

## ORIGINAL ARTICLE

**Outbreak of *Streptococcus equi* subsp. *zooepidemicus* infection in a group of rhesus monkeys (*Macaca mulatta*)**K. Mätz-Rensing<sup>1</sup>, J. Winkelmann<sup>1</sup>, T. Becker<sup>1</sup>, I. Burckhardt<sup>2\*</sup>, M. van der Linden<sup>2</sup>, S. Köndgen<sup>3</sup>, F. Leendertz<sup>3</sup> & F.-J. Kaup<sup>1</sup><sup>1</sup> Department of Pathology, German Primate Center, Göttingen, Germany<sup>2</sup> Department of Medical Microbiology, German National Reference Center for Streptococci, University Hospital RWTH Aachen, Aachen, Germany<sup>3</sup> Robert Koch Institute Berlin, Research Group Emerging Zoonoses, Berlin, Germany**Keywords**non-human primate – purulent  
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**Abstract****Background** A severe upper respiratory tract infection occurred in a breeding group of rhesus monkeys housed together in one of six indoor/outdoor corals of the German Primate Center. The clinical signs of the disease included severe purulent conjunctivitis, rhinitis, pharyngitis, respiratory distress and lethargy. Six of 45 animals died within a few days after developing signs of infection.**Methods and results** Histopathologic and microbiologic examinations of the dead animals were consistent with a severe fibrinopurulent bronchopneumonia. Microbiology revealed a Lancefield group C streptococcus identified as *Streptococcus equi* subsp. *zooepidemicus* as the causative agent of infection.**Conclusions** The infection was passed on from animal to animal but did not spread to the other five breeding groups nearby. Extensive diagnostic testing failed to reveal the consisting presence of copathogens in individual cases. A visitor with upper respiratory disease was suspected as source of infection.**Introduction**

*Streptococcus (S.) equi* subspecies *zooepidemicus* belongs to the  $\beta$ -hemolytic group C streptococci. It is able to cause disease both in animals and humans. *Streptococcus equi* subsp. *zooepidemicus* primarily causes equine infections. The agent may be found in the nasopharynx, on the tonsils, in the respiratory tract and on the genital mucous membranes of healthy horses and cattle. It is an important cause of respiratory tract infections in foals and young horses and it is involved in uterine infections in mares. The agent has also been associated with a wide variety of infections including mastitis in pigs, sheep, cows, goats and several other mammalian species [8]. These hosts can be a reservoir for human infections. The clinical manifestation includes pharyngitis, septicemia, meningitis,

purulent arthritis and endocarditis. The source of human infection is often traced back to contact with domestic animals, especially horses, or ingestion of unpasteurized milk or milk products. Streptococci are colonizers of mucous membranes and are transmitted through droplets or direct contact.

*Streptococcus equi* subsp. *zooepidemicus* has sporadically been described in non-human primates. One outbreak occurred in the National Zoological Park, Washington D.C., leading to the death of several callitrichids after contact to infected horse meat fed to armadillos kept in the same exhibition area [4, 7]. Another outbreak of group C streptococcal infection with a high mortality rate occurred in a group of wanderoos (*Macaca silenus*) of the Zoological Garden of Rheine/Germany. The source was suspected to be a human being [1]. The Russian Primate Center in Sochi

reported an outbreak of septicaemia caused by *S. equi* subsp. *zooepidemicus* in representatives of five species of lower monkeys [2]. In 1994, an outbreak among the pig and monkey population was reported from the island of Bali, Indonesia. The infection spread from pigs to monkeys and infected animals died within a few days with signs of bronchopneumonia, pleuritis, epicarditis, endocarditis and meningitis [6]. However, most of the reported disease outbreaks were characterized by symptoms of an enteric infection. The present case report describes an outbreak of respiratory diseases among rhesus monkeys induced by *S. equi* subsp. *zooepidemicus*.

### Materials and methods

The outbreak involved one of six breeding colonies housed in an indoor–outdoor facility. Each unit was composed of an indoor area, a heated and roofed outdoor room and a large fenced outdoor enclosure. Contact to any other animal species except wild birds was not possible. The animals could move freely between the different compartments of the unit. They were fed twice a day with a primate specific diet, composed of standard commercial monkey pellets (ssniff, Spezialdiäten GmbH, Soest, Germany), fresh fruits and vegetables. Water was available *ad libitum*. The animals are kept in accordance with the guidelines of the European Union for the accommodation and care of animals used for experimental and other scientific purposes (2007/526/EG). The primate husbandry is controlled by local and regional veterinary authorities in accordance with the German Animal Protection Law. All procedures are supervised by an animal welfare officer and the ethical committee for experiments using animals in the federal state of Lower Saxony.

A total of 12 animals died in one of these breeding colonies in a short time period. At the moment of the outbreak, the group was composed of 49 animals; most of them were adult females with their offspring and one adult male. Necropsies were performed on all carcasses and samples for histology, parasitology, microbiology and molecular diagnostics were taken. For histopathology, samples were taken from all thoracic, abdominal and pelvic organs, fixed in 10% neutrally buffered formalin, embedded in paraffin and stained with hematoxylin and eosin. For parasitological diagnosis fresh samples were examined microscopically.

For microbiological investigations swabs from liver, spleen, kidney, heart and lung were streaked immediately onto Columbia blood agar plates. Swabs recovered from the intestine were swabbed onto Columbia agar,

EMB, McConkey and Skirrows campylobacter agar plates and inoculated into Rapaport and Skirrow's enrichment broth. Biochemical characterization of the isolated bacterial colonies was achieved using the Crystel-system from Becton Dickinson. Bacterial isolates were sent to the German National Reference Center for Streptococci for further characterization.

Pulsed field gel electrophoresis (PFGE) of bacterial DNA was carried out on a BIO-RAD CHEF-DR III system using the BIO-RAD (BIO-RAD Laboratories GmbH, Munich, Germany) Gene Path Gel Kit and the restriction enzyme *Sma*I.

### Results

The outbreak started at the end of October 2007. Two adult females were found seriously ill with signs of severe upper respiratory tract infection and severe purulent conjunctivitis (Fig. 1). Antibiotic therapy with Baytril® (dosage 2.5 mg/kg) and Aviapen® (dosage 40.000–80.000 IE/kg) was started but both animals died within 2 days after onset of the first symptoms (animals No. 1, 2). One month later, four more female animals developed severe upper respiratory tract infection. Two died suddenly without obvious clinical findings (animals No. 3, 4), one of these was a pregnant one (animal No. 3). The other two died after 10 days on antibiotic therapy with the antibiotics mentioned above (animals No. 5, 6).

Necropsy findings were similar for all six animals. The main findings were severe inflammatory alterations of the upper respiratory tract and the lung (Table 1). All six animals showed a severe subacute tonsillitis and



**Fig. 1** Rhesus monkey, case No. 1. Severe fibrinopurulent conjunctivitis and rhinitis.

**Table 1** Signalment, clinical signs of disease, and pathohistologic findings in rhesus monkeys with streptococcosis due to *Streptococcus equi* subsp. *zooepidemicus* infection

Monkey No.	Sex/age (year)	Clinical observation	Main pathohistologic findings	Culture-positive organs
1 (7488)	Female/10	Upper respiratory tract infection severe fibrinopurulent conjunctivitis	Severe purulent tonsillitis and pharyngitis severe purulent lymphadenitis severe subacute fibrinopurulent pleuropneumonia, epi- and pericarditis mild diffuse purulent hepatitis and splenitis severe cardiomegaly	Lung, heart, liver, spleen, kidney, CNS, eye, tonsil
2 (7490)	Female/20	Upper respiratory tract infection severe fibrinopurulent conjunctivitis	Severe purulent tonsillitis and pharyngitis severe purulent lymphadenitis severe subacute fibrinopurulent pleuropneumonia, epi- and pericarditis mild diffuse purulent hepatitis and splenitis severe cardiomegaly	Lung, heart, liver, spleen, kidney, CNS, eye, tonsil
3 (7502)	Female/9	Sudden death without obvious findings	Severe acute purulent endometritis severe fibrinopurulent serositis severe purulent lymphadenitis moderate subacute fibrinopurulent pleuropneumonia mild diffuse purulent hepatitis and splenitis Mild diffuse purulent hepatitis and splenitis	Lung, heart, liver, spleen, kidney, CNS, tonsil, uterus, placenta
3a (7503)	Fetus from animal No. 3	–		Lung, heart, liver, spleen, kidney, CNS
4 (7508)	Female/8	Sudden death without obvious findings	Moderate purulent tonsillitis and pharyngitis severe subacute fibrinopurulent pleuropneumonia, epi- and pericarditis mild diffuse purulent hepatitis and splenitis severe fibrinopurulent serositis	Lung, heart, liver, spleen, kidney, CNS, tonsil
5 (7515)	Female/23	Severe upper respiratory tract infection cardiac murmur	Severe purulent lymphadenitis severe subacute fibrinopurulent pleuropneumonia severe fibrinopurulent epi- and pericarditis severe cardiomegaly	Lung, heart, liver, spleen, kidney, CNS
6 (7516)	Female/8	Severe upper respiratory tract infection antibiotic treatment of two weeks	Severe purulent lymphadenitis severe subacute fibrinopurulent pleuropneumonia severe cardiomegaly	Lung, heart, liver, spleen, kidney, CNS, tonsil
7 (7519)	Female/14	Sudden death without obvious findings after fighting	moderate multifocal purulent encephalitis Severe acute purulent tonsillitis severe acute purulent lymphadenitis	Tonsil
8 (7520)	Female/4	Sudden death without obvious findings after fighting	Moderate acute purulent tonsillitis	Tonsil
9 (7522)	Female/7	Sudden death without obvious findings after fighting	Moderate acute purulent tonsillitis moderate acute purulent lymphadenitis	Tonsil

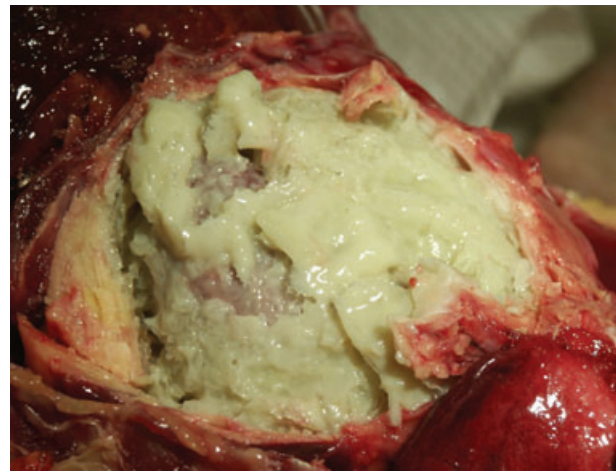
**Table 1** Continued

Monkey No.	Sex/age (year)	Clinical observation	Main pathohistologic findings	Culture-positive organs
10 (7523)	Female/2	Sudden death without obvious findings after fighting	-	-
11 (7524)	Female/4	Sudden death without obvious findings after fighting	-	-
12 (7525)	Male/2	Sudden death without obvious findings after fighting	-	-

IHC, immunohistochemistry; PCR, polymerase chain reaction; CNS, central nervous system.

pharyngitis and a severe subacute fibrinopurulent pleuritis and pneumonia. The inflammation spread to the heart and developed into a fibrinopurulent epi- and pericarditis with exception of animal No. 3 (Figs 2 and 3). Variably sized and poorly demarcated atelectatic areas were present in all lung lobes.

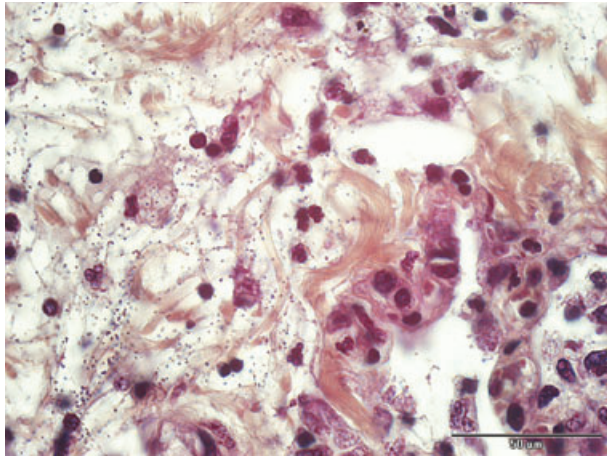
Histologically, there was a fibrinosuppurative pneumonia with extensive to lobular obliteration of the airways and alveolar spaces with neutrophils (Fig. 5A). Large and medium sized airways were unaffected but terminal airways were intensely altered and showed signs of a purulent bronchopneumonia (Fig. 5B). The inflammatory process extended from the pleura to the



**Fig. 2** Heart; rhesus monkey, case No. 6. Severe fibrinopurulent epi- and pericarditis.



**Fig. 3** Heart; rhesus monkey, case No. 4. Severe fibrinopurulent epi- and pericarditis.

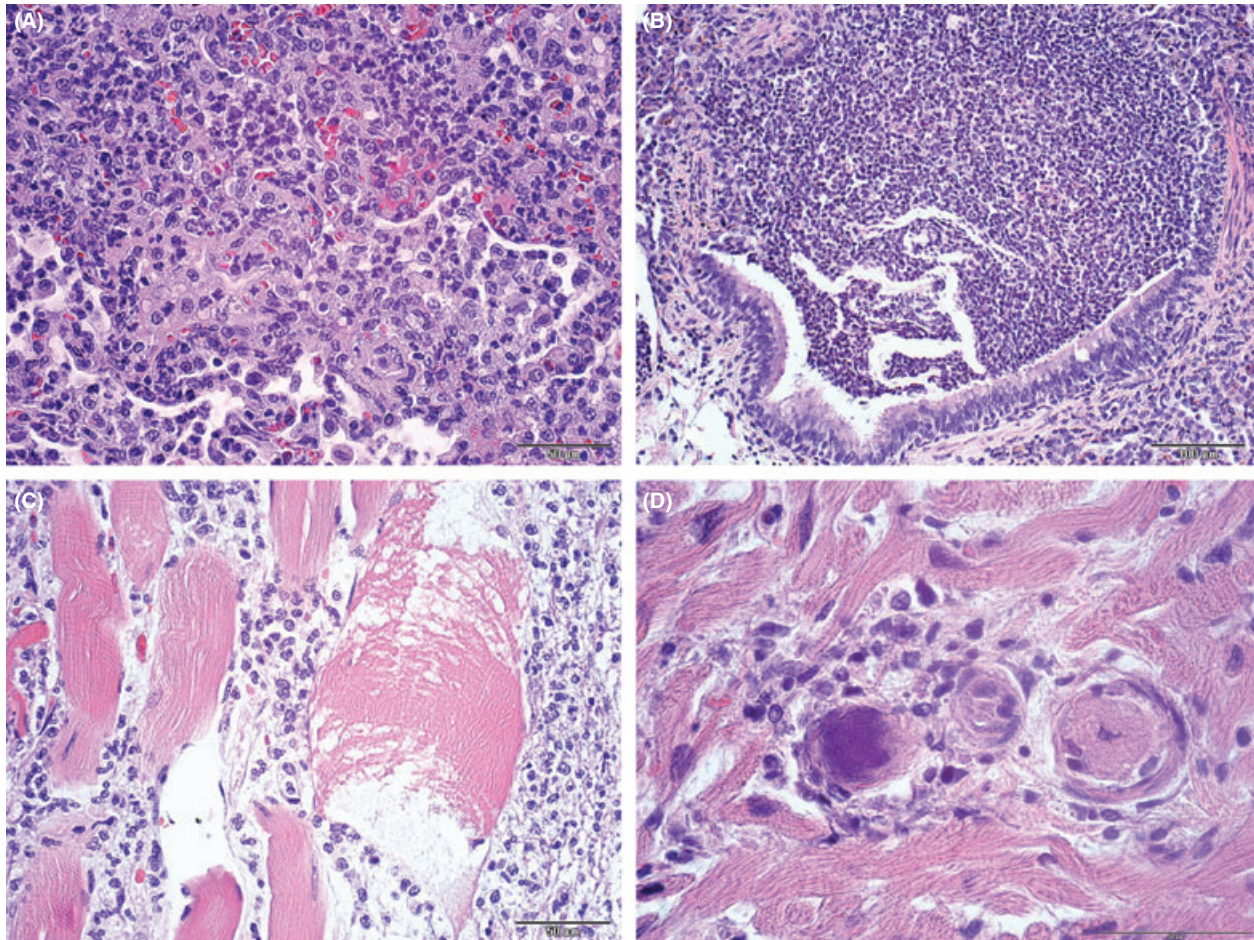


**Fig. 4** Lung; rhesus monkey, case No. 1. Purulent pneumonia with clusters and chains of gram-positive cocci Gram-stain; Scale bar = 50 μm.

diaphragm leading to severe purulent myositis (Fig. 5C). One pregnant animal additionally developed a severe purulent endometritis. The aborted fetus was nearly fully developed and showed signs of a purulent hepatitis and splenitis (animal No. 3a). Splenitis and hepatitis were mild side effects in four more animals. Only one animal developed a purulent encephalomeningitis (animal No. 6).

Clusters and chains of gram-positive cocci were present within the altered organs. They were found within the cytoplasm of macrophages and free within the extracellular spaces laying in pairs or chains (Fig. 4). Septic thromboemboli were less consistently observed (Fig. 5D). Septicemia was attributed as cause of death in all cases.

Septicemia and bacteriemia were confirmed by microbiological investigations on samples taken during necropsy. Pure cultures of large colonies of  $\beta$ -hemo-



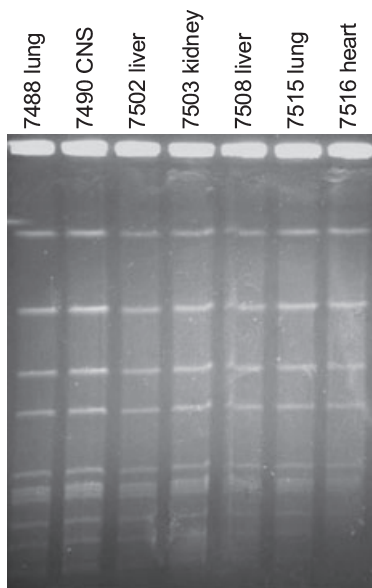
**Fig. 5** (A) Lung; rhesus monkey, case No. 1. Severe fibrinopurulent pneumonia. H&E stain. Scale bar = 50 μm. (B) Lung; rhesus monkey, case No. 1. Severe fibrinopurulent bronchopneumonia. H&E stain. Scale bar = 100 μm. (C) Muscle; rhesus monkey, case No. 1. Severe purulent myositis of the diaphragm. H&E stain. Scale bar = 50 μm. (D) Heart; rhesus monkey, case No. 1. mild myocarditis with septic thromboemboli. H&E stain. Scale bar = 50 μm.

lytic streptococci were grown not only from the respiratory tract, respectively tonsils and lung, but also from nearly all internal organs including the central nervous system. Bacterial characterization was based on the observation of gram-positive, catalase-negative chain forming cocci which fermented lactose and trehalose. Agglutination tests showed that they belong to Lancefield group C.

Pulsed field gel electrophoresis patterns were identical for all seven isolates analyzed, showing that the infections were caused by the same strain in all cases (Fig. 6).

Polymerase chain reaction based on amplification of common primate respiratory pathogens was performed on the lung tissue of the necropsied animals to exclude the presence of other important viral pathogens. The methods used are summarized in Köndgen et al. 2008 [3]. All samples were negative for influenza A and B, parainfluenza, rhinovirus, coronavirus, adenovirus, human metapneumovirus, respiratory syncytial virus and enterovirus.

The six dead animals were high ranked closely related animals. Their loss induced severe ranking fights, a problem which is well known in rhesus monkey colonies. As a consequence of these heavy fights six more animals (animal No. 7–12) died due to severe injuries, stress and shock-symptomatic. *Streptococcus equi* subsp. *zooepidemicus* was isolated from the tonsils of three of these animals (animal No. 7–9).



**Fig. 6** Pulsed field gel electrophoresis of seven different *Streptococcus equi* subsp. *zooepidemicus* isolates from a *Macaca mulatta* colony in the German primate center in Göttingen.

## Discussion

A fatal outbreak of streptococcus infection occurred in one of six breeding colonies housed in an indoor/outdoor facility.

The source of infection with *S. equi* subsp. *zooepidemicus* in the present outbreak remains unknown. Contact to horsemeat, equines or domestic animals could reliably be excluded. Instead a human to animal transmission was suspected. Generally *S. equi* subsp. *zooepidemicus* can be transmitted by aerosols, via the oral route or through wound contamination. Aerosol transmission is most likely in the present case. It was assumed that contact to a visitor with upper respiratory disease led to the initial infection of two elderly and closely related animals.

Streptococci of Lancefield group C can be recovered from the pharynx of 1.5% of normal humans. Infections of the upper respiratory tract in humans are rare but may occur after contact with infected horses [5].

It was assumed that the infection was subsequently transmitted from animal to animal because ill and dead animals were closely related to each other. There was a high degree of relationship and close contact among them. Close contact seems to be necessary for the transmission of this agent. None of the animals in adjacent corrals situated at a distance of 2–3 m became infected. None of the staff members was ill before or during the time of the outbreak.

In horses *S. equi* subsp. *zooepidemicus* is a commonly isolated mucosal commensal that opportunistically invades after virus infections, transport stress or other immunosuppressive conditions. In the described monkey group the situation seems to be different. Bacteriologic investigation of tonsillar swabs indicates that there was no colonization or asymptomatic infection with *S. equi* subsp. *zooepidemicus* among the group members which could have been activated by a stressor. The only exceptions were three animals (animal No. 7–9) that died in the consequence of heavy position fights. In these animals streptococci were found in the tonsils but they developed no clinical findings. The three animals belonged to the same family and had been in close contact to the dead animals. They developed a purulent tonsillitis and it is speculative if these animals may have established a latent infection comparable to horses if they survived the attacks.

The streptococcus outbreak among rhesus monkeys clearly demonstrates the high susceptibility of these animals. This is confirmed by data from the literature describing outbreaks caused by *S. equi* subsp. *zooepidemicus* in non-human primates marked by sudden, explosive appearance and high fatality rate [1, 4, 7].

This observation should lead to a critical discussion of housing primates and other animal species in close vicinity of each other as is often the case in zoological gardens. Particularly equines should not be kept in the vicinity of primate facilities and there should be a strict separation between the different units regarding animal keepers and other staff members.

The problem of the described outbreak for the breeding colony was not only the initial fatal infection but also the death of the first six animals. The loss of these animals caused ranking fights, injuries and death of six more animals. As a consequence the group had to be split up and newly composed. The outbreak ended after the dramatic depopulation of the group and the intense cleaning and disinfection of the facility.

The disease caused heavy losses; not only the loss of the animals, but also a reduced reproductive rate and high costs for therapy. The experience from this outbreak leads to the conclusion that contact of important and valuable breeding colonies to visitors should be strictly avoided.

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