





Genome Sequences of 12 Phages That Infect Klebsiella pneumoniae

Trever L. Thurgood, Ruchira Sharma, Jackson J. Call, Joshua D. Chronis, Daniel D. Dawson, Zachary K. Finnegan, a Kent W. Foster, Tyler Meek, Emily Potts, Michael R. Sirrine, Alex D. Atkinson, Jacob D. Fairholm, Yoga A. Handoko, b Kai Li Ong, a Olivia B. Tateoka, a Daniel K. Arens, a Liam Johnson, a Jared L. Kruger, a Emily Loertscher, a Daniel W. Thompson, a Jamison K. Walker, a Richard A. Robison, a Sherwood R. Casjens, c,d DJulianne H. Grosea

- ^aDepartment of Microbiology and Molecular Biology, Brigham Young University, Provo, Utah, USA
- ^bDepartment of Agrotechnology, Satya Wacana Christian University, Salatiga, Central Java, Indonesia
- cSchool of Biological Sciences, University of Utah, Salt Lake City, Utah, USA
- ^aDivision of Microbiology and Immunology, Department of Pathology, School of Medicine, University of Utah, Salt Lake City, Utah, USA

ABSTRACT Klebsiella pneumoniae is a pathogen responsible for significant proportions of nosocomial and health care-associated infections and is known to acquire multiple antibiotic resistance genes. Here, we announce the full genome sequences of 12 K. pneumoniae bacteriophages from samples collected in wastewater treatment facilities across the western United States.

Plebsiella pneumoniae is a pathogenic member of the Enterobacteriaceae family that is responsible for a significant proportion (>10%) of hospital-acquired infections annually, as well as many community-acquired infections in the United States (3 to 5%) (1). K. pneumoniae is also known to be involved in the dissemination of a major class of carbapenemase genes (K. pneumoniae carbapenemases [KPCs]), which has contributed to the global spread of bacterial antibiotic resistance (2, 3). Classic treatments for K. pneumoniae infections are losing efficacy in the face of rising rates of antibiotic resistance. Therefore, the study of alternative treatments such as bacteriophage therapy could be beneficial in the future.

Here, we report the complete genome sequences of 12 K. pneumoniae bacteriophages isolated from wastewater in the western United States. All phages were propagated on Klebsiella pneumoniae ATCC 13883. Phages were amplified from enrichment cultures using LB medium at 37°C, plated on LB top agar at 37°C, and purified through a minimum of three successive single-plaque isolations (4). Phage genomic DNA was isolated from high-titer lysates using the phage DNA isolation kit from Norgen Biotek (Canada). The Illumina TruSeg DNA Nano kit was used for genomic library preparation with unique barcodes, followed by sequencing on the Illumina HiSeq 2500 platform (250-bp paired-end reads) at the Brigham Young University DNA Sequencing Center (Provo, UT). All contigs were assembled de novo using Geneious (5) version R11 and were annotated using DNA Master (6) and GeneMarkS (7) gene prediction software; all software was used with default settings. These 12 phages circularized upon assembly, and base pair 1 was called by alignment with the closest published phage relative that was reported as a complete genome, using BLASTn (8).

The 12 phage genomes can be placed into three previously established Caudovirales clusters, or groups of phages having homology over >50% of the genome (9, 10), according to our previous cluster definitions for Enterobacteriaceae phages (11). The largest cluster, populated by vB_KpnS_Domnhall, vB_KpnS_IMGroot, vB_ KpnS_KingDDD, vB_KpnS_Call, vB_KpnS_SegesCirculi, vB_KpnS_Alina, and vB_ KpnS_Penguinator, shows at least 85% average nucleotide identity (ANI) (as determined by Kalign [12]) among all seven Siphoviridae phages (average genome size,

Citation Thurgood TL, Sharma R, Call JJ, Chronis JD, Dawson DD, Finnegan ZK, Foster KW, Meek T, Potts E, Sirrine MR, Atkinson AD, Fairholm JD, Handoko YA, Ong KL, Tateoka OB, Arens DK, Johnson L, Kruger JL, Loertscher E, Thompson DW, Walker JK, Robison RA, Casjens SR, Grose JH. 2020. Genome sequences of 12 phages that infect Klebsiella pneumoniae. Microbiol Resour Announc 9:e00024-20. https://doi.org/10.1128/MRA.00024-20.

Editor John J. Dennehy, Queens College Copyright © 2020 Thurgood et al. This is an

open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Julianne H. Grose, julianne grose@byu.edu.

Received 11 January 2020 Accepted 24 March 2020 Published 16 April 2020

Thurgood et al.

TABLE 1 Basic properties and accession numbers of 12 K. pneumoniae phages

	GenBank	SRA	Total no.	Fold coverage (range	Genome	No. of		GC content
Phage name	accession no.	accession no.	of reads	[avg read depth])	length (bp)	ORFs ^a	Taxonomy ^b	(%) ^c
vB_KpnP_Emp27	MN013074	SAMN13072788	3,139	183-430 (281)	38,603	45	Α	50.7
vB_KpnS_Domnhall	MN013075	SAMN12752290	441,095	41-250 (219.1)	54,438	90	W	51.6
vB_KpnS_IMGroot	MN013076	SAMN13155540	338,553	452-1,559 (988.7)	52,866	88	W	51.3
vB_KpnS_KingDDD	MN013078	SAMN13072790	35,976	110-357 (172)	51,562	83	W	51.6
vB_KpnS_Call	MN013079	SAMN13228337	208,987	630-1,495 (899.1)	51,487	82	W	51.5
vB_KpnS_SegesCirculi	MN013080	SAMN13228338	365,333	1,123-5,463 (1,764)	50,713	80	W	51.1
vB_KpnM_Potts1	MN013081	SAMN12752291	92,223	183-430 (281)	169,384	298	T	40.7
vB_KpnP_Sibilus	MN013082	SAMN13072791	45,438	2-287 (170.3)	40,171	53	Α	51.2
vB_KpnS_Alina	MN013083	SAMN13072792	22,399	44-199 (99.5)	51,780	83	W	51.6
vB_KpnP_NahiliMali	MN013085	SAMN13072794	45,628	117-732 (173.7)	39,556	52	Α	51.2
vB_Kpn_Chronis	MN013086	SAMN13072795	93,145	2-30 (14.6)	45,702	73	Р	52.3
vB_KpnS_Penguinator	MN013087	SAMN12752292	11,098	5–43 (21.6)	51,678	87	W	51.5

^a ORFs, open reading frames in the current annotation, including 8 tRNAs for vB_KpnM_Potts1 and 1 tRNA for vB_Kpn_Chronis.

52,075 \pm 1,219 bp), which are T1-like (11) phages. The second largest cluster consists of three T7-like (11) podovirus phages, vB_KpnP_Sibilus, vB_KpnP_NahiliMali, and vB_KpnP_Emp27, the latter of which represents its own subcluster, sharing 63% ANI with the former two phages, which share 92% ANI with each other (average genome size, 39,442 \pm 790 bp). The third cluster consists of one T4-like (11) *Myoviridae* phage, vB_KpnM_Potts1 (genome size, 169,384 bp). Phage vB_Kpn_Chronis is an unclassified temperate phage, with close relatives in many *K. pneumoniae* genomes, that forms a new cluster in the lambda-like supercluster (11). The division of these 12 phages into four clusters is consistent with the classifications outlined by the International Committee on Taxonomy of Viruses (ICTV) (13) (Table 1). T1-like and T4-like phages have been shown previously to package DNA by a headful mechanism and T7-like phages through direct terminal repeats, which is consistent with the apparently circular genomes achieved during phage assembly (14, 15).

Data availability. The accession numbers for all 12 phages are found in Table 1.

ACKNOWLEDGMENTS

We extend special thanks to the Howard Hughes Medical Institute Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) for support and training in phage analysis. We appreciate the help of Ed Wilcox (Brigham Young University DNA Sequencing Center).

This work was graciously funded by the Department of Microbiology and Molecular Biology and the College of Life Sciences at Brigham Young University, as well as by a private donation to the Brigham Young University Phage Hunters program.

REFERENCES

- Ashurst JV, Dawson A. 2020. Klebsiella pneumonia. StatPearls, Treasure Island, FL.
- Meyer KS, Urban C, Eagan JA, Berger BJ, Rahal JJ. 1993. Nosocomial outbreak of *Klebsiella* infection resistant to late-generation cephalosporins. Ann Intern Med 119:353–358. https://doi.org/10.7326/0003-4819 -119-5-199309010-00001.
- Munoz-Price LS, Poirel L, Bonomo RA, Schwaber MJ, Daikos GL, Cormican M, Cornaglia G, Garau J, Gniadkowski M, Hayden MK, Kumarasamy K, Livermore DM, Maya JJ, Nordmann P, Patel JB, Paterson DL, Pitout J, Villegas MV, Wang H, Woodford N, Quinn JP. 2013. Clinical epidemiology of the global expansion of *Klebsiella pneumoniae* carbapenemases. Lancet Infect Dis 13:785–796. https://doi.org/10.1016/S1473-3099(13) 70190-7.
- 4. Sauder AB, Quinn MR, Brouillette A, Caruso S, Cresawn S, Erill I, Lewis L,

- Loesser-Casey K, Pate M, Scott C, Stockwell S, Temple L. 2016. Genomic characterization and comparison of seven *Myoviridae* bacteriophage infecting *Bacillus thuringiensis*. Virology 489:243–251. https://doi.org/10.1016/j.virol.2015.12.012.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics 28:1647–1649. https://doi.org/10.1093/bioinformatics/ hts199.
- Lawrence J. 2007. DNA Master. http://cobamide2.bio.pitt.edu/computer .htm.
- 7. Besemer J, Lomsadze A, Borodovsky M. 2001. GeneMarkS: a self-training method for prediction of gene starts in microbial genomes: implications

Volume 9 Issue 16 e00024-20 mra.asm.org **2**

^b The following abbreviations are used for taxonomy, which is provided by whole-genome BLASTN (9) at >95% identity for species taxonomy and >50% identity for genus taxonomy, as recommended by the Bacterial and Archaeal Viruses Subcommittee of the ICTV: A, *Podoviridae*, *Autographivirinae*, unclassified *Teseptimavirus*; W, *Siphoviridae*, *Tunavirinae*, *Webervirus*; T, *Myoviridae*, *Tevenvirinae*, unclassified *Tevenvirinae*; P, unclassified *Podoviridae*. All 12 phages belong to the superkingdom of viruses and the order *Caudovirales*.

^c GC content for the genome.



- for finding sequence motifs in regulatory regions. Nucleic Acids Res 29:2607–2618. https://doi.org/10.1093/nar/29.12.2607.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic Local Alignment Search Tool. J Mol Biol 215:403–410. https://doi.org/10 .1016/S0022-2836(05)80360-2.
- Hatfull GF. 2010. Mycobacteriophages: genes and genomes. Annu Rev Microbiol 64:331–356. https://doi.org/10.1146/annurev.micro.112408 134233
- Hatfull GF, Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) Program, KwaZulu-Natal Research Institute for Tuberculosis and HIV (K-RITH) Mycobacterial Genetics Course, University of California-Los Angeles Research Immersion Laboratory in Virology, Phage Hunters Integrating Research and Education (PHIRE) Program. 2013. Complete genome sequences of 63 mycobacteriophages. Genome Announc 1:e00847-13. https://doi.org/10 .1128/genomeA.00847-13.
- 11. Grose JH, Casjens SR. 2014. Understanding the enormous diversity of

- bacteriophages: the tailed phages that infect the bacterial family *Enter-obacteriaceae*. Virology 468 470:421 443. https://doi.org/10.1016/j.virol.2014.08.024.
- 12. Lassmann T, Sonnhammer EL. 2005. Kalign: an accurate and fast multiple sequence alignment algorithm. BMC Bioinformatics 6:298. https://doi.org/10.1186/1471-2105-6-298.
- Lefkowitz EJ, Dempsey DM, Hendrickson RC, Orton RJ, Siddell SG, Smith DB. 2018. Virus taxonomy: the database of the International Committee on Taxonomy of Viruses (ICTV). Nucleic Acids Res 46:D708–D717. https://doi.org/10.1093/nar/gkx932.
- Casjens SR, Gilcrease EB. 2009. Determining DNA packaging strategy by analysis of the termini of the chromosomes in tailed-bacteriophage virions. Methods Mol Biol 502:91–111. https://doi.org/10.1007/978-1 -60327-565-1_7.
- Merrill BD, Ward AT, Grose JH, Hope S. 2016. Software-based analysis of bacteriophage genomes, physical ends, and packaging strategies. BMC Genomics 17:679. https://doi.org/10.1186/s12864-016-3018-2.

Volume 9 Issue 16 e00024-20 mra.asm.org **3**