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Low-dose Perampanel rescues cortical gamma dysregulation associated with parvalbumin interneuron GluA2 upregulation in epileptic *Syngap1*^{+/-} mice

Brennan J. Sullivan¹, Simon Ammanuel², Pavel A. Kipnis¹, Yoichi Araki³, Richard L. Haganir³, Shilpa D. Kadam^{1,4,*}

¹Neuroscience Laboratory, Hugo Moser Research Institute at Kennedy Krieger, Baltimore, MD, 21205, USA

²School of Medicine, University of California, San Francisco, 505 Parnassus Avenue, San Francisco, CA, 94143, USA.

³Department of Neuroscience, Kavli Neuroscience Discovery Institute, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA

⁴Department of Neurology, Johns Hopkins University School of Medicine, Baltimore, MD, 21205, USA;

Abstract

BACKGROUND: Loss-of-function *SYNGAP1* mutations cause a neurodevelopmental disorder characterized by intellectual disability and epilepsy. *SYNGAP1* is a Ras-GTPase-activating protein that underlies the formation and experience-dependent regulation of postsynaptic densities. The mechanisms that contribute to this proposed monogenic cause of intellectual disability and epilepsy remain unresolved.

METHODS: Here, we establish the phenotype of the epileptogenesis in a *Syngap1*^{+/-} mouse model using 24h video electroencephalogram/electromyogram (vEEG/EMG) recordings at advancing ages. We administered an acute low-dose of Perampanel, an FDA approved AMPAR antagonist, during a follow-on 24h vEEG to investigate the role of AMPARs in *SYNGAP1* haploinsufficiency. To determine the region- and location-specific differences in the expression of the GluA2 AMPAR subunit, immunohistochemistry was performed.

***Corresponding Author:** Shilpa D. Kadam, PhD, Hugo Moser Research Institute at Kennedy Krieger; Department of Neurology, Johns Hopkins University School of Medicine, 707 North Broadway, 400H; Baltimore, MD 21205, Phone: 443-923-2688, Fax: 443-923-2695, kadam@kennedykrieger.org.

Author Contributions

S.D.K. designed the experiments. S.D.K. and R.L.H. conceptualized the project. S.D.K. and B.J.S. performed experiments. Y.A. managed mouse colonies. B.J.S., S.A., and P.A.K. analyzed data. S.D.K. and B.J.S. interpreted the data. S.D.K. and B.J.S. wrote the manuscript. S.D.K., B.J.S., and P.A.K. edited and reviewed the manuscript.

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Competing Interests

The authors report no biomedical financial interests or potential conflicts of interest.

RESULTS: A progressive worsening of the epilepsy with emergence of multiple seizure phenotypes, interictal spike frequency, sleep dysfunction, and hyperactivity was identified in *Syngap1*^{+/-} mice. Interictal spikes emerged predominantly during NREM in 24h vEEG of *Syngap1*^{+/-} mice. Myoclonic seizures occurred at behavioral-state transitions both in *Syngap1*^{+/-} mice and during an overnight EEG from a child with *SYNGAP1* haploinsufficiency. In *Syngap1*^{+/-} mice, EEG spectral power analyses identified a significant loss of gamma power modulation during behavioral-state transitions. A significant region-specific increase of GluA2 AMPAR subunit expression in the somas of parvalbumin-positive (PV+) interneurons was identified.

CONCLUSIONS: Acute dosing with Perampanel significantly rescued behavioral-state dependent cortical gamma homeostasis, identifying a novel mechanism implicating Ca²⁺ impermeable AMPARs on PV+ interneurons underlying circuit dysfunction in *SYNGAP1* haploinsufficiency.

Keywords

Myoclonic seizures; gamma oscillations; parvalbumin interneurons; Perampanel; GluA2; intellectual disability

INTRODUCTION

SYNGAP1 codes for synaptic Ras GTPase activating protein 1 (SYNGAP1), a RasGAP critical for the formation of postsynaptic densities (PSDs) and the experience-dependent AMPA receptor (AMPA) insertion that underlies synaptic plasticity (1–5). Mutations in *SYNGAP1* are prevalent in patients with schizophrenia, intellectual disability (ID), and autism spectrum disorder (6–8). Loss-of-function *SYNGAP1* mutations result in haploinsufficiency and cause mental retardation type 5 (MRD5, OMIM#612621), a severe distinct generalized developmental and epileptic encephalopathy with ID, ataxia, severe behavioral problems, and a risk for autism (6–12).

The majority of patients with MRD5 have refractory epilepsy (12). Poor quality sleep is highly prevalent in patients with neurodevelopmental disorders (NDDs) and epilepsy (13,14). The relationship between epilepsy and sleep is a major focus of ongoing clinical and pre-clinical research (13–15). For example, Rett syndrome (RTT) is an NDD with epilepsy and sleep dysfunction (16,17). Research in both pre-clinical models of RTT and patients with RTT has identified translatable quantitative EEG (qEEG) biomarkers (17,18). There is an urgent need to develop robust biomarkers for NDDs, as bench-to-bedside therapeutic strategies are severely hindered without validated quantitative outcome measures.

Patients with MRD5 predominantly present with myoclonic, absence, or tonic-clonic seizures (8,12). Children with epileptic encephalopathies have a slow developmental regression that is primarily due to seizures, interictal spikes (IISs), or cortical dysrhythmia identified on EEG (19,20). Case reports indicate that adult patients demonstrate gradual decline in cognitive abilities (21). Perampanel (PMP), a recently FDA approved AMPAR antagonist has shown significant promise for multiple seizure types in idiopathic generalized

epilepsies including absence (22). Its use in infants with epilepsy is in clinical trial (23). However, the role of AMPAR antagonists such as PMP in *SYNGAP1* haploinsufficiency related seizures and ID is unknown.

Mice with *Syngap1* haploinsufficiency (*Syngap1*^{+/-}) are relevant translational models presenting with learning and memory deficits, abnormal dendritic spine dynamics, cortical hyperexcitability, and precocious unsilencing of thalamocortical synapses during development (24–26). Currently, no studies have investigated the epileptogenesis, sleep, and underlying mechanisms associated with MRD5. An understanding of the mechanisms that promote spontaneous seizures and network dysfunction can help accelerate the discovery of novel therapeutic strategies. For this purpose, a *Syngap1*^{+/-} loss-of-function mouse model (26) (exon 7 and 8 deletions) underwent 24h video-vEEG/EMG recordings at advancing ages (P60-P120). A subsequent 24h EEG investigated the effect of low-dose PMP on EEG-identified biomarkers.

METHODS AND MATERIALS

Animals

All animal care and procedures were carried out in accordance with the recommendations in the Guide for Care and Use of Laboratory Animals of the National Institutes of Health. All protocols used in this study were approved by the Committee on the Ethics of Animals Experiments of the Johns Hopkins University. In *Syngap1*^{+/-} mice, exons 7 and 8 (C2 domain) were replaced with a neomycin resistance gene in the opposite direction to eliminate all possible splice variants (26). All mice were single-housed at P46 (7d before electrode implantation surgeries) in polycarbonate cages with food and water provided *ad libitum*, on a 12h light-dark cycle. All efforts were made to minimize animal suffering and the total number of animals used.

Surgery and 24h Video-EEG/EMG

All surgical procedures implemented in this study are as previously published (27). Briefly, subdural EEG and suprascapular EMG electrode implantation was performed under isoflurane anesthesia (4–1.5%). EEG screw electrodes utilized coordinates from bregma for consistent placement (Supplemental Fig. 1), as previously published (18). After recovering from electrode implantation surgery (7±1 day), mice were placed in a recording chamber with food and water provided *ad libitum*. WT^{+/+} and *Syngap1*^{+/-} mice underwent 24h vEEG/EMG recording at younger (P60; WT^{+/+} N=4 mice and Het^{+/-} N=6 mice) and older (P90–120; WT^{+/+} N=4 mice and Het^{+/-} N=7 mice). For further details of EEG acquisition parameters see Supplementary Methods.

Hypnogram and EEG Power Analysis

EEG data were manually scored for Wake, NREM, and REM states for every 10s epoch by a scorer blinded to genotype and sex. All EEG artifacts identified on EEG underwent video confirmation and were removed from all further analyses. All EEG power analysis protocols used in this study are as previously published (27,28). For further details see Supplementary Methods.

Seizure and IIS Scoring

Spontaneous seizures were identified by manual review of all 24h EEGs by a scorer blinded to genotype and sex. IISs were visually analyzed on raw EEG using conservative parameters obtained from obvious pre- and post-ictal spikes for every 5s EEG epoch. For further details on seizure and ISS scoring parameters see Supplementary Methods.

Analysis of Human EEG recording

Continuous overnight EEGs are being acquired with parent consent under approved IRB protocol (29). The EEG channels F4, C4 and O2 were re-referenced to A1 or A2 and exported in the European Data File (EDF) format, de-identified and assigned an ID number. Each epoch was manually scored as Wake, NREM, or REM, as previously published (17). Seizure scoring was performed separately from sleep scoring. For further details, see Supplementary Methods.

Immunohistochemistry

One week after the final EEG recordings (P127), mice were anesthetized with chloral hydrate (300mg/ml; IP) followed by transcardiac perfusion with ice-cold PBS and fixation with PBS buffered formalin (10%). Brains were subsequently cryoprotected and stored at -80°C before cryosectioning. All coronal sections were cut at $14\mu\text{m}$ and mounted onto Permafast™ microscope slides before staining. Fluorescent images were acquired at 10X and 40X using Z-stacks with Axiovision Software 4.6 and Apotome® (Carl Zeiss, Jena, Germany). All data analyses were performed using the ImageJ software (30). All data were analyzed by a scorer blinded to genotype and sex. For further details, see Supplementary Methods.

Statistical Analyses

Statistical tests were performed using Prism 8.3 (GraphPad Software, La Jolla, CA, USA). All 1-way and 2-way ANOVAs were performed with Bonferroni's post-hoc corrections. All independent *t*-tests and *Nested t*-tests were two-tailed. In this study, all data are reported as means \pm 1 S.E.M. For further details, see Supplementary Methods.

RESULTS

***Syngap1*^{+/-} mice have recurrent spontaneous seizures with multiple phenotypes**

Continuous 24h vEEG/EMG recordings at temporally advancing ages identified spontaneous seizures in 50% (n/n mice; 4/8) of *Syngap1*^{+/-} mice (Fig. 1). At P60 all seizures were myoclonic (n/n seizures; 5/5); 80% (4/5) of these seizures started in non-rapid eye movement (NREM) at transitions from NREM to wake (Fig. 1A). Myoclonic seizures were $34.4\pm 5.9\text{s}$ in duration with rhythmic spike-wave discharges occurring $\sim 3\text{--}4\text{Hz}$ (Fig. 1A), similar to clinical reports (12). The myoclonic seizures were distinguished by their time-locked myoclonic jerks recorded on EMG with video confirmation of head, neck, and upper shoulder myoclonic jerks (see Supplemental Video 1).

Multiple seizure phenotypes were identified in P120^{+/-} mice, consistent with the latest clinical reports (12,31). Myoclonic, generalized tonic-clonic, and electrographic seizures

were observed (Fig. 1B–C and Supplemental Fig. 2). All myoclonic (n/n seizures; 3/11 seizures) and tonic-clonic seizures (1/11) occurred during NREM in P120^{+/-} mice, whereas all electrographic seizures occurred during wake (7/11 seizures; Fig 1C3). Electrographic (subclinical) seizures were distinguished by their abnormal EEG patterns that were apparent only on EEG and not associated with any behavioral change. Myoclonic seizures had a propensity to emerge during NREM regardless of age. From P60 to P120, the average seizure duration significantly increased from 27±0.6 to 106±22.7s (Fig. 1C1). This increase in seizure duration was driven by the emergence of electrographic seizures at P120.

Recurrent generalized seizures in human *SYNGAP1* haploinsufficiency show affinity for transition-states and clustering in NREM.

To evaluate the translational value of 24h qEEG analysis in *Syngap1*^{+/-} mice, two EEG recordings from children diagnosed with *SYNGAP1* haploinsufficiency related epilepsy were acquired (29), and analyzed. A 20h continuous EEG captured an entire cycle of overnight sleep in a 3-year-old boy with *SYNGAP1* haploinsufficiency (Fig. 2). Global seizures were identified as bursts of 3–4Hz spike and slow waves that lasted ~0.5–3s (Fig. 2A). All ictal events observed during sleep occurred in NREM (Fig. 2B), and an affinity for clustering of ictal events (i.e. inter-ictal durations <5min) during NREM was noted (Fig. 2C–D). Further, ictal events predominantly occurred during Wake-NREM and NREM-REM transitions. A 50min clinical EEG recorded from a 5-year-old girl diagnosed with *SYNGAP1* haploinsufficiency showed a similar clustering of generalized ~3–4Hz short-duration seizures during transitions from Wake to first NREM (Supplemental Fig. 3). These observations identified a previously undescribed clustering of ictal-events during NREM, and the propensity of the ictal clusters to occur during transitions between behavioral-states.

Progressive alterations in macro-sleep architecture

Sleep dysfunction is a common feature of NDDs and is reported in patients with *SYNGAP1* haploinsufficiency (12,21). Quantitative assessments of the sleep problems in patients with *SYNGAP1* haploinsufficiency are lacking, therefore future assessments will be informative (29). In this study, the WT^{+/+} macro-sleep architecture was ultradian with 15.38±2.2 sleep cycles occurring over 24h (Fig. 3A). All WT^{+/+} mice regardless of age demonstrated a consistent sleep architecture (Supplemental Fig. 4). WT^{+/+} mice spent ~80% of the time awake during the dark phase (Fig. 3A–B2), demonstrating the nocturnal predominance of activity in WT^{+/+} mice. Between groups, P60^{+/-} and P120^{+/-} mice spent more time asleep than WT^{+/+} mice over 24h (Fig. 3B). Compared to WT^{+/+} mice, P120^{+/-} mice spent significantly more time asleep during the light phase (Fig. 3B1). In WT^{+/+} mice, there were significantly fewer sleep/wake cycles during the dark phase (10.37±2.0 cycles) compared to the light phase (22.13±3.3 cycles, Fig. 3C). In P60^{+/-} and P120^{+/-} mice, the difference between the numbers of sleep/wake cycles during light vs. dark phase was not significant (Fig. 3C). In contrast to the significant differences in macro-sleep architecture, all mice maintained a sleep microarchitecture of ~80% NREM and ~20% REM during the 24h recordings (Fig. 3C1–C2).

IISs predominantly occurred during NREM sleep and were progressive with age.

Uncontrolled epilepsy is at least partially responsible for cognitive regression in children with epileptic encephalopathies (19,20). All *Syngap1*^{+/-} mice presented IISs during both sleep and wake (Fig. 4A–B). IIS frequency significantly increased at P120^{+/-} when compared to WT^{+/+} mice (1-way ANOVA, $F_{2,20}=4.278$, $p=0.0284$). This significant increase was driven by IISs that occurred during NREM sleep, specifically during light-phase sleep (Fig. 4C–D). Furthermore, even though the total duration of sleep increased significantly in P120^{+/-} mice, (Fig. 3B) the rate of IIS occurrence per hour of NREM sleep was significantly greater in P120^{+/-} mice compared to WT^{+/+} (Fig. 4D).

Emerging research utilizing intracranial recordings from epilepsy patients has demonstrated that regulation of brain activity occurs on long timescales(32). Daily patterns of seizure occurrence demonstrate circadian rhythmicity and clustering organization(32). In this study, we report the relationship between seizure occurrence and frequency distribution of IISs over 24h for all mice (Fig. 4E–F). Grouped data of IIS frequency averaged over 24h showed an escalation in IIS frequency at the beginning of the light phase, and was associated with an increased incidence of seizures in P120^{+/-} mice. Antagonizing AMPARs acutely with PMP (2mg/kg I.P. at 10AM and 6PM) prevented seizures during the 24h EEG, but did not significantly reduce IIS frequency (Supplemental Fig. 5). In summary, IIS occurrence was greatest during NREM in *Syngap1*^{+/-} mice.

Hyperactivity worsened with age

Video tracking software allows for high specificity and sensitivity when tracking mouse motor activity over the 24h recordings. Total distance traveled by each mouse over a 24h period was used to identify hyperactivity in *Syngap1*^{+/-} mice (Fig. 5A1–A3). Compared to WT^{+/+} mice (267.92±46.8m), P120^{+/-} mice traveled greater distances (479.75±46.1m), which was not detected in the P60^{+/-} mice (288.22±32m; Fig. 5A2–A3). Since P120^{+/-} mice spent more time asleep than WT^{+/+} mice (Fig. 3), the distance traveled by each mouse was normalized to the duration of time spent asleep. The running average of mice in each group highlighted increased activity at the end of the dark phase, when the majority of nesting behavior occurs (33) (Fig. 5B). The normalized distance traveled by P120^{+/-} mice was significantly greater than WT^{+/+} and P60^{+/-} mice (Fig. 5C). P120^{+/-} mice spent more time asleep, but were hyperactive when awake, as they travelled twice the distance compared to WT^{+/+} mice. These results are consistent with recent reports of home cage hyperactivity in male *Syngap1*^{+/-} mice (34,35).

Activity-dependent theta modulation is impaired during wake in P120^{+/-}

Cross-frequency coupling occurs between theta and gamma oscillations(36,37). Specifically, gamma oscillations are nested within theta oscillations(38,39) and are critical for memory retrieval (40–42). To determine differences in theta and gamma power during transitions from stationary- to active-wake, 10min epochs were analyzed (Fig. 6A). During stationary-wake, high theta and low gamma power were apparent in WT^{+/+} mice (Fig. 6A–B). After the transition to active-wake, WT^{+/+} mice had high gamma and low theta power. The state-dependent (stationary- vs. active-wake) modulation of theta power was not apparent in P120^{+/-} mice (Fig. 6B–C).

In WT^{+/+} mice, theta and gamma spectral power demonstrated an inverse relationship during wake-sleep state transitions (Supplemental Fig. 6A). During wake, the theta-gamma ratio (TGR) was low in WT^{+/+} mice, while during sleep the TGR was high (Supplemental Fig. 6A) (43,44). This relationship between theta and gamma power was lost in P120^{+/-} mice (Supplemental Fig. 6A–B).

Loss of behavioral-state homeostasis of cortical gamma

High cognitive load during wakefulness (45–47) is associated with increased gamma power. Accordingly, in WT^{+/+} mice gamma power was high during wake and low during NREM sleep (Fig. 7A). P120^{+/-} mice did not demonstrate this behavioral state-dependent gamma power modulation during Wake-NREM transitions, as gamma power remained high during NREM (Fig. 7B). PMP-treated *Syngap1*^{+/-} mice showed a significant rescue of the gamma dysregulation, high gamma during wake, and low gamma during NREM similar to WT^{+/+} mice (Fig. 7C). In WT^{+/+} mice the slopes of gamma power during Wake-NREM transitions over 24h were negative, as gamma power decreased during NREM. During NREM-Wake transitions, the slope of gamma power was positive in WT^{+/+} mice as gamma power increased during wake (Fig. 7D1). These negative slopes of Wake-NREM transitions and the positive slopes of NREM-Wake transitions were significantly different from one another in WT^{+/+} mice (Fig. 7D1). P120^{+/-} mice failed to show significant differences between gamma slopes (Fig. 7D2). PMP significantly rescued the transition state-dependent differences between the slopes of gamma power in the *Syngap1*^{+/-} mice (Fig. 7D3). The average gamma slopes for both Wake-NREM and NREM-Wake transitions were compared between groups (Supplemental Fig. 7). Mean gamma slopes between NREM-Wake and Wake-NREM transitions in WT^{+/+} mice were significantly different, and were lost in P120^{+/-} mice. PMP rescued this loss in the P120^{+/-} mice (Supplemental Fig. 7). Further, low-dose PMP did not significantly alter cortical gamma during transitions between Wake and Sleep in the WT^{+/+} mice that were administered PMP (Supplemental Fig. 8).

Additional analyses of binned gamma power were undertaken as spectral frequency analysis identified that the significant behavioral-state alterations in P120^{+/-} gamma power modulation between NREM and Wake were predominantly driven within the 35–45Hz range (Supplemental Fig. 9). Power in the 35–45Hz bin was significantly higher during Wake compared to NREM in WT^{+/+} mice (Supplemental Fig. 9), which was lost and reversed in P120^{+/-} mice (Supplemental Fig. 9). In summary, *Syngap1*^{+/-} mice showed a loss of state-dependent gamma modulation.

During NREM the majority of brain activity is synchronous slow wave activity with low gamma power (45–47). In contrast, gamma power increases during the asynchronous activity of REM (45–47), hence its description as *paradoxical sleep*. In WT^{+/+} mice, NREM-REM transitions underwent modulation in gamma, theta, and delta powers (Fig. 8A & B). A significantly lower percent change in gamma power during NREM-REM transitions was apparent in P120^{+/-} mice compared to WT^{+/+}. This impairment in gamma power modulation from NREM to REM was significantly rescued by PMP (Fig. 8B).

Deficits in PV+IN innervation in prefrontal cortex

The majority of perisomatic inhibition is mediated by parvalbumin-positive interneurons (PV+INs) (45), which have a significant role in the modulation of cortical gamma oscillations(48). Our novel finding of consistently high gamma power in *Syngap1*^{+/-} mice directed the investigation of PV+IN density distribution in the cerebral cortex, hippocampus, and prefrontal cortices. PV+IN distributions did not show significant differences in overall densities in either the dorsal pallidum cortex (DP; WT^{+/+} vs. Het^{+/-}: *Nested t-test*, $t_{14}=0.6823$, $p=0.51$) or the prelimbic cortex (PrL; WT^{+/+} vs. Het^{+/-}: *Nested t-test*, $t_{14}=0.5755$, $p=0.46$). The quantification of total and perisomatic PV+ puncta counts identified significant deficits in the DP cortex of *Syngap1*^{+/-} mice (Fig. 9A–B), but not in the PrL cortex (PrL Total PV counts WT^{+/+} vs. Het^{+/-}: *Nested t-test* $t_{14}=0.055$, $p=0.95$ and PrL Soma PV counts WT^{+/+} vs. Het^{+/-}: $t_{14}=1.434$, $p=0.1734$). GluA2 fluorescence intensity on PV+IN somas (putative Ca²⁺ impermeable AMPARs) and non-PV+ neurons showed no significant differences in DP (Fig. 9C–D; for antibody verification see Supplemental Fig. 10), or PrL. These findings indicate that deficits in PV+IN innervation were region-specific within the prefrontal cortex in *Syngap1*^{+/-} mice.

These findings warranted the analysis of PV+ puncta counts in other ROIs. Unlike the prefrontal cortex, no significant differences in PV+ puncta count were noted in either the barrel (BCX) or motor (MCX; Fig. 10A1) cortices. A layer-specific investigation of BCX and MCX demonstrated that PV+ puncta counts were not significantly different between layers for both the total counts, and the non-PV+ somas (Fig. 10A1–B1). In the hippocampus, PV+ puncta counts were not significantly different in CA1 or CA3 (Fig. 10C1). Normalized PV+ puncta counts were not significantly different between the SP, SR, or SO of WT^{+/+} and P120^{+/-} mice in CA1 and CA3. In summary, deficits in PV+ innervation were only observed in the DP cortex.

Increased GluA2 expression on PV+ soma in somatosensory but not motor cortex

A rich diversity of GABAergic neurons shape the spatiotemporal dynamics of cortical circuit outputs by elegant inhibitory control mechanisms(49,50). Previous research reports that compared to pyramidal neurons, PV+INs have a lower expression profile for the GluA2 subunit(51). PV+INs in the both L2–3 and L5–6 of the BCX showed significant upregulation of GluA2 (Fig. 10A2), which was absent in the MCX (Fig. 10B2). The mean GluA2 fluorescence intensity on PV+INs was also significantly higher in the CA3 but not in the CA1 (Fig. 10C2). No significant differences in GluA2 fluorescence intensities were noted in the non-PV+ cells in any ROI investigated (Supplemental Fig. 11). Since all *Syngap1*^{+/-} mice were epileptic, the hippocampi were investigated for mossy fiber sprouting. No evidence for mossy fiber sprouting was detected in this model (Supplemental Fig. 12). In summary, a significant region-specific upregulation of GluA2 expression in PV+INs, a population of INs known to generate cortical gamma oscillations (48), was detected in *Syngap1*^{+/-} mice.

DISCUSSION

24h continuous EEG monitoring and qEEG protocols (27) identified novel biomarkers underlying epileptogenesis in a mouse model of *Syngap1* haploinsufficiency. A progressive worsening in epileptogenesis, IIS frequency, hyperactivity, and an impaired behavioral-state dependent modulation of cortical gamma activity during both Wake- and Sleep-state transitions was identified. Myoclonic ~3Hz seizures in the *Syngap1*^{+/-} mice arose during NREM sleep. Human overnight and clinical EEG analyses identified ictal clusters of 3Hz spike-wave discharges during NREM that were aggravated at transitions from Wake to Sleep and Sleep to Wake. Currently, it is unknown if any other models of *Syngap1* haploinsufficiency that differ by mutation site or mutation strategy (i.e. germ-line vs. Cre-recombinase) show these clinically relevant 3Hz seizures with progression to multiple other phenotypes. Additionally, the rescue of impaired cortical gamma homeostasis during behavioral-state transitions by PMP identified the critical role of AMPARs in during such transitions. Induced increase of GluA2 expression in INs resulted in severe disruption of gamma oscillations (52). This research implicated the Ca²⁺ impermeable AMPAR subunit, which confers a slow EPSP in excitatory neurons, and usually expressed at low levels in GABAergic interneurons (52), in playing a critical role in gamma power during behavioral-state transitions. Immuno-labelling identified excessive GluA2-AMPA expression on the somas of PV+INs in *Syngap1*^{+/-} mice that was location-specific, supporting our hypothesis.

In this study, 50% of the *Syngap1*^{+/-} mice presenting with seizures during the 24 recording period indicating a high incidence of seizures. Progressive worsening of the sleep dysfunction and increasing NREM IIS frequency in 100% of the mice is also a common feature of several developmental disorders associated with epilepsy (53–59).

Wakefulness is associated with net-synaptic potentiation, whereas sleep favors global synaptic depression thereby preserving an overall balance of synaptic strength (60). AMPAR levels are high during wakefulness and low during sleep. The global reduction of AMPARs during sleep may be disrupted in *Syngap1*^{+/-} mice as haploinsufficiency could result in the inability to regulate Ras and Rap, two proteins that are involved in the regulation of surface AMPARs (3,4,61–64). The inability to downregulate surface AMPARs during transition-state synaptic homeostasis could lead to chronic overexpression of AMPARs in highly active neurons, such as fast spiking PV+INs.

The significant loss in cortical gamma homeostasis associated with behavioral-state transitions in *Syngap1*^{+/-} mice could critically impede cortical information processing. Gamma oscillations (52,65) are known to depend upon AMPAR kinetics in interneurons, especially for long-range synchrony. The significant rescue of the cortical gamma homeostasis by acute dosing of low-dose PMP when compared to the relatively weaker effect on IIS suppression may indicate a novel role for PMP beyond its FDA approved anti-seizure properties. New trials evaluating PMP efficacy in infants (23) are underway. In the clinic, PMP dosing is gradually ramped-up starting with the low-dose of 2mg/kg in weekly increments reaching maintenance doses of 8–12 mg/day based on tolerability. The acute low-dose protocol used here did not show significant effects on IIS suppression; however no seizure events were noted with PMP treatment. PMP anti-seizure efficacy for SYNGAP1-

related seizures are currently unknown, and a recent cohort study with 57 patients reported only one patient having received PMP treatment (12). The findings indicate the necessity to develop GluA2-AMPA selective antagonists, which currently do not exist. These analyses also identified poor PV+IN innervation in the prefrontal cortex supported by previously published reports(66), and was not detected in the other ROIs.

Increasingly, brain oscillations are being used to understand complex neuropsychiatric disorders. Gamma (35–50Hz) oscillations have warranted special attention due to their association with higher order cognitive processes including sensory processing, attention, working memory, and executive functioning. Activation of GABAergic interneuron networks has been shown to produce gamma oscillations (~40Hz) in both the hippocampal and neocortical networks. Given the critical role of PV+INs in cortical gamma oscillations (48,67) and the role of AMPARs in homeostatic synaptic plasticity (68), the loss of gamma homeostasis identified for *Syngap1* haploinsufficiency during periods commonly associated with intense synaptic plasticity (69) provide novel insights into the associated intellectual disabilities. AMPA-mediated currents rise and decay faster in interneurons partly because of these differences in subunit profiles (70,71). PV+INs receive convergent excitatory input from principal neurons, and inhibitory input primarily from other PV+INs. Experiments that have causally tested stimulation of PV+ cells *in vivo* in the BCX by ontogenetic manipulation selectively amplified gamma oscillations (72). Importantly, this activation suppressed the sensory processing in nearby excitatory neurons within the BCX. Findings of increased GluA2 AMPARs on PV+ soma both in layers 2/3 and 5/6 of the BCX would predict similar suppression of sensory processing in the BCX (73). An increase of GluA2 expression in PV+INs in the BCX could implicate recruitment of excessive PV+IN mediated inhibition resulting in cortical hyperexcitability via rebound excitation. This hypothesis may underlie the clinical reports of reflex seizures in MRD5 emerging during activities associated with increased cortical gamma synchronicity (12,31), a role in which electrically-coupled projection PV+IN are significantly involved.

Morphology of PV+INs has been identified as unipolar vs. multipolar (74) and little is known about the differences in AMPA-mediated currents in these subtypes. GluA2-lacking AMPARs promote anti-Hebbian long-term plasticity (75) which is critical for projection interneurons that functionally connect spatially distant circuits during development. Increase of GluA2-AMPA in PV+IN soma could be one novel mechanism underlying autistic-like behaviors, ID, and seizures in *SYNGAP1* haploinsufficiency. Little is known of the developmental and regional expression profiles of SYNGAP1 isoforms, however it is known that they can exert opposing effects on synaptic strength (76).

To summarize, SYNGAP1 regulates synaptic plasticity, and this process is heavily involved in both epileptogenesis and sleep homeostasis. Investigation of circuit function in intact brains using qEEG biomarkers as they transition from active exploratory, inactive-Wake, and REM/NREM sleep states is an exciting frontier. These behavioral-state transitions are the most reliant upon synaptic plasticity.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Abbreviations list:

AMPAR	AMPA receptor
PV+IN	parvalbumin interneuron
IIS	interictal spike
BCX	barrel cortex
MCX	motor cortex
PMP	Perampanel
CA1	cornu Ammonis 1
CA3	cornu Ammonis 3
Sr	stratum radiatum
SO	stratum oriens
SP	stratum pyramidale
DP	dorsal pallidum
PrL	prelimbic cortex
MRD5	mental retardation type 5
PSD	postsynaptic density
NREM	non-rapid eye movement
REM	rapid eye movement
EEG	electroencephalogram
EMG	electromyogram
NDD	neurodevelopmental disorder

TGR	theta-gamma ratio
IHC	immunohistochemical
FFT	fast Fourier transform

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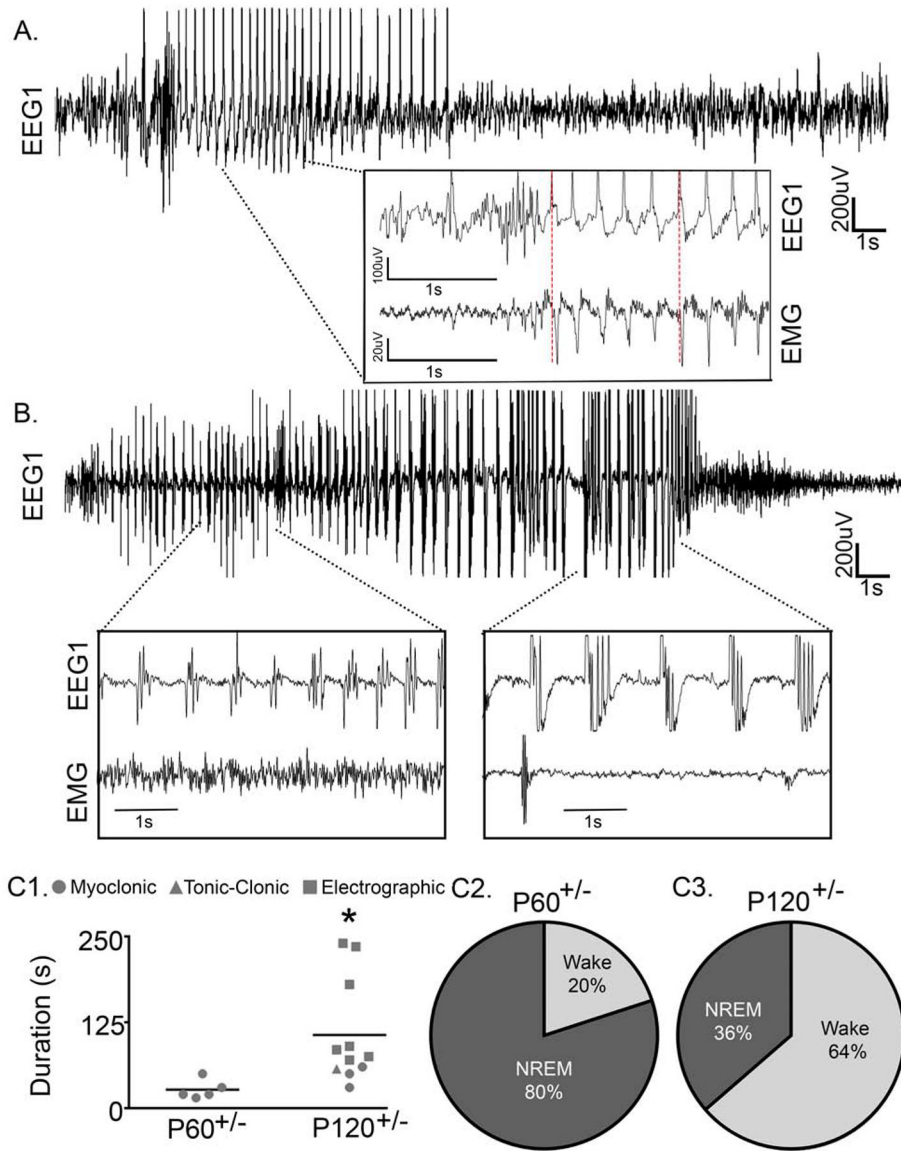


Fig. 1. *Syngap1*^{+/-} mice had recurrent spontaneous seizures.

(A) Representative EEG trace (0.5–50 Hz) of a spontaneous myoclonic seizure at P60 during NREM sleep (P60 EEG recordings N=6 *Syngap1*^{+/-} mice). Prototypical time-locked myoclonic jerks were recorded on EMG (see Supplementary Video 1). (B) Representative spontaneous tonic-clonic seizure at P120 during NREM sleep (P120 EEG recordings N=7 *Syngap1*^{+/-} mice). (C1) Multiple seizure phenotypes emerged at P120, and the average duration of seizures significantly increased from P60 to P120 (*t*-test, $t_{14}=2.307$, $p=0.037$). (C2) At P60 all seizures were myoclonic and the majority (4/5 seizures; N=2 mice) occurred during NREM. (C3) At P120, all myoclonic (3/11 seizures; N=3 mice) and tonic-clonic (1/11 seizures; N=1 mouse) seizures occurred during NREM, whereas all electrographic seizures occurred during wake (7/11 seizures; N=3 mice) (for the electrographic seizure trace see Supplementary Fig. 2). ($p<0.05$ *, post-hoc Bonferroni).

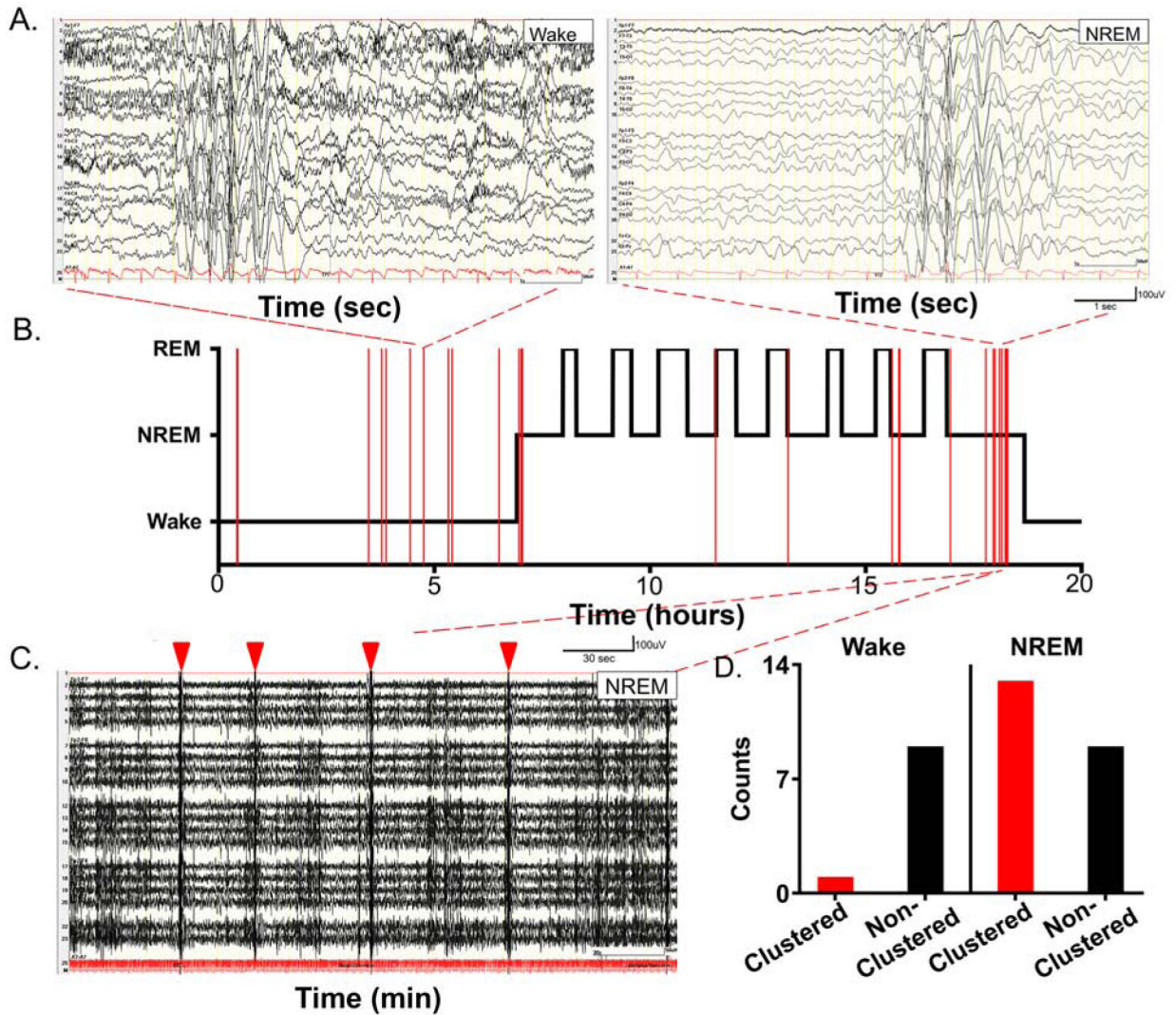


Fig. 2. Ictal events during a 20h overnight EEG recording in a 3-year-old boy with *SYNGAPI* haploinsufficiency.

(A) Identical 3Hz short duration epileptiform events during wake and NREM sleep. (B) Hypnogram for the 20h period, with clustering of events identified during NREM sleep. (C) 5min EEG trace showed four clustered events (red arrowheads) occurring within a 5min period during the last NREM cycle before wake. Note ictal events occurring at transition points between Wake-NREM and REM-NREM (For a 50min EEG recorded from a 5-year-old girl with *SYNGAPI* haploinsufficiency see Supplemental Fig. 3) (D) Bar graphs grouping clustered vs. non-clustered events for Wake and NREM showed affinity for ictal clustering during NREM (N=1; a 3-year-old-boy with *SYNGAPI* haploinsufficiency). Seizure rates normalized per hour of EEG recording in the raster were Wake=1.27/h; NREM=2.68/h and REM=0/h.

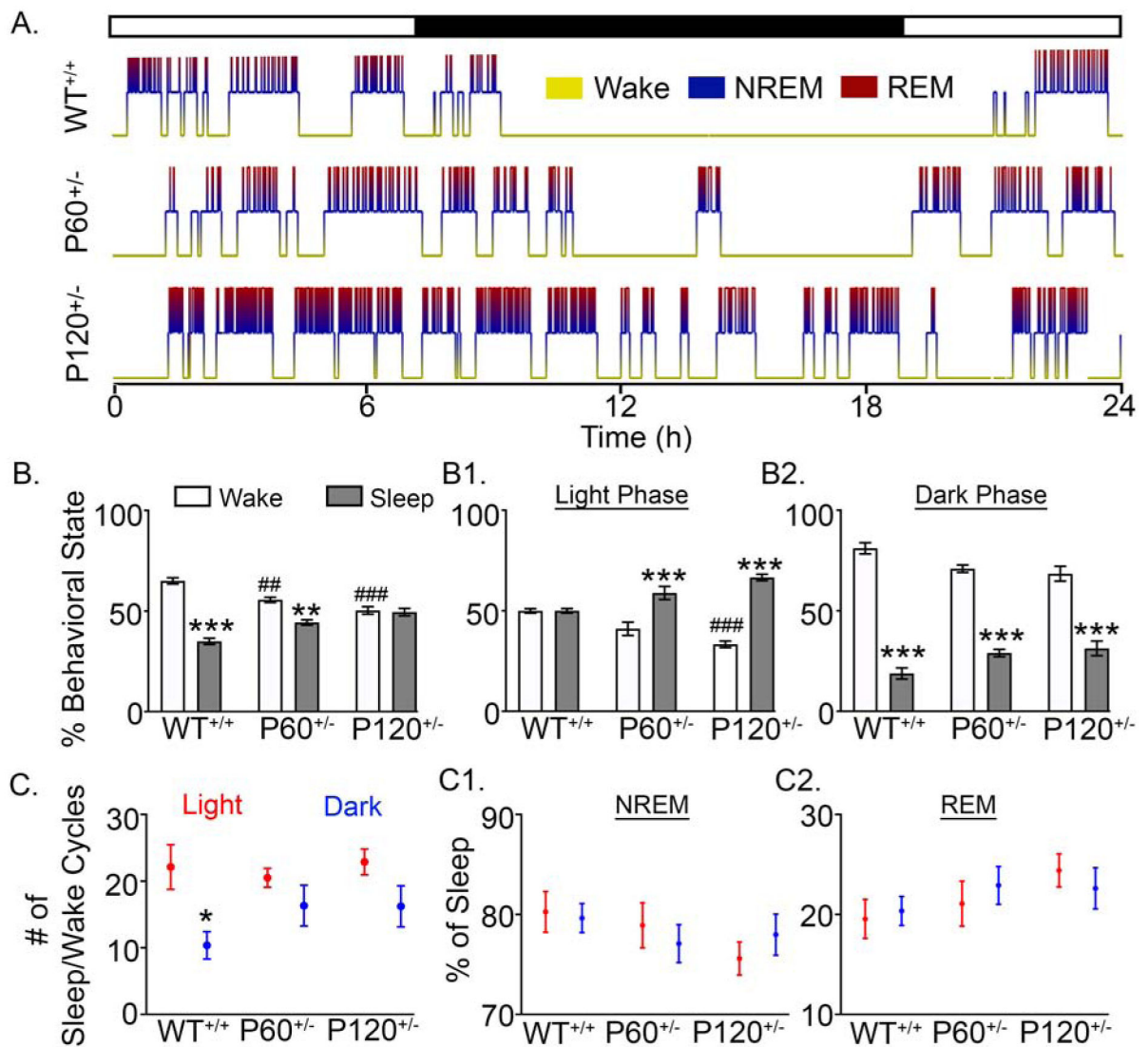


Fig. 3. Progressive alterations in macro-sleep architecture.

(A) 24h hypnograms of nocturnal ultra-radian rhythms in WT^{+/+}, P60^{+/-}, and P120^{+/-} mice. Regardless of age, all WT^{+/+} mice had consistent macro and micro-sleep architectures (Supplemental Fig. 4). (B) WT^{+/+} mice spent significantly less time asleep than awake (WT^{+/+} Wake vs. WT^{+/+} Sleep: 2-way ANOVA, $F_{2,40}=42.01$, $p<0.0001$). P60^{+/-} mice also spent significantly less time asleep than awake (P60^{+/-} Wake vs. P60^{+/-} Sleep: 2-way ANOVA, $F_{2,40}=42.01$, $p=0.002$). In contrast, P120^{+/-} mice lacked any significant differences between durations of Wake and Sleep. Between groups, time spent awake significantly decreased in P60^{+/-} (WT^{+/+} Wake vs. P60^{+/-} Wake: 2-way ANOVA, $F_{2,40}=42.01$, $p=0.009$) and P120^{+/-} mice (WT^{+/+} Wake vs. P120^{+/-} Wake: 2-way ANOVA, $F_{2,40}=42.01$, $p<0.0001$). (B1) During the 12h light phase, in contrast to WT^{+/+}, both P60^{+/-} and P120^{+/-} mice spent more time asleep than awake (P60^{+/-} Light Wake vs. P60^{+/-} Light Sleep: 2-way ANOVA, $F_{2,40}=40.07$, $p<0.0001$; P120^{+/-} Light Wake vs. P120^{+/-} Light Sleep, $p<0.0001$). The amount of time P120^{+/-} mice spent awake was significantly lower than WT^{+/+} (WT^{+/+} Light Wake vs. P120^{+/-} Light Wake: $F_{2,40}=40.0$, $p<0.0001$). (B2) WT^{+/+}, P60^{+/-}, and P120^{+/-} mice all spent

significantly less time asleep than awake during the dark phase (WT^{+/+} Dark Wake vs. WT^{+/+} Dark Sleep: 2-way ANOVA, $F_{2,40}=9.617$, $p<0.0001$; P60^{+/-} Dark Wake vs. P60^{+/-} Dark Sleep: $p<0.0001$, P120^{+/-} Dark Wake vs. P120^{+/-} Dark Sleep: $p<0.0001$). (C) Only WT^{+/+} mice had significantly fewer cycles (transitions between sleep and wake) during the dark phase than the light phase (WT^{+/+} Light cycles vs. WT^{+/+} dark cycles: 2-way ANOVA, $F_{2,40}=0.998$, $p=0.04$). (C1 & C2) Although the significant diurnal difference in the number of sleep/wake cycles detected in WT^{+/+} did not occur in P60^{+/-} and P120^{+/-} mice, the microarchitecture of both NREM and REM cycles during sleep was not significantly different. WT^{+/+} N=6 mice, P60^{+/-} N=6 mice, and P120^{+/-} N=7 mice ($p<0.05$ *, $p<0.01$ **; $p<0.001$ ***; $p<0.05$ #, $p<0.01$ ##; $p<0.001$ ###, post-hoc Bonferroni).

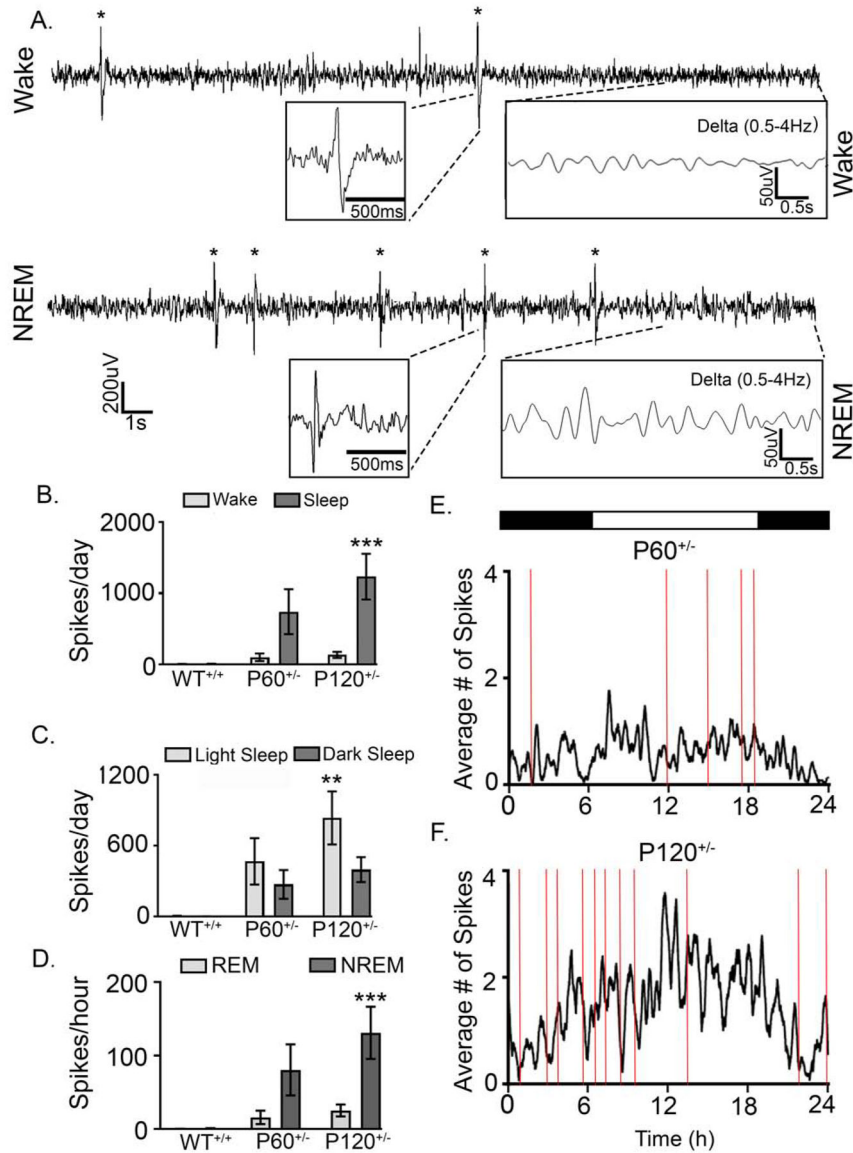


Fig. 4. Progressive interictal spiking during NREM.

(A) 30s EEG trace (0.5–50Hz) during wake of interictal spikes ($n=2$ spikes; black asterisks) with low amplitude delta power (0.5–4Hz). During NREM sleep, a 30s EEG trace demonstrated interictal spikes ($n=5$ spikes; black asterisks) with high amplitude delta power (0.5–4Hz). (B) Number of spikes during sleep significantly increased in P120^{+/-} compared to WT mice (WT^{+/+} Sleep Spikes vs. P120^{+/-} Sleep Spikes: 2-way ANOVA, $F_{2,40}=4.80$, $p=0.0003$) (C) Number of spikes significantly increased during light sleep in P120^{+/-} mice (WT^{+/+} vs. P120^{+/-}: 2-way ANOVA, Light Sleep Spikes: $F_{2,40}=1.33$, $p=0.0012$). (D) Interictal spikes significantly increased with age during NREM (WT^{+/+} NREM spikes vs. P120^{+/-} NREM spikes: 2-way ANOVA, $F_{2,40}=3.508$, $p=0.0006$) (E & F) Interictal spikes (black traces) and seizure frequency (red lines) over 24h. WT^{+/+} N=6 mice, P60^{+/-} N=6 mice, and P120^{+/-} N=7 mice ($p<0.05$ *, $p<0.01$ **, $p<0.001$ ***; post-hoc Bonferroni).

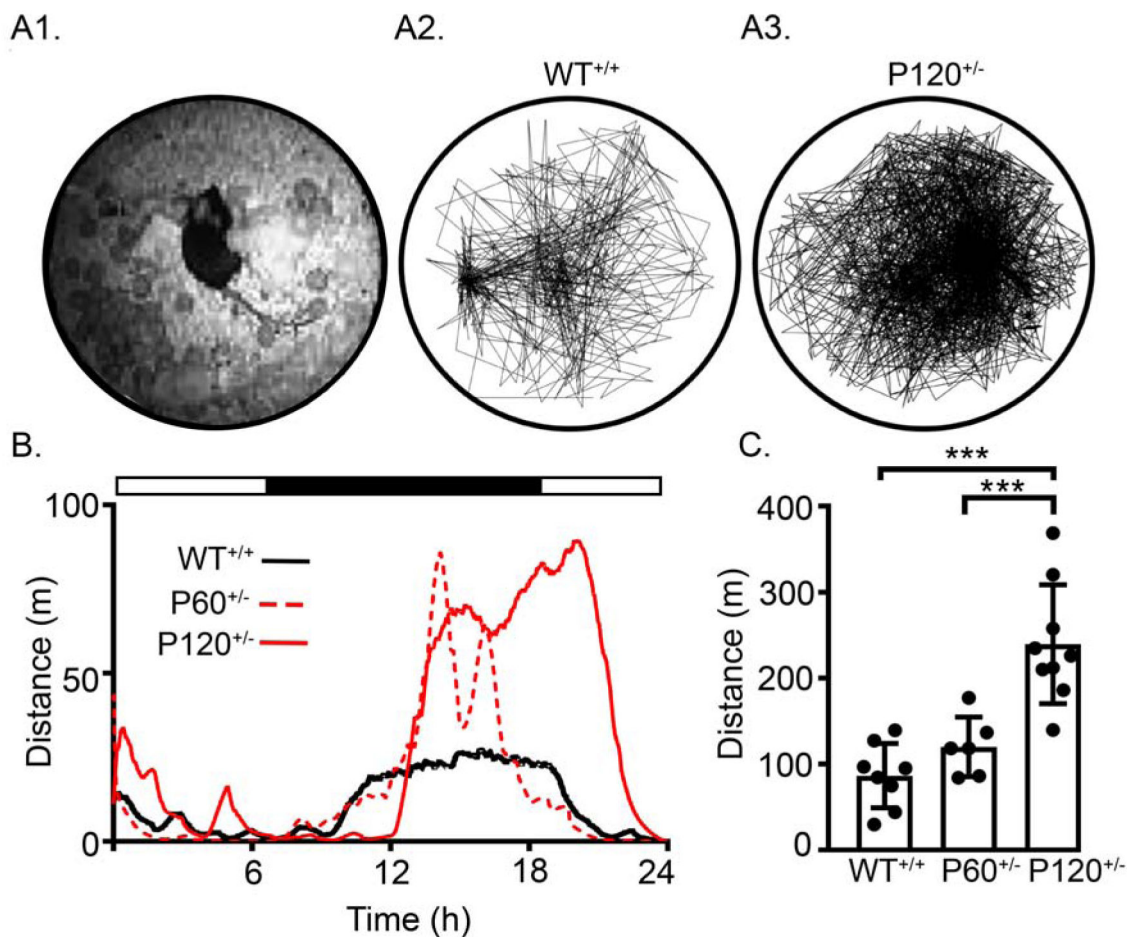


Fig. 5. Progressive increase in activity during 24h recordings.

(A1) Image from the infrared top-view camera utilized for motion tracking. (A2 & A3) Representative traces of WT^{+/+} and P120^{+/-} activity over 24h within individual recording chambers. (B) A representative local regression for distance traveled over 24h by a WT^{+/+}, P60^{+/-}, and P120^{+/-} mouse, all mice showed increased activity during the exploration and nesting phase. (C) P120^{+/-} mice showed significantly higher activity than WT^{+/+} and P60^{+/-} mice (WT^{+/+} vs. P120^{+/-}: 1-Way ANOVA; $F_{2,20}=20.18$, $p<0.001$, P120^{+/-} vs. P60^{+/-}: $p<0.001$). WT^{+/+} N=6 mice, P60^{+/-} N=6 mice, and P120^{+/-} N=7 mice. ($p<0.05$ *, $p<0.01$ **, $p<0.001$ ***; post-hoc Bonferroni).

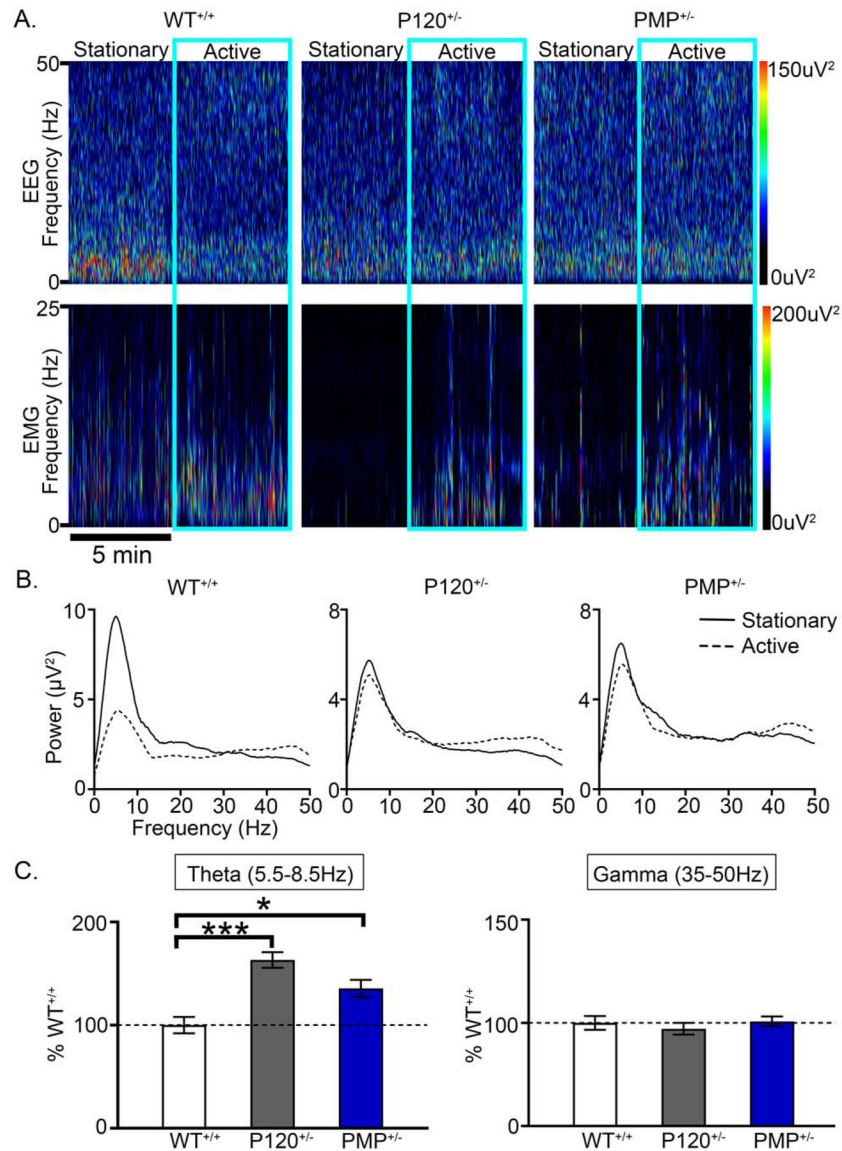


Fig. 6. Progressive impairment of theta power during transitions from inactive to active wake. (A) EEG and EMG spectral power heat maps of activity-dependent transitions from stationary to active wake in WT^{+/+} mice. P120^{+/-} mice failed to demonstrate an increase in gamma with concurrent decrease in theta. (B) Representative spectral frequency traces in WT^{+/+} and P120^{+/-} mice during stationary to active transitions. (C) The lack of theta modulation in P120^{+/-} mice during transitions from inactive to active wake was significant (WT^{+/+} theta vs. P120^{+/-} theta %: 1-way ANOVA, $F_{2,22}=13.51$, $p<0.0001$). PMP administration (2mg/kg; I.P.) at 10am and 6pm failed to significantly rescue theta modulation with acute dosing (WT^{+/+} theta vs. PMP^{+/-} theta %: 1-way ANOVA, $F_{2,22}=13.51$, $p=0.017$). During transitions from inactive to active wake, gamma modulation was not significantly impaired. WT^{+/+} N=6 mice, P60^{+/-} N=6 mice, P120^{+/-} N=7 mice, and PMP^{+/-} N=4 mice. ($p<0.05$ *, $p<0.01$ **, $p<0.001$ ***; post-hoc Bonferroni).

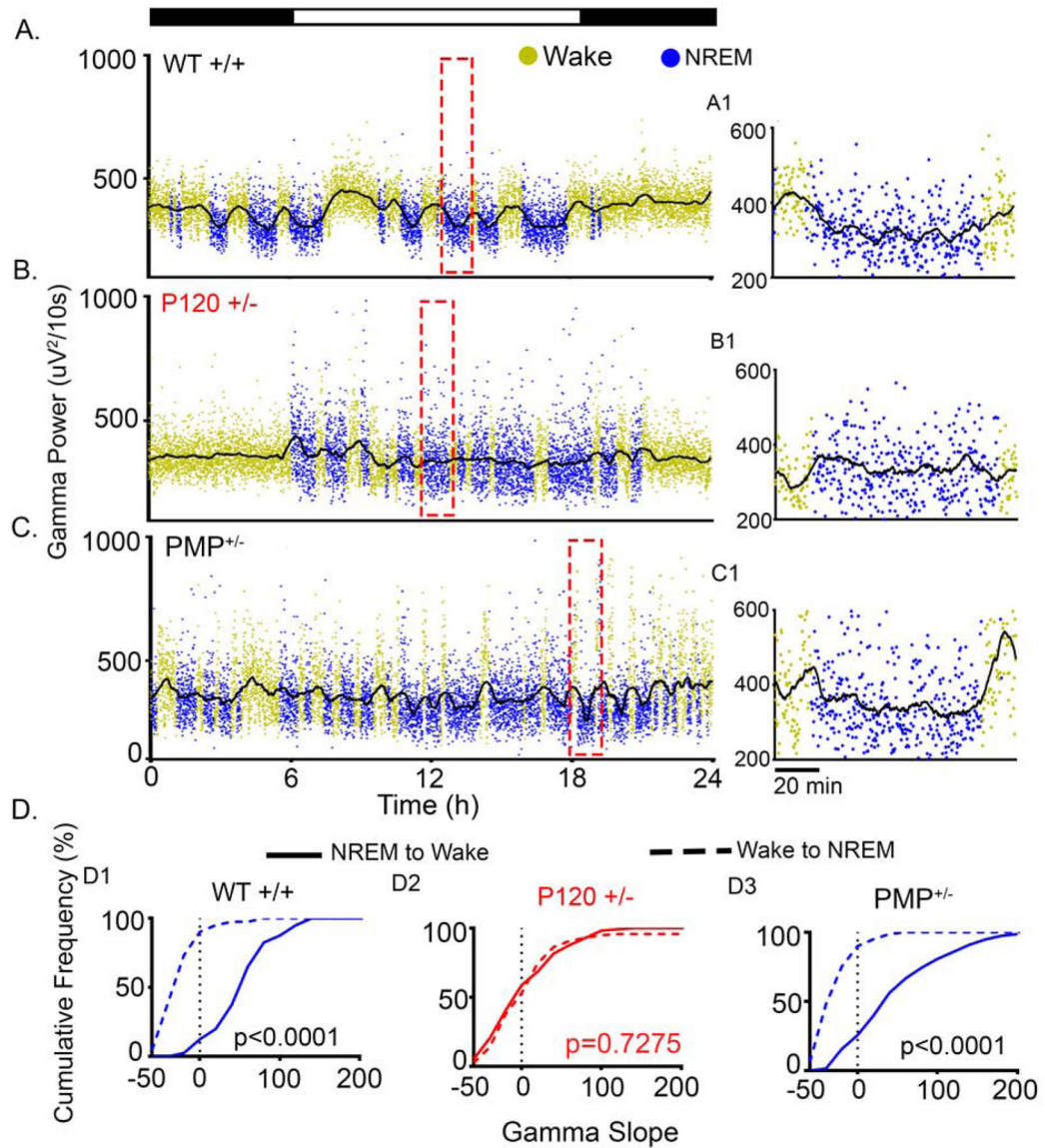


Fig. 7. Gamma during transitions between wake and NREM behavioral state modulation of EEG gamma spectral power.

(A) 24h WT $^{+/+}$ gamma traces had high gamma during wake and low gamma during NREM. (A1) Expanded time scales (red boxes in A-C indicate location of expanded timescales in A1–C1) show a gradual fall of gamma from wake to NREM and gradual rise of gamma from NREM to Wake. (B) Gamma power in the P120 $^{+/-}$ failed to transition to NREM levels, expanded time scale (B1) demonstrates the failure of gamma attenuation during a NREM cycle. (C) In PMP-administered P120 $^{+/-}$ (PMP $^{+/-}$), behavioral-state transitions in gamma power were significantly rescued (C1). (D) Cumulative frequency graphs of positive NREM to Wake slopes and negative wake to NREM slopes in WT $^{+/+}$. (D1) These slopes were significantly different (t -test, $t_{275}=10.59$, $p < 0.001$) between the two independent transition states. (D2) No behavioral-state dependent modulation of gamma power occurred in P120 $^{+/-}$ (t -test, $t_{327}=0.3488$, $p=0.7275$). (D3) PMP administration restored gamma power behavioral-

state dependent modulation (*t-test*, $t_{156} = 9.854$, $p < 0.0001$). WT^{+/+} N=6 mice, P60^{+/-} N=6 mice, P120^{+/-} N=7 mice, and PMP^{+/-} N=4 mice.

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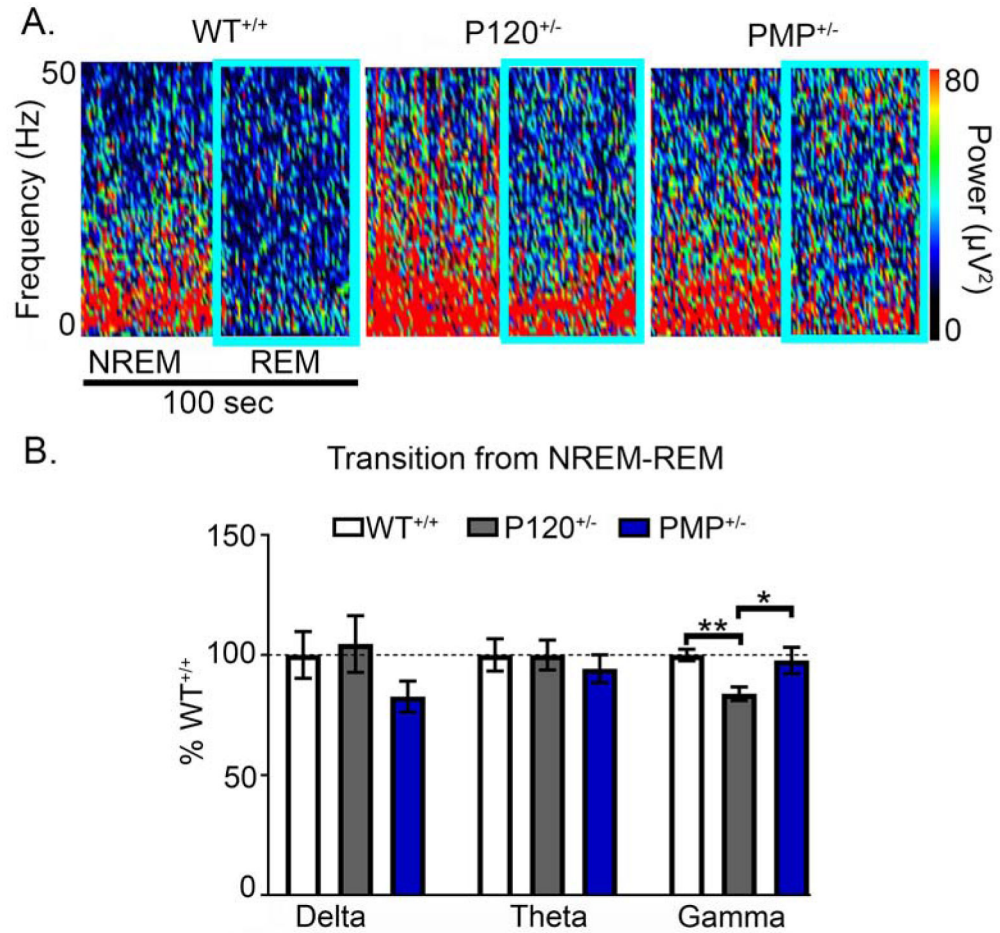


Fig. 8. PMP rescued gamma modulation during transitions from NREM to REM.

(A) Spectral power heat maps during NREM to REM transitions in WT^{+/+}, P120^{+/-}, and PMP^{+/-} mice. (B) A significant decrease in gamma power during NREM to REM transitions was demonstrated in P120^{+/-} mice (WT^{+/+} Gamma vs. P120^{+/-} Gamma % 1-way ANOVA, $F_{2,105}=6.11$, $p=0.005$), but was significantly rescued by PMP administration (P120^{+/-} Gamma vs. PMP Gamma: 1-way ANOVA, $F_{2,105} = 6.11$, $p=0.026$). WT^{+/+} N=6 mice, P60^{+/-} N=6 mice, P120^{+/-} N=7 mice, and PMP^{+/-} N=4 mice. ($p<0.05$ * and $p<0.01$ **; post-hoc Bonferroni).

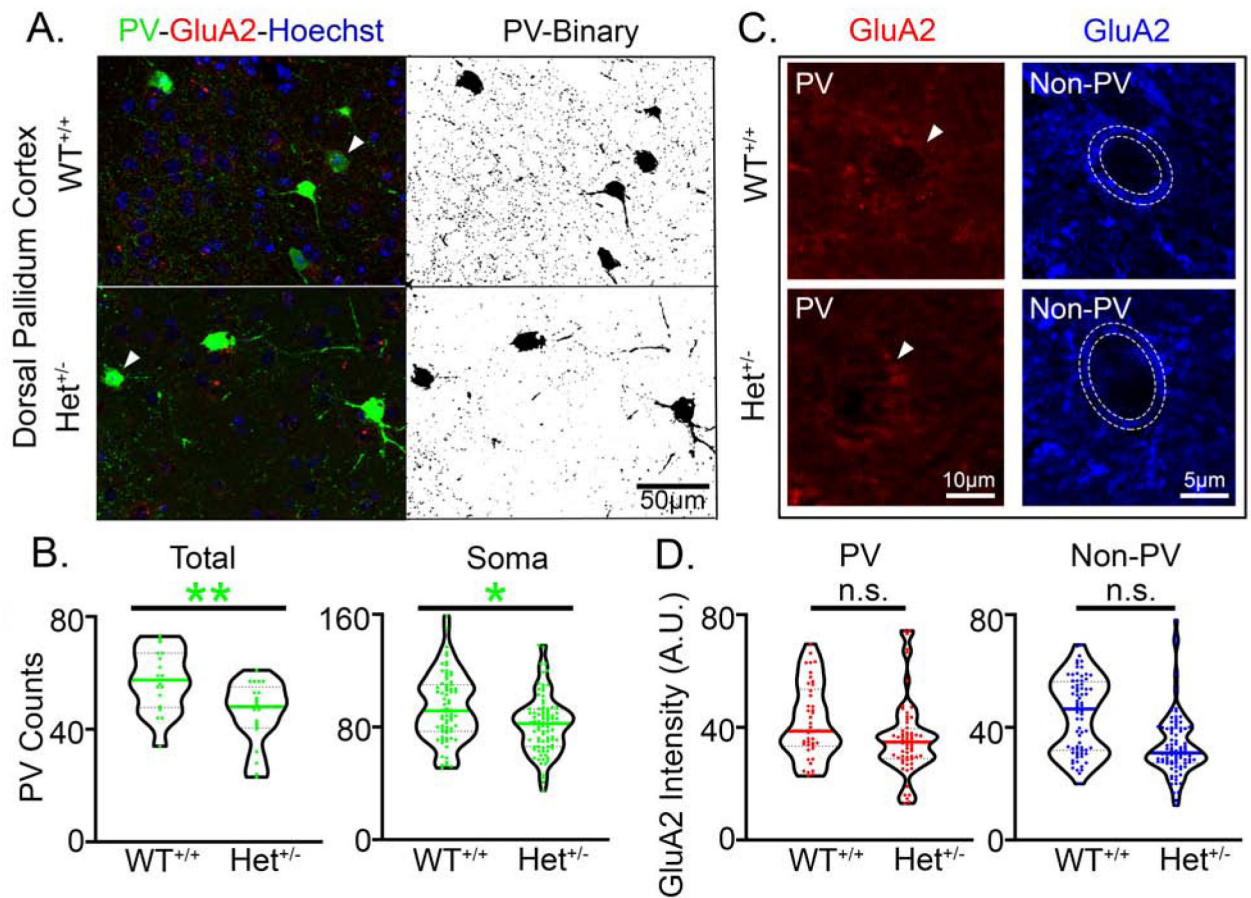


Fig. 9. PV+IN deficits in pre-frontal cortex.

(A) Representative 40X Z-Stack image from the dorsal pallidum (DP) cortex in WT^{+/+} and *Syngap1*^{+/-} mice (Het^{+/-}) mice (white arrowheads indicate PV+INs shown in C). (B) PV immunofluorescence represented by total PV+ puncta in the 40X image showed a significantly lower putative presynaptic puncta in the Het^{+/-} mice (*Nested t-test*, $t_{37}=3.24$, $p=0.0026$). PV+ puncta counts onto non-PV somas were also significantly lower (*Nested t-test*, $t_{14}=2.398$, $p=0.03$). (C) Representative images of GluA2 fluorescence on PV (red) and non-PV (blue) somas in the DP cortex in WT^{+/+} and Het^{+/-} mice (white arrowheads indicate soma of respective PV+INs from A). (D) There were no significant differences in GluA2 expression on PV+IN (*Nested t-test*, $t_{14}=1.230$, $p=0.2391$) or non-PV (*Nested t-test*, $t_{14}=1.883$, $p=0.0806$) neurons in the DP cortex. N=8 *Syngap1*^{+/-} mice and N=8 WT^{+/+} mice ($p<0.05$ *, $p<0.01$ **).

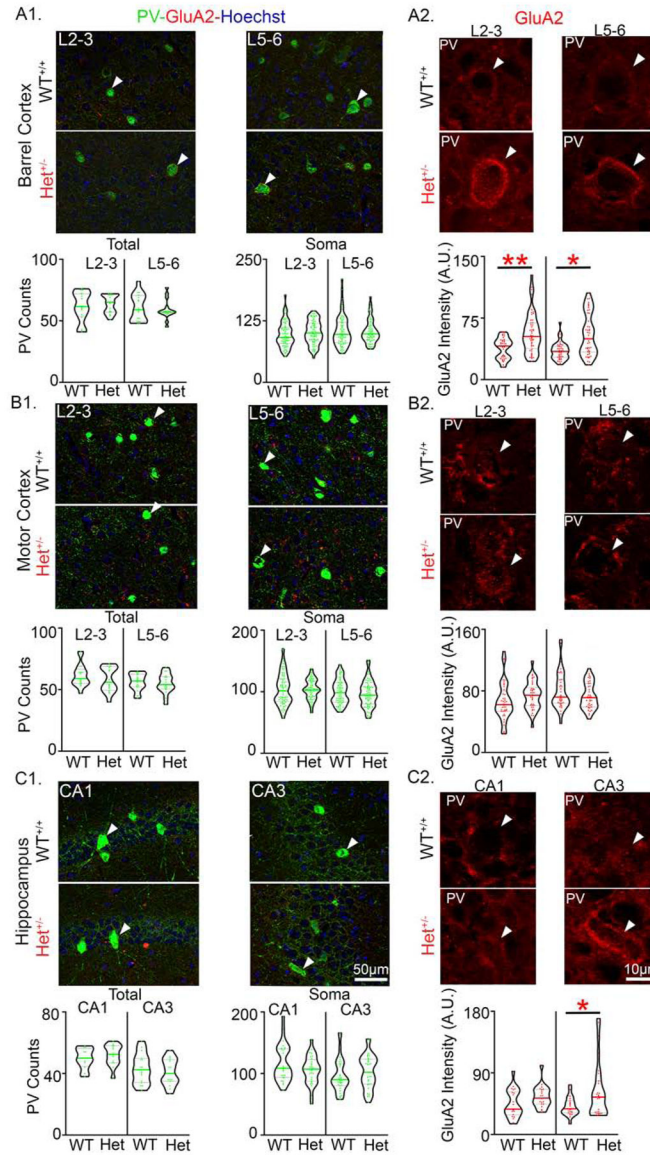


Fig. 10. Increased GluA2 expression on PV+IN soma were region specific

(A1) Representative 40X Z-stack images of WT^{+/+} and *Syngap1*^{+/-} mice (Het^{+/-}) from barrel cortex (BCX) layers 2–3 and 5–6 (L2–3 and L5–6). White arrowheads indicate PV +INs shown in A2 for BCX L2–3 and L5–6, respectively. Total PV+ puncta counts and somatic PV+ puncta counts on non-PV soma were not significantly different in the BCX. (A2) Representative Z-stack images of GluA2 immunofluorescence on PV+INs in L2–3 and L5–6. Het^{+/-} GluA2 expression was significantly higher on PV+INs in both BCX L2–3 (*Nested t-test*, $t_{12}=3.467$ $p=0.0047$) and L5–6 (*Nested t-test*, $t_{12}=2.361$, $p=0.036$). White arrowheads indicate soma of respective PV+INs from A1. (B1) Representative 40X Z-stack images from motor cortex (MCX) L2–3 and L5–6. Total PV+ puncta counts and somatic PV+ puncta counts on non-PV soma were not significantly different in the MCX. White arrowheads indicate PV+INs shown in B2 for MCX L2–3 and L5–6, respectively. (B2) Representative Z-stack images of GluA2 expression was not significantly different for PV

+INs in the MCX L2/3 and L5/6. White arrowheads indicate soma of respective PV+INs from **B1**. **(C1)** Representative 40X Z-stack images from hippocampal CA1 and CA3. White arrow heads indicate PV+INs show in C2 for CA1 and CA3, respectively. Total PV+ puncta counts and somatic PV+ puncta counts on non-PV soma were not significantly different in CA1 or CA3. **(C2)** Representative images of GluA2 immunofluorescence on PV+INs. CA3 Het^{+/-} PV+INs had a significant increase in GluA2 expression (*Nested t-test*, $t_{12}=2.331$, $p=0.038$). White arrowheads indicate soma of respective PV+INs from **C1**. N=8 *Syngap1*^{+/-} mice and N=8 WT^{+/+} mice ($p<0.05$ *, $p<0.01$ **).

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