



# Draft Genome Sequences of Four *Saccharibacter* sp. Strains Isolated from Native Bees

Eric A. Smith,<sup>a</sup> Hoang Q. Vuong,<sup>b</sup> Delaney L. Miller,<sup>a</sup> Audrey J. Parish,<sup>a</sup> Quinn S. McFrederick,<sup>b</sup>  Irene L. G. Newton<sup>a</sup>

<sup>a</sup>Department of Biology, Indiana University, Bloomington, Indiana, USA

<sup>b</sup>Department of Entomology, University of California, Riverside, California, USA

**ABSTRACT** The genus *Saccharibacter* is currently understudied, with only one described species, *Saccharibacter floricola*, isolated from a flower. In an effort to better understand the microbes that come in contact with native bee pollinators, we isolated and sequenced four additional strains of *Saccharibacter* from native bees in the genera *Melissodes* and *Anthophora*. These genomes range in size from 2,104,494 to 2,316,791 bp (mean, 2,246,664 bp) and contain between 1,860 and 2,167 (mean, 2,060) protein-coding genes.

The genus *Saccharibacter* currently comprises only one described species, isolated from the pollen of Japanese flowers (1). *Saccharibacter floricola* is an acetic acid bacterium (AAB). AAB have been shown to be associated with diverse insects that rely on sugar-rich diets (2, 3). As the genus *Saccharibacter* has been isolated from flowers, it may also come in contact with insect pollinators and play a role in their utilization of nectar as a food source. To determine whether this is the case and to bolster the available data for this potentially important genus, we isolated and sequenced the genomes of four *Saccharibacter* sp. strains isolated from native bee pollinators.

Four strains of *Saccharibacter* were isolated from native bees. Strains EH60, EH70, and E611 were isolated from bees in the genus *Anthophora*, and strain 17.LH.SD was isolated from a bee in the genus *Melissodes*, collected from Yosemite National Park, CA, and The Dalles, OR, respectively. Initial samples were streaked on MRS plus 2% fructose agar plates, after which single colonies were picked and grown in liquid culture. Strains were grown in yeast-peptone-glucose medium at 30°C with aeration. Total DNA was extracted using the DNeasy blood and tissue kit (Qiagen, Hilden, Germany), and sequencing library preparation was performed using the NEBNext Ultra II DNA library preparation kit (NEB, Ipswich, MA), both according to the manufacturers' protocols. The resulting libraries were subjected to 250-bp paired-end sequencing on the Illumina NextSeq 500 platform (version 2 chemistry) (strain 17.LH.SD was sequenced on the Illumina MiSeq platform [version 2 chemistry] with 300-bp paired-end sequencing) at the Indiana University Center for Genomics and Bioinformatics (Bloomington, IN). These sequencing runs generated 104,267 to 941,404 (mean, 551,094) read pairs.

Initial *de novo* assembly of these strains was performed using MaSuRCA version 3.2.8 with default settings (4). Reads were not subjected to quality control (QC) prior to assembly, as MaSuRCA performs internal QC during the assembly process. Additionally, reads were randomly subsampled down to approximately 50× coverage prior to assembly (100× for strain EH70). The completeness of the assemblies was assessed using both BUSCO (version 3; alphaproteobacteria lineage data set) (5) and CheckM (version 1.1.1) (6). Both of these tools indicated that the assemblies were >99% complete.

The assemblies resulted in 13 to 30 (mean, 23) contigs comprising 2,104,494 to 2,316,791 bp (mean, 2,246,664 bp), with an  $N_{50}$  contig length of 297,436 to 461,108 bp

**Citation** Smith EA, Vuong HQ, Miller DL, Parish AJ, McFrederick QS, Newton ILG. 2020. Draft genome sequences of four *Saccharibacter* sp. strains isolated from native bees. *Microbiol Resour Announc* 9:e00022-20. <https://doi.org/10.1128/MRA.00022-20>.

**Editor** Julie C. Dunning Hotopp, University of Maryland School of Medicine

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Address correspondence to Irene L. G. Newton, [irnewton@indiana.edu](mailto:irnewton@indiana.edu).

**Received** 9 January 2020

**Accepted** 14 February 2020

**Published** 5 March 2020

**TABLE 1** Relevant assembly information and statistics

Strain	GenBank accession no.	No. of contigs	Genome size (Mb)	% GC content	$N_{50}$ (bp)	$L_{50}$	Host genus	Isolation location <sup>a</sup>
EH60	<a href="#">WVHQ00000000</a>	30	2.32	51.47	301,953	4	<i>Anthophora</i>	Yosemite NP, CA
EH611	<a href="#">WVHP00000000</a>	25	2.28	51.47	303,728	4	<i>Anthophora</i>	Yosemite NP, CA
EH70	<a href="#">WVHN00000000</a>	26	2.29	50.70	297,436	4	<i>Anthophora</i>	Yosemite NP, CA
17.LH.SD	<a href="#">WVHO00000000</a>	13	2.10	48.61	461,108	2	<i>Melissodes</i>	The Dalles, OR

<sup>a</sup> NP, National Park.

(mean, 341,556 bp). The GC content of these strains was 48.61 to 51.47% (mean, 50.56%). Annotation was carried out with NCBI's Prokaryotic Genome Annotation Pipeline (PGAP) (7), which predicted between 1,860 and 2,167 (mean, 2,060) protein-coding genes and 1 to 4 (mean, 2) rRNAs. Assembly statistics are summarized in Table 1.

**Data availability.** These whole-genome shotgun projects have been deposited at DDBJ/ENA/GenBank under the accession numbers [WVHN00000000](#), [WVHO00000000](#), [WVHP00000000](#), and [WVHQ00000000](#). The versions described in this paper are versions WVHN01000000, WVHO01000000, WVHP01000000, and WVHQ01000000. Sequencing reads have been deposited in the Sequence Read Archive (SRA) under the accession numbers [SRR10728926](#) to [SRR10728929](#).

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