

Multifaceted effects of soluble human CD6 in experimental cancer models

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ABSTRACT

Background CD6 is a lymphocyte surface co-receptor physically associated with the T-cell receptor (TCR)/CD3 complex at the center of the immunological synapse. There, CD6 assists in cell-to-cell contact stabilization and modulation of activation/differentiation events through interaction with CD166/ALCAM (activated leukocyte cell adhesion molecule), its main reported ligand. While accumulating evidence is attracting new interest on targeting CD6 for therapeutic purposes in autoimmune disorders, little is known on its potential in cancer. In an attempt to elucidate the *in vivo* relevance of blocking CD6-mediated interactions in health and disease, we explored the consequences of expressing high circulating levels of a soluble form CD6 (sCD6) as a decoy receptor.

Methods High sCD6 serum levels were achieved by using transgenic C57BL/6 mice expressing human sCD6 under the control of lymphoid-specific transcriptional elements (shCD6LckEμTg) or wild type either transduced with hepatotropic adeno-associated virus coding for mouse sCD6 or undergoing repeated infusions of recombinant human sCD6 protein. Characterization of sCD6-induced changes was performed by *ex vivo* flow cytometry and functional analyses of mouse lymphoid organ cells. The *in vivo* relevance of those changes was explored by challenging mice with subcutaneous or metastatic tumors induced by syngeneic cancer cells of different lineage origins.

Results Through a combination of *in vitro* and *in vivo* studies, we show that circulating sCD6 expression induces defective regulatory T cell (Treg) generation and function, decreased CD166/ALCAM-mediated tumor cell proliferation/migration and impaired galectin-induced T-cell apoptosis, supporting the fact that sCD6 modulates antitumor lymphocyte effector function and tumorigenesis. Accordingly, sCD6 expression *in vivo* resulted in delayed subcutaneous tumor growth and/or reduced metastasis on challenge of mice with syngeneic cancer cells.

Conclusions Evidence is provided for the disruption of CD6 receptor–ligand interactions as a feasible immunomodulatory approach in cancer.

BACKGROUND

Immunomodulation is gaining ground for the treatment of several disorders involving either excessive (eg, autoimmune) or

defective (eg, cancer) immune responses by targeting the on/off molecular switches that set the intensity and/or duration of ongoing immune responses. These include co-stimulatory (eg, CD28) or inhibitory (eg, CTLA-4) membrane-bound lymphocyte receptors present at the immunological synapse (IS), a supramolecular activation complex at the interface between antigen-presenting cells (APCs) and T cells.¹

CD6 is a lymphocyte surface receptor physically associated with the T-cell receptor (TCR)/CD3 complex at the center of the IS, with roles in the stabilization of this structure and in T cell activation and differentiation signaling.² CD6 variants have been identified as susceptibility or disease modifier markers for multiple sclerosis, inflammatory bowel disease, Behçet's disease, psoriasis, and rheumatoid arthritis, and an anti-CD6 monoclonal antibody (mAb) is now under therapeutic consideration in some autoimmune disorders.³ These reports suggest that CD6 is a suitable candidate for immunomodulatory targeting.

CD6 is expressed on thymocytes and mature T cells, a subset of B (B1a) and natural killer (NK) cells, and on certain hematopoietic precursors and brain regions.² Its extracellular region is composed of three tandem scavenger receptor cysteine-rich (SRCR) domains, from which the membrane-proximal domain (D3) binds to the amino-terminal domain (D1) of activated leukocyte cell adhesion molecule (ALCAM), a member of the immunoglobulin (Ig) superfamily of cell adhesion molecules present on activated lymphocytes, professional APCs (dendritic cells, macrophages and B cells), thymic epithelial cells, endothelial cells, and brain cells.⁴ Additional interactions mediated by the CD6 ectodomain include galectin (Gal)-1 and Gal-3, conserved bacterial cell

wall components, and more recently, CD318/CDCP1 (CUB domain-containing protein 1).^{5,6} The intracellular domain of CD6 lacks intrinsic catalytic activity but can be phosphorylated by Ser/Thr and Tyr kinases to mediate signal transduction. The signaling pathway used by CD6 is only partially known and involves activation of mitogen-activated protein kinases, and recruitment of intracellular effectors such as Syntenin-1, GADS/SLP-76 and TSA.²

In addition to its co-stimulatory lymphocyte receptor function, data from *cd6* knockout (*cd6*^{-/-}) mice reveal that CD6 downregulates TCR signaling in immature thymocytes, with an impact on peripheral effector/memory (T_{EM}) and regulatory (Treg) T cell function.⁷ Recent *in vitro* evidence from *cd6*^{-/-} mice shows that CD6 is also a negative regulator of activation and activation-induced cell death for peripheral T cells.⁸

As for a large number of other lymphocyte surface receptors, low circulating levels (picomolar) of soluble human CD6 (shCD6) generated mainly by proteolytic cleavage on TCR/CD3-mediated activation can be detected in normal serum, and in serum from patients with systemic autoimmune or inflammatory disorders.⁹⁻¹¹ Recombinant shCD6 inhibits T-cell proliferation *in vitro* to an extent comparable with CD6-blocking antibodies or recombinant ALCAM-Fc protein, suggesting that CD6 and ALCAM are a key receptor–ligand pair in APC–T-cell interactions.¹²⁻¹⁴

To gain further insight into the role of CD6 in immune responses, we studied the effects of increasing the serum concentration of shCD6 in mice under normal and disease-inducing conditions. We hypothesized that shCD6 could abolish CD6 function by acting as a decoy receptor in mice and blocking homophilic and/or heterophilic interactions between CD6 ligands. Indeed, the same interface of ALCAM is involved in heterophilic (CD6–ALCAM) and homophilic (ALCAM–ALCAM) interactions, where heterophilic interactions show higher affinity and better conservation between mouse and human.^{15,16} To test our hypothesis, we generated a new transgenic mouse line (shCD6LckE μ Tg) that expresses shCD6 under the control of lymphoid-specific transcriptional elements active from early T and B cell developmental stages.¹⁷ Transgenic expression of solubilized receptors (eg, CTLA-4, IL-6R, TNFRp55, CD14, VEGFR-3 or CR1/CD35) has proved useful to understand their *in vivo* function and therapeutic potential.¹⁸⁻²² This strategy also favors eventual translation to the clinical practice through established and approved protein delivery systems.

The shCD6LckE μ Tg mice exhibited delayed growth of non-orthotopic tumors, which was reproduced in wild-type (WT) mice by infusion of recombinant shCD6 protein. *In vitro* studies showed that mechanisms operating at the level of lymphocyte effector function and tumorigenicity were engaged in the presence of shCD6. This provides the first proof-of-concept therapeutic potential of shCD6 in cancer.

MATERIALS AND METHODS

Animals

Transgenic shCD6E μ LckTg mice were generated in C57BL/6N background. Non-transgenic (NonTg) mice used for comparative purposes came from the same common heterozygous ancestors as the shCD6E μ LckTg mice and were kept under the same housing conditions as the latter. The shCD6LckE μ Tg mice were viable and fertile, and their lifespan and behavior were equivalent to those of NonTg mice. WT C57BL/6N mice were purchased from Charles River Laboratories. All animals were kept under specific pathogen-free (SPF) conditions and used at 7–8 weeks of age.

Cells and proteins

Spleen, inguinal lymph nodes (LNs) and thymus cell suspensions were obtained by pressing with a syringe plunger through a 40 μ m cell strainer (Biologix Research), and erythrocyte-depleted by using red blood cell lysis buffer (eBioscience).

C57BL/6-derived B16.F0, MCA205, EL-4, and RMA-S tumor cell lines were obtained from Dr R. Alemany (Institut Català d'Oncologia, L'Hospitalet de Llobregat, Spain) and Dr P. Lauzurica (Instituto de Salud Carlos III, Madrid, Spain). B16.F0 cells were grown in Dulbecco's Modified Eagle medium (DMEM, Invitrogen) supplemented with 10% fetal bovine serum (FBS) and 100 IU/mL penicillin/streptomycin. MCA205, EL-4, and RMA-S cells were grown in Rosswell Park Medium Institut (RPMI) 1640 medium (Lonza) supplemented with 10% FBS, 10 mM HEPES, 2 mM glutamine, 1 mM sodium pyruvate and 100 IU/mL penicillin/streptomycin.

Production and purification of endotoxin-free purified recombinant shCD6 and shCD5 proteins (in PBS with 10% glycerol, pH 7.4) from SURE CHO-M cell line clones was done following previously reported methods.²³ Human (HSA) and bovine (BSA) serum albumin were from Sigma-Aldrich.

Generation of shCD6LckE μ transgenic mice

The codon-optimized cDNA sequence encoding the whole ectodomain of human CD6 (GenBank X60992.1) from the initiation M1 to R373 was synthesized at GenScript (New Jersey, USA) and cloned into the *Bam*HI site of the p1026x vector (kindly provided by Dr R.M. Perlmutter), just downstream of the *lck* proximal promoter and the Ig heavy chain enhancer (E μ).¹⁷ Transgenic mice were generated at PolyGene (Rümlang, Switzerland) by injecting the purified *NotI*–*NotI* fragments into zygotes from superovulated C57BL/6N female mice mated to C57BL/6N breeder males (Charles River). Injected zygotes were cultured overnight and transferred into pseudopregnant B6xCBAF₁ females (Charles River). PCR screening for transgene integration identified 2 positive out of 14 pups. One of them was bred to homozygosity and maintained in parallel with WT mice from the same original cross. Tail genomic DNA was extracted by the KAPA Mouse Genotyping Kit (Kapa Biosystems) and subjected to PCR amplification with primer pairs specific

for the CD6 ectodomain (Fw 5'-ACC TGA CCA GCT CAA CAC-3' and Rv 5'-GTC ATC GCA CAC TGA TCC-3') and the MHC class II invariant Ii chain (Fw 5'-TCA CTC AAG GCA ACC TTC CTGC-3' and Rv 5'-CGA CCT CAT CTC TAA CCA TGA ACAG-3'), resulting in 495 bp and 150 bp fragments, respectively.

Construction and production of recombinant adeno-associated virus (AAV)

Construction and production of recombinant AAV8 virus expressing soluble mouse CD6 (AAV-smCD6) or luciferase (AAV-Luc) proteins under the control of a chimerical liver-specific (EalbAAT) promoter was carried out as previously reported.²³

ELISA assays

The plasma levels of shCD6 and smCD6 were quantified with the human soluble CD6 PicoKine ELISA Kit (Boster Bio) and the DuoSet mouse CD6 kit (R&D Systems), respectively, following manufacturer's instructions.

Flow cytometry analyses

Surface stainings were performed by blocking cells in phosphate-buffered saline (PBS) plus 10% FBS for 30 min at 4°C with rat anti-mouse CD16/32 (2.4G2; Mouse BD Fc Block) and further incubation with the following labeled mAb and appropriate isotype controls: biotin-labeled anti-BP-1 (6C3, eBioscience); fluorescein isothiocyanate (FITC)-labeled anti-CD4 (RM4-5, Tonbo Biosciences), anti-B220 (Ra3-6B2, Tonbo), anti-CD44 (IM7, BD Biosciences), and anti-CD24 (M1/69, BD Pharmingen); phycoerythrin-labeled anti-IgD (11-26, SouthernBiotech), anti-NK1.1 (PK136, BD Biosciences), anti-CD8 (53-6.7, BioLegend), anti-FoxP3 (3G3, Tonbo), anti-CD19 (1D3, Tonbo), and anti-ALCAM (eBioALC48, eBiosciences); PerCP-Cy5.5-labeled anti-CD3 (145-2 C11, Tonbo), anti-CD8 (53-6.7, Tonbo), anti-CD44 (IM7, Tonbo), and streptavidin (BD Biosciences); allophycocyanin-labeled anti-CD3 (145-2 C11, Tonbo), anti-CD62L (MEL-14, Tonbo), anti-CD3e (145-2 C11, Tonbo), anti-IgM (II/41, eBioscience), anti-LAP (TW7-16B4, Biolegend), and anti-CD25 (PC61.5, eBiosciences); APC-Cy7-labeled anti-CD25 (PC61, BD Pharmingen); Pe-Cy7-labeled anti-CD5 (53-7.3, eBiosciences), anti-CD19 (1D3, Tonbo), and anti-CD4 (GK1.5, BioLegend); violetFluor 450-labeled anti-B220 (RA3-6B2, Tonbo) and anti-CD8 (53-6.7, Tonbo). FoxP3 intracellular staining was performed with the phycoerythrin-labeled anti-mouse/rat Treg Staining Kit (eBioscience), according to the manufacturer's instructions. Data were analyzed in a FACSCanto II flow cytometer (Becton Dickinson) using FlowJo software (Tree Star).

Bromodeoxyuridine incorporation and Treg suppression assays

Detection of in vivo proliferating cells with bromodeoxyuridine (BrdU) was performed as reported elsewhere.²⁴ Treg isolation and suppressive activity assays were performed as previously reported.²⁵

CD4⁺ T cell polarization assays

Naïve CD4⁺ T cells (CD4⁺CD25[−]CD62L^{hi}CD44^{lo}; 1×10⁵) from C57BL/6/J spleens sorted with Dynabeads Untouched Mouse CD4 cells Kit (Thermo Fisher Scientific) were activated for 72 hours in triplicate in 96-well U-bottom plates with coated anti-CD3 (2 µg/mL) plus soluble anti-CD28 (0.5 µg/mL) mAb under standard Treg, Th1, Th2 or Th17 polarization conditions (see <https://BioLegend.com/en-us/technical-protocols>) in the presence or absence of shCD6 protein (0–10 µg/mL). Cells were stained for surface CD4 (RM4-5) and intracellular FoxP3 (3G3) expression for Treg analysis. For Th1, Th2 and Th17 analysis, cells were restimulated for 5 hours with phorbol myristate acetate (PMA, 80 nM) and ionomycin (1 µg/mL) in the presence of 2 µM monensin followed by surface CD4 and intracellular IFN-γ (XMG1.2, Tonbo), IL-4 (11B11, BD Biosciences) or IL-17 (eBio17B7, eBioscience) staining, respectively.

In vivo tumor cell growth and metastasis assays

Assessment of local subcutaneous growth and lung metastatization of syngeneic tumor cells in vivo was performed as reported elsewhere.^{25,26}

Quantitative real-time PCR analyses

Total mRNA (0.5 µg) extracted from cell lines by using the PureLink RNA Mini Kit (Ambion, Life Technologies) was retrotranscribed into cDNA by using the high-capacity cDNA Kit (Life Technologies). Galectin mRNA levels were assessed by quantitative real-time PCR (qRT-PCR) analysis (Taqman Fast Universal PCR Master Mix; Life Technologies) using a 7900HT fast real-time PCR System (Applied Biosystems) and the following Taqman probes (Thermo Fisher): Gal-1 (Mm00839408_g1), Gal-3 (Mm00802901_m1) and GAPDH (Mm99999915_g1). Galectin mRNA expression was adjusted by GAPDH expression as 2^{ΔCt}, where ΔCt=Ct (GAPDH) – Ct (gene of interest).

Galectin-induced apoptosis assays

C57BL/6J splenocytes (1×10⁵) activated for 72 hours in 12-well plates with coated anti-CD3 (145-2 C11; 1 µg/mL, Tonbo) plus soluble anti-CD28 (37.51; 5 µg/mL, Tonbo) mAb were further suspended in FBS-free RPMI medium for 30 min at 37°C in the presence of 100 µg/mL of endotoxin-free glutathione S-transferase (GST) and fusion GST-Gal1 and GST-Gal3 proteins, produced and purified as reported elsewhere.⁵

Apoptosis assays

Apoptotic cells were detected using the Annexin V-FITC apoptosis detection kit following the manufacturer's instructions (ImmunoStep).

In vitro tumor cell growth and migration assays

Tumor cell proliferation was monitored as reported elsewhere.²⁷ Wound healing technique was used to assess tumor cell migration.²⁸

Statistical analyses

Statistical analyses and data representation were performed with GraphPad Prism V.6 Software.

RESULTS

Generation and immunophenotypic analysis of shCD6LckE μ Tg mice

CD6-mediated interactions were investigated *in vivo* by generating a transgenic mouse line (shCD6LckE μ Tg) expressing the soluble ectodomain of human CD6 (shCD6) in a C57BL/6N background. The transgene was under control of lymphocyte-specific transcriptional elements (*lck* proximal promoter and E μ enhancer) (figure 1A), which drive shCD6 expression mainly to primary and secondary lymphoid organs. Identification of transgenic mice was performed by PCR amplification of a 495 bp fragment corresponding to the human CD6 ectodomain (figure 1B), and ELISA measurement of plasma shCD6 levels, which yielded 79.2 \pm 19.7 and 38.7 \pm 6.3 ng/mL for homozygous and heterozygous mice, respectively (figure 1C).

Primary lymphoid organ analyses (online supplementary figure S1A and S1B) showed no differences between shCD6LckE μ Tg and NonTg mice regarding thymus and bone marrow total cell numbers, as well as the frequency of all B-cell developmental subsets (figure 1D, left and middle). In contrast, shCD6LckE μ Tg presented significantly higher fractions of double-negative (DN) and CD8 single-positive (CD8SP) thymocytes (figure 1D, right), suggesting a role for CD6-mediated interactions in T cell but not B-cell developmental process.

The analysis of peripheral lymphoid organs revealed decreased total cell numbers in spleen and LN from shCD6LckE μ Tg mice compared with NonTg controls (figure 1E, left). Annexin V and 7-AAD staining of total spleen and LN cells showed no differences between shCD6LckE μ Tg and NonTg mice regarding cell apoptosis levels (figure 1E, right). *In vivo* BrdU-incorporation assays revealed a decrease in the percentage of BrdU⁺ cells in spleen and LN from shCD6LckE μ Tg mice (figure 1F). A more detailed analysis (online supplementary figure S1C and S1E) showed that lower BrdU incorporation affected the B cell (CD19⁺) compartment in both spleen and LN. These results indicate that a lower proliferative capacity of B cells is behind the reduced cellularity found in the peripheral lymphoid organs of shCD6LckE μ Tg mice.

Another consistent finding in peripheral (spleen and LN) but not central (thymus) lymphoid organs from shCD6LckE μ Tg mice was the presence of decreased percentages of CD25⁺FoxP3⁺ cells within the CD4⁺ subset (Treg) (figure 2A and online supplementary figure S2). Moreover, Treg cells from shCD6LckE μ Tg mice showed lower expression levels of latency-associated peptide (LAP) (figure 2B), a well-known surface marker directly correlating with Treg functionality.²⁹ In agreement with this finding, *in vitro* suppression assays confirmed that CD4⁺CD25^{+/hi} T cells (Treg) from shCD6LckE μ Tg

mice exhibited lower suppressive activity on activated CD4⁺CD25⁻ T cells (Tconv) at high Tconv:Treg (8:1) ratios than those from NonTg mice (figure 2C).

The putative effects of shCD6 on T cell polarization were further explored *in vitro* by culturing naïve CD4⁺ T cells (CD4⁺CD25⁻CD62L^{hi}CD44^{lo}) from WT mice under Th1, Th2, Th17 and Treg cell polarization conditions in the presence of increasing doses of shCD6 (0.1, 1 and 10 μ g/mL). As illustrated in figure 2D, dose-dependent reduction in the percentage of polarized Treg but not Th1, Th2 or Th17 cells was observed. Moreover, a reduction in the surface expression levels of LAP was observed in *in vitro* polarized Treg cells in the presence of shCD6 (10 μ g/mL) (figure 2E), pointing to a role for shCD6 on the reduced frequency and/or functionality of peripheral Treg cells in shCD6LckE μ Tg and shCD6-treated mice.

To assess the *in vivo* functional relevance of our immunophenotypic observations, shCD6LckE μ Tg mice were challenged by subcutaneous injection with syngeneic cancer cells. As shown in figure 3 (left), growth of melanoma B16.F0 tumor cells was slower in shCD6LckE μ Tg mice than in NonTg controls. Tumor weight at the end of the follow-up period (day 20) was not statistically significant. Similar analyses performed with syngeneic tumor cells of fibrosarcoma (MCA205) and lymphoma (RMA-S) origin also showed reduced tumor growth and weight for shCD6LckE μ Tg mice (figure 3, middle and right), indicating an enhanced and non-cell lineage-specific anti-tumor response.

Infusion of shCD6 to WT mice induces immunophenotypical changes in peripheral lymphoid organs and reduced growth of subcutaneously implanted syngeneic cancer cells

In order to rule out transgenesis artifacts, immunophenotypical analyses were performed following repeated administration of shCD6 protein (1.25 mg/kg, every-other-day intraperitoneal for 15 days) to WT mice. Thymus analyses showed no differences between shCD6-treated and vehicle-treated mice regarding total cell numbers (figure 4A) and the frequency of T-cell developmental subsets (online supplementary figure S3). In the periphery, statistically significant reductions of total cell numbers were detected in spleen but not LN from shCD6-treated mice (figure 4A). The percentage of CD25⁺FoxP3⁺ cells within the CD4⁺ subset was also found significantly decreased in LN but not spleen samples from shCD6-treated mice (figure 4B). Thus, infusion of shCD6 on alternate days for a limited period of time reproduced some of the immunophenotypical changes observed in shCD6LckE μ Tg.

To investigate whether infusion of shCD6 protein also delays tumor growth *in vivo*, WT mice were subcutaneously implanted with B16.F0 cells and then intraperitoneally infused with 1.25 mg/kg shCD6 every-other-day from the day of tumor implantation. As illustrated in figure 5, shCD6-treated mice showed a statistically significant reduction in B16.F0 tumor cell growth compared with vehicle-treated mice. Similar results were observed when

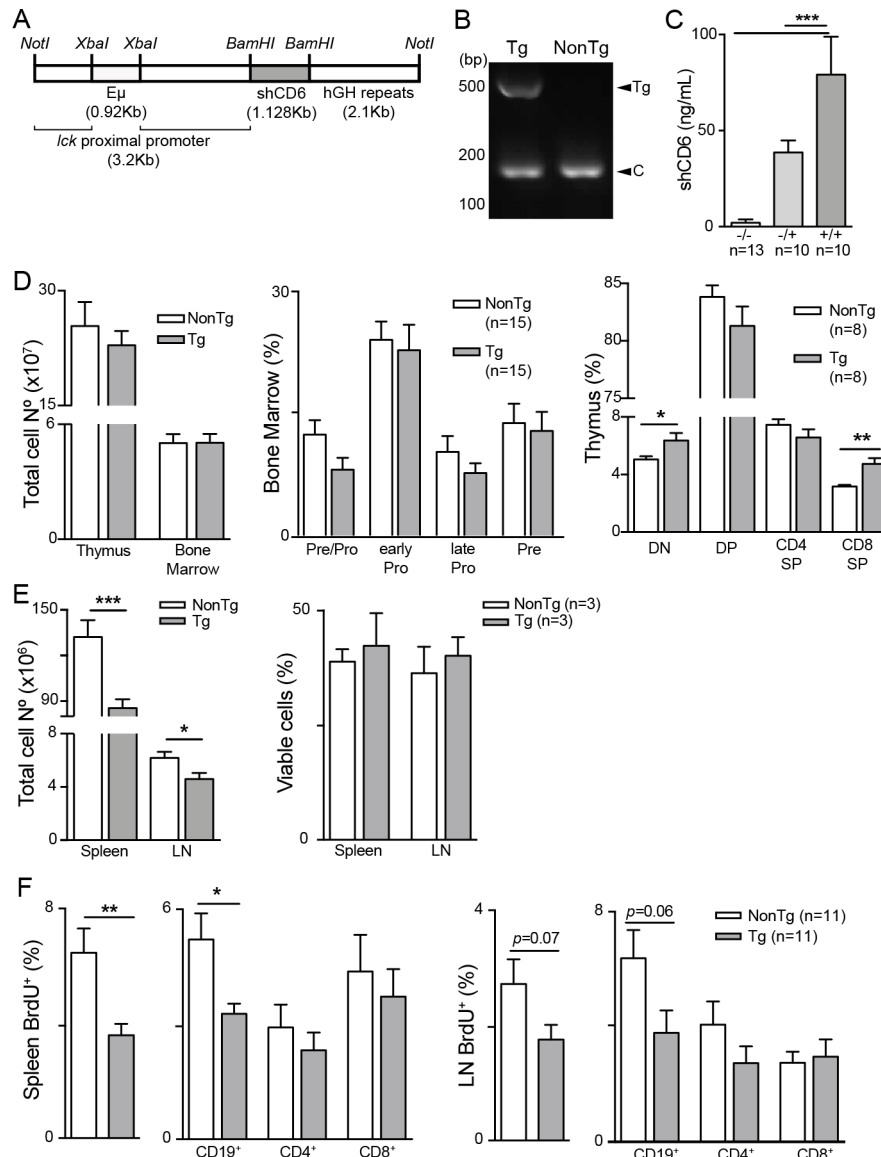


Figure 1 Generation and immunophenotypical characterization of shCD6LckEμTg mice. (A) Schematic representation of the transgene coding for soluble human CD6 (shCD6). (B) PCR screening of genomic DNA from homozygous shCD6LckEμTg and non-transgenic (NonTg) mice. Bands correspond to transgene (Tg) and internal PCR control (C). (C) ELISA quantification of plasma shCD6 levels from NonTg (-/-), and heterozygous (-/+) and homozygous (+/+) shCD6EμLckTg (Tg) mice. Represented are cumulative data from two independent experiments, expressed as mean±SEM. ***p<0.001; two-tailed Student's t-test. (D) Immunophenotypical characterization of primary lymphoid organs from shCD6LckEμTg mice. Left: total lymphoid cell numbers from thymus and bone marrow of shCD6LckEμTg (n=21 and n=13, respectively) and NonTg (n=15 and n=16, respectively) mice. Middle: percentage of CD24^{low}BP1⁻ (pre-pro B cells), CD24⁺BP1⁻ (early pro B cells), CD24⁺BP1⁺ (late pro B cells) and CD24^{high}BP1⁺ (pre B cells) within gated CD43⁺B220⁺ bone marrow cells from shCD6LckEμTg and NonTg mice. Right: percentage of CD4⁻CD8⁻ (DN), CD4⁺CD8⁺ (DP), CD4⁺CD8⁻ (CD4SP) and CD4⁺CD8⁺ (CD8SP) thymocytes from shCD6LckEμTg and NonTg mice. Represented are cumulative data from at least two independent experiments, expressed as mean±SEM. *p<0.05; **p<0.01; ***p<0.001; two-tailed Student's t-test. (E) Immunophenotypical characterization of secondary lymphoid organs from shCD6LckEμTg mice. Left: total lymphoid cell numbers in spleen, and lymph node (LN) from shCD6LckEμTg (n=50 and n=23, respectively) and NonTg (n=61 and n=28, respectively) mice. Right: percentage of viable cells (7AAD⁻AnnexinV⁻) from spleen and LN of shCD6LckEμTg and NonTg mice. One of two independent experiments is shown. (F) In vivo Bromodeoxyuridine (BrdU)-incorporation assays in secondary lymphoid organs from shCD6LckEμTg mice. Percentage of spleen (left) and LN (right) BrdU⁺ lymphoid cells represented as total cells and as cells gated on CD19⁺, CD4⁺ and CD8⁺. Represented are cumulative data from at least two independent experiments, expressed as mean±SEM. *p<0.05; **p<0.01; ***p<0.001; two-tailed Student's t-test.

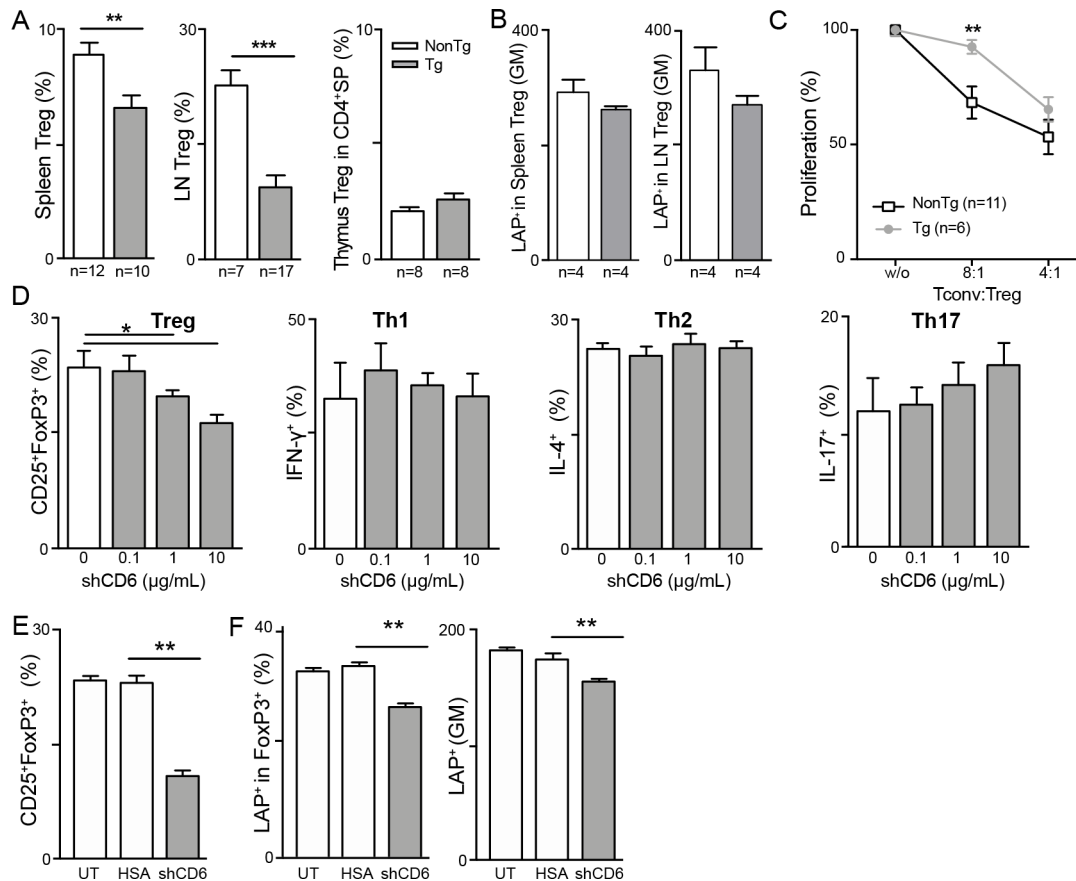


Figure 2 Effects of soluble human CD6 (shCD6) on the frequency and functionality of Treg cells. (A) Percentage of CD25⁺FoxP3⁺ in CD4⁺ gated cells (Treg) from spleen, lymph node (LN) and thymus of shCD6LckEμTg and non-transgenic (NonTg) mice. (B) Geometric mean fluorescence intensity (GM) for latency-associated peptide (LAP) expression on spleen and LN Treg cells (defined as in a) from shCD6LckEμTg and NonTg mice. (C) Percentage of proliferation of CD4⁺CD25⁻ (Tconv) cells from NonTg mice stimulated for 72 hours with anti-CD3 mAb plus anti-CD28 monoclonal antibody (mAb) in the absence (w/o) or presence of different ratios of CD4⁺CD25⁺ (Treg) cells from shCD6LckEμTg or NonTg mice. (D) Sorted naïve CD4⁺CD25⁻CD62L^{hi}CD44^{lo} splenocytes (1×10⁵) from C57BL/6J (n=5) were activated in triplicate for 72 hours with anti-CD3 (2 μg/mL) and anti-CD28 (0.5 μg/mL) mAbs under Treg, Th1, Th2, and Th17 polarization conditions, all in the presence or absence of different amounts of shCD6 (0, 0.1, 1 and 10 μg/mL). Percentage of Treg, Th1, Th2, and Th17 in CD4⁺ cells as deduced from intracellular FOXP3, IFN-γ, IL-4 and IL-17 co-staining, respectively, are represented as mean±SEM. **p<0.01; ***p<0.001; unpaired t-test with Welch's correction. (E) In vitro polarized Treg cells generated as in (D) and cultured alone (UT) or in the presence of 10 μg/mL HSA or shCD6 (left) were then subjected to flow cytometry analysis for surface LAP expression. percentage (%) and geometric mean fluorescence intensity (GM) of LAP⁺ Treg cells is shown (middle and right, respectively). *p<0.05; **p<0.01; ***p<0.001; two-tailed Student's t-test.

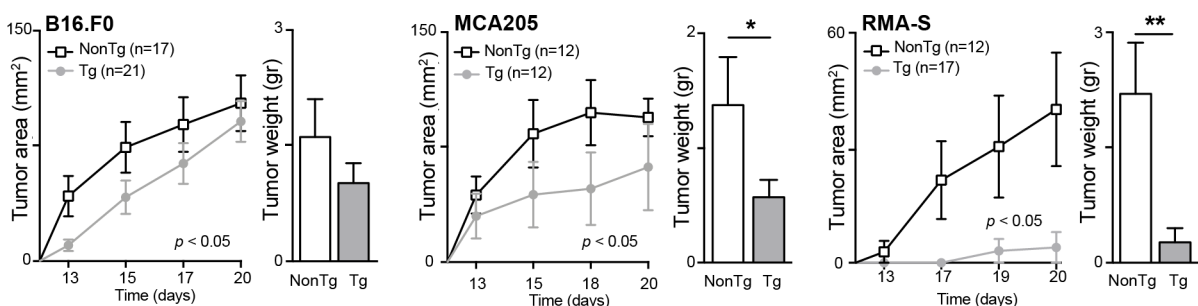


Figure 3 Growth curves of subcutaneously implanted syngeneic cancer cells in shCD6LckEμTg mice. B16.F0 (5×10⁴), MCA205 (5×10⁴) and RMA-S (2.5×10⁴) cells were injected subcutaneously into shCD6LckEμTg and non-transgenic (NonTg) mice. Tumor growth (area in mm²) and weight (GR) were measured every other day and at sacrifice, respectively. Cumulative data from two or three independent experiments are represented. Two-way analysis of variance and Mann-Whitney tests were used for statistical analysis of tumor growth and weight, respectively. Data are presented as mean±SEM.

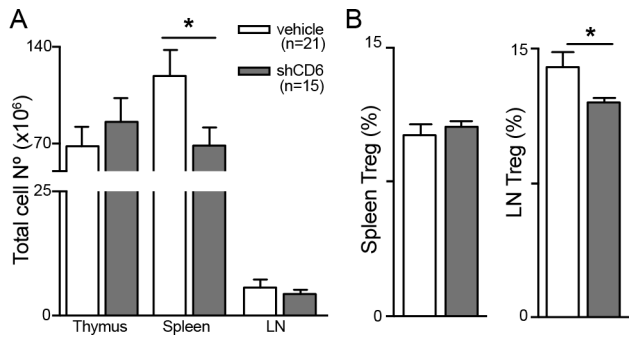


Figure 4 Immunophenotypical analysis of lymphoid organs from wild-type (WT) mice infused with recombinant soluble human CD6 (shCD6) protein. (A) Total lymphoid cell numbers in thymus, spleen, lymph node (LN) and peritoneum of WT mice after intraperitoneal administration of shCD6 (1.25 mg/kg) or vehicle (PBS 10% glycerol) every 48 hours for 14 days. (B) Percentage of CD25⁺FoxP3⁺ in gated CD4⁺T cells (Treg) from spleen and LN of same mice as in (A) is shown. Data represent cumulative results from two independent experiments expressed as mean±SEM. **p*<0.05; two-tailed Student's *t*-test.

shCD6-treated mice were challenged with MCA205, EL-4 or RMA-S cells (figure 5), indicating a role for circulating shCD6 in delayed tumor growth in shCD6LckEμTg mice in vivo.

As an approximation to clinical conditions, we explored the effect of every-other-day shCD6 infusion to WT mice bearing ~5–9 mm² B16.F0 tumors (~day 9 post subcutaneous implantation). Again, reduced B16.F0 tumor growth was observed in mice therapeutically treated with shCD6 protein (online supplementary figure S4).

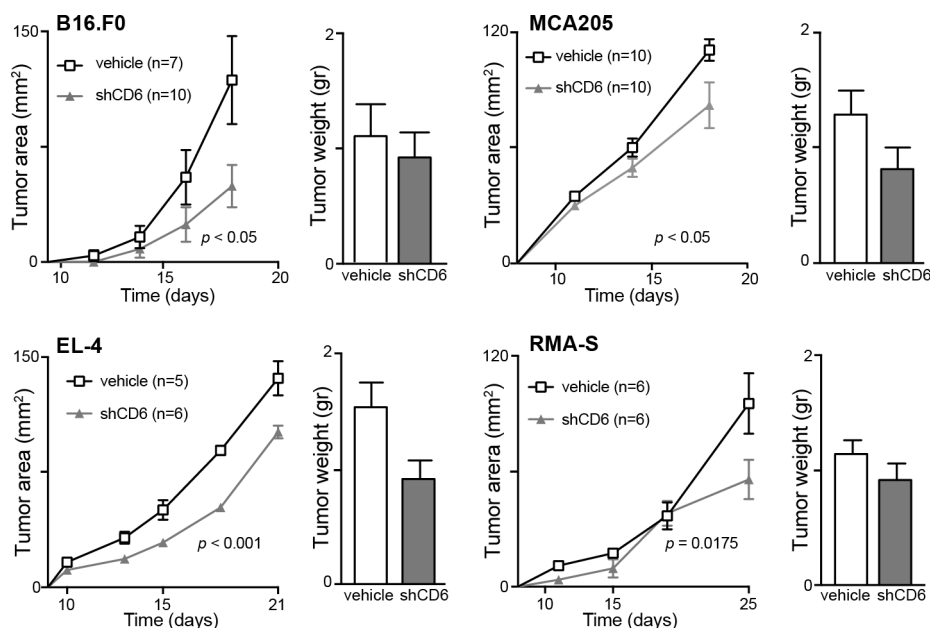


Figure 5 Growth curves of subcutaneously implanted syngeneic cancer cells into wild-type (WT) mice prophylactically infused with soluble human CD6 (shCD6) protein. B16.F0 (5×10^4), MCA205 (5×10^4), EL-4 (5×10^4), and RMA-S (5×10^4) cells were subcutaneously injected into WT mice receiving intraperitoneal injections of shCD6 (1.25 mg/kg) or vehicle (PBS 10% glycerol) every other day from tumor implantation. Tumor growth (area in mm²) and weight (GR) were measured every other day and at sacrifice, respectively. Two-way analysis of variance and Mann-Whitney tests were used for statistical analysis of tumor growth and weight, respectively. Data are presented as mean±SE.

AAV-mediated gene transfer of soluble mouse CD6 (smCD6) leads to reduced metastasis

The antitumor responses of WT mice transduced with hepatotropic AAV for sustained circulating smCD6 protein levels were next explored. Previous data from our group show that AAV-smCD6 transduced WT mice present detectable smCD6 levels from day 10 on, which peak at days 15 to 20, and remain sustained for longer than 3 months.²³ We reasoned that smCD6 would be less immunogenic than shCD6 in mice and its long-term expression and function would not be impaired by newly appearing neutralizing antibodies. Mice transduced with optimal AAV-smCD6 doses (10^{11} viral genome (v.g.)/mouse), as determined by dose–response experiments (online supplementary figure S5A), do not display delayed growth of subcutaneously implanted B16.F0 tumors compared with control mice (AAV-Luc) (figure 6A). In contrast, significantly lower number of lung metastases and improved survival was observed when AAV-smCD6-treated mice were challenged intravenously (tail vein) with B16.F0 cells (figure 6B and C). As illustrated in figure 6D, the effects of AAV-smCD6 mouse transduction on lung metastatization were dose-dependent, with optimal results at 10^{11} v.g./mouse. AAV-smCD6 treatment also resulted in reduced numbers of liver metastases when B16.F0 cells were injected into spleen (online supplementary figure S5B).

The possibility that AAV-mediated effects could be in part responsible for the reduced metastatization observed was further ruled out by performing lung metastases assays in which only purified sCD6 protein was infused. In order

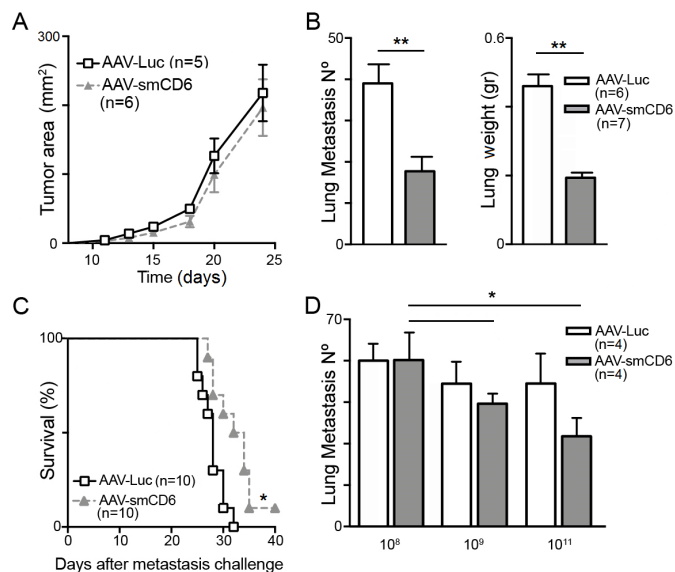


Figure 6 Local growth and metastatization of B16.F0 cells in wild-type (WT) mice transduced with adeno-associated virus (AAV) encoding smCD6 protein. (A) C57BL/6J mice administered viral genome with 10^{11} AAV-smCD6 or AAV-Luc viral genome (v.g./mouse) were challenged 2 weeks later with subcutaneous B16.F0 (5×10^4) cells. Tumor area (mm^2) monitored twice a week is represented. (B) C57BL/6J mice administered intravenously with 10^{11} AAV-smCD6 or AAV-Luc v.g./mouse were challenged intravenously (tail vein) 2 weeks later with B16.F0 (2×10^5) cells. Number of lung metastasis and lung weight (GR) was determined 3 weeks after challenge. (C) Kaplan-Meier survival curve of an independent experiment performed under the same conditions as in (B). (D) C57BL/6J mice transduced intravenously with 10^8 , 10^9 , 10^{11} AAV-smCD6 or AAV-Luc v.g./mouse were challenged 2 weeks later with B16.F0 (2×10^5) cells intravenously injected in the tail. Number of lung metastasis determined 3 weeks after challenge is represented. All data represent cumulative results from two independent experiments expressed as mean \pm SEM. * $p < 0.05$; ** $p < 0.01$; two-tailed Student's t-test.

to reproduce as much as possible the conditions used in the AAV experiments (prophylactic mode and high circulating sCD6 levels), WT mice were infused intraperitoneally with 1.25 mg/kg shCD6 or HSA 1 hour before and after intravenous challenge with 2×10^5 B16.F0 cells preincubated for 15 min with shCD6 or HSA. Under such experimental conditions, lower number of lung metastases was observed in shCD6-treated mice compared with HSA-treated controls (online supplementary figure S6).

shCD6 protein reduces in vitro galectin-induced T-cell apoptosis and tumor cell growth and migration

Abnormal tumor expression of Gal-1 and/or Gal-3 is known to influence tumor development, progression and metastasis through different mechanisms, one of them involving T-cell apoptosis.³⁰ All tumor cell lines used in this work express Gal-1 and/or Gal-3 at different levels, as deduced from mRNA analyses (figure 7A). Since CD6 is also known to interact with Gal-1 and Gal-3,⁵ the influence of shCD6 on galectin-induced T-cell apoptosis was further investigated. To this end, 72-hour activated spleen T cells

were exposed to GST-Gal-1 and GST-Gal-3 in the presence of increasing concentrations of shCD6 (10 and 20 $\mu\text{g}/\text{mL}$). As shown in figure 7B, dose-dependent reduction in Gal-1 and Gal-3-induced apoptotic T cells was induced by shCD6, indicating that shCD6 may enhance resistance of tumor-specific effector T cells to apoptosis mediated by either tumor or stroma cell-derived galectins.³¹

Extensive evidence indicates that ALCAM functions as a risk factor in cancer, with its different functional isoforms (soluble and membrane-bound) correlating with specific clinical outcomes depending on cancer type.³² Moreover, the interference of homophilic ALCAM-ALCAM interactions by mAbs or chimeric proteins has been shown to reduce tumor cell growth in vitro and in vivo.^{27 33 34} In light of these data, we analyzed the effect of shCD6 on in vitro growth of the tumor cell lines used in this study, which show different surface ALCAM levels (figure 7C and online supplementary figure S7). As illustrated in figure 7D, dose-dependent effects were induced by shCD6 but not HSA on the proliferation of tumor cell lines expressing higher ALCAM surface levels (B16.F0, EL-4, and MC205) compared with RMA-S cells, whose proliferation remained unaffected. Similar inhibitory effects were observed for shCD6 when B16.F0 cell migration was explored by performing wound healing assays (figure 7E).

DISCUSSION

Recent preclinical and clinical information is attracting new interest on targeting the CD6 lymphocyte surface receptor for therapeutic purposes.^{2 3} Genetically modified mouse models greatly contribute to increase our knowledge on the role of CD6 in vivo, as no human CD6 deficiencies have been reported. Accordingly, recent data from *cd6*^{-/-} mice are uncovering unsuspected CD6 activity in health and disease.^{7 8 24 35-37} However, gene knockdown results do not always reflect *bona fide* receptor-ligand interaction blockade. This is well illustrated by the role of ALCAM in neuroinflammation, where *alcam*^{-/-} mice show increased neuroinflammatory responses as opposed to that of mice treated with anti-ALCAM blocking mAbs.³⁸ In light of this, we reasoned that a soluble form CD6 acting as a decoy receptor in homotypic and/or heterotypic interactions between known (ALCAM, Gal-1 and Gal-3, and CD318/CDCP1)—and hitherto unknown CD6 ligand/s—would provide further information on CD6-ligand interaction blockade in vivo.

Our hypothesis was first tested by analyzing the new shCD6E μ LckTg transgenic mouse line. One advantage of such experimental approach is its translatability to the clinical practice by infusion of soluble recombinant forms of the subject receptor. The latter represents our second experimental approach: the treatment of WT animals with repeated infusions of shCD6 protein. Intermittent protein dosage, even if it does not allow for sustained circulating levels, has served us to exclude transgenesis artifacts in our findings using shCD6E μ LckTg mice.

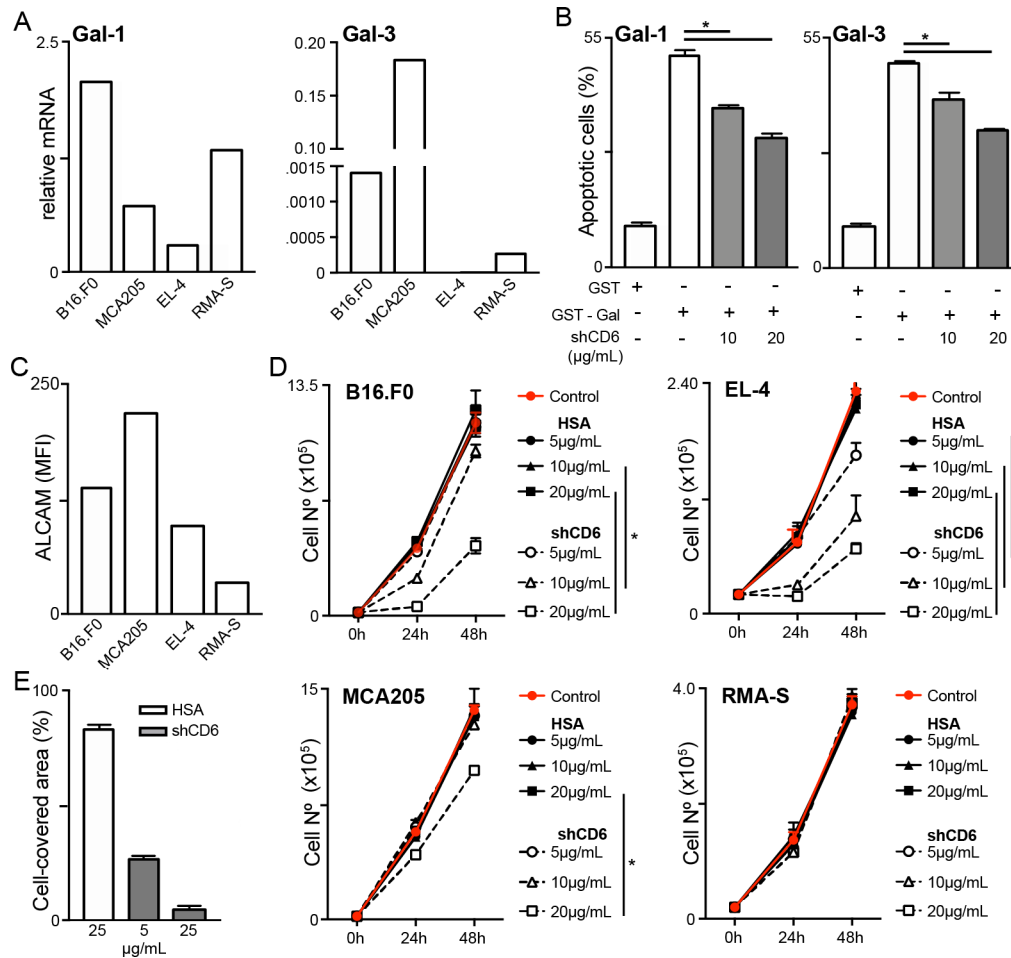


Figure 7 Inhibition of galectin-induced T-cell apoptosis and tumor cell growth and migration by soluble human CD6 (shCD6). (A) Relative Gal-1 and Gal-3 mRNA expression in the different tumor cell lines analyzed in this study, as assessed by qRT-PCR. (B) Splenocytes from wild-type (WT) C57BL/6J mice were activated for 72 hours with anti-CD3 (1 µg/mL) and anti-CD28 (5 µg/mL) monoclonal antibodies (mAbs) and then incubated for 30 min with 100 µg/mL of endotoxin-free glutathione S-transferase (GST), GST-Gal-1 or GST-Gal-3 in the presence of growing concentrations (10 and 20 µg/mL) of shCD6 protein (n=4/group). Percentage of apoptotic cells was determined by flow cytometry. (C) Indicated tumor cells were stained for surface ALCAM expression and mean fluorescence intensity (MFI) was determined by flow cytometry. (D) Tumor cells (2×10^4) were cultured with increasing concentrations (5, 10 and 20 µg/mL; n=4/group) of shCD6 or HSA and their numbers counted at 24-hour intervals. (E) Wound healing assays were used to measure the migratory activity of B16.F0 cells (5×10^5) seeded onto 12-well plates for 24 hours, followed by 2-hour exposure to mitomycin-C (50 µg/mL) and further 12-hour incubation with increasing concentrations (5 and 25 µg/mL) of shCD6 (n=4/group) or HSA control (n=3). Data represent percentage of cell-covered area. In all cases, cumulative results from two independent experiments expressed as mean \pm SEM are presented. *p<0.05; two-tailed Student's t-test. ALCAM, activated leukocyte cell adhesion molecule; HSA, human serum albumin.

The immunophenotypical analysis of primary lymphoid organs from shCD6EµLckTg mice under disease-free conditions did not reveal gross alterations. The only noteworthy finding was an increased frequency of DN and CD8SP thymocytes in conjunction with non-significant reductions in that of DP and CD4SP. This indicates that effective interaction between CD6 and its ligand/s expressed on the surface of developing thymocytes and/or thymic stromal cells (ie, ALCAM) would favor progression from DN to DP developmental stage, and limit that of DP to CD8SP. Interestingly, this thymic phenotype does not match the results from *cd6*^{-/-} C57BL/6 mice, where an increased frequency of DP undergoing selection concomitant with reduction in recently selected CD4SP and CD8SP thymocytes is observed.⁷ The disparity

between shCD6EµLckTg and *cd6*^{-/-} mice could relate to the inability of the latter to (1) interact with CD6 ligand/s and (2) transduce ligand-independent intracellular signals.

The immunophenotypical analysis of secondary lymphoid organs from shCD6EµLckTg mice revealed decreased total cell numbers in both spleen and LN, while this was only observed in spleens from shCD6-treated mice. This could be ascribed to a decrease in the in vivo homeostatic proliferative capability of mainly B cells. This again contrasts with analyses performed with *cd6*^{-/-} C57BL/6 mice, which revealed no reductions in total cell numbers from spleen and LN, and no defects in B cell proliferative responses in vitro.⁷ However, significantly decreased B1a cell population has been reported

cd6^{-/-} mice of DBA/1 background.³⁶ The fact that B cells (in contrast to T cells) constitutively express ALCAM³⁹ and that ALCAM-mediated cell-to-cell homotypic interaction is necessary to sustain proper proliferative responses in different cell types^{40–42} may be behind our observations.

A relevant finding was the reduced frequency of Treg cells in peripheral lymphoid organs from shCD6E μ LckTg and shCD6-treated mice, which also showed decreased suppressive activity in vitro. In vitro polarization experiments of naïve CD4⁺ T cells from WT mice confirmed that shCD6 reduces the frequency and functionality of Treg cells, contrary to observations from *cd6*^{-/-} C57BL/6 mice, which show increased frequency of peripheral Treg with lower suppressive activity.⁷ Since Treg cell function has been linked to SLP-76-mediated signaling^{43 44} and CD6 constitutes a key element of the SLP-76 interactome^{43 44} inappropriate recruitment of SLP-76 may be behind the Treg observations made in shCD6E μ LckTg and shCD6-treated mice.

The functional relevance of the immunophenotypical changes detected in shCD6E μ LckTg mice was explored by subcutaneous challenge with different syngeneic tumor cell lines. In all cases, tumor growth was significantly decreased in the shCD6E μ LckTg and/or shCD6-treated WT mice. The lower tumor growth in both experimental settings may result from the lower levels of peripheral Treg cells with reduced suppressive activity. However, *cd6*^{-/-} C57BL/6 mice show increased frequency of dysfunctional Treg cells⁷ but do not significantly differ from WT C57BL/6 mice regarding antitumor responses (online supplementary figure S8). Thus, alternative mechanisms may also contribute to the observed decrease in tumor growth. By using different experimental in vitro systems we demonstrate that shCD6 interferes with certain tumorigenicity-related events such as galectin-induced T-cell apoptosis, and ALCAM-mediated tumor cell proliferation and migration. Indeed, preliminary experiments with patient-derived melanoma cells (A375) engrafted into immunocompromised NSG mice indicate that shCD6 effects do not require a fully preserved immune system (online supplementary figure S9).

Galectins (namely, Gal-1 and Gal-3) are overexpressed in a broad range of cancers (by both stromal or malignant cells) and known to induce T-cell apoptosis contributing to immunosuppression within the tumor microenvironment.^{31 45–47} The tumorigenic properties of galectins are, however, not limited to immunosuppression but to promotion of cancer cell proliferation, migration, and invasion,⁴⁷ which could be also interfered by shCD6.

The most membrane-proximal SRCR ectodomain of CD6 (D3) is directly involved in the interaction with ALCAM.^{15 16} This heterophilic interaction is conserved across species and competes with homophilic ALCAM interactions due to its higher binding affinity.^{13 15} ALCAM overexpression has been detected in numerous malignancies (eg, melanoma, gastric cancer, esophageal squamous cell carcinoma, endometrial cancer, lung cancer, and pancreatic cancer), and relates to poor prognosis,

tumor progression, invasion and metastasis.^{41 48–51} The contribution of ALCAM to tumor progression through the fine-tuning of homophilic adhesive cell-to-cell interactions between cancer cells and with stromal and/or endothelial cells has been reported.^{34 48} However, heterophilic ALCAM interactions with Gal-1, Gal-3 and Gal-8^{5 52} or the Ig-like transcript 3 (ILT3)—an inhibitory receptor expressed by cells of myelomonocytic origin and a marker of tolerogenic dendritic cells²⁷—may also play a role. Similar to shCD6, chimeric soluble ILT3 (ILT3.Fc) inhibited in vitro T cell proliferation and differentiation to Treg, and also tumor cell growth and migration in an ALCAM-dependent manner cells.²⁷ These observations support the targeting of ALCAM for immunotherapeutic purposes in cancer. On this basis, we tested the efficacy of sustained smCD6 delivery by means of AAV-based gene transfer—a successful strategy in several preclinical models and in recent clinical cancer trials.⁵³ Contrary to expectations, implanted subcutaneous B16.F0 melanoma maintained local tumor growth rates in WT mice previously transduced with either hepatotropic AAV-smCD6 or control AAV-Luc, while lower lung and liver metastatization were observed after intravenous B16.F0 cell infusion of WT mice transduced with AAV-smCD6 *v.p.* In our view, infection with AAVs triggers a non-specific immune-boosting effect (eg, through innate recognition of viral capsid and DNA) irrespective of shCD6 expression.⁵⁴ This alone reduces local tumor growth, and overshadows the anti-tumor effects of AAV-smCD6. In the metastasis model, the AAV immune-boost is overridden by ALCAM-dependence of tumor cell adhesion to endothelia. High circulating smCD6 levels achieved with AAV-smCD6 infection are highly effective in that regard, highlighting its role in metastasis prevention.

A general concern with therapeutic approaches that potentiate antitumor responses is the development of autoimmunity as a secondary effect. Preliminary results addressed to assess potential autoimmune responses generated by shCD6 show that shCD6E μ LckTg mice present a less severe outcome in two different models of induced autoimmune disease (collagen-induced arthritis, CIA; experimental autoimmune encephalitis, EAE), compared with NonTg controls (online supplementary figure S10A and S10B). Importantly, a similar attenuated autoimmune disease was observed when WT mice were infused every other day with recombinant shCD6 from day 0 of EAE induction (online supplementary figure S10C). These results are in line with published data showing attenuated EAE disease severity in *cd6*^{-/-} mice of DBA/1 and C57BL/6 background and in CD6 humanized mice treated with a non-depleting mouse anti-human CD6 mAb.^{8 55} The beneficial effect of a shCD6 decoy receptor on both tumor and autoimmune contexts may relate to (1) differences in their respective pathophysiological effectors, and/or (2) the multiligand properties of CD6, since drivers of tumorigenic microenvironment in cancer such as galectins and ALCAM are also known to favor autoimmune inflammation.^{39 56}

CONCLUSIONS

In summary, the shCD6 μ LckTg model reveals novel therapeutic potential of impairing CD6–ligand/s interaction/s by infusion of shCD6 protein. Enhancement of antitumor responses following ALCAM blockade with single chain fragment variable (scFv) fragments or whole antibodies has been reported to disturb cell-to-cell and cell-extracellular matrix interactions important for primary tumor growth and/or leukocyte trafficking (including that of Treg cells).^{33 39 57} These mechanisms may also explain the therapeutic effects we report for shCD6 as a proof-of-concept. Future studies are required to reveal the full immunotherapeutic applications of shCD6 in relevant human disorders.

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Competing interests None declared.

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Data availability statement All data relevant to the study are included in the article or uploaded as supplementary information.

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