

Comparable *In Vitro* Activities of Second-Generation HIV-1 Integrase Strand Transfer Inhibitors (INSTIs) on HIV-1 Clinical Isolates with INSTI Resistance Mutations

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ABSTRACT Second-generation HIV-1 integrase strand transfer inhibitors (INSTIs) dolutegravir (DTG), bictegravir (BIC), and cabotegravir (CAB) showed a high genetic barrier to resistance and limited cross-resistance with first-generation INSTIs raltegravir (RAL) and elvitegravir (EVG). In this study, DTG, BIC, and CAB demonstrated a comparable activity on a panel of INSTI-resistant strains isolated from patients exposed to RAL, EVG, and/or DTG, with a significantly reduced susceptibility only with the pathway Q148H/K/R plus one to two additional INSTI mutations.

KEYWORDS HIV-1 resistance, integrase inhibitors, phenotypic assay, dolutegravir, cabotegravir, bictegravir

Exposure to first-generation HIV-1 integrase strand transfer inhibitors (INSTIs), such as raltegravir (RAL) and elvitegravir (EVG), is frequently associated with the selection of resistance-associated mutations (RAMs) at virological failure. Recently developed second-generation INSTIs, such as dolutegravir (DTG; approved by FDA and European Medicines Agency [EMA] in 2013 and 2014, respectively), bictegravir (BIC; approved by FDA and EMA in 2018 as part of a single tablet regimen, including tenofovir alafenamide [TAF] and emtricitabine [FTC]), and cabotegravir (CAB; currently under phase III clinical investigation) have demonstrated a superior genetic barrier to resistance and a variable activity against viruses harboring INSTI resistance mutations selected by RAL and EVG. However, predicting the activity of secondgeneration INSTIs on HIV-1 variants with different combination of INSTI RAMs is not straightforward. This study aimed at clarifying cross-resistance to DTG, BIC, and CAB in a panel of INSTI-resistant strains isolated from patients previously exposed to RAL, EVG, and/or DTG.

Plasma samples from 19 patients were collected during routine drug resistance testing under virological failure of an INSTI regimen, based on the detection of at least one major INSTI RAM (T66A, E92Q, E138K/T, G140A/C/S, Y143C/R, S147G, Q148H/R, N155H, and R263K). Virological failure was defined at the individual clinics as confirmed viral load of >50 HIV-1 RNA copies/ml or one viral load of >1,000 HIV-1 RNA copies/ml. Access to residual samples for research purposes was regulated by informed consent approved by the Southeast Tuscany Ethics Committee. Plasma RNA was extracted by the DSP virus kit using the EZ1 workstation (Qiagen, Hilden, Germany). Population sequencing of the integrase region was performed by homebrew technology using primers previously described (1). For *in vitro* susceptibility testing, the integrase coding region was amplified through a 2-step PCR protocol, and the resulting amplicon was used for the generation of chimeric viruses by homologous recombination with an integrase-deleted pNL4-3 vector in 293T cells (2). Integrase sequences of recombinant viruses were confirmed by Sanger sequencing.

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TABLE 1 Stanford HIVdb major and accessory	INSTI RAMs and DTG,	BIC, and CAB IC ₅₀ and FC	2 values in a panel of 1	9 samples from
patients failing INSTI-based therapy ^a				

Sample	Subtype	INSTI exposure ^b	Major mutation(s)	Accessory mutation	DTG		BIC		САВ		
					$IC_{50} \pm SD nM$	FC	$IC_{50} \pm SD nM$	FC	$IC_{50} \pm SD nM$	FC	Group
NL4-3	В	Wild type	None	None	0.8 ± 0.4	1	0.5 ± 0.1	1	1.8 ± 0.9	1	Wild type
151277	F1	RAL	Y143R	T97A	0.8 ± 0.1	1.0	1.1 ± 0.4	2.4	$2.1~\pm~1.0$	1.2	Y143C/H/R
151648	G	RAL	Y143C	S230R	2.6 ± 1.7	3.5	1.2 ± 0.1	2.5	$4.0~\pm~0.5$	2.2	
150678	В	RAL	Q148R	None	0.4 ± 0.3	0.5	0.4 ± 0.3	0.8	2.1 ± 0.1	1.2	Q148H/K/R
146943	В	RAL	G140C, Q148R	None	6.0 ± 4.9	8.0	3.8 ± 2.4	8.0	121.3 ± 40.9	66.3	Q148H/K/R plus
147026	В	EVG	E138K, Q148R	None	0.8 ± 0.9	1.0	1.5 ± 0.3	3.2	18.2 ± 8.0	9.9	one RAM
147086	В	RAL	G140A, Q148R	None	5.5 ± 5.7	7.3	2.5 ± 0.5	5.4	39.6 ± 28.8	21.7	
149357	В	RAL, DTG	E138K, G140S, Q148H, N155H	None	>100	>100	>100	>100	>200	>100	Q148H/K/R plus two
151377	В	RAL, DTG	E138T, G140S, Q148H	T97A	>100	>100	>100	>100	>200	>100	RAMs
151236	В	RAL, DTG	E138K, G140A, S147SG, Q148R	T97A	>100	>100	>100	>100	>200	>100	
150953	CRF03_AB	RAL	N155H	E157Q	1.6 ± 1.2	2.1	0.9 ± 0.2	1.8	3.0 ± 0.7	1.6	N155H
150336	G	RAL	N155H	T97A	3.0 ± 0.2	4.0	0.7 ± 0.2	1.5	5.4 ± 4.2	3.0	
149789	В	RAL, EVG	N155H	None	2.0 ± 0.9	2.7	0.8 ± 0.4	1.7	2.3 ± 0.9	1.3	
150077	В	EVG	N155H	None	3.3 ± 0.7	4.4	1.6 ± 0.8	3.4	4.3 ± 1.2	2.3	
149865	В	RAL	N155H	None	3.4 ± 1.4	4.5	1.2 ± 0.3	2.5	4.5 ± 2.8	2.5	
150348	В	RAL	N155H	None	2.3 ± 2.0	3.1	0.8 ± 0.4	1.7	4.2 ± 0.2	2.3	
149570	CRF06_cpx	EVG	E92Q	None	0.9 ± 0.1	1.2	0.2 ± 0.1	0.4	1.4 ± 0.5	0.8	Others
148640	В	RAL, DTG	E92Q	None	1.9 ± 0.3	2.6	0.7 ± 0.6	1.4	2.3 ± 1.0	1.3	
150129	В	EVG	T66A, S147SG	G163K	0.5 ± 0.1	0.7	0.1 ± 0.1	0.2	1.4 ± 0.4	0.8	
151422	F1	RAL, DTG	R263K	None	2.7 ± 0.2	3.5	0.5 ± 0.1	1.1	6.5 ± 2.3	3.5	
Median (IQR)					2.6 (1.4–3.9)	3.5 (1.2–7.3)	1.1 (0.7–1.8)	2.4 (1.4–5.4)	4.3 (2.3–39.6)	2.3 (1.3–21.7)	

^aINSTI, integrase strand transfer inhibitor; RAM, resistance-associated mutation; DTG, dolutegravir; BIC, bictegravir; CAB, cabotegravir; IC₅₀, half maximal inhibitory concentration; FC, fold change; SD, standard deviation; IQR, interquartile range; HIVdb, HIV database.

^bWhere two INSTIs are indicated, the former was used before and the latter was in use at the time of genotyping; therapy based on the first INSTI failed in all these cases except for cases 149789 and 149357, where the second INSTI-based therapy was started as simplification while the patient was virologically suppressed. Where only one INSTI is indicated, this was in use at the time of genotyping.

Susceptibility to DTG, BIC, and CAB was assessed through a TZM-bl cell line-based phenotypic assay, shown to correlate well with the *de facto* reference PhenoSense assay by Monogram Biosciences and expressed as fold change (FC) with respect to the reference wild type NL4-3 virus (2). Differences in FC values among groups were tested by Friedman's ANOVA, and multiple comparisons were performed by the Dunn's test when appropriate.

The patients had been exposed to RAL only (n = 9), EVG only (n = 4), RAL and DTG (n = 5), or RAL and EVG (n = 1). By querying the COMET HIV-1 subtyping tool (3), sequences were assigned to subtype B (n = 13), subtype F1 (n = 2), subtype G (n = 2), CRF03_AB (n = 1), and CRF06_cpx (n = 1). Sequences were grouped according to the presence of major INSTI RAMs at codon 143, 148, or 155, alone or in combination with other INSTI RAMs. Four samples with INSTI RAMs E92Q (n = 2), T66A plus S147SG, and R263K were grouped together as "others" (Table 1). Median (interquartile range [IQR]) FC values for DTG, BIC, and CAB were 3.5 (1.2 to 7.3), 2.4 (1.4 to 5.4), and 2.3 (1.3 to 21.7), respectively, without statistically significant differences between groups. Median (IQR) DTG, BIC, and CAB FC values from patients not exposed to DTG were 2.9 (1.0 to 4.4), 2.1 (1.3 to 3.3), and 2.3 (1.2 to 4.7), respectively, while exposure to RAL and then DTG resulted in a FC of >100 in three cases (Q148H/K/R plus 2 INSTI RAMs) and FC of <3.5 in two cases (samples with E92Q and with R263K) for all three drugs (Fig. 1A). Median (IQR) DTG, BIC, and CAB FC values were consistently, but not significantly, higher following exposure to RAL only versus EVG only (3.5 [1.6 to 5.9], 2.4 [1.6 to 3.9], and 2.3 [1.4 to 12.3] versus 1.1 [0.8 to 3.6], 1.8 [0.3 to 3.3], and 1.6 [0.8 to 8.0], respectively) (Fig. 1B). According to the three major INSTI resistance pathways, median (IQR) DTG, BIC, and CAB FC values were 2.3 (1.0 to 3.5), 2.5 (2.4 to 2.5), and 1.7 (1.2 to 2.2), respectively, with Y143R/C alone (n = 2); 3.6 (2.6 to 4.4), 1.8 (1.7 to 2.7), and 2.3 (1.5 to 2.6) with N155H alone (n = 6); 0.5, 0.8, and 1.2 with Q148R alone (n = 1); 7.3 (1.0 to 8.0), 5.4 (3.2 to 8.0), and 21.7 (9.9 to 66.3) with Q148R plus one additional INSTI RAM (n = 3); and >100 for all the three drugs with Q148H/K/R plus two additional INSTI RAMs (n = 3) (Fig. 1C). In the N155H group, the BIC and CAB FC values were significantly lower than the DTG FC values (P = 0.005). Among samples with other INSTI RAMs (two with E92Q, one with



FIG 1 Fold change (FC) dolutegravir (DTG), bictegravir (BIC), and cabotegravir (CAB) susceptibility values stratified by exposure to DTG (A), exposure to RAL only or EVG only (B), and mutational pathway (C). ns, *P* value not significant; **, *P* = 0.005.

R263K, and one with T66A plus S147S/G), FC values of \leq 3.5 were measured for all the three drugs. In particular, one sample from a failing DTG-based regimen harboring R263K had DTG and CAB FCs of 3.5, while BIC FC was 1.1.

These data are in line with and yet add further information to those reported with specific laboratory clones and with a few clinical samples (4–7). In the largest data set analyzed comprising 47 patient-derived HIV-1 isolates, BIC appeared to retain more activity against INSTI-resistant mutants than DTG (4). The median FC value for BIC was also lower than that for DTG in our case file; however, the difference was statistically significant only for the N155H pathway and yet not for the data overall. The Q148H/K/R plus G140S pathway was confirmed to be the most challenging for all second-generation INSTIs, with the addition of a third RAM (particularly at codon 138) substantially increasing the FC value, as shown also in previous studies (4–7). Contrary to the results generated with molecular clones (7), we did not detect better activity of DTG than with CAB, likely due to the overrepresentation of N155H single mutants in our data set.

In summary, DTG, BIC, and CAB retain comparable activity against major INSTI RAMs; however, adding one and, particularly, two additional INSTI RAMs to the Q148H/K/R pathway results in a substantial loss of susceptibility for all three drugs. The full spectrum of additional mutations playing such a role in different HIV-1 subtypes remains to be elucidated. More extensive *in vivo* data are required to fully define the correlation between HIV-1 genotype, FC values, and response to treatment with second-generation INSTIs.

Data availability. Integrase sequences of study samples were submitted to Gen-Bank under accession numbers MN389749 to MN389767.

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