



# **Surveillance of Omadacycline Activity Tested against Clinical Isolates from the United States and Europe: Report from the SENTRY Antimicrobial Surveillance Program, 2016 to 2018**

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**ABSTRACT** Omadacycline is a broad-spectrum aminomethylcycline approved in October 2018 by the U.S. Food and Drug Administration for treating acute bacterial skin and skin structure infections and community-acquired pneumonia as both an oral and intravenous once-daily formulation. In this report, the activities of omadacycline and comparators were tested against 49,000 nonduplicate bacterial isolates collected prospectively during 2016 to 2018 from medical centers in Europe (24,500 isolates, 40 medical centers [19 countries]) and the United States (24,500 isolates, 33 medical centers [23 states and all 9 U.S. census divisions]). Omadacycline was tested by broth microdilution following the methods in Clinical and Laboratory Standards Institute document M07 (Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically; Approved Standard, 11th ed., 2018). Omadacycline  $(MIC<sub>50/90</sub>, 0.12/0.25 mg/liter)$  inhibited 98.6% of Staphylococcus aureus isolates at  $\leq$ 0.5 mg/liter, including 96.3% of methicillin-resistant S. aureus isolates and 99.8% of methicillin-susceptible S. aureus isolates. Omadacycline potency was comparable for Streptococcus pneumoniae (MIC<sub>50/90</sub>, 0.06/0.12 mg/liter), viridans group streptococci (MIC<sub>50/90</sub>, 0.06/0.12 mg/liter), and beta-hemolytic streptococci (MIC<sub>50/90</sub>, 0.12/ 0.25 mg/liter), regardless of species and susceptibility to penicillin, macrolides, or tetracycline. Omadacycline was active against all Enterobacterales tested (MIC $_{50/90}$ , 1/8 mg/liter; 87.5% of isolates were inhibited at  $\leq$ 4 mg/liter) except Proteus mirabilis (MIC<sub>50/90</sub>, 16/>32 mg/liter) and indole-positive Proteus spp. (MIC<sub>50/90</sub>, 8/32 mg/liter) and was most active against Escherichia coli (MIC $_{50/90}$ , 0.5/2 mg/liter), Klebsiella oxytoca (MIC<sub>50/90</sub>, 1/2 mg/liter), and Citrobacter spp. (MIC<sub>50/90</sub>, 1/4 mg/liter). Omadacycline inhibited 92.4% of Enterobacter cloacae species complex and 88.5% of Klebsiella pneumoniae isolates at ≤4 mg/liter. Omadacycline was active against Haemophilus influenzae (MIC<sub>50/90</sub>, 0.5/1 mg/liter), regardless of  $\beta$ -lactamase status, and against  $M$ oraxella catarrhalis (MIC $_{50/90'}$   $\leq$ 0.12/0.25 mg/liter). The potent activity of omadacycline against Gram-positive and -negative bacteria indicates that omadacycline merits further study in serious infections in which multidrug resistance and mixed Grampositive and Gram-negative bacterial infections may be a concern.

**KEYWORDS** tetracycline, omadacycline, SENTRY, surveillance, susceptibility

**T**he tetracycline class of antimicrobial agents has been an important element in the outpatient treatment of acute bacterial skin and skin structure infection and community-acquired bacterial pneumonia [\(1,](#page-19-0) [2\)](#page-19-1). Unfortunately, heavy usage of the tetracyclines has gradually eroded the activity of this class of agents due to the development of resistance [\(1](#page-19-0)[–](#page-19-2)[4\)](#page-19-3). Tetracycline-resistant pathogens most commonly express efflux and ribosomal protection mechanisms that greatly reduce the utility of this class against many of the clinically relevant pathogens that were previously **Citation** Pfaller MA, Huband MD, Shortridge D, Flamm RK. 2020. Surveillance of omadacycline activity tested against clinical isolates from the United States and Europe: report from the SENTRY Antimicrobial Surveillance Program, 2016 to 2018. Antimicrob Agents Chemother 64:e02488-19. [https://doi.org/10.1128/AAC](https://doi.org/10.1128/AAC.02488-19) [.02488-19.](https://doi.org/10.1128/AAC.02488-19)

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aAbbreviations: S, susceptible; I, intermediate; R, resistant; ABSSSI, acute bacterial skin and skin structure infection; CABP, community-acquired bacterial pneumonia. bDetermined with a 30- $\mu$ g omadacycline disk.

c Klebsiella pneumoniae and Enterobacter cloacae only.

dOmadacycline is not active in vitro against Morganella spp., Proteus spp., and Providencia spp.

eThe Streptococcus anginosus group includes S. anginosus, S. intermedius, and S. constellatus.

gHaemophilus species includes H. influenzae and H. parainfluenzae.

covered by these agents [\(2](#page-19-1)[–](#page-19-3)[5\)](#page-19-4). The search for a safe and effective oral agent that is active against bacteria expressing tetracycline resistance mechanisms has led to the development of omadacycline [\(3](#page-19-2)[–](#page-19-5)[7\)](#page-19-6).

Omadacycline is a semisynthetic derivative of minocycline and the first member of the novel class of aminomethylcyclines [\(5,](#page-19-4) [8](#page-19-7)[–](#page-19-8)[11\)](#page-19-9). Omadacycline remains active against tetracycline-resistant bacterial isolates expressing ribosomal protection and efflux resistance mechanisms  $(3, 5, 9-11)$  $(3, 5, 9-11)$  $(3, 5, 9-11)$  $(3, 5, 9-11)$  $(3, 5, 9-11)$  $(3, 5, 9-11)$  $(3, 5, 9-11)$ . Omadacycline has potent in vitro activity against difficult-to-treat pathogens, such as methicillin-resistant Staphylococcus aureus, vancomycin-resistant enterococci, extended-spectrum  $\beta$ -lactamase (ESBL)-producing Enterobacteriaceae, carbapenem-resistant Enterobacteriaceae, and multidrug-resistant (resistant to  $\geq$ 3 classes of agents) strains of Acinetobacter spp. and Stenotrophomonas maltophilia [\(5,](#page-19-4) [11,](#page-19-9) [12\)](#page-19-11). Omadacycline is approved by the U.S. Food and Drug Administration (FDA) for the treatment of community-acquired bacterial pneumonia and acute bacterial skin and skin structure infections by susceptible organisms [\(Table 1\)](#page-1-0) [\(6,](#page-19-5) [13,](#page-19-12) [14\)](#page-19-13). Phase 2 studies for the use of omadacycline to treat uncomplicated urinary tract infections (UTIs; [ClinicalTrials.gov](http://ClinicalTrials.gov) registration number NCT03425396) and acute pyelonephritis [\(ClinicalTrials.gov](http://ClinicalTrials.gov) registration number NCT03757234) in women have been completed [\(11,](#page-19-9) [15,](#page-19-14) [16\)](#page-19-15). In both studies, omadacycline showed levels of clinical success generally comparable to those of either nitrofurantoin or levofloxacin, as determined by the investigator's assessment of the clinical response at the posttreatment evaluation. However, the microbiological responses were generally lower than those of the comparators (Paratek, data on file).

Due to the exploratory intent and the small numbers of subjects enrolled in each dose group in these phase 2 studies, the sponsor has identified dose regimens that require additional investigation before determining any future development plans for these indications. Additional analyses are ongoing, including the pathogen-specific level of efficacy and the relationships of both clinical and microbiological responses to urinary pharmacokinetic (PK) data.

In the present study, we evaluated the antimicrobial activities of omadacycline and comparator agents against 49,000 isolates of Gram-positive cocci (GPC) and Gramnegative bacilli collected between 2016 and 2018 from 73 individual academic and/or tertiary care medical centers in the United States (33 medical centers and all 9 census divisions) and Europe (40 medical centers from 19 countries) as part of the SENTRY Antimicrobial Surveillance Program. Evaluations of resistant organism subsets were included in the analysis, when available.

f Klebsiella pneumoniae only.

## **RESULTS**

**Number of organisms and key resistance phenotypes.** The 49,000 isolates tested included 10,016 S. aureus isolates, 1,437 coagulase-negative staphylococci, 2,506 Enterococcus isolates (including 1,577 Enterococcus faecalis isolates and 851 Enterococcus faecium isolates), 3,153 Streptococcus pneumoniae isolates, 615 viridans group streptococci, 216 Streptococcus anginosus group isolates, 2,141 beta-hemolytic streptococci (including 1,030 Streptococcus pyogenes isolates, 776 Streptococcus agalactiae isolates, and 335 isolates of other beta-hemolytic streptococci), 20,028 Enterobacterales (including 8,749 Escherichia coli isolates, 4,220 Klebsiella pneumoniae isolates, 939 Klebsiella oxytoca isolates, 1,666 Enterobacter cloacae species complex isolates, 837 Citrobacter isolates, 1,144 Proteus mirabilis isolates, 750 indole-positive Proteus isolates, and 865 Serratia marcescens isolates), 892 Acinetobacter baumannii-Acinetobacter calcoaceticus species complex isolates, 4,564 Pseudomonas aeruginosa isolates, 604 S. maltophilia isolates, 1,886 Haemophilus influenzae isolates, 71 Haemophilus parainfluenzae isolates, and 984 Moraxella catarrhalis isolates [\(Tables 2](#page-3-0) and [3\)](#page-5-0).

Isolates with key resistant phenotypes included 3,326 (33.2%) methicillin-resistant S. aureus (MRSA) isolates, 880 (61.2%) methicillin-resistant (MR) coagulase-negative staphylococci, 330 (38.8%) vancomycin-nonsusceptible E. faecium isolates, 336 (10.7%) penicillin-resistant S. pneumoniae isolates, 1,784 (20.4%) extended-spectrum- $\beta$ lactamase-phenotype E. coli isolates, 13 (0.1%) carbapenem-resistant E. coli isolates, 1,383 (32.8%) extended-spectrum- $\beta$ -lactamase-phenotype K. pneumoniae isolates, 388 (9.2%) carbapenem-resistant K. pneumoniae isolates, and 478 (28.7%) ceftazidimenonsusceptible E. cloacae species complex isolates [\(Table 2\)](#page-3-0). A total of 11,729 isolates were tetracycline resistant, including 514 S. aureus isolates (5.1% of all S. aureus isolates), 177 coagulase-negative staphylococci (12.3% of all coagulase-negative staphylococci), 1,178 E. faecalis isolates (74.7% of all E. faecalis isolates), 486 E. faecium isolates (57.1% of all E. faecium isolates), 656 S. pneumoniae isolates (20.8% of all S. pneumoniae isolates), 191 viridans group streptococci (31.1% of all viridans group streptococci), 923 beta-hemolytic streptococci (43.1% of all beta-hemolytic streptococci), 6,870 Enterobacteriaceae (34.3% of all Enterobacteriaceae), 544 A. baumannii isolates (61.0% of all A. baumannii isolates), and 15 H. influenzae isolates (0.8% of all H. influenzae isolates) (data not shown).

**Overall omadacycline activity.** The MIC distributions for omadacycline and each organism or organism group from the 73 participating medical centers are shown in [Table 2.](#page-3-0) Omadacycline demonstrated essentially identical activity against the key target pathogens from the United States and Europe [\(Table 3\)](#page-5-0). As such, the data sets were combined for further comparison.

Omadacycline had potent activity against S. aureus (10,016 isolates tested;  $MIC_{50/90}$ 0.12/0.25 mg/liter) [\(Table 2\)](#page-3-0). A subset of 9,880 (98.6%) isolates were inhibited by  $\leq$ 0.5 mg/liter of omadacycline, including 99.8% of methicillin-susceptible S. aureus isolates and 96.3% of methicillin-resistant S. aureus isolates [\(Table 2\)](#page-3-0). The omadacycline MIC<sub>50/90</sub> values for all coagulase-negative staphylococci were 0.12/0.5 mg/liter [\(Table 2\)](#page-3-0). Overall, 92.4% of coagulase-negative staphylococci were inhibited by  $\leq$ 0.5 mg/liter of omadacycline. Tetracycline resistance had little effect on omadacycline MIC values against S. aureus (omadacycline MIC<sub>50/90</sub>, 0.12/0.5 mg/liter) or coagulase-negative staphylococci (omadacycline MIC<sub>50/90</sub>, 0.25/1 mg/liter). Among the tetracycline-resistant isolates, 95.1% of S. aureus isolates and 89.2% of coagulase-negative staphylococci were inhibited by  $\leq$ 0.5 mg/liter of omadacycline (data not shown).

Omadacycline was as active against E. faecium (MIC<sub>50/90</sub>, 0.06/0.12 mg/liter; 97.3% of isolates were inhibited at  $\leq$ 0.25 mg/liter) as it was against *E. faecalis* (MIC<sub>50/90</sub>, 0.12/ 0.25 mg/liter; 97.6% of isolates were inhibited at  $\leq$ 0.25 mg/liter), and its activity was not adversely affected by resistance to vancomycin or tetracycline when tested against these organism groups [\(Table 2\)](#page-3-0).

Omadacycline potency was comparable for S. pneumoniae (MIC<sub>50/90</sub>, 0.06/0.12 mg/ liter), viridans group streptococci (MIC<sub>50/90</sub>, 0.06/0.12 mg/liter), and beta-hemolytic

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 $\epsilon$ The extended-spectrum  $\beta$ -lactamase phenotype was defined as having a MIC value of

2 mg/liter for ceftazidime, ceftriaxone, or aztreonam (confirmatory testing was not performed).

<span id="page-5-0"></span>**TABLE 3** Summary of omadacycline activity stratified by geographic region



The extended-spectrum  $\beta$ -lactamase (ESBL) phenotype was defined as having a MIC value of  $\geq 2$  mg/liter for ceftazidime, ceftriaxone, or aztreonam (confirmatory testing was not performed).

streptococci (MIC<sub>50/90</sub>, 0.12/0.25 mg/liter), regardless of species and susceptibility to penicillin, macrolides (beta-hemolytic streptococci only), or tetracycline [\(Table 2\)](#page-3-0). All but two S. pneumoniae isolates (99.9%), five viridans group streptococci (99.2%), and two Streptococcus pyogenes isolates (99.8%) were inhibited by <0.5 mg/liter of omadacycline. Omadacycline had activity against most of the 20,028 Enterobacterales isolates tested (MIC<sub>50/90</sub>, 1/8 mg/liter; 87.5% of isolates were inhibited at  $\leq$ 4 mg/liter) and was most potent against E. coli (MIC<sub>50/90</sub>, 0.5/2 mg/liter; 99.1% of isolates were inhibited at  $\leq$ 4 mg/liter), *K. oxytoca* (MIC<sub>50/90</sub>, 1/2 mg/liter; 97.0% of isolates were inhibited at  $\leq$ 4 mg/liter), and C*itrobacter* spp. (MIC<sub>50/90</sub>, 1/4 mg/liter; 94.9% of isolates were inhibited at  $\leq$ 4 mg/liter) [\(Table 1\)](#page-1-0). Omadacycline lacked activity against the *Enterobacterales* species P. mirabilis (MIC<sub>50/90</sub>, 16/>32 mg/liter) and indole-positive Proteus spp. (MIC<sub>50/90</sub>, 8/32 mg/ liter) [\(Table 1\)](#page-1-0).

Omadacycline also had potent activity against resistant subsets of Enterobacteriaceae, including extended-spectrum- $\beta$ -lactamase-phenotype (MIC<sub>50/90</sub>, 1/2 mg/ liter [98.0% of isolates were inhibited at  $\leq$ 4 mg/liter]), non-extended-spectrum- $\beta$ lactamase-phenotype (MIC $_{50/90}$ , 0.5/2 mg/liter [99.4% of isolates were inhibited at  $\leq$ 4 mg/liter]), and carbapenem-resistant (MIC<sub>50/90</sub>, 1/2 mg/liter [100.0% of isolates

were inhibited at  $\leq$ 4 mg/liter]) strains of *E. coli*. The extended-spectrum- $\beta$ lactamase phenotype is based on an MIC value of  $\geq 2$  mg/liter for ceftazidime, ceftriaxone, or aztreonam. Confirmatory susceptibility testing was not performed. Omadacycline was less active against extended-spectrum- $\beta$ -lactamase-phenotype K. pneumoniae (MIC $_{50/90'}$  4/8 mg/liter [76.4% of isolates were inhibited at  $\leq$ 4 mg/ liter]) and carbapenem-resistant K. pneumoniae (MIC<sub>50/90</sub>, 4/8 mg/liter [72.4% of isolates were inhibited at  $\leq$ 4 mg/liter]) isolates than against non-extended-spectrum- $\beta$ -lactamase-phenotype K. pneumoniae isolates (MIC<sub>50/90</sub>, 1/4 mg/liter [94.4% of isolates were inhibited at ≤4 mg/liter]). Against ceftazidime-nonsusceptible E. cloacae species complex isolates (MIC,  $\geq$ 8 mg/liter; AmpC-derepressed phenotype isolates), omadacycline was less active (MIC<sub>50/90</sub>, 2/8 mg/liter; 84.9% of isolates were inhibited at  $\leq$ 4 mg/liter) than it was against ceftazidime-susceptible isolates (MIC<sub>50/90</sub>, 2/4 mg/liter; 95.5% of isolates were inhibited at  $\leq$ 4 mg/liter) [\(Table 2\)](#page-3-0).

The MIC<sub>50/90</sub> values for omadacycline against tetracycline-resistant strains of E. coli and K. pneumoniae were 1/4 mg/liter and 4/16 mg/liter, respectively, and 97.6% of tetracycline-resistant E. coli isolates and 63.2% of tetracycline-resistant K. pneumoniae isolates were inhibited by  $\leq$ 4 mg/liter of omadacycline (data not shown). Among the tetracycline-resistant E. cloacae species complex isolates, 56.7% were inhibited by omadacycline at  $\leq$ 4 mg/liter (MIC<sub>50/90</sub>, 4/16 mg/liter) (data not shown). Among the tetracycline-resistant strains of Citrobacter spp., omadacycline activity was decreased (MIC<sub>50/90</sub>, 4/8 mg/liter [70.0% of isolates were inhibited at  $\leq$ 4 mg/liter]) (data not shown) compared to that against all Citrobacter isolates (MIC<sub>50/90</sub>, 1/4 mg/liter [94.9% of isolates were inhibited at  $\leq$ 4 mg/liter]) [\(Table 2\)](#page-3-0). In contrast, tetracycline-resistant strains of S. marcescens remained mostly susceptible to omadacycline (MIC $_{50/90}$ , 4/8 mg/liter [84.3% of isolates were inhibited at ≤4 mg/liter]) (data not shown) when their omadacycline susceptibility was compared to that of all S. marcescens isolates (MIC<sub>50/90</sub>, 4/8 mg/liter [86.6% of isolates were inhibited at  $\leq$ 4 mg/liter]) [\(Table 2\)](#page-3-0).

Omadacycline (MIC<sub>50/90</sub>, 4/8 mg/liter) inhibited 76.6% of A. baumannii isolates at  $\leq$ 4 mg/liter [\(Table 2\)](#page-3-0). Omadacycline retained some activity against tetracyclineresistant A. baumannii isolates (MIC<sub>50/90</sub>, 4/8 mg/liter; 61.9% of isolates were inhibited at ≤4 mg/liter). Omadacycline demonstrated good *in vitro* activity against S. *maltophilia* (MIC $_{50/90'}$  4/8 mg/liter [79.6% of isolates were inhibited at  $\leq$ 4 mg/liter]) and had limited to no activity at the concentrations tested against *P. aeruginosa* (MIC<sub>50/90</sub>, 32/>32 mg/ liter) [\(Table 2\)](#page-3-0).

Omadacycline was equally active against  $\beta$ -lactamase-negative (MIC<sub>50/90</sub>, 0.5/1 mg/ liter) and  $\beta$ -lactamase-positive (MIC<sub>50/90</sub>, 0.5/1 mg/liter) isolates of H. influenzae [\(Table](#page-3-0) [2\)](#page-3-0). Omadacycline was also very active against *M. catarrhalis* isolates (MIC<sub>50/90</sub>,  $\leq$ 0.12/ 0.25 mg/liter) [\(Table 2\)](#page-3-0).

**Activity of omadacycline and comparators against acute bacterial skin and skin structure infection isolates.** The activity of omadacycline and comparators against key pathogens from patients with acute bacterial skin and skin structure infection is shown in [Table 4.](#page-7-0)

Greater than 90.0% of S. aureus (99.0% susceptible), Staphylococcus lugdunensis (98.1% susceptible), E. faecalis (97.4% susceptible), S. anginosus group (100.0% susceptible), and S. pyogenes (98.2% susceptible) isolates from acute bacterial skin and skin structure infection were susceptible to omadacycline at the U.S. Food and Drug Administration-assigned breakpoints [\(Tables 1](#page-1-0) and [4\)](#page-7-0). Notably, 94.0% of coagulasenegative staphylococci from acute bacterial skin and skin structure infection were inhibited at the S. aureus breakpoint of  $\leq$ 0.5 mg/liter, whereas only 66.7% were inhibited at the S. lugdunensis breakpoint (data not shown). Notably, 94.0% of coagulase-negative staphylococci isolates from acute bacterial skin and skin structure infection were inhibited at the S. aureus breakpoint of  $\leq$ 0.5 mg/liter, whereas only 66.7% were inhibited at the S. lugdunensis breakpoint of  $\leq$ 0.12 mg/liter (data not shown).

Methicillin-resistant S. aureus accounted for 31.0% of S. aureus isolates from acute bacterial skin and skin structure infection, and 97.1% of those methicillin-resistant S.

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<span id="page-7-0"></span>**TABLE 4** Activity of omadacycline and comparator antimicrobial agents when tested against bacterial isolates from skin and skin structure infections in the United States and Europe, SENTRY Program, 2016 to 2018



## **TABLE 4** (Continued)



## **TABLE 4** (Continued)



## **TABLE 4** (Continued)



aCriteria published by the Clinical and Laboratory Standards Institute [\(33\)](#page-20-0) and The European Committee on Antimicrobial Susceptibility Testing [\(34\)](#page-20-1). S, susceptibility; R, resistance; —, not applicable.

bIn the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for S. aureus (acute bacterial skin and skin structure infection) were applied [\(33,](#page-20-0) [35,](#page-20-2) [36\)](#page-20-3).

Corganisms include Staphylococcus capitis (n = 15), Staphylococcus caprae (n = 7), Staphylococcus cohnii (n = 1), Staphylococcus epidermidis (n = 153), Staphylococcus haemolyticus (n = 35), Staphylococcus hominis (n = 9), Staphylococcus lugdunensis (n = 103), Staphylococcus saprophyticus (n = 4), Staphylococcus schleiferi (n = 4), Staphylococcus simulans ( $n = 9$ ), and Staphylococcus warneri ( $n = 8$ ).

dIn the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for S. lugdunensis acute bacterial skin and skin structure infection) were applied [\(33,](#page-20-0) [35,](#page-20-2) [36\)](#page-20-3).

eOrganisms include Streptococcus agalactiae (n = 258), Streptococcus canis (n = 3), Streptococcus dysgalactiae (n = 155), and Streptococcus pyogenes (n = 544). <sup>n</sup>n the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for S. pyogenes (were applied [\(33,](#page-20-0) [35,](#page-20-2) [36\)](#page-20-3).  $9$ Organisms include Streptococcus anginosus (n = 31), S. anginosus group (n = 19), Streptococcus constellatus (n = 13), Streptococcus gallolyticus (n = 4), Streptococcus gordonii (n = 2), Staphylococcus intermedius (n = 4), Streptococcus mitis group (n = 15), Streptococcus mutans (n = 1), Streptococcus parasanguinis (n = 2), Streptococcus salivarius/Streptococcus vestibularis ( $n = 4$ ), and Streptococcus sanguinis ( $n = 2$ ).

hIn the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for the S. anginosus group (acute bacterial skin and skin structure infection) were applied [\(33,](#page-20-0) [35,](#page-20-2) [36\)](#page-20-3).

In the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for E. faecalis (acute bacterial skin and skin structure infection) were applied [\(33,](#page-20-0) [35,](#page-20-2) [36\)](#page-20-3).

The extended-spectrum β-lactamase (ESBL) phenotype was defined as having a MIC value of ≥2 mg/liter for ceftazidime, ceftriaxone, or aztreonam (confirmatory testing was not performed).

kIn the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for the Enterobacteriaceae (Enterobacter cloacae and Klebsiella pneumoniae only; acute bacterial skin and skin structure infection) were applied [\(33,](#page-20-0) [35,](#page-20-2) [36\)](#page-20-3).

'Organisms include Enterobacter cloacae ( $n = 227$ ), E. cloacae species complex ( $n = 172$ ), Enterobacter asburiae ( $n = 6$ ), and Enterobacter kobei ( $n = 2$ ).

aureus isolates were susceptible to omadacycline [\(Tables 1](#page-1-0) and [4\)](#page-7-0). Comparable activity was seen with linezolid (99.9% susceptible), daptomycin (100.0% susceptible), trimethoprim-sulfamethoxazole (97.4% susceptible), and vancomycin (100.0% susceptible). Erythromycin, clindamycin, levofloxacin, and tetracycline were all considerably less active against methicillin-resistant S. aureus than against methicillin-susceptible S. aureus [\(Table 4\)](#page-7-0). Omadacycline was active against 92.8% of tetracycline-resistant S. aureus isolates [\(Table 4\)](#page-7-0). Linezolid, daptomycin, and vancomycin were the most active

agents against coagulase-negative staphylococci from acute bacterial skin and skin structure infection.

Among the streptococci isolated from patients with acute bacterial skin and skin structure infection, beta-hemolytic streptococci were generally susceptible to the agents tested, with these isolates showing less than 90.0% susceptibility only to erythromycin (74.1% susceptible), clindamycin (86.0% susceptible), and tetracycline (61.2% susceptible) [\(Table 4\)](#page-7-0). Penicillin, linezolid, daptomycin, and vancomycin were the most active agents tested against beta-hemolytic streptococci, with all isolates (100.0%) being susceptible at the Clinical and Laboratory Standards Institute (CLSI) breakpoints. Omadacycline (MIC $_{50/90}$ , 0.06/0.12; 98.2% susceptible) was more active than tetracycline (MIC $_{50/90'}$   $\leq$ 0.25/ $>$ 4; 82.1% susceptible) against S. pyogenes isolates from acute bacterial skin and skin structure infection [\(Table 4\)](#page-7-0). Omadacycline (95.9% of isolates were inhibited using the S. anginosus breakpoint of  $\leq$ 0.12 mg/liter), ceftriaxone (95.9% susceptible), levofloxacin (96.9% susceptible), linezolid (99.0% susceptible), daptomycin (100.0% susceptible), and vancomycin (100.0% susceptible) were all very active against acute bacterial skin and skin structure infection isolates of viridans group streptococci [\(Table 4\)](#page-7-0). Only erythromycin and tetracycline showed less than 80.0% activity against viridans group streptococci. Omadacycline (MIC $_{50/90}$ , 0.06/0.12; 100.0% susceptible) was more active than tetracycline (MIC $_{50/90'}$  0.5/ $>$ 4 mg/liter; 65.2% susceptible) against S. anginosus group isolates from acute bacterial skin and skin structure infection [\(Table 4\)](#page-7-0).

Omadacycline was active against 97.4% of E. faecalis isolates from acute bacterial skin and skin structure infection [\(Tables 1](#page-1-0) and [4\)](#page-7-0). Similarly, 96.8% of E. faecium isolates from acute bacterial skin and skin structure infection were inhibited by  $\leq$ 0.25 mg/liter of omadacycline. The only comparator agents with activity against E. faecium isolates were daptomycin (100.0% susceptible) and linezolid (100.0% susceptible) [\(Table 4\)](#page-7-0). Among the vancomycin-nonsusceptible enterococci from acute bacterial skin and skin structure infection, only omadacycline (93.5% of isolates were inhibited at  $\leq$ 0.25 mg/ liter), linezolid (100.0% susceptible), and daptomycin (100.0% susceptible at an MIC of  $\leq$ 4 mg/liter) retained activity (data not shown).

Omadacycline and imipenem were the most active agents tested against acute bacterial skin and skin structure infection isolates of E. coli, K. pneumoniae, and the E.  $c$ loacae species complex, including isolates of the extended-spectrum- $\beta$ -lactamase and ceftazidime-nonsusceptible phenotypes [\(Table 4\)](#page-7-0). Only 3.6% of S. marcescens isolates were susceptible to tetracycline, but 86.8% were inhibited by  $\leq$ 4 mg/liter of omadacycline [\(Table 4\)](#page-7-0). Ceftazidime (99.4% susceptible) and gentamicin (99.4% susceptible) were the most active agents tested against S. marcescens isolates.

A. baumannii isolates associated with acute bacterial skin and skin structure infection showed a characteristic resistance profile, with the isolates displaying a rate of susceptibility of greater than 70.0% only to doxycycline (71.2% susceptible) [\(Table 4\)](#page-7-0). There are no interpretive criteria for omadacycline against A. baumannii; however, 82.1% of the 156 acute bacterial skin and skin structure infection isolates tested were inhibited at  $\leq$ 4 mg/liter.

**Activity of omadacycline and comparators against community-acquired bacterial pneumonia isolates.** The activity of omadacycline and comparators against key pathogens from patients with community-acquired bacterial pneumonia is shown in [Table 5.](#page-12-0) Using U.S. Food and Drug Administration community-acquired bacterial pneumonia susceptibility breakpoints for S. pneumoniae and Haemophilus spp. [\(Table 1\)](#page-1-0), omadacycline was highly active against S. pneumoniae (MIC<sub>50/90</sub>, 0.06/0.12 mg/liter; 98.7% susceptible; [Tables 2](#page-3-0) and [5\)](#page-12-0), regardless of the penicillinresistant (omadacycline MIC<sub>50/90</sub>, 0.06/0.12 mg/liter; 97.9% susceptible) or tetracyclineresistant (omadacycline MIC<sub>50/90</sub>, 0.06/0.12 mg/liter; 97.0% susceptible) phenotype [\(Table 4\)](#page-7-0). Omadacycline activity against penicillin-resistant (MIC,  $\geq$ 2 mg/liter) S. pneumoniae isolates (MIC<sub>50/90</sub>, 0.06/0.12 mg/liter; 97.9% susceptible) was comparable to levofloxacin activity (MIC<sub>50/90</sub>, 1/2 mg/liter; 97.9% susceptible) and slightly less than vancomycin activity (MIC<sub>50/90</sub>, 0.25/0.5 mg/liter; 100.0% susceptible) and linezolid ac-

<span id="page-12-0"></span>



aCriteria published by the Clinical and Laboratory Standards Institute [\(33\)](#page-20-0) and The European Committee on Antimicrobial Susceptibility Testing [\(34\)](#page-20-1). S, susceptibility; R, resistance; —, not applicable.

<sup>b</sup>In the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for S. pneumoniae community-acquired bacterial pneumonia were applied [\(35,](#page-20-2) [36\)](#page-20-3).

c Using nonmeningitis breakpoints.

dCriteria published by the Clinical and Laboratory Standards Institute (2019) for penicillin (oral penicillin) [\(33\)](#page-20-0).

eIn the absence of Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for Haemophilus spp. were applied [\(35,](#page-20-2) [36\)](#page-20-3).

tivity (MIC<sub>50/90</sub>, 1/2 mg/liter; 100.0% susceptible). Most of the other agents tested showed suboptimal activity against penicillin-resistant S. pneumoniae isolates (range, 20.1 to 72.3% susceptible). Aside from S. pneumoniae, omadacycline was active against other pathogens frequently recovered from patients with community-acquired bacterial pneumonia, such as H. influenzae (MIC<sub>50/90</sub>, 0.5/1 mg/liter; 99.6% susceptible) and M.  $\,$ catarrhalis (MIC $_{50/90'}$   $\leq$ 0.12/0.25 mg/liter; 98.8% of isolates were inhibited at  $\leq$ 0.25 mg/ liter), regardless of  $\beta$ -lactamase production or geographic region [\(Tables 2,](#page-3-0) [3,](#page-5-0) and [5\)](#page-12-0). These isolates were generally susceptible to most agents tested, as well.

**Activity of omadacycline and comparators against urinary tract infection iso**lates. Among the Gram-positive cocci causing urinary tract infection, S. aureus and Enterococcus spp. were the most common [\(Table 6\)](#page-14-0). Omadacycline (MIC $_{50/90}$ , 0.12/ 0.25 mg/liter; 98.2% of isolates were inhibited at  $\leq$ 0.5 mg/liter) was comparable in activity to linezolid (MIC<sub>50/90</sub>, 1/2 mg/liter; 100.0% susceptible) and daptomycin  $(MIC<sub>50/90</sub>$ , 0.25/0.5 mg/liter; 100.0% susceptible) against S. aureus (including methicillin-resistant S. aureus isolates) [\(Table 6\)](#page-14-0). Omadacycline was active against E. faecalis (2.2% vancomycin-resistant) and E. faecium (52.4% vancomycin-resistant) isolates from urinary tract infection, inhibiting 97.4% of E. faecalis isolates and 97.6% of *E. faecium* isolates at ≤0.25 mg/liter [\(Table 6\)](#page-14-0). Linezolid (MIC<sub>50/90</sub>, 1/2 mg/ liter; 98.8% to 100.0% susceptible) and daptomycin (MIC<sub>50/90</sub>, 0.5 to 1/1 to 2 mg/ liter; 100.0% susceptible) were the only comparator agents with activity against >90.0% of both E. faecalis and E. faecium isolates.

Among the E. coli ( $n = 2,872$ ) and K. pneumoniae ( $n = 911$ ) isolates from urinary tract infections, 17.1% and 27.0%, respectively, expressed an extended-spectrum- $\beta$ -lactamase phenotype [\(Table 6\)](#page-14-0). Omadacycline (MIC<sub>50/90</sub>, 0.5/2 mg/liter; 99.4% of isolates were inhibited at  $\leq$ 4 mg/liter), imipenem (MIC $_{50/90'}$   $\leq$ 0.12/ $\leq$ 0.12 mg/liter; 99.9% susceptible), and piperacillin-tazobactam (MIC $_{50/90}$ , 2/8 mg/liter; 96.1%/93.5% susceptible [CLSI/European Committee on Antimicrobial Susceptibility Testing {EUCAST} breakpoints]) were the most active agents tested against E. coli, including those with an extended-spectrum- $\beta$ -lactamase phenotype (98.8% of isolates were inhibited at ≤4 mg/liter, 99.4% susceptible, and 85.5% susceptible, respectively) [\(Table 6\)](#page-14-0). Similarly, omadacycline and imipenem were the most active agents tested against K. pneumoniae  $(92.1\% \text{ of isolates were inhibited at } \leq 4 \text{ mg/liter and } 94.8\%$  were susceptible, respectively), including those with an extended-spectrum- $\beta$ -lactamase phenotype (81.3% of isolates were inhibited at  $\leq$ 4 mg/liter and 81.3% were susceptible, respectively).

The most active agents against urinary tract infection isolates of the Enterobacter cloacae species complex were omadacycline (MIC $_{50/90}$ , 2/8 mg/liter; 85.2% of isolates were inhibited at an MIC of  $\leq$ 4 mg/liter), imipenem (MIC<sub>50/90</sub>, 0.25/1 mg/liter; 96.8% susceptible), and gentamicin (MIC<sub>50/90</sub>, 0.25/8 mg/liter; 89.7% susceptible) [\(Table 6\)](#page-14-0). Although all tested agents showed decreased activity against ceftazidime-nonsusceptible (MIC,  $\geq$ 8 mg/liter) isolates, omadacycline (74.0% of isolates were inhibited at  $\leq$ 4 mg/liter [data not shown]), imipenem (93.2% susceptible), and gentamicin (75.3% susceptible) were the most active against these AmpC-derepressed-phenotype isolates [\(Table 6\)](#page-14-0). Omadacycline inhibited 95.5% of Citrobacter urinary tract infection isolates at  $\leq$ 4 mg/liter, activity exceeded only by that of imipenem (98.5% susceptible).

## **DISCUSSION**

Antimicrobial resistance is a growing problem worldwide [\(17\)](#page-19-16). Active surveillance and antimicrobial stewardship efforts are essential to combat this threat to patient safety across all health care settings [\(18,](#page-19-17) [19\)](#page-19-18). The SENTRY Antimicrobial Surveillance Program has conducted surveillance of antimicrobial-resistant pathogens globally for more than 20 years [\(20\)](#page-19-19) and for omadacycline-resistant pathogens since 2009 [\(12,](#page-19-11) [21](#page-19-20)[–](#page-19-21)[26\)](#page-19-22). The present study documents the in vitro activity of omadacycline against 49,000 bacterial isolates collected in the United States and Europe during the 2016 to 2018 SENTRY survey.

Overall, omadacycline provided broad coverage against Gram-positive cocci and Gram-negative bacilli, including those isolated from patients with acute bacterial skin <span id="page-14-0"></span>**TABLE 6** Activity of omadacycline and comparator antimicrobial agents when tested against bacterial isolates from urinary tract infections in the United States and Europe, SENTRY Program, 2016 to 2018



# **TABLE 6** (Continued)



### **TABLE 6** (Continued)



aCriteria published by the Clinical and Laboratory Standards Institute [\(33\)](#page-20-0) and The European Committee on Antimicrobial Susceptibility Testing [\(34\)](#page-20-1). S, susceptibility; R, resistance; —, not applicable.

 $b$ The extended-spectrum  $\beta$ -lactamase (ESBL) phenotype was defined as having a MIC value of  $\geq$ 2 mg/liter for ceftazidime, ceftriaxone, or aztreonam (confirmatory testing was not performed).

<sup>c</sup>Organisms include Citrobacter amalonaticus (n = 1), C. amalonaticus/Citrobacter farmeri (n = 4), Citrobacter braakii (n = 1), Citrobacter freundii (n = 28), C. freundii species complex ( $n = 80$ ), Citrobacter koseri ( $n = 83$ ), and Citrobacter sedlakii ( $n = 1$ ).

and skin structure infection, community-acquired bacterial pneumonia, and urinary tract infection [\(Tables 2](#page-3-0) and [4](#page-7-0) to [6\)](#page-14-0). Omadacycline was active against methicillinresistant S. aureus isolates, MR coagulase-negative staphylococci, vancomycin-resistant enterococci, viridans group streptococci, beta-hemolytic streptococci, and penicillinand macrolide-resistant S. pneumoniae isolates [\(Table 2\)](#page-3-0). Tetracycline-resistant Grampositive strains remained susceptible to omadacycline. Omadacycline was active against extended-spectrum- $\beta$ -lactamase-phenotype and carbapenem-resistant strains of E. coli and was less active against extended-spectrum- $\beta$ -lactamase-phenotype K. pneumoniae, carbapenem-resistant K. pneumoniae, and ceftazidime-nonsusceptible E. cloacae strains. Tetracycline-resistant Enterobacteriaceae strains were slightly less susceptible to omadacycline than tetracycline-susceptible strains. Omadacycline demonstrated activity against Acinetobacter spp. and S. maltophilia as well as against respiratory isolates of H. influenzae and M. catarrhalis. Omadacycline was not active (MIC $_{00}$ ,  $\geq$ 32 mg/liter) at the concentrations tested against Proteus spp., indole-positive Proteus spp., and P. aeruginosa.

Omadacycline was among the most active agents tested against pathogens from each of the key clinical indications, acute bacterial skin and skin structure infection, community-acquired bacterial pneumonia, and urinary tract infection. Among the most common Gram-positive cocci isolated from patients with acute bacterial skin and skin structure infection, more than 90.0% (97.4% to 100.0% susceptible) of S. aureus, S. lugdunensis, S. pyogenes, S. anginosus group, and E. faecalis isolates were susceptible to omadacycline at the approved U.S. Food and Drug Administration breakpoints [\(Table](#page-1-0) [1\)](#page-1-0). Methicillin-resistant S. aureus isolates accounted for 31.0% of S. aureus isolates from acute bacterial skin and skin structure infection, and 97.1% were susceptible to omadacycline [\(Table 4\)](#page-7-0). Comparable activity was seen with linezolid (99.9% susceptible), daptomycin (100.0% susceptible), trimethoprim-sulfamethoxazole (97.4% susceptible), and vancomycin (100.0% susceptible). In addition to omadacycline (98.2 to 100.0% susceptible), all S. pyogenes and S. anginosus group isolates from acute bacterial skin and skin structure infection were susceptible to linezolid, daptomycin, and vancomycin. Omadacycline was the most active agent by the  $MIC_{50/90}$  against acute bacterial skin and skin structure infection isolates of E. faecalis and E. faecium, including vancomycinnonsusceptible strains.

Omadacycline and imipenem were the most active agents tested against acute bacterial skin and skin structure infection isolates of E. coli, K. pneumoniae, and the E.  $cloacae$  species complex, including extended-spectrum- $\beta$ -lactamase-phenotype and ceftazidime-nonsusceptible isolates [\(Table 4\)](#page-7-0). Likewise, omadacycline and doxycycline

were the most active agents tested against A. baumannii isolates from acute bacterial skin and skin structure infection.

The major pathogens associated with community-acquired bacterial pneumonia included S. pneumoniae, H. influenzae, and M. catarrhalis [\(Table 5\)](#page-12-0). Omadacycline was active against these organisms, including penicillin-resistant, macrolide-resistant, and tetracycline-resistant strains of S. pneumoniae. Omadacycline, levofloxacin, linezolid, and vancomycin were the most active agents against S. pneumoniae. H. influenzae and M. catarrhalis were generally susceptible to most agents tested.

Urinary tract infections (UTIs) commonly occur in the community and health care settings [\(27\)](#page-19-23). The microbial spectrum of uncomplicated cystitis and pyelonephritis consists mainly (75 to 95%) of E. coli, with occasional species of other Enterobacterales, such as Klebsiella pneumoniae and Proteus mirabilis, being responsible [\(27\)](#page-19-23). Grampositive cocci (GPC; methicillin-resistant Staphylococcus aureus [MRSA] and enterococci) are common in more complicated catheter-associated urinary tract infections (cUTIs) but are rarely isolated in uncomplicated urinary tract infections (uUTIs) [\(27\)](#page-19-23). Omadacycline and imipenem were the most active agents tested against Enterobacteriaceae isolates from UTIs, including extended-spectrum- $\beta$ -lactamase-phenotype isolates of E. coli and K. pneumoniae and ceftazidime-nonsusceptible isolates of the E. cloacae species complex. The Gram-positive cocci causing urinary tract infection were most often S. aureus and Enterococcus spp. [\(Table 6\)](#page-14-0). Omadacycline, linezolid, and daptomycin were among the most active agents tested against both S. aureus and enterococci, including the methicillin-resistant S. aureus and vancomycin-resistant enterococcus subsets. In view of the broad range of activity of omadacycline against Gram-positive cocci and Gram-negative bacilli [\(Tables 2](#page-3-0) and [6\)](#page-14-0), it would seem to be a useful choice in treating cUTIs, in which mixed Gram-positive cocci and Gram-negative bacilli may occur and pathogens resistant to older antimicrobials are common [\(3,](#page-19-2) [5,](#page-19-4) [25\)](#page-19-21).

It has been established that the ratio of the area under the concentration-time curve (AUC) to the MIC (AUC/MIC) is the pharmacokinetic (PK)/pharmacodynamic (PD) parameter that best correlates with antibacterial efficacy for the tetracycline class of antibiotics [\(28\)](#page-19-24). Studies by Lepak et al. of omadacycline in S. aureus neutropenic mouse models of pneumonia [\(29\)](#page-19-25) and thigh infection [\(30\)](#page-19-26) have confirmed this to be true for omadacycline as well. Based on the MIC<sub>90</sub> value of 0.25 mg/liter for omadacycline and S. aureus, as shown in numerous large surveillance studies, including SENTRY [\(10,](#page-19-8) [12,](#page-19-11) [21,](#page-19-20) [22,](#page-19-27) [24\)](#page-19-28), the clinical doses of omadacycline would produce exposures that would exceed all 1-log-kill targets for epithelial lining fluid (ELF) and plasma in the pneumonia model and the plasma stasis target in the thigh infection model. Bhavnani et al. [\(31\)](#page-19-29) used the in vivo PD targets for the S. aureus thigh infection model in Monte Carlo simulations to predict the probability of target attainment in patients with acute bacterial skin and skin structure infection. The omadacycline MIC distribution for S. aureus was simulated using clinical isolate data from the SENTRY Surveillance Program ( $MIC<sub>90</sub>$ , 0.25 mg/liter) [\(25\)](#page-19-21). They found that at the MIC<sub>90,</sub> the predicted target attainment for bacteriostasis exceeded 90% for dosing regimens of either 100 mg or 200 mg intravenously (i.v.) twice daily on day 1 followed by 100 mg i.v. once on day 2 and 300 mg orally on day 3. These data support the FDA clinical breakpoint of  $\leq$ 0.5 mg/liter for S. aureus and acute bacterial skin and skin structure infection [\(29\)](#page-19-25).

These data build upon information from the SENTRY Program beginning in 2009 [\(12,](#page-19-11) [21](#page-19-20)[–](#page-19-21)[26\)](#page-19-22) and demonstrate a consistency in the spectrum of omadacycline activity over time and across geographic regions. In addition, we document the excellent activity of omadacycline against key pathogens from the U.S. Food and Drug Administration indications of acute bacterial skin and skin structure infection and community-acquired bacterial pneumonia as well as those from the indication of urinary tract infection, for which clinical trials have recently completed. Surveillance is ongoing.

#### **MATERIALS AND METHODS**

**Organisms.** A total of 49,000 nonduplicate bacterial isolates were collected prospectively from 73 medical centers located in the United States (33 sites [23 states and all 9 United States Census Divisions], 24,500 isolates) and Europe (40 sites [19 countries], 24,500 isolates) for the 2016 to 2018 SENTRY Antimicrobial Surveillance Program. European isolates were obtained from Belarus (1 site), Belgium (1 site), The Czech Republic (1 site), France (5 sites), Germany (5 sites), Greece (1 site) Hungary (1 site), Ireland (2 sites), Israel (1 site), Italy (4 sites), Poland (1 site), Portugal (1 site), Romania (1 site), Russia (3 sites), Slovenia (1 site), Spain (3 sites), Sweden (3 sites), Turkey (2 sites), and the United Kingdom (3 sites). All organisms were isolated from hospitalized patients with bloodstream infections (12,758 isolates), community-acquired respiratory tract infections (5,135 isolates), hospital-associated respiratory tract infections (10,225 isolates), acute bacterial skin and skin structure infections (11,013 isolates), intra-abdominal infections (2,829 isolates), complicated urinary tract infections (5,914 isolates), and other types of infections (1,126 isolates). Isolates were identified to the species level at each participating medical center, and isolate identity was confirmed by the monitoring laboratory (JMI Laboratories, North Liberty, IA, USA), using standard microbiology methods and matrixassisted laser desorption ionization–time of flight technology mass spectrometry (Bruker, Billerica, Massachusetts, United States) when necessary.

**Antimicrobial susceptibility testing.** MIC values were determined using the reference Clinical and Laboratory Standards Institute broth microdilution method [\(32\)](#page-19-30). Quality control and results interpretation were performed in accordance with the Clinical and Laboratory Standards Institute M100 document [\(33\)](#page-20-0) and European Committee on Antimicrobial Susceptibility Testing (EUCAST) [\(34\)](#page-20-1) guidelines. In the absence of omadacycline Clinical and Laboratory Standards Institute breakpoints, U.S. Food and Drug Administration breakpoints for acute bacterial skin and skin structure infection and community-acquired bacterial pneumonia [\(Table 1\)](#page-1-0) were applied [\(35\)](#page-20-2). E. coli and K. pneumoniae were grouped as being of extended-spectrum- $\beta$ -lactamase phenotype based on the Clinical and Laboratory Standards Institute screening criteria for potential extended-spectrum- $\beta$ -lactamase production, i.e., a MIC of  $\geq$ 2 mg/liter for ceftazidime, ceftriaxone, or aztreonam [\(33\)](#page-20-0). Although other  $\beta$ -lactamases, such as AmpC and KPC, may also produce an extended-spectrum- $\beta$ -lactamase phenotype, these strains were grouped because they demonstrate resistance to various broad-spectrum  $\beta$ -lactam compounds. Carbapenem-resistant *Enterobacteriaceae* were defined as having an imipenem MIC value of  $\geq$ 4 mg/liter. E. cloacae species complex isolates were classified as ceftazidime susceptible (MIC,  $\leq$ 4 mg/liter) and ceftazidime nonsusceptible (MIC,  $\geq$ 8 mg/liter). Other isolates with resistant phenotypes tested included methicillin-resistant S. aureus (oxacillin MIC,  $\geq$ 4 mg/liter, or cefoxitin MIC,  $\geq$ 8 mg/liter); vancomycin-nonsusceptible enterococci (MIC,  $\geq$ 8 mg/liter); tetracycline-resistant Enterobacteriaceae, A. baumannii, staphylococci, and enterococci (all MIC values,  $\geq$ 16 mg/liter) and S. pneumoniae (MIC,  $\geq$ 4 mg/liter); and macrolide-resistant beta-hemolytic streptococci (erythromycin MIC,  $\geq$  1 mg/liter). Quality control strains were tested concurrently and included E. coli ATCC 25922 and 35218, S. aureus ATCC 29213, P. aeruginosa ATCC 27853, E. faecalis ATCC 29212, and S. pneumoniae ATCC 49619. All quality control results were within published ranges.

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#### <span id="page-19-0"></span>**REFERENCES**

- 1. Chopra I. 2002. New developments in tetracycline antibiotics: glycylcyclines and tetracycline efflux pump inhibitors. Drug Resist Updat 5:119 –125. [https://doi.org/10.1016/s1368-7646\(02\)00051-1.](https://doi.org/10.1016/s1368-7646(02)00051-1)
- <span id="page-19-2"></span><span id="page-19-1"></span>2. Roberts MC. 2003. Tetracycline therapy: update. Clin Infect Dis 36: 462– 467. [https://doi.org/10.1086/367622.](https://doi.org/10.1086/367622)
- <span id="page-19-3"></span>3. Gallagher JC. 2019. Omadacycline: a modernized tetracycline. Clin Infect Dis 69:S1–S5. [https://doi.org/10.1093/cid/ciz394.](https://doi.org/10.1093/cid/ciz394)
- 4. Grossman TH. 2016. Tetracycline antibiotics and resistance. Cold Spring Harb Perspect Med 6:a025387. [https://doi.org/10.1101/](https://doi.org/10.1101/cshperspect.a025387) [cshperspect.a025387.](https://doi.org/10.1101/cshperspect.a025387)
- <span id="page-19-4"></span>5. Karlowsky JA, Steenbergen J, Zhanel GG. 2019. Microbiology and preclinical review of omadacycline. Clin Infect Dis 69:S6 –S15. [https://doi](https://doi.org/10.1093/cid/ciz395) [.org/10.1093/cid/ciz395.](https://doi.org/10.1093/cid/ciz395)
- <span id="page-19-6"></span><span id="page-19-5"></span>6. Chambers HF. 2019. Omadacycline—the newest tetracycline. N Engl J Med 380:588 –589. [https://doi.org/10.1056/NEJMe1900188.](https://doi.org/10.1056/NEJMe1900188)
- 7. Tanaka SK, Steenbergen J, Villano S. 2016. Discovery, pharmacology, and clinical profile of omadacycline, a novel aminomethylcycline antibiotic. Bioorg Med Chem 24:6409 – 6419. [https://doi.org/10.1016/j.bmc.2016](https://doi.org/10.1016/j.bmc.2016.07.029) [.07.029.](https://doi.org/10.1016/j.bmc.2016.07.029)
- <span id="page-19-7"></span>8. Draper MP, Weir S, Macone A, Donatelli J, Trieber CA, Tanaka SK, Levy SB. 2014. Mechanism of action of the novel aminomethylcycline antibiotic omadacycline. Antimicrob Agents Chemother 58:1279 –1283. [https://doi](https://doi.org/10.1128/AAC.01066-13) [.org/10.1128/AAC.01066-13.](https://doi.org/10.1128/AAC.01066-13)
- <span id="page-19-10"></span>9. Honeyman L, Ismail M, Nelson ML, Bhatia B, Bowser TE, Chen J, Mechiche R, Ohemeng K, Verma AK, Cannon EP, Macone A, Tanaka SK, Levy S. 2015. Structure-activity relationship of the aminomethylcyclines and the discovery of omadacycline. Antimicrob Agents Chemother 59: 7044 –7053. [https://doi.org/10.1128/AAC.01536-15.](https://doi.org/10.1128/AAC.01536-15)
- <span id="page-19-8"></span>10. Macone AB, Caruso BK, Leahy RG, Donatelli J, Weir S, Draper MP, Tanaka SK, Levy SB. 2014. In vitro and in vivo antibacterial activities of omadacycline, a novel aminomethylcycline. Antimicrob Agents Chemother 58:1127–1135. [https://doi.org/10.1128/AAC.01242-13.](https://doi.org/10.1128/AAC.01242-13)
- <span id="page-19-9"></span>11. Villano S, Steenbergen J, Loh E. 2016. Omadacycline: development of a novel aminomethylcycline antibiotic for treating drug-resistant bacterial infections. Future Microbiol 11:1421–1434. [https://doi.org/10.2217/fmb](https://doi.org/10.2217/fmb-2016-0100) [-2016-0100.](https://doi.org/10.2217/fmb-2016-0100)
- <span id="page-19-11"></span>12. Pfaller MA, Huband MD, Rhomberg PR, Flamm RK. 2017. Surveillance of omadacycline activity tested against clinical isolates from a global (North America, Europe, Latin America, and Asia-Western Pacific) collection (2010-2011). Antimicrob Agents Chemother 61:e00018-17. [https://doi](https://doi.org/10.1128/AAC.00018-17) [.org/10.1128/AAC.00018-17.](https://doi.org/10.1128/AAC.00018-17)
- <span id="page-19-12"></span>13. O'Riordan W, Green S, Overcash JS, Puljiz I, Metallidis S, Gardovskis J, Garrity-Ryan L, Das AF, Tzanis E, Eckburg PB, Manley A, Villano SA, Steenbergen JN, Loh E. 2019. Omadacycline for acute bacterial skin and skin-structure infections. N Engl J Med 380:528 –538. [https://doi.org/10](https://doi.org/10.1056/NEJMoa1800170) [.1056/NEJMoa1800170.](https://doi.org/10.1056/NEJMoa1800170)
- <span id="page-19-13"></span>14. Stets R, Popescu M, Gonong JR, Mitha I, Nseir W, Madej A, Kirsch C, Das AF, Garrity-Ryan L, Steenbergen JN, Manley A, Eckburg PB, Tzanis E, McGovern PC, Loh E. 2019. Omadacycline for community-acquired bacterial pneumonia. N Engl J Med 380:517–527. [https://doi.org/10.1056/](https://doi.org/10.1056/NEJMoa1800201) [NEJMoa1800201.](https://doi.org/10.1056/NEJMoa1800201)
- <span id="page-19-15"></span><span id="page-19-14"></span>15. ClincalTrials.gov. 2017. Oral omadacycline vs. oral nitrofurantoin for the treatment of cystitis. [https://clinicaltrials.gov/ct2/show/NCT03425396.](https://clinicaltrials.gov/ct2/show/NCT03425396) Accessed June 2018.
- <span id="page-19-16"></span>16. ClinicalTrials.gov. 2018. IV or IV/PO omadacycline vs. IV/PO levofloxacin for the treatment of acute pyelonephritis. [https://clinicaltrials.gov/ct2/](https://clinicaltrials.gov/ct2/show/NCT03757234) [show/NCT03757234.](https://clinicaltrials.gov/ct2/show/NCT03757234) Accessed June 2018.
- <span id="page-19-17"></span>17. Friedman ND, Temkin E, Carmeli Y. 2016. The negative impact of antibiotic resistance. Clin Microbiol Infect 22:416 – 422. [https://doi.org/10](https://doi.org/10.1016/j.cmi.2015.12.002) [.1016/j.cmi.2015.12.002.](https://doi.org/10.1016/j.cmi.2015.12.002)
- <span id="page-19-18"></span>18. Perez F, Villegas MV. 2015. The role of surveillance systems in confronting the global crisis of antibiotic-resistant bacteria. Curr Opin Infect Dis 28:375–383. [https://doi.org/10.1097/QCO.0000000000000182.](https://doi.org/10.1097/QCO.0000000000000182)
- 19. Schuts EC, Hulscher ME, Mouton JW, Verduin CM, Stuart JW, Overdiek HW, van der Linden PD, Natsch S, Hertogh CM, Wolfs TF, Schouten JA, Kullberg BJ, Prins JM. 2016. Current evidence on hospital antimicrobial

stewardship objectives: a systematic review and meta-analysis. Lancet Infect Dis 16:847– 856. [https://doi.org/10.1016/S1473-3099\(16\)00065-7.](https://doi.org/10.1016/S1473-3099(16)00065-7)

- <span id="page-19-19"></span>20. Fuhrmeister A, Jones RN. 2019. The importance of antimicrobial resistance monitoring worldwide and the origins of SENTRY Antimicrobial Surveillance Program. Open Forum Infect Dis 6:S1–S4. [https://doi.org/](https://doi.org/10.1093/ofid/ofy346) [10.1093/ofid/ofy346.](https://doi.org/10.1093/ofid/ofy346)
- <span id="page-19-20"></span>21. Carvalhaes CG, Huband MD, Reinhart HH, Flamm RK, Sader HS. 2019. Antimicrobial activity of omadacycline tested against clinical bacterial isolates from hospitals in mainland China, Hong Kong, and Taiwan: results from the SENTRY Antimicrobial Surveillance Program (2013 to 2016). Antimicrob Agents Chemother 63:e02262-18. [https://doi.org/10](https://doi.org/10.1128/AAC.02262-18) [.1128/AAC.02262-18.](https://doi.org/10.1128/AAC.02262-18)
- <span id="page-19-27"></span>22. Huband MD, Pfaller MA, Shortridge D, Flamm RK. 2019. Surveillance of omadacycline activity tested against clinical isolates from the United States and Europe: results from the SENTRY Antimicrobial Surveillance Programme, 2017. J Glob Antimicrob Resist 19:56 – 63. [https://doi.org/](https://doi.org/10.1016/j.jgar.2019.02.017) [10.1016/j.jgar.2019.02.017.](https://doi.org/10.1016/j.jgar.2019.02.017)
- 23. Pfaller MA, Rhomberg PR, Huband MD, Flamm RK. 2017. Activities of omadacycline and comparator agents when tested against Staphylococcus aureus isolates from a surveillance program conducted in North America and Europe. Antimicrob Agents Chemother 61:e02411-16. [https://doi.org/10.1128/AAC.02411-16.](https://doi.org/10.1128/AAC.02411-16)
- <span id="page-19-28"></span>24. Pfaller MA, Rhomberg PR, Huband MD, Flamm RK. 2018. Activity of omadacycline tested against Streptococcus pneumoniae from a global surveillance program (2014). Diagn Microbiol Infect Dis 90:143–147. [https://doi.org/10.1016/j.diagmicrobio.2017.10.010.](https://doi.org/10.1016/j.diagmicrobio.2017.10.010)
- <span id="page-19-21"></span>25. Pfaller MA, Rhomberg PR, Huband MD, Flamm RK. 2018. Activity of omadacycline tested against Enterobacteriaceae causing urinary tract infections from a global surveillance program (2014). Diagn Microbiol Infect Dis 91: 179 –183. [https://doi.org/10.1016/j.diagmicrobio.2018.01.019.](https://doi.org/10.1016/j.diagmicrobio.2018.01.019)
- <span id="page-19-22"></span>26. Pfaller MA, Huband MD, Shortridge D, Flamm RK. 2018. Surveillance of omadacycline activity tested against clinical isolates from the United States and Europe: report from the SENTRY Antimicrobial Surveillance Program, 2016. Antimicrob Agents Chemother 62:e02327-17. [https://doi](https://doi.org/10.1128/AAC.02327-17) [.org/10.1128/AAC.02327-17.](https://doi.org/10.1128/AAC.02327-17)
- <span id="page-19-23"></span>27. Gupta K, Hooton TM, Naber KG, Wullt B, Colgan R, Miller LG, Moran GJ, Nicolle LE, Raz R, Schaeffer AJ, Soper DE, Infectious Diseases Society of America, European Society for Microbiology and Infectious Diseases. 2011. International clinical practice guidelines for the treatment of acute uncomplicated cystitis and pyelonephritis in women: a 2010 update by the Infectious Diseases Society of America and the European Society for Microbiology and Infectious Diseases. Clin Infect Dis 52:e103– e120. [https://doi.org/10.1093/cid/ciq257.](https://doi.org/10.1093/cid/ciq257)
- <span id="page-19-24"></span>28. Gotfried MH, Horn K, Garrity-Ryan L, Villano S, Tzanis E, Chitra S, Manley A, Tanaka SK, Rodvold KA. 2017. Comparison of omadacycline and tigecycline pharmacokinetics in the plasma, epithelial lining fluid, and alveolar cells of healthy adult subjects. Antimicrob Agents Chemother 61:e01135-17. [https://doi.org/10.1128/AAC.01135-17.](https://doi.org/10.1128/AAC.01135-17)
- <span id="page-19-25"></span>29. Lepak AJ, Zhao M, Marchillo K, VanHecker J, Andes DR. 2020. In vivo pharmacodynamic evaluation of omadacycline against Staphylococcus aureus in the neutropenic mouse pneumonia model. Antimicrob Agents Chemother 64:e02058-19. [https://doi.org/10.1128/AAC.02058-19.](https://doi.org/10.1128/AAC.02058-19)
- <span id="page-19-26"></span>30. Lepak AJ, Zhao M, Marchillo K, VanHecker J, Andes DR. 2019. In vivo pharmacodynamics of omadacycline against Staphylococcus aureus in the neutropenic murine thigh infection model. Antimicrob Agents Chemother 63:e00624-19. [https://doi.org/10.1128/AAC.00624-19.](https://doi.org/10.1128/AAC.00624-19)
- <span id="page-19-29"></span>31. Bhavnani SM, Hammel JP, Lakota EA, Liolios K, Rubino CM, Steenbergen JN, Friedrich L, Tzanis E, Ambrose PA. 2019. Assessment of pharmacokineticspharmacodynamics to support omadacyline dosing regimens for the treatment of patients with acute bacterial skin and skin structure infections (ABSSSI), abstr P1944. Abstr European Congress of Clinical Microbiology and Infectious Disease (ECCMID), Amsterdam, Netherlands.
- <span id="page-19-30"></span>32. Clinical and Laboratory Standards Institute. 2018. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard, 11th ed. M07Ed11. Clinical and Laboratory Standards Institute, Wayne, PA.
- <span id="page-20-0"></span>33. Clinical and Laboratory Standards Institute. 2019. Performance standards for antimicrobial susceptibility testing: 29th informational supplement. M100Ed29. Clinical and Laboratory Standards Institute, Wayne, PA.
- <span id="page-20-1"></span>34. European Committee on Antimicrobial Susceptibility Testing. 2019. Breakpoint tables for interpretation of MICs and zone diameters, version 9.0. [http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/](http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_tables/v_9.0_Breakpoint_Tables.pdf) [Breakpoint\\_tables/v\\_9.0\\_Breakpoint\\_Tables.pdf.](http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_tables/v_9.0_Breakpoint_Tables.pdf) Accessed January 2019.
- <span id="page-20-2"></span>35. U.S. Food and Drug Administration. 2018. Recognized antimicrobial susceptibility test interpretive criteria for omadacycline. [https://www](https://www.fda.gov/drugs/development-resources/antibacterial-susceptibility-test-interpretive-criteria) [.fda.gov/drugs/development-resources/antibacterial-susceptibility-test](https://www.fda.gov/drugs/development-resources/antibacterial-susceptibility-test-interpretive-criteria) [-interpretive-criteria.](https://www.fda.gov/drugs/development-resources/antibacterial-susceptibility-test-interpretive-criteria) Accessed 15 October 2018.
- <span id="page-20-3"></span>36. Paratek Pharmaceuticals. 2018. NuzyraTM (omadacycline) package insert. [https://www.accessdata.fda.gov/drugsatfda\\_docs/label/2018/209816](https://www.accessdata.fda.gov/drugsatfda_docs/label/2018/209816_209817lbl.pdf) [\\_209817lbl.pdf.](https://www.accessdata.fda.gov/drugsatfda_docs/label/2018/209816_209817lbl.pdf) Accessed November 2019.