





# Distinct Mechanisms of Dissemination of NDM-1 Metallo- $\beta$ -Lactamase in *Acinetobacter* Species in Argentina

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**ABSTRACT** A 4-year surveillance of carbapenem-resistant *Acinetobacter* spp. isolates in Argentina identified 40 strains carrying *bla*<sub>NDM-1</sub>. Genome sequencing revealed that most were *Acinetobacter baumannii*, whereas seven represented other *Acinetobacter* spp. The *A. baumannii* genomes were closely related, suggesting recent spread. *bla*<sub>NDM-1</sub> was located in the chromosome of *A. baumannii* strains and on a plasmid in non-*A. baumannii* strains. A resistance gene island carrying *bla*<sub>PER-7</sub> and other resistance determinants was found on a plasmid in some *A. baumannii* strains.

**KEYWORDS** *Acinetobacter baumannii*, NDM-1, carbapenem resistance, *Acinetobacter*, carbapenems

The predominant species expressing the NDM-1 carbapenemase are *Klebsiella pneumoniae* and *Escherichia coli*. However, *Acinetobacter* spp. isolates are recognized as intermediate reservoirs for the *bla*<sub>NDM-1</sub> resistance determinant (1, 2). *bla*<sub>NDM</sub> is a metallo- $\beta$ -lactamase (MBL) generally found on a plasmid or other mobile element that carries resistance determinants for other antibiotic classes, rendering many NDM-positive isolates extensively drug resistant. Infections caused by carbapenem-resistant *Acinetobacter baumannii* isolates are associated with mortality rates as high as 60% (3, 4).

The Argentina National Reference Laboratory (NRL) identified an increase in the prevalence of NDM-containing *Acinetobacter* spp. isolates beginning in 2015. Of the 20,028 clinical isolates screened since 2010, 15,621 were carbapenem resistant and 144 had an MBL phenotype (5–7). PCR assays for common MBLs confirmed that 68 (47%), i.e., 40 *bla*<sub>NDM</sub> and 28 *bla*<sub>IMP</sub>, of 144 isolates were producers; whereas in the remaining strains, the MBL-like phenotype observed in the institution of origin was due to the presence of *bla*<sub>OXA-23</sub> or *bla*<sub>OXA-58</sub>.

NDM-producing strains were recovered from 19 hospitals in nine cities and seven

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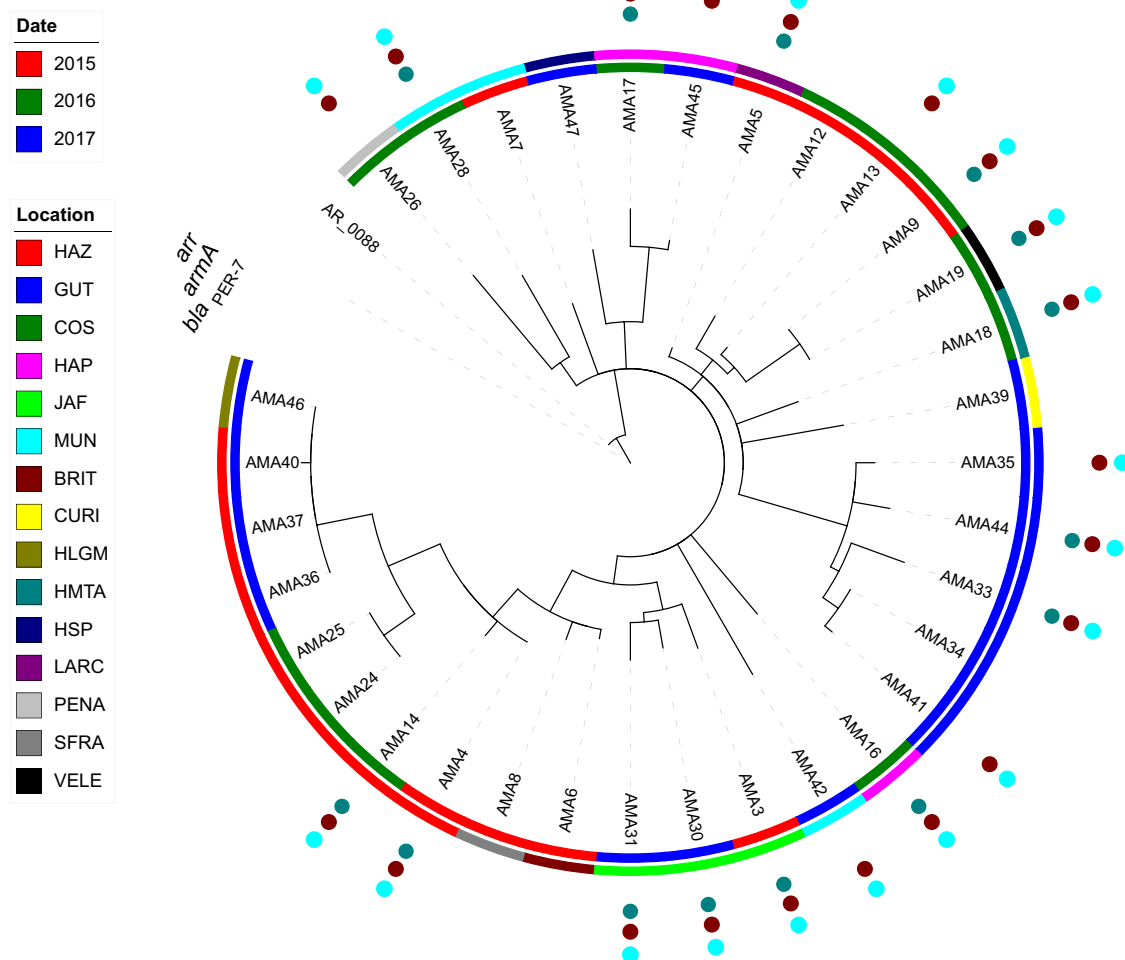
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Tree scale: 10



**FIG 1** Phylogenetic tree of *A. baumannii* genomes. A neighbor-joining tree was constructed using shared SNP and IS insertion sites. Inner circle shows year of isolation. Outer circle shows hospital in which the isolate was recovered. See Table S1 for further information on the location of each hospital. Isolates that are positive for components of RI-PER-7 are denoted by colored circles outside of the tree. The scale bar represents the combined number of SNPs and IS insertion events. The AR\_0088 genome was used as the out group, but the branch length is not shown to highlight the relationships of the AMA strains. The figure was created using iTOL (24).

provinces in Argentina (see Fig. S1 in the supplemental material). *A. baumannii* isolates had more extensive antimicrobial resistance profiles than the non-*baumannii* isolates (see Table S1 in the supplemental material). Genome sequences were obtained on an Illumina NextSeq 500 and assembled using Velvet (8). A BLASTN search at NCBI classified seven genomes as representing five different non-*baumannii* *Acinetobacter* species. (see Table S1). MLST analysis showed that all 33 *A. baumannii* isolates belonged to sequence type ST25 (9). The ST25 genomes were most closely related to isolates found throughout the world, including HEU3 (Honduras), HWBA8 (South Korea), NM3 (United Arab Emirates), and two genomes with no geographic origin provided, i.e., AR\_0088 and AB5256. The AR\_0088 genome has been completely sequenced (GenBank accession no. CP027530.1) and was used as the reference genome for single nucleotide polymorphisms (SNPs) and insertion element (IS) annotation.

The *A. baumannii* genomes differed by only 14 to 36 sequence variants, suggesting recent divergence. Patterns of shared SNPs (10), IS locations (11), and epidemiological data were highly concordant (Fig. 1). Genomes were predominantly clustered by isolation location, with a few exceptions. For example, AMA19 from Hospital General de Agudos Vélez Sarsfield (VELE) was essentially identical to AMA9 from Hospital Dr. Cosme Argerich (COS) and on the same branch as other COS isolates. These hospitals

belong to the public care sector in the Buenos Aires capital district, where patient exchange is frequent, suggesting a possible transmission event.

The AR\_0088 reference genome contained two plasmids, i.e., pAR\_0088\_1 (GenBank accession no. CP027531.1) and pAR\_0088\_2 (GenBank accession no. CP027532.1). pAR\_0088\_2 carries the *bla*<sub>NDM-1</sub> gene in Tn125, and this plasmid is the likely location of the *bla*<sub>NDM-1</sub> gene in the non-*baumannii* *Acinetobacter* genomes (1, 12, 13). pAR\_0088\_2 sequences were not present in the *A. baumannii* ST25 genomes. To ascertain the location of Tn125 in the *A. baumannii* genomes, we identified the locations of IS*Aba*125 insertions, which flank the transposon, in the draft genome assembly of a representative strain (AMA16). Six IS*Aba*125 insertion sites were inferred, all in the chromosome, based on alignment of flanking sequences to the AR\_0088 reference sequence. PCR and Sanger sequencing were used to demonstrate that Tn125 was inserted at base 3,921,386 of the AR\_0088 genome in AMA16 and all other *A. baumannii* isolates, interrupting a gene encoding a hypothetical protein, AM467\_RS18915 (see Table S2 in the supplemental material).

A large resistance gene island (RI) was identified in 14 of the ST25 strains (RI-PER-7). This ~23.8-kb sequence is bound by a pair of IS26 elements in direct-repeat orientation (Fig. 2). The island carries genes encoding resistance to aminoglycosides (*armA*), rifampin (*arr*), cephalosporins (*bla*<sub>PER-7</sub>), and fosfomycin (GST) (Fig. 2). This RI also carries two copies of IS*CR1* (IS91-like) and three other IS elements that are found predominantly in *Enterobacteriaceae* isolates but are rare in *Acinetobacter* isolates, i.e., IS10*A*, ISEc28, and ISEc29. The 7.5 kb at the 3' end of the island is found in several non-*Acinetobacter* genomes. The complete structure was identified in seven accessions in GenBank, mostly plasmid sequences. Five of the ST25 genomes harbored a shorter version of the RI, lacking the *bla*<sub>PER-7</sub> gene. Genetic structures similar to RI-PER-7 were identified in other species, including *K. pneumoniae*, *Proteus mirabilis*, and *E. coli*. In these structures, *bla*<sub>PER-7</sub> was not present, but other  $\beta$ -lactamases were found (Fig. 2).

RI-PER-7 is not present in AR\_0088, but it was found on one of the plasmids in HWBA8 (14). pHWBA8\_1 (GenBank accession no. CP020596.1) is 195,838 bases, and a large portion of this sequence is present in the ST25 *A. baumannii* genomes, including the complete RI-PER-7. Interestingly, pHWBA8\_1 is also similar to pAR\_0088\_1, but the latter plasmid lacks the RI and another ~17.5-kb segment and has extensive rearrangements covering another ~20 kb of the sequence. The location of RI-PER-7 in the plasmid was confirmed by PCR amplification and Sanger sequencing of the junction regions (see Table S2). This large plasmid carries five additional AR genes located outside RI-PER-7.

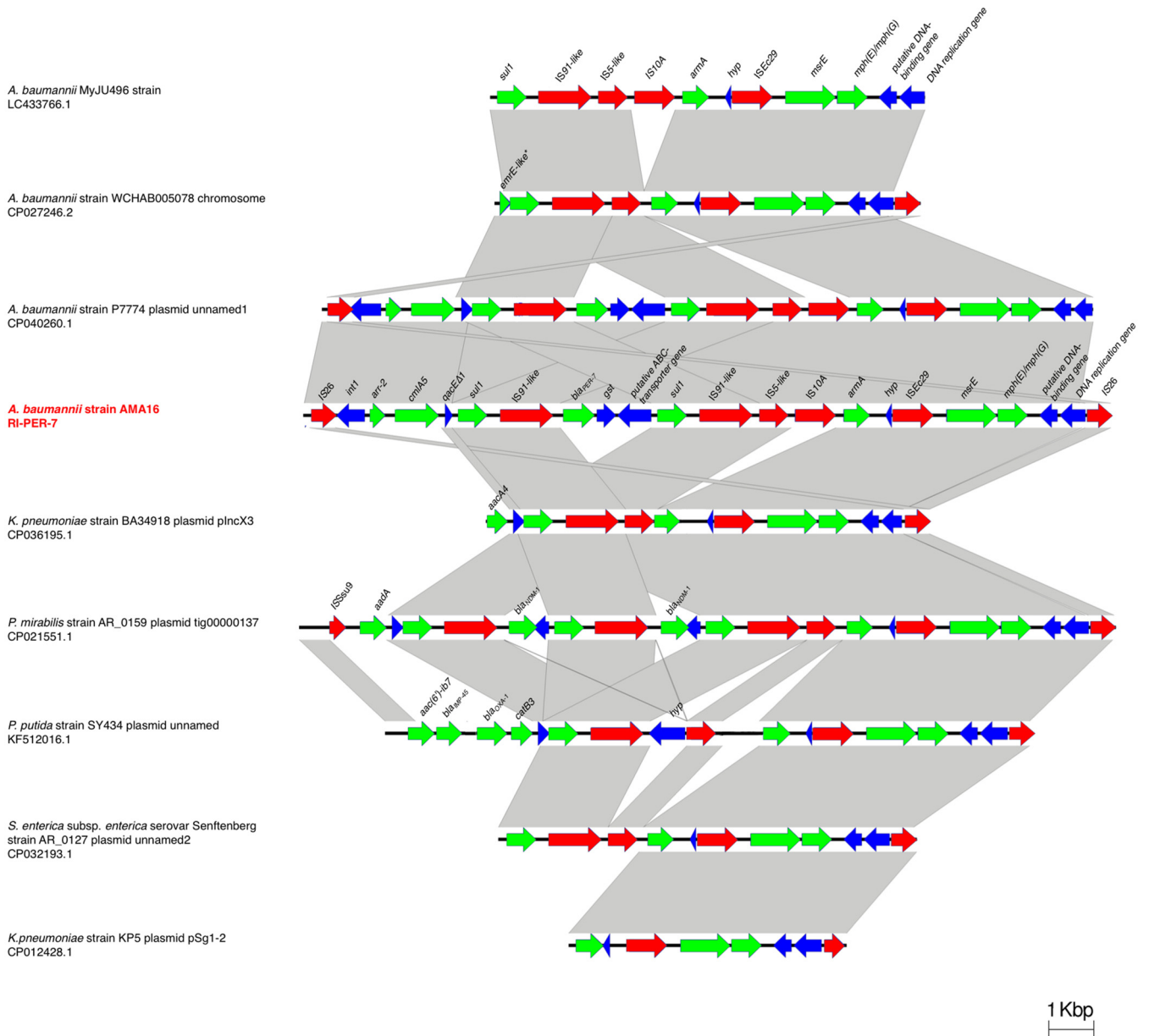
The *comM* gene, a common location for insertion of RIs in *A. baumannii* isolates, is intact in ST25 strains. IS elements were not found upstream of the chromosomal *bla*<sub>ADC</sub> or *bla*<sub>OXA</sub> genes in this study. The ST25 strains carry S84L and S81L substitutions in the *parC* and *gyrA* genes, consistent with their nonsusceptibility to ciprofloxacin.

The NRL also confirmed the presence of *bla*<sub>NDM</sub>-producing *Enterobacteriaceae* in 12 of 19 hospitals (data not shown). In 8 of the 12 hospitals, this emergence occurred 1 to 4 months after the first detection of NDM-producing *Acinetobacter* isolates. Additional work is needed to determine whether *Acinetobacter* spp. could have played a role in the interspecies dissemination of NDM in these institutions.

Although reports of *bla*<sub>NDM</sub> in Latin America are limited, sentinel investigations have described the presence of Tn125 in *Acinetobacter* spp. (6, 15–17). An ST25 *A. baumannii* strain from a patient with an abdominal infection in Honduras was determined to have Tn125 on a plasmid (18), but this plasmid was not present in the ST25 strains analyzed here. Tn125 has been reported in the chromosome of *Acinetobacter* spp. as well (19, 20).

ST25 strains in Argentina typically carry *bla*<sub>OXA-23</sub>  $\beta$ -lactamase (17). According to available data, the cooccurrence of *bla*<sub>NDM-1</sub> and *bla*<sub>OXA-23/58</sub> seems to be an uncommon event (12, 21–23), a scenario that may change due to the increasing emergence of NDM-1 in *A. baumannii* isolates.

In summary, the genetic context of *bla*<sub>NDM-1</sub> differs between *A. baumannii* and non-*baumannii* isolates in Argentina, with non-*baumannii* strains mostly retaining



**FIG 2** Comparison of genetic structure of RI-PER-7 genomic island. Gray bars, regions shared between isolates; red arrows, IS elements; green arrows, antibiotic resistances genes. The figure was created using EasyFig, version 2.2.2.

susceptibility to some antibiotics. We also describe an RI in a subset of *A. baumannii* genomes that likely contributes substantially to the MDR phenotype of these strains. The escalating number of reports of NDM-1 among *A. baumannii* isolates suggests a switch regarding the genetic basis of carbapenem resistance in this species, and intensive tracking of patient contacts is warranted for ST25.

**Data availability.** This Whole Genome Shotgun project has been deposited at DDBJ/ENA/GenBank under BioProject accession no. [PRJNA562922](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA562922). Contig sequences for each genome are available under GenBank accession nos. [VYSH00000000](https://www.ncbi.nlm.nih.gov/nuccore/VYSH00000000) to [VYTU00000000](https://www.ncbi.nlm.nih.gov/nuccore/VYTU00000000).

**SUPPLEMENTAL MATERIAL**

Supplemental material is available online only.  
**SUPPLEMENTAL FILE 1**, PDF file, 0.1 MB.

**SUPPLEMENTAL FILE 2**, XLSX file, 0.02 MB.

**SUPPLEMENTAL FILE 3**, XLSX file, 0.01 MB.

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## REFERENCES

- Bontron S, Nordmann P, Poirel L. 2016. Transposition of Tn125 encoding the NDM-1 carbapenemase in *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 60:7245–7251. <https://doi.org/10.1128/AAC.01755-16>.
- Toleman MA, Spencer J, Jones L, Walsh TR. 2012. *bla*<sub>NDM-1</sub> is a chimera likely constructed in *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 56:2773–2776. <https://doi.org/10.1128/AAC.06297-11>.
- Jamulitrat S, Arunpan P, Phainuphong P. 2009. Attributable mortality of imipenem-resistant nosocomial *Acinetobacter baumannii* bloodstream infection. *J Med Assoc Thai* 92:413–419.
- Kumar SH, De AS, Baveja SM, Gore MA. 2012. Prevalence and risk factors of metallo beta-lactamase producing *Pseudomonas aeruginosa* and *Acinetobacter* species in burns and surgical wards in a tertiary care hospital. *J Lab Physicians* 4:39–42. <https://doi.org/10.4103/0974-2727.98670>.
- Fiorilli G, Faccione D, Lopardo H, Callejo R, Rapoport M, Prieto M, Galas M, Pasteran F. 2010. Emergence of metallo-beta-lactamases in *Acinetobacter* spp clinical isolates from Argentina. *Rev Esp Quimioter* 23:100–102.
- Faccione D, Martino F, Pasteran F, Albornoz E, Biondi E, Vazquez M, Rapoport M, Rodrigo V, De Belder D, Gomez S, Corso A. 2019. Multiple clones of metallo-beta-lactamase-producing *Acinetobacter ursingii* in a children hospital from Argentina. *Infect Genet Evol* 67:145–149. <https://doi.org/10.1016/j.meegid.2018.11.011>.
- Pasteran F, Mora MM, Albornoz E, Faccione D, Franco R, Ortellado J, Melgarejo N, Gomez S, Riquelme I, Matheu J, Ramon-Pardo P, Corso A. 2014. Emergence of genetically unrelated NDM-1-producing *Acinetobacter pittii* strains in Paraguay. *J Antimicrob Chemother* 69:2575–2578. <https://doi.org/10.1093/jac/dku139>.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <https://doi.org/10.1101/gr.074492.107>.
- Diancourt L, Passet V, Nemec A, Dijkshoorn L, Brisse S. 2010. The population structure of *Acinetobacter baumannii*: expanding multiresistant clones from an ancestral susceptible genetic pool. *PLoS One* 5:e10034. <https://doi.org/10.1371/journal.pone.0010034>.
- Poplin R, Ruano-Rubio V, DePristo MA, Fennell TJ, Carneiro MO, Van der Auwera GA, Kling DE, Gauthier LD, Levy-Moonshine A, Roazen D, Shakir K, Thibault J, Chandran S, Whelan C, Lek M, Gabriel S, Daly MJ, Neale B, MacArthur DG, Banks E. 2018. Scaling accurate genetic variant discovery to tens of thousands of samples. *bioRxiv* <https://doi.org/10.1101/201178:201178>.
- Adams MD, Bishop B, Wright MS. 2016. Quantitative assessment of insertion sequence impact on bacterial genome architecture. *Microb Genom* 2:e000062. <https://doi.org/10.1099/mgen.0.000062>.
- Krizova L, Bonnin RA, Nordmann P, Nemec A, Poirel L. 2012. Characterization of a multidrug-resistant *Acinetobacter baumannii* strain carrying the *bla*<sub>NDM-1</sub> and *bla*<sub>OXA-23</sub> carbapenemase genes from the Czech Republic. *J Antimicrob Chemother* 67:1550–1552. <https://doi.org/10.1093/jac/dks064>.
- Poirel L, Bonnin RA, Boulanger A, Schrenzel J, Kaase M, Nordmann P. 2012. Tn125-related acquisition of *bla*<sub>NDM-like</sub> genes in *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 56:1087–1089. <https://doi.org/10.1128/AAC.05620-11>.
- Yoon EJ, Kim JO, Yang JW, Kim HS, Lee KJ, Jeong SH, Lee H, Lee K. 2017. The *bla*<sub>OXA-23</sub>-associated transposons in the genome of *Acinetobacter* spp. represent an epidemiological situation of the species encountering carbapenems. *J Antimicrob Chemother* 72:2708–2714. <https://doi.org/10.1093/jac/dkx205>.

15. Brasiliense D, Cayo R, Streling AP, Nodari CS, Barata RR, Lemos PS, Massafra JM, Correa Y, Magalhaes I, Gales AC, Sodre R. 2019. Diversity of metallo-beta-lactamase-encoding genes found in distinct species of *Acinetobacter* isolated from the Brazilian Amazon region. *Mem Inst Oswaldo Cruz* 114:e190020. <https://doi.org/10.1590/0074-02760190020>.
16. Pillonetto M, Arend L, Vespero EC, Pelisson M, Chagas TP, Carvalho-Assef AP, Asensi MD. 2014. First report of NDM-1-producing *Acinetobacter baumannii* sequence type 25 in Brazil. *Antimicrob Agents Chemother* 58:7592–7594. <https://doi.org/10.1128/AAC.03444-14>.
17. Rodriguez CH, Nastro M, Famiglietti A. 2018. Carbapenemases in *Acinetobacter baumannii*. Review of their dissemination in Latin America. *Rev Argent Microbiol* 50:327–333. <https://doi.org/10.1016/j.ram.2017.10.006>.
18. Waterman PE, McGann P, Snesrud E, Clifford RJ, Kwak YI, Munoz-Urbizo IP, Tabora-Castellanos J, Milillo M, Preston L, Aviles R, Sutter DE, Lesho EP. 2013. Bacterial peritonitis due to *Acinetobacter baumannii* sequence type 25 with plasmid-borne New Delhi metallo- $\beta$ -lactamase in Honduras. *Antimicrob Agents Chemother* 57:4584–4586. <https://doi.org/10.1128/AAC.00275-13>.
19. Jiang L, Yu Y, Zeng W, Guo J, Lv F, Wang X, Liu X, Zhao Z. 2019. Whole-genome analysis of New Delhi metallo- $\beta$ -lactamase-1-producing *Acinetobacter haemolyticus* from China. *J Glob Antimicrob Resist* 20:204–208. <https://doi.org/10.1016/j.jgar.2019.05.012>.
20. Krahn T, Wibberg D, Maus I, Winkler A, Bontron S, Sczyrba A, Nordmann P, Pühler A, Poirel L, Schlüter A. 2016. Intraspecies transfer of the chromosomal *Acinetobacter baumannii* *bla*<sub>NDM-1</sub> carbapenemase gene. *Antimicrob Agents Chemother* 60:3032–3040. <https://doi.org/10.1128/AAC.00124-16>.
21. Chen Z, Qlu S, Wang Y, Wang Y, Liu S, Wang Z, Du X, Wang L, Guo J, Wang Z, Liu N, Yuan J, Song H, Huang L. 2011. Coexistence of *bla*<sub>NDM-1</sub> with the prevalent *bla*<sub>OXA23</sub> and *bla*<sub>IMP</sub> in pan-drug resistant *Acinetobacter baumannii* isolates in China. *Clin Infect Dis* 52:692–693. <https://doi.org/10.1093/cid/ciq231>.
22. El-Mahdy TS, Al-Agamy MH, Al-Qahtani AA, Shibl AM. 2017. Detection of *bla*<sub>OXA-23-like</sub> and *bla*<sub>NDM-1</sub> in *Acinetobacter baumannii* from the eastern region, Saudi Arabia. *Microb Drug Resist* 23:115–121. <https://doi.org/10.1089/mdr.2015.0304>.
23. Karthikeyan K, Thirunarayan MA, Krishnan P. 2010. Coexistence of *bla*<sub>OXA-23</sub> with *bla*<sub>NDM-1</sub> and *armA* in clinical isolates of *Acinetobacter baumannii* from India. *J Antimicrob Chemother* 65:2253–2254. <https://doi.org/10.1093/jac/dkq273>.
24. Letunic I, Bork P. 2019. Interactive Tree of Life (iTOL) v4: recent updates and new developments. *Nucleic Acids Res* 47:W256–W259. <https://doi.org/10.1093/nar/gkz239>.