

In-Solution IgG Titer Determination in Fermentation Broth Using Affibodies and Flow-Induced Dispersion Analysis

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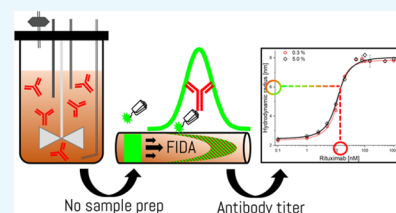


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Supporting Information

ABSTRACT: Biopharmaceuticals such as protein and peptide-based drugs are often produced by fermentation processes where it is necessary to monitor the amount and quality of the product expressed during fermentation and for release testing of the final drug product. Standard procedures involve surface-based ligand binding technologies such as enzyme-linked immunosorbent assay and biolayer interferometry, or extensive purification using, e.g., preparative chromatography followed by spectrophotometric protein quantification. The multistep nature of these methodologies leads to lengthy protocols and renders real-time process control impractical. Recently, flow-induced dispersion analysis (FIDA) was introduced as a novel in-solution ligand binding technology, requiring only nano/microliter sample volumes. FIDA is based on Taylor dispersion analysis in narrow fused silica capillaries and provides the hydrodynamic radius of the binding ligand and complex in addition to the detailed binding characterization. Here, we demonstrate the use of FIDA for quantification of monoclonal IgG antibodies (rituximab) directly in mammalian cell fermentation broth with only 4 min of analysis time. The FIDA assay utilizes a small anti-IgG affibody, conjugated to a fluorophore, as a selective rituximab binder. The apparent change in the hydrodynamic radius of the affibody, as it interacts with known concentrations of rituximab, is used for generating a binding curve in a blank fermentation medium, and hence determining the dissociation constant and complex size. Finally, the binding curve is utilized for quantifying the rituximab titer concentration in clarified fermentation broth samples.



INTRODUCTION

The vast majority of biopharmaceuticals are produced by fermentation processes via genetically modified cell lines based, primarily, on mammalian, bacterial, or yeast cells.^{1–3} These cells are exploited for their inherent protein synthesis apparatus, which is harnessed for assembling, folding, and expressing protein-based drugs. The cells require complex media for sustainable growth and satisfactory protein expression. Furthermore, the cells express host cell proteins (HCPs), as part of their metabolism, and may release cellular debris components (DNA, cell wall, and HCPs) into the growth medium upon apoptosis. Consequently, the growth medium, i.e., fermentation broth, is, at the end of a fermentation process, a complex solution. This presents a substantial challenge to most analytical assays; thus, purification steps are typically implemented prior to analysis,^{4–7} which impede fast and inexpensive selection of clones and further cell line optimization. Currently, surface-based methodologies such as biolayer interferometry (BLI) and enzyme-linked immunosorbent assay (ELISA) are the preferred methods for assessing IgG titers via immobilization of selective ligands (e.g., protein A).^{8,9} However, these methods rely on surface-based interactions and thus may not report the actual in-solution titer due to nonspecific surface adsorption.

Flow-induced dispersion analysis (FIDA) has emerged as a new immobilization-free ligand binding technology with abilities for in-solution characterization of protein size,

stability, and binding affinity.^{10–14} Briefly, FIDA utilizes Taylor dispersion analysis (TDA) in narrow capillaries under a laminar flow for measuring the change in the apparent hydrodynamic radius of a selective ligand (termed the indicator) as it interacts with the analyte of interest. The apparent hydrodynamic radius (i.e., size) of the indicator is measured at different analyte concentrations and thereby forms the basis for generation of a binding curve and determination of the dissociation constant (K_d) and complex size. Furthermore, the binding curve serves as a standard curve for selectively quantifying analyte concentrations in unknown samples based on the apparent indicator size measured.

Here, we present a FIDA-based approach for quantifying the rituximab titer directly in clarified Chinese hamster ovary (CHO) cell fermentation broth using an anti-IgG affibody as the indicator. The FIDA assay was developed in an uninoculated fermentation medium using known quantities of rituximab and subsequently utilized for quantifying the rituximab titer in CHO cell fermentation broth samples. The titers measured by the FIDA assay are in good agreement with

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traditional measurements performed using UV–vis absorption spectroscopy, following a protein A-based purification procedure.

■ EXPERIMENTAL SECTION

Equipment. The FIDA experiments were conducted on a FIDA 1 instrument employing light-emitting diode (LED)-induced fluorescence detection (FIDA Biosystems ApS, Copenhagen, Denmark) with an excitation wavelength of 480 nm and a high-pass emission filter (515 nm cutoff).

TDA experiments were performed on a PrinCE NEXT 870 instrument (PrinCE Technologies, Emmen, The Netherlands) employing an ActiPix D100 UV imaging system (Paraytec Ltd., York, United Kingdom) for detection at 214 nm.

PEG-coated fused silica capillaries (inner diameter: 75 μm , outer diameter: 375 μm , length total: 100 cm, length to detection window: 84 cm) from FIDA Biosystems were used.

Materials and Chemicals. Rituximab (lot H0156B04, 144,544 kDa) from Roche (MabThera, Basel, Switzerland), clarified (without cells) CHEF1 CHO cell fermentation broths from two parallel runs (TF262, PM01, and PM02 respectively), and uninoculated fermentation media were obtained from AGC Biologics (Soeborg, Denmark). The anti-IgG affibody (cat. no. ab31900) was purchased from Abcam (Cambridge, UK). Alexa Fluor 488 C₅-maleimide was purchased from Thermo Fisher (Waltham, Massachusetts, USA). Dithiothreitol (DTT) and glycine were purchased from Sigma Aldrich (St Louis, Missouri, USA). Sodium dihydrogen phosphate monohydrate and disodium hydrogen phosphate dodecahydrate were purchased from Merck (Darmstadt, Germany). Dimethyl sulfoxide (DMSO) was obtained from EMP Biotech (Berlin, Germany). Tris ultrapure was obtained from ICN Biomedical (Aurora, Ohio, USA). Ultrapure water (18.2 M Ω -cm at 25 °C) was tapped from a water purification system (SG Ultraclear, SG Water, Barsbüttel, Germany).

Assay Buffer. A 67 mM phosphate buffer, pH 7.4, was prepared with ultrapure water, filtered through a Q-max 0.45 μm nylon syringe filter (Frisenette, Knebel, Denmark), and used as the assay buffer.

Fluorophore Labelling of the Anti-IgG Affibody. The anti-IgG affibody (200 μg at 1 mg/mL) was reduced in 0.02 M DTT for 2 h at room temperature. Subsequently, the DTT was removed by buffer exchange into the assay buffer utilizing a pre-equilibrated PD SpinTrap G-25 column (GE Healthcare, Marlborough, Massachusetts, USA). Alexa Fluor 488 C₅-maleimide was dissolved in DMSO to a stock concentration of 10 mM and then added to the affibody solution with a molar ratio of 19:1 (Alexa Fluor/affibody) immediately after the buffer exchange. The labelling reaction was incubated for 2 h at room temperature and subsequently purified in the assay buffer utilizing a pre-equilibrated SpinTrap G-25 column.

The concentration of the affibody–alexa488 conjugate was determined using a microvolume spectrophotometer (NanoDrop 2000C, Thermo Fisher Scientific, Wilmington, Delaware, USA), using the molar absorption coefficient of alexa488 ($\epsilon = 72,000 \text{ cm}^{-1} \text{ M}^{-1}$) at 493 nm¹⁵ since the anti-IgG affibody does not contain any tyrosine or tryptophan residues.

Protein A Purification Buffers. The equilibration and binding buffers were a 20 mM phosphate buffer, pH 7.0; the elution buffer was 0.1 M glycine–HCl, pH 2.7; and the neutralizing buffer was 1 M Tris–HCl, pH 9.0. All buffers were prepared with ultrapure water.

FIDA and TDA Procedures. The PEG-coated capillary was flushed and equilibrated prior to sample analysis with the 67 mM phosphate buffer, pH 7.4, at 1500 mbar for 5 min. The analyte sample was injected at 1500 mbar for 45 s followed by injection of the indicator sample at 50 mbar for 10 s (39 nL, corresponding to 1% of the capillary volume). Subsequently, the injected indicator sample was mobilized toward the LED detector with the analyte sample at 400 mbar for 240 s. The capillary, samples, and buffer vials were kept in a temperature-controlled environment at 25 °C inside the instrument. All samples were analyzed in triplicate.

Development of the FIDA Assay. The hydrodynamic radius of the unconjugated anti-IgG affibody was measured using TDA at a concentration of 1 mg/mL in a neat phosphate buffer.

Subsequently, the indicator, anti-IgG affibody–alexa488, was diluted to a fixed indicator concentration of 20 nM in varying concentrations (0–10% v/v) of CHO cell fermentation broth (TF262, PM02). The fermentation broth was diluted with the assay buffer. All samples were preincubated for >10 min to attain equilibrium prior to the FIDA measurements.

Standard Curves for Rituximab Quantification in the Fermentation Broth. Standard curves were established in 0.3 and 5.0% v/v uninoculated fermentation media. The indicator, anti-IgG affibody–alexa488, was added to provide a fixed indicator concentration of 20 nM in the medium with varying and known concentrations of rituximab (0–2000 nM). All samples were preincubated for >10 min to attain equilibrium prior to the FIDA measurements.

Determination of the Rituximab Titer by FIDA. Quantification of rituximab in two parallel CHO cell fermentation broths (TF262 PM01 and TF262 PM02) was performed for three different concentrations of fermentation broth (0.05, 0.075, and 0.10% v/v). The indicator, anti-IgG affibody–alexa488, was added to provide a fixed indicator concentration of 20 nM, and an uninoculated fermentation medium was added to reach a final medium concentration of 5% v/v (i.e., 4.95, 4.925, and 4.90% v/v, respectively). All samples were preincubated for >10 min to attain equilibrium prior to the FIDA measurements.

Determination of the Rituximab Titer by Protein A Purification and UV Spectrophotometry. The rituximab content was also determined using disposable protein A HP SpinTrap columns from GE Healthcare, following the supplier instructions (#28-9031-32). Three parallel purification runs were performed for each of the two fermentations (i.e., TF262 PM01 and TF262 PM02), and every column was loaded with 300 μL of fermentation broth. After purification, the concentration of rituximab was determined utilizing a micro-volume spectrophotometer (NanoDrop 2000C) and the extinction coefficient of rituximab ($\epsilon = 1.7 \text{ cm}^{-1} (\text{mg/mL})^{-1}$) at 280 nm.¹⁶

Data Analysis. The Taylorgrams were processed using the FIDA software, version 1.03 (FIDA Biosystems ApS, Copenhagen, Denmark), for calculating the apparent hydrodynamic radius of the indicator (anti-IgG affibody–alexa-488) at each analyte concentration. The Taylorgram fraction setting was consistently fixed to 75% for all data points, and minor fluctuations in sample viscosity were corrected according to a reference measurement of the indicator in a neat phosphate buffer, as previously described.¹⁰

Typically, the following binding isotherm is used for FIDA assays, assuming an indicator concentration below the

dissociation constant, 1:1 binding stoichiometry, and equilibrium¹⁰

$$R_{\text{app}} = \frac{1 + \frac{1}{K_d}[A]}{\left((R_I)^{-1} - (R_{IA})^{-1}\right) + \left(1 + \frac{1}{K_d}[A]\right)(R_{IA})^{-1}} \quad (1)$$

where R_{app} , R_I , R_{IA} are the apparent, indicator, and complex hydrodynamic radii, respectively; K_d is the dissociation constant; and $[A]$ is the formal analyte concentration.

However, in the cases where the binding affinity between the indicator and analyte is very high (low nM – pM range), the indicator concentration cannot be assumed to be less than K_d . Therefore, the excess indicator binding isotherm applies

$$R_{\text{app}} = \left[\begin{aligned} & (R_{IA})^{-1} \cdot \left(\frac{(C_I + C_A + K_d) - \sqrt{(C_I + C_A + K_d)^2 - 4C_A \cdot C_I}}{2C_I} \right) \\ & + (R_I)^{-1} \cdot \left(\frac{(C_I - C_A - K_d) + \sqrt{(C_I + C_A + K_d)^2 - 4C_A \cdot C_I}}{2C_I} \right) \end{aligned} \right]^{-1} \quad (2)$$

where C_I and C_A are the formal concentrations of the indicator and analyte, respectively. The details of the excess indicator binding isotherm are described in the Supporting Information.

RESULTS AND DISCUSSION

Development of the FIDA Assay. Protein quantification using FIDA is based on an indicator molecule (anti-IgG affibody) interacting selectively with the analyte protein (rituximab, IgG) in the sample matrix. An apparent change in the indicator size due to the interaction with the analyte present is used for the quantification. An anti-IgG affibody was selected as the indicator due to an expected selectivity towards the Fc region of rituximab and, hence, not interacting with other components of the sample matrix. The affibody was fluorescently labelled at the dedicated C-terminal cysteine for single-point modification,¹⁷ ensuring selective and high sensitivity measurements in the complex samples. Furthermore, the affibody was considered a promising indicator due to the small size (7 kDa) relative to rituximab (145 kDa), thus potentially providing high sensitivity and a wide dynamic range. Other anti-IgG specific ligands, e.g., protein A (similar specificity as the affibody used in the present work), protein G, aptamers, or nanobodies,¹⁸ might be suitable for FIDA-based rituximab quantification as well. The specificity of the methods is directly linked to the specificity of the interaction between the affibody and the constant region of the antibody. As the affibody recognizes the constant region of the antibody, the same methodology is applicable to the detection of any human IgG antibody.

The hydrodynamic radii of the unconjugated dimeric affibody (14 kDa) and monomeric affibody–alexa488 conjugate (7 kDa) were determined in a neat phosphate buffer by TDA and FIDA to 3.27 ± 0.05 and 2.24 ± 0.10 nm, respectively. These values were slightly higher than expected for globular proteins,¹⁹ thereby suggesting a relatively loose conformation.

Subsequently, a feasibility study was performed to assess whether the anti-IgG affibody could be measured directly in a clarified fermentation broth containing rituximab (TF262 PM02). The apparent hydrodynamic radius of the anti-IgG affibody was measured in increasing fermentation broth concentrations using FIDA (Figure 1). The apparent hydrodynamic radius of the anti-IgG affibody was found to increase from 2.2 to a plateau at ~ 8 nm (Figure 1), thereby indicating affibody binding to rituximab.

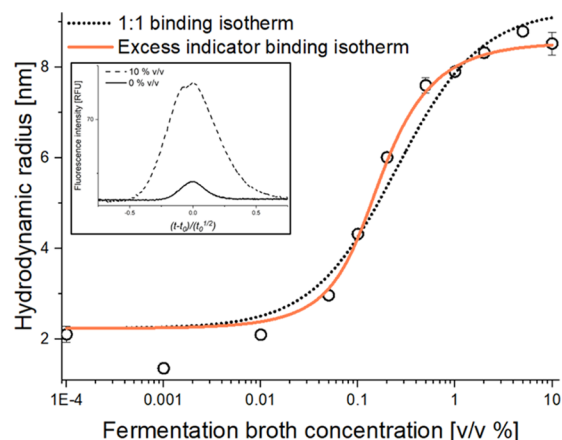


Figure 1. Apparent hydrodynamic radius of the 20 nM anti-IgG affibody–alexa488 as a function of fermentation broth concentration (PM02 (containing rituximab), 0–10% v/v) determined by FIDA at 25 °C (black open circles, $n = 3$, error bars represent standard deviation). The dotted black line and solid orange line represents fitting to 1:1 ($R^2 = 0.980$) and excess indicator ($R^2 = 0.990$) binding isotherms, respectively. (Insert) visual comparison of viscosity-corrected Taylorgrams at 0 and 10% v/v fermentation broth (solid and dashed line, respectively).

It was attempted to fit the data in Figure 1 to the binding isotherm assuming a 1:1 binding stoichiometry (eq 1). However, the binding curve appeared steeper than anticipated, which could point to the affibody concentration being higher than the K_d at low rituximab concentrations. Therefore, the data was also fitted to an excess indicator binding isotherm (eq 2), resulting in a significantly better fit (Figure 1, orange line). This was further investigated by a second set of experiments in which the affibody concentration was reduced from 20 to 5 nM. Here, a shift in the binding isotherm was observed (Figure S1 in the Supporting Information), indicating that the affibody was indeed in excess at low rituximab concentrations. This scenario has previously been described theoretically for FIDA experiments.¹³

The Taylorgram overlay at the 0 and 10% v/v fermentation broth in Figure 1 (insert, corrected for viscosity-related differences²⁰) revealed a noteworthy change in the peak area, thereby indicating adsorption of the indicator (i.e., affibody) to labware surfaces (e.g., vials and capillary) at low fermentation broth concentrations. Here, the fermentation broth was simply added to prevent indicator (and potentially rituximab) adsorption to the labware surfaces. Taylorgrams of indicator peak areas depicted as a function of fermentation broth concentration (Figure S2) showed increased peak areas and minimized adsorption at broth concentrations above 0.05% v/v. This initial set of experiments demonstrated that the FIDA assay was capable of characterizing interactions directly in a

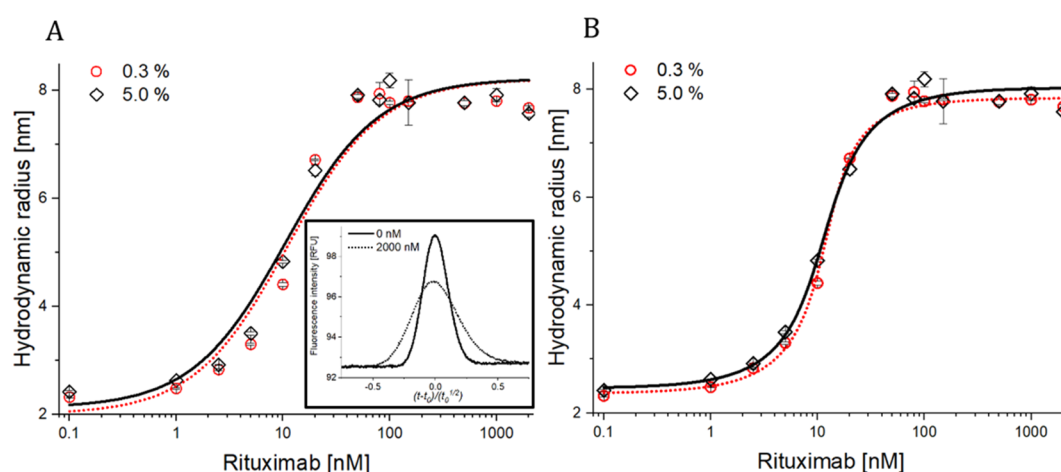


Figure 2. Apparent hydrodynamic radius of the 20 nM anti-IgG affibody–alexa488 as a function of rituximab concentration (0–2000 nM) determined by FIDA at 25 °C. The black open diamonds and red open circles represent measurements in 0.3 and 5.0% v/v uninoculated fermentation media, respectively ($n = 3$, error bars represent standard deviation). The solid black line and dotted red line represent fitting to (A) 1:1 and (B) excess indicator binding isotherms, respectively. Insert: visual comparison of viscosity-corrected Taylorgrams in a 5.0% v/v uninoculated fermentation medium at 0 and 2000 nM rituximab (solid and dashed line, respectively).

Table 1. Fitted Binding Parameters (Standard Curves) in 0.3 and 5% Uninoculated Fermentation Medium, Representing Both 1:1 Binding Stoichiometry and Excess Indicator Models

medium concentration	0.3% v/v	5% v/v	0.3% v/v	5% v/v
binding isotherm model	1:1 (eq 1)	1:1 (eq 1)	excess (eq 2)	excess (eq 2)
indicator size (R_I) (nm)	2.00	2.12	2.37	2.46
indicator-complex size (R_{IA}) (nm)	8.22	8.23	7.84	8.03
dissociation constant, K_d (nM)	2.66	2.70	0.66	1.12
coefficient of determination (R^2)	0.965	0.970	0.999	0.995

Table 2. Quantification of Rituximab in CHO Cell Fermentation Broth Samples TF262 PM01 and TF262 PM02^a

fermentation broth concentration [% v/v]	TF262 PM01			TF262 PM02		
	0.05	0.075	0.10	0.05	0.075	0.10
rituximab concentration by FIDA [mg/mL]	1.34 ± 0.14	1.28 ± 0.07	1.38 ± 0.06	1.46 ± 0.02	1.34 ± 0.06	1.28 ± 0.04
rituximab concentration by UV spectrophotometry following protein A purification	1.22 ± 0.08			1.41 ± 0.04		

^aThe rituximab concentration was found to be within the expected titer for a typical CHO cell fermentation process.¹

clarified fermentation broth. We did not observe any nonspecific binding to matrix components.

Standard Curves for Rituximab in Fermentation Media. Rituximab standard curves (0–2000 nM) were established in 0.3 and 5% v/v uninoculated (i.e., blank) fermentation media in order to define optimal conditions for quantification. The apparent hydrodynamic radius of the anti-IgG affibody increased with an increasing rituximab concentration, as expected (Figure 2), and a plateau was reached around 8 nm (Figure 2), indicating full binding, in line with the initial experiments (Figure 1).

As expected from the assay development experiment (Figure 1), the excess indicator isotherm (eq 2) was most suitable for the binding curves (Figure 2). This was further verified by the binding parameters summarized in Table 1. Here, the R^2 -values for the excess ligand binding isotherm were superior to the 1:1 in terms of describing the data. The affinity was notably high, and using the 1:1 binding model (eq 1), the apparent dissociation constants (K_d) were in the low nM range, indicating that the indicator concentration is not lower than the K_d . While eq 1 provides an apparent K_d for the interaction,

it would, however, require lower indicator concentrations to quantify the true K_d using this model. In this case, it is therefore more accurate to apply the excess indicator model (eq 2). For quantification, the binding curve is used as a standard curve linking the apparent size to antibody concentration. When the limit of quantification (LOQ) is defined as 10 times the relative standard deviation of the blank sample, an LOQ of 0.9 nM or 135 ng/mL is determined, which is about two orders of magnitude lower than HPLC procedures.²¹ In the FIDA methodology, there is a direct link between the apparent size, fraction unbound, and ultimately concentration. There is no requirement for linearity for connecting the apparent size, fraction bound, and concentration.

The standard curves in 0.3 and 5% fermentation media were highly similar (Figure 2). However, it was decided to perform the rituximab quantification in the 5% v/v fermentation medium since this concentration was superior in terms of eliminating nonspecific surface adsorption (Figure S2). The standard curve covers rituximab quantification in the 5% v/v fermentation medium, thus the samples (PM01 and PM02)

were diluted with the uninoculated fermentation medium in order to be within the dynamic range of the binding curve since the interaction was fully saturated at 5% broth as seen in Figure 1. The excess indicator fitting model was chosen since it fitted the data with the highest accuracy (Table 1).

Quantification of Rituximab in CHO Cell Fermentation Broth with the Anti-IgG Affibody Using FIDA. The samples (PM01 and PM02) were diluted to fermentation broth concentrations of 0.05, 0.075, and 0.10% v/v with a blank fermentation medium and assay buffer to a final medium concentration of 5% v/v. This approach was applied in order to be within the dynamic range of the binding curve (Figure 2) and eliminate nonspecific adsorption. The selected dilutions were also chosen for verifying the concentration measurements in the dynamic range of the standard curve. The apparent hydrodynamic radius of the 20 nM anti-IgG affibody–alexa488 was measured in each dilution and related to an apparent rituximab concentration using the standard curve (5% v/v, Figure 2), which led to the results in Table 2 upon correction for dilution.

Quantification of Rituximab in CHO Cell Fermentation Broth by Protein A Purification and UV Spectrophotometry. The content of rituximab in the fermentation broth samples (PM01 and PM02) was quantified using a conventional method involving protein A-based solid phase purification followed by quantification using UV absorbance at 280 nm.^{21,22} Although we employed spin columns (see the Experimental Section), HPLC methodologies are also applicable.²¹ Whereas an HPLC-based procedure often requires sample preparation, a recent study reported a fast (less than 5 min) HPLC-based procedure but with a detection limit two orders of magnitude higher than the methodology reported in the present work.²¹ The results are depicted in Table 2. The rituximab titers determined by this approach are in good agreement with the FIDA measurements. This demonstrates that the developed FIDA method is compatible with specific measurements in a clarified fermentation broth. Furthermore, FIDA can measure selectively in a clarified fermentation broth, thus eliminating any requirement for purification prior to sample analysis, which, in combination with a fully automated instrument, allows for a considerably high throughput of samples with few procedural steps. The simple FIDA methodology enables assessment of the in-solution IgG titer in less than 4 min, thereby allowing nearly real-time measurements of IgG titer during fermentation processes.

CONCLUSIONS

The present work shows that FIDA is a viable technology for direct (no sample pretreatment) quantification of the antibody titer in a CHO cell fermentation broth (nanomolar sensitivity) and for characterization of in-solution binding to monoclonal antibodies. The developed FIDA assay utilized a selective anti-IgG affibody as indicator and thereby accurately determining the rituximab concentration without any preceding purification process. This demonstrates that FIDA is characterized by being highly tolerant to the sample matrix, hence, little or no sample pretreatment is required. Furthermore, the developed method characterized the affibody–rituximab interaction with regards to determining the dissociation constant (K_d), complex size (R_n), and binding stoichiometry.

This protocol may be extended to include a full functional assessment of the antibody by assessment of specific target

binding. We thus envision that the FIDA methodology will find applications in development and production of therapeutic proteins.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acsomega.0c00791>.

Additional experimental details (PDF)

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Author Contributions

All authors have given approval to the final version of the manuscript.

Notes

The authors declare the following competing financial interest(s): HJ, JO, and MEP have commercial interests in FIDA Biosystems ApS. MEP and HJ are employees of FIDA Biosystems ApS.

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