 strain causing the outbreak of haemolytic uraemic syndrome in Germany, May to June 2011. *Euro Surveill.* 16 (24), 19889.
- Schneeberger, P.H., Becker, S.L., Pothier, J.F., Duffy, B., N'Goran, E.K., Beuret, C., et al., 2016. Metagenomic diagnostics for the simultaneous detection of multiple pathogens in human stool specimens from Côte d'Ivoire: a proof-of-concept study. *Infect. Genet. Evol.* 40, 389–397.
- Seck, A.M., Isanaka, S., Guindo, O., Langendorf, C., Plikaytis, B.D., Sayinzoga-Makombe, N., et al., 2017. Efficacy of a low-cost, heat-stable oral rotavirus vaccine in Niger: a randomised, controlled trial. *F1000Research* 6.
- Serrano, E., Millán, J., 2014. What is the price of neglecting parasite groups when assessing the cost of co-infection? *Epidemiol. Infect.* 142 (7), 1533–1540.
- Shah, M.P., Tate, J.E., Mwenda, J.M., Steele, A.D., Parashar, U.D., 2017. Estimated reductions in hospitalizations and deaths from childhood diarrhoea following implementation of rotavirus vaccination in Africa. *Expert Rev. Vaccine* 16 (10), 987–995.
- Siregar, A.Y., Pitriyan, P., Walters, D., 2018. The annual cost of not breastfeeding in Indonesia: the economic burden of treating diarrhoea and respiratory disease among children (<24mo) due to not breastfeeding according to recommendation. *Int. Breastfeed. J.* 13 (1), 10.
- Sirinavin, S., Techasaeansiri, C., Okascharoen, C., Nuntrarumit, P., Tonsuttakul, S., Pongsuwan, Y., 2006. Neonatal astrovirus gastroenteritis during an inborn nursery outbreak. *J. Hosp. Infect.* 64 (2), 196–197.
- Smits, S.L., Rahman, M., Schapendonk, C.M., van Leeuwen, M., Faruque, A.S., Haagmans, B.L., et al., 2012. Calicivirus from novel Recovirus genogroup in human diarrhoea, Bangladesh. *Emerg. Infect. Dis.* 18 (7), 1192.
- Soares-Weiser, K., Bergman, H., Henschke, N., Pitani, F., Cunliffe, N., 2019. Vaccines for preventing rotavirus diarrhoea: vaccines in use. *Cochrane Database Syst. Rev.* 10.
- Somji, S.S., Dhingra, P., Dhingra, U., Dutta, A., Devi, P., Kumar, J., et al., 2019. Effect of dose reduction of supplemental zinc for childhood diarrhoea: study protocol for a double-masked, randomised controlled trial in India and Tanzania. *BMJ Paediatr. Open* 3 (1).
- Spitz, M.A., Nguyen, M.A., Roche, S., Heron, B., Milh, M., De Lonlay, P., et al., 2016. Chronic diarrhoea in L-amino acid decarboxylase (AADC) deficiency: a prominent clinical finding among a series of ten French patients. In: *JIMD Reports*, 31. Springer, Berlin, Heidelberg, pp. 85–93.
- Squire, S.A., Ryan, U., 2017. Cryptosporidium and Giardia in Africa: current and future challenges. *Parasites Vectors* 10 (1), 195.
- Stevens, G.A., Bennett, J.E., Hennocq, Q., Lu, Y., De-Regil, L.M., Rogers, L., et al., 2015. Trends and mortality effects of vitamin A deficiency in children in 138 low-income and middle-income countries between 1991 and 2013: a pooled analysis of population-based surveys. *Lancet Global Health* 3 (9), e528–e536.
- Sun, G., Zang, Q., Gu, Y., Niu, G., Ding, C., Zhang, P., 2016. Viral metagenomics analysis of picobirnavirus-positive faeces from children with sporadic diarrhoea in China. *Arch. Virol.* 161 (4), 971–975.
- Tagbo, B.N., Chukwubike, C.M., Ifeyinwa, R.I.R., Ani, E.O., 2019. Adenovirus and rotavirus associated diarrhoea in fewer than 5 children from enugu rural communities, South east Nigeria. *World J. Vaccine* 9 (3), 71.
- Tajeddin, E., Hasani, Z., Ganji, L., Gholam Mostafaei, F.S., Azimirad, M., Torabi, P., et al., 2019. Shiga toxin-producing bacteria as emerging enteric pathogens associated with outbreaks of foodborne illness in the Islamic Republic of Iran. *East. Mediterr. Health J.* 25.
- Tarr, G.A., Chui, L., Lee, B.E., Pang, X.L., Ali, S., Nettell-Aguirre, A., et al., 2018. Performance of stool-testing recommendations for acute gastroenteritis when used to identify children with 9 potential bacterial enteropathogens. *Clin. Infect. Dis.*
- Tennant, S.M., Steele, A.D., Pasetti, M.F., 2016. Highlights of the 8th international conference on vaccines for enteric diseases: the Scottish encounter to defeat diarrhoeal diseases. *Clin. Vaccine Immunol.* 23 (4), 272–281.
- Thakur, N., Jain, S., Changotra, H., Shrivastava, R., Kumar, Y., Grover, N., Vashist, J., 2018. Molecular characterization of diarrhoeagenic *Escherichia coli* pathotypes: association of virulent genes, serogroups, and antibiotic resistance among moderate-to-severe diarrhea patients. *J. Clin. Lab. Anal.* 32 (5), e22388.
- Thiagarajan, J.R., Donowitz, M., Verkman, A.S., 2015. Secretory diarrhoea: mechanisms and emerging therapies. *Nat. Rev. Gastroenterol. Hepatol.* 12 (8), 446.
- Thu, H.M., Myat, T.W., Win, M.M., Thant, K.Z., Rahman, S., Umeda, K., et al., 2017. Chicken egg yolk antibodies (IgY) for prophylaxis and treatment of rotavirus diarrhea in human and animal neonates: a concise review. *Kor. J. Food Sci. Anim. Resour.* 37 (1), 1.
- Tian, L., Zhu, X., Chen, Z., Liu, W., Li, S., Yu, W., et al., 2016. Characteristics of bacterial pathogens associated with acute diarrhea in children under 5 years of age: a hospital-based cross-sectional study. *BMC Infect. Dis.* 16 (1), 253.
- Tickell, K.D., Brander, R.L., Atlas, H.E., Pernica, J.M., Wilson, J.L., Pavlinac, P.B., 2017. Identification and management of Shigella infection in children with diarrhoea: a systematic review and meta-analysis. *Lancet Global Health* 5 (12), e1235–e1248.

- Troeger, C.E., Khalil, I.A., Blacker, B.F., Biehl, M.H., Albertson, S.B., Zimsen, S.R., et al., 2020. Quantifying risks and interventions that have affected the burden of diarrhoea among children younger than 5 years: an analysis of the Global Burden of Disease Study 2017. *Lancet Infect. Dis.* 20 (1), 37–59.
- Troeger, C., Blacker, B.F., Khalil, I.A., Rao, P.C., Cao, S., Zimsen, S.R., et al., 2018. Estimates of the global, regional, and national morbidity, mortality, and aetiologies of diarrhoea in 195 countries: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet Infect. Dis.* 18 (11), 1211–1228.
- Troeger, C., Forouzanfar, M., Rao, P.C., Khalil, I., Brown, A., Reiner Jr., R.C., et al., 2017. Estimates of global, regional, and national morbidity, mortality, and aetiologies of diarrhoeal diseases: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet Infect. Dis.* 17 (9), 909–948.
- Udoh, E.E., Meremikwu, M.M., 2017. Antibiotic prescriptions in the case management of acute watery diarrhea in under-fives. *Int. J. Contemp. Pediatr.* 4 (3), 691–695.
- Ugboko, H.U., Nwinyi, C.O., Oranusi, S.U., Fatoki, T.H., Akindutu, P.A., Enibukun, J.M., 2019. In silico screening and analysis of broad-spectrum molecular targets and lead compounds for childhood diarrhoea therapy. *Bioinf. Biol. Insights* 12, 1–11.
- UNICEF, WHO, 2009. Why Children Are Still Dying and what Can Be Done. The United Nations Children's Fund (UNICEF). World Health Organization (WHO), New York.
- United Nations International Children's Emergency Fund, 2018. <http://www.data.unicef.org/topic/child-health/diarrhoeal-disease/>. (Accessed 20 May 2018).
- Usman, A., Sarkinfada, F., Mufunda, J., Nyarango, P., Mansur, K., Daiyabu, T.M., 2005. Recurrent cholera epidemics in Kano-northern Nigeria. *Cent. Afr. J. Med.* 51 (3-4), 34–38.
- Van Der Kam, S., Salse-Ubach, N., Roll, S., Swarthout, T., Gayton-Toyoshima, S., Jiya, N.M., et al., 2016. Effect of short-term supplementation with ready-to-use therapeutic food or micronutrients for children after illness for prevention of malnutrition: a randomised controlled trial in Nigeria. *PLoS Med.* 13 (2), e1001952.
- Vubil, D., Balleste-Delpierre, C., Mabunda, R., Acácio, S., Garrine, M., Nhampossa, T., et al., 2018. Antibiotic resistance and molecular characterization of shigella isolates recovered from children aged less than 5 years in Manhiça, Southern Mozambique. *Int. J. Antimicrob. Agents* 51 (6), 881–887.
- Walker, C.L.F., Perin, J., Aryee, M.J., Boschi-Pinto, C., Black, R.E., 2012. Diarrhoea incidence in low-and middle-income countries in 1990 and 2010: a systematic review. *BMC Publ. Health* 12 (1), 220.
- Walker, C.L.F., Rudan, I., Liu, L., Nair, H., Theodoratou, E., Bhutta, Z.A., et al., 2013. Global burden of childhood pneumonia and diarrhoea. *Lancet* 381 (9875), 1405–1416.
- Wang, J., Sun, H., Qi, L., Shi, X.H., Zhou, Y., Min, K.Y., et al., 2019. The surveillance of Yersiniosis among children in central area of Beijing from 2011 to 2018. *Zhonghua yu fang yi xue za zhi [Chin. J.Prev. Med.]* 53 (10), 1027–1031.
- Wasihun, A.G., Dejene, T.A., Teferi, M., Marugán, J., Negash, L., Yemane, D., McGuigan, K.G., 2018. Risk factors for diarrhoea and malnutrition among children under the age of 5 years in the Tigray Region of Northern Ethiopia. *PloS One* 13 (11), e0207743.
- Weill, F.X., Domman, D., Njamkepo, E., Almesbahi, A.A., Naji, M., Nasher, S.S., et al., 2019. Genomic insights into the 2016–2017 cholera epidemic in Yemen. *Nature* 565 (7738), 230–233.
- Weldegebriel, G., Mwenda, J.M., Chakauya, J., Daniel, F., Masresha, B., Parashar, U.D., Tate, J.E., 2018. Impact of rotavirus vaccine on rotavirus diarrhoea in countries of East and Southern Africa. *Vaccine* 36 (47), 7124–7130.
- Wessells, K.R., Brown, K.H., Kounnavong, S., Barffour, M.A., Hinnouho, G.M., Sayasone, S., et al., 2018. Comparison of two forms of daily preventive zinc supplementation versus therapeutic zinc supplementation for diarrhoea on young children's physical growth and risk of infection: study design and rationale for a randomized controlled trial. *BMC Nutrition* 4 (1), 39.
- Willey, J., Sherwood, L., Wolverton, C., 2013. *Prescott's Microbiology*, ninth ed. McGraw-Hill, New York, pp. 51–84.
- Williams, P.C., Berkley, J.A., 2018. Guidelines for the treatment of dysentery (shigellosis): a systematic review of the evidence. *Paediatr. Int. Child Health* 38 (sup1), S50–S65.
- Wolf, J., Hunter, P.R., Freeman, M.C., Cumming, O., Clasen, T., Bartram, J., et al., 2018. Impact of drinking water, sanitation and handwashing with soap on childhood diarrhoeal disease: updated meta-analysis and meta-regression. *Trop. Med. Int. Health* 23 (5), 508–525.
- World Health Organization, 2002. *Infant and Young Child Nutrition: Global Strategy on Infant and Young Child Feeding*. Geneva, Switzerland. Available at: [http://apps.who.int/gb/archive/pdf\\_files/WHA55/ea5515.pdf?ua=1](http://apps.who.int/gb/archive/pdf_files/WHA55/ea5515.pdf?ua=1). (Accessed 23 July 2019).
- World Health Organization, 2007. *Diarrhoeal Disease*. <https://www.who.int/news-room/fact-sheets/detail/diarrhoeal-disease>. (Accessed 30 January 2020).
- Yinda, C.K., Vanhulle, E., Conceição-Neto, N., Beller, L., Deboutte, W., Shi, C., et al., 2019. Gut virome analysis of Cameroonian reveals high diversity of enteric viruses, including potential interspecies transmitted viruses. *mSphere* 4 (1) e00585-18.
- Yodmeeklin, A., Khamrin, P., Chuachaona, W., Kumthip, K., Kongkaew, A., Vachirachewin, R., et al., 2017. Analysis of complete genome sequences of G9P [19] rotavirus strains from human and piglet with diarrhea provides evidence for whole-genome interspecies transmission of nonreassorted porcine rotavirus. *Infect. Genet. Evol.* 47, 99–108.
- Youmans, B.P., Ajami, N.J., Jiang, Z.D., Campbell, F., Wadsworth, W.D., Petrosino, J.F., et al., 2015. Characterization of the human gut microbiome during travelers' diarrhea. *Gut Microb.* 6 (2), 110–119.
- Yu, F., Chen, X., Zheng, S., Han, D., Wang, Y., Wang, R., et al., 2018. Prevalence and genetic diversity of human diarrhoeagenic *Escherichia coli* isolates by multilocus sequence typing. *Int. J. Infect. Dis.* 67, 7–13.
- Yu, F., Li, D., Chang, Y., Wu, Y., Guo, Z., Jia, L., et al., 2019. Molecular characterization of three intestinal protozoans in hospitalized children with different disease backgrounds in Zhengzhou, central China. *Parasites Vectors* 12 (1), 543.
- Yu, J., Jing, H., Lai, S., Xu, W., Li, M., Wu, J., et al., 2015. Etiology of diarrhea among children under the age five in China: results from a five-year surveillance. *J. Infect.* 71 (1), 19–27.
- Zhang, S.X., Zhou, Y.M., Tian, L.G., Chen, J.X., Tinoco-Torres, R., Serrano, E., et al., 2018. Antibiotic resistance and molecular characterization of diarrheagenic *Escherichia coli* and non-typhoidal *Salmonella* strains isolated from infections in Southwest China. *Infect. Dis. Poverty* 7 (1), 53.
- Zhang, S.X., Zhou, Y.M., Xu, W., Tian, L.G., Chen, J.X., Chen, S.H., et al., 2016. Impact of co-infections with enteric pathogens on children suffering from acute diarrhoea in southwest China. *Infect. Dis. Poverty* 5 (1), 64.
- Zhao, W., Torabi, B., Duncan, L., Ke, D., Cohen, C.G., Ben-Shoshan, M., Mazer, B.D., 2019. Serological profile of children undergone cow's milk oral immunotherapy. *J. Allergy Clin. Immunol.* 143 (2), AB253.
- Zhou, Y., Wylie, K.M., El Feghaly, R.E., Mihindukulasuriya, K.A., Elward, G.A., et al., 2016. Metagenomic approach for identification of the pathogens associated with diarrhoea in stool specimens. *J. Clin. Microbiol.* 54 (2), 368–375.