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Immune imbalance and activation are associated with lower lung function in youth with perinatally-acquired HIV

Engi F. ATTIA, MD MPH^a, Denise JACOBSON, PhD MPH^b, Wendy YU, MPH^b, Claudia S. CROWELL, MD MPH^{a,c}, Elizabeth MALECHE-OBIMBO, MBChB MMed MPH^d, Paige L. WILLIAMS, PhD MS^b, T. Eoin WEST, MD MPH^a, Sandra K. BURCHETT, MD MSc^{e,f}, Meyer KATTAN, MD^g, Andrew A. COLIN, MD^h, Sherry ESKANDER, MBBCh MScⁱ, Michael H. CHUNG, MD MPH^a, Kristina CROTHERS, MD^a, William T. SHEARER, MD PhD^{j,*†} **Pediatric HIV/AIDS Cohort Study**

^aUniversity of Washington, Seattle, WA, US

^bHarvard T.H. Chan School of Public Health, Boston, MA, US

^cSeattle Children's Hospital, Seattle, WA, US

^dUniversity of Nairobi, Nairobi, Kenya

^eBoston Children's Hospital, Boston, MA, US

^fHarvard Medical School, Boston, MA, US

^gColumbia University Medical Center, New York City, NY, US

^hUniversity of Miami, Miami, FL, US

ⁱCoptic Hospital and Coptic Hope Center for Infectious Diseases, Nairobi, Kenya

Corresponding Author: Engi F. Attia, MD MPH, 325 Ninth Avenue, Campus Box 359762, Harborview Medical Center, Seattle, WA 98144, Phone: +1 206 744 3244, Fax: +1 206 744 8584, eattia@uw.edu.

*This work is dedicated to the memory of Dr. William T. Shearer. He was the senior mentor for this manuscript and participated with great joy in conducting the study and composing early drafts. It is of great solace to us all that he spent his last days working on this, and other, passion projects.

†Deceased: October 9, 2018.

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Baylor College of Medicine and Texas Children's Hospital, Houston, TX, US

Capsule Summary:

High CD8 T-cells and low CD4/CD8 are associated with lower lung function among youth living with perinatally-acquired HIV, despite antiretroviral therapy and CD4 preservation/reconstitution. Understanding these underlying mechanisms is critical to mitigate lung function impairment.

Keywords

Adolescents; youth; perinatally-acquired HIV; chronic lung disease; immune imbalance; immune activation; CD8 T-cells; CD4/CD8 ratio; lung function; FEV₁

To the Editor:

Contemporary reports indicate a high burden of impaired lung function among youth living with perinatally-acquired HIV (YLP HIV) worldwide. The prevalence of impaired lung function is estimated to be 33% among YLP HIV in the U.S.-based Pediatric HIV/AIDS Cohort Study (PHACS),¹ and up to 45% among YLP HIV in several sub-Saharan African countries.^{2,3}

Little is known about the pathophysiologic mechanisms underlying impaired lung function among YLP HIV. Persistent immune imbalance and activation, reflected by a low (reversed) CD4/CD8 ratio, are linked with chronic lung disease (CLD) among adults living with HIV despite antiretroviral therapy (ART) and CD4 T-cell reconstitution.⁴ In this study, we determined whether immune imbalance and activation based on a high CD8 T-cell count and low CD4/CD8 ratio were associated with lower lung function, as measured by forced expiratory volume in one second (FEV₁), among YLP HIV and explored whether FEV₁ differed between YLP HIV living in the U.S. and Kenya.

We performed a cross-sectional analysis of 10-21 year-old YLP HIV in the multi-site U.S.-based pulmonary sub-study of the PHACS Adolescent Master Protocol¹ and in a cohort study in Nairobi, Kenya.³ Both studies collected pre- and post-bronchodilator spirometry, CD4 and CD8 T-cell counts, demographics and anthropometric data. We obtained spirometry in both cohorts according to American Thoracic Society standards and used NHANES III reference equations to calculate percent-predicted values for FEV₁ (%FEV₁). Institutional review boards at Baylor College of Medicine, Harvard T.H. Chan School of Public Health, University of Washington, University of Nairobi/Kenyatta National Hospital, and participating U.S. sites approved the studies. Participants and guardians provided written consent/assent.

We compared characteristics between 188 U.S. and 49 Kenyan YLP HIV, using Chi-square and Wilcoxon rank-sum tests for categorical and continuous variables. We fit linear regression models to evaluate mean differences in %FEV₁ among U.S. compared to Kenyan youth, adjusted for age and sex. Because immunologic markers may differ by ethnicity/race and be potential confounders of the relationship between setting (U.S. vs Kenya) and FEV₁, we additionally adjusted for CD8 T-cells ("CD8" from here forward) or CD4/CD8 in

separate models.⁵ Conversely, CD8 and CD4/CD8 may be on the causal pathway between setting and FEV₁ by way of the association of these immunologic markers with environmental exposures and timing of ART initiation. To further evaluate the relationship of CD8 and CD4/CD8 with FEV₁, we calculated Spearman partial correlations of CD8 and CD4/CD8 with pre- and post-bronchodilator %FEV₁, adjusted for age and sex in U.S. and Kenyan youth separately. In sensitivity analyses, we restricted models to black U.S. and Kenyan YLPHIV. Because immune imbalance and activation may be triggered by infections, we also stratified the CD4/CD8 ratio by prior pulmonary tuberculosis.

U.S. youth were older, on average, than Kenyan youth (17 vs 13 years); 42% of U.S. youth and 51% of Kenyan youth were male (Table I). Active tobacco use was reported by 10% of U.S. youth, but Kenyan youth reported more exposure to passive smoking and nearly universal indoor combustible fuel use. Prior pulmonary tuberculosis was less common among U.S. compared to Kenyan youth. Although U.S. youth initiated ART at a median age of three years compared to eight years among Kenyan youth, U.S. and Kenyan youth had similar nadir and recent CD4, and similar proportions were receiving ART. Nonetheless, the median [interquartile range] CD4/CD8 was lower in Kenyan than U.S. youth (0.65 [0.42, 0.94] vs 0.91 [0.55, 1.25], $p=0.004$), driven by higher CD8 in Kenyan compared to U.S. youth (1098 cells/ μ L [849, 1473] vs 722 [514, 930], $p<0.001$).

Pre- and post-bronchodilator %FEV₁ were significantly lower among Kenyan compared to U.S. youth. On average, Kenyan YLPHIV had 9.6% (95% CI 3.8, 15.4) lower pre-bronchodilator %FEV₁ and 8.3% (2.7, 14.0) lower post-bronchodilator %FEV₁ than U.S. YLPHIV, after adjusting for age and sex. These differences were attenuated in models also adjusted for CD8 (pre-bronchodilator: -5.2 [-11.6 , 1.2], $p=0.11$; post-bronchodilator: -4.5 [-10.7 , 1.8], $p=0.16$) or CD4/CD8. Differences persisted in analyses restricted to black U.S. and Kenyan youth.

In U.S. youth, higher CD8 correlated with lower pre- and post-bronchodilator %FEV₁ ($r=-0.19$, $p=0.01$; $r=-0.16$, $p=0.03$) after adjusting for age and sex; when restricted to black youth, correlations remained negative at $r=-0.11$ ($p=0.23$) and $r=-0.13$ ($p=0.14$), respectively. In Kenyan youth, correlations were in the same direction ($r=-0.22$, $p=0.15$ and $r=-0.24$, $p=0.12$). Correlations of lower CD4/CD8 with lower %FEV₁ were similar. The distribution of CD4/CD8 was similar among youth with and without prior pulmonary tuberculosis across settings.

In summary, Kenyan YLPHIV had lower %FEV₁ than U.S. YLPHIV. Higher CD8 and lower CD4/CD8 were associated with lower pre-bronchodilator and post-bronchodilator %FEV₁, even in the setting of CD4 preservation/reconstitution.

Lower lung function, especially lower post-bronchodilator %FEV₁, may represent CLD that manifests with irreversible airflow limitation. Imaging studies among YLPHIV in sub-Saharan Africa suggest that obliterative bronchiolitis may represent a unique etiology of CLD.^{2,3} Bronchiectasis is also common, especially in light of prior pulmonary infections.² Asthma also contributes to CLD, potentially via mechanisms linked to immune

reconstitution or CD8 subsets; asthma-chronic obstructive pulmonary disease overlap may also play a role.^{6,7}

HIV infection and associated chronic immune activation during critical periods of immune system and organ development are likely to influence end-organ injury in YLPHIV, including CLD.² Although most experience CD4 reconstitution and decline in CD8 with contemporary ART, some maintain persistently elevated CD8 and depressed CD4/CD8. As CD8 counts can be higher in alveolar fluid than in peripheral circulation, persistent activation of residual pulmonary CD8 T-cells may be important in CLD.⁸ This may be particularly relevant for youth who initiate ART at older ages and do not experience a sufficient decline in CD8. Further, low peripheral CD4/CD8 is associated with CLD among adults living with HIV,⁴ and was correlated with lower %FEV₁ in our cohort of YLPHIV.

Importantly, Kenyan YLPHIV had lower %FEV₁ than U.S. YLPHIV. Measured risk factors, such as pulmonary infections, tobacco and combustible fuel smoke exposure, and age at ART initiation, as well as unmeasured factors, including disease severity, delayed HIV diagnosis/treatment, nutritional status and socioeconomic status likely contribute to observed differences in %FEV₁. However, these factors may also impact CD8 and, consequently, the CD4/CD8 ratio. Notably, we found no difference in CD4/CD8 among youth with and without prior pulmonary tuberculosis, suggesting that HIV-related immune imbalance and activation may play a bigger role.⁹ Although adjusting for CD8 or CD4/CD8 attenuated differences in %FEV₁, even in models restricted to black youth, this should be interpreted cautiously because the mechanistic pathway between these immunologic parameters and FEV₁ remains unclear.

Despite the additional inherent limitations of the small sample size, cross-sectional nature of our study, use of NHANES reference equations and lack of an HIV-uninfected comparison group, a substantial strength is that we leveraged data from existing, well-characterized prospective cohorts that collected comparable exposure and outcome data in distinctly different settings. Our findings suggest that chronic immune imbalance and activation may contribute to CLD in YLPHIV despite ART and CD4 reconstitution/preservation. As more children living with HIV are surviving into adolescence and adulthood, it is imperative to understand risk factors and mechanisms of lung function impairment among youth living with HIV to guide efforts to mitigate the global CLD burden.

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Abbreviations:

ART	antiretroviral therapy
CLD	chronic lung disease
FEV₁	forced expiratory volume in one second
%FEV₁	percent-predicted forced expiratory volume in one second
NHANES III	National Health and Nutrition Examination Survey III
PHACS	Pediatric HIV/AIDS Cohort Study
YLP HIV	youth living with perinatally-acquired human immunodeficiency virus
U.S.	United States of America

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Table I.

Characteristics of YLPHIV in U.S. vs Kenyan cohort *

	U.S. YLPHIV (n=188)	Kenya YLPHIV (n=49)	p-value
Age (years)	17 (14 – 19)	13 (11 – 15)	<0.001
Male	79 (42)	25 (51)	0.26
Black race/ethnicity	138 (73)	49 (100)	--
Tobacco Exposure			
Active tobacco use	16 (9)	0 (0)	0.01
Passive smoking	18 (10)	11 (22)	
No smoking	144 (81)	38 (78)	
Indoor combustible fuel use	--	40 (82)	--
Prior pulmonary infection			
Bacterial pneumonia	87 (46)	19 (39)	0.35
Tuberculosis	3 (2)	14 (29)	<0.001
Self-reported asthma (ever)	63 (34)	2 (4)	<0.001
HIV-related variables[^]			
CD4 T-cell count (cells/ μ L)	635 (458 – 783)	667 (453 – 843)	0.45
CD8 T-cell count (cells/ μ L)	722 (514 – 930)	1098 (849 – 1473)	<0.001
CD4/CD8 ratio	0.91 (0.55 – 1.25)	0.65 (0.42 – 0.94)	0.004
Nadir CD4 T-cell count (cells/ μ L)	335 (148 – 473)	269 (193 – 482)	0.85
HIV-1 RNA 400 copies/mL	127 (69)	--	--
Currently receiving ART	165 (89)	46 (94)	0.33
Duration of ART (years)	13 (10 – 14)	5 (3 – 8)	<0.001
Age at ART initiation (years)	3 (1 – 5)	8 (5 – 11)	<0.001
Percent-predicted FEV₁ (%FEV₁) results			
Pre-bronchodilator %FEV ₁	98 (85 – 109)	86 (76 – 100)	0.003
Post-bronchodilator %FEV ₁ [^]	100 (88 – 113)	93 (83 – 104)	0.006

* Results presented as median (interquartile range) or n (%).

[^] HIV-related variables available for 185 U.S. youth; post-bronchodilator %FEV₁ available for 183 U.S. youth; no missing data points for Kenyan youth.

ART: antiretroviral therapy, %FEV₁: percent-predicted forced expiratory volume in one second; YLPHIV: youth living with perinatally-acquired human immunodeficiency virus,