



# A Comprehensive Study on Thiadiazole-Based Anticancer Agents Inducing Cell Cycle Arrest and Apoptosis/Necrosis Through Suppression of Akt Activity in Lung Adenocarcinoma and Glioma Cells

Akciğer Adenokarsinom ve Glioma Hücrelerinde Akt Aktivitesinin Bastırılması Yoluyla Hücre Döngüsü Arrestini ve Apoptozu İndükleyen Tiyadiazol Türevi Antikanser Ajanlar Üzerine Kapsamlı Bir Çalışma

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## ABSTRACT

**Objectives:** Akt is considered as an attractive target for anticancer drug discovery and development and therefore extensive efforts have been devoted to the discovery of new potent anticancer agents targeting Akt.

**Materials and Methods:** Due to the importance of thiadiazoles for anticancer drug discovery, herein eight 1,3,4-thiadiazole derivatives were investigated for their cytotoxic effects on C6 rat glioma and A549 human lung adenocarcinoma cell lines using the MTT assay. The effects of the most promising anticancer agents on apoptosis, caspase-3 activation, mitochondrial membrane potential, and cell cycle arrest were determined on a BD FACSAria (I) flow cytometer. Akt activity was measured in the C6 and A549 cell lines using an ELISA colorimetric method. Schrödinger's Maestro molecular modeling package was used to explore the possible binding modes of compounds **3** and **8** in the active site of Akt enzyme (PDB code: 3OW4).

**Results:** *N*-(4-Chlorophenyl)-2-[(5-((4-nitrophenyl)amino)-1,3,4-thiadiazol-2-yl)thio]acetamide (**3**) and *N*-(6-nitrobenzothiazol-2-yl)-2-[(5-((4-nitrophenyl)amino)-1,3,4-thiadiazol-2-yl)thio]acetamide (**8**) induced apoptosis and cell cycle arrest in the C6 cell line through inhibition of Akt activity (92.36% and 86.52%, respectively). The docking results of compounds **3** and **8** indicated that  $\pi$ - $\pi$  interactions, H bonds, and salt-bridge formation were responsible for the observed Akt inhibitory activity.

**Conclusion:** According to *in vitro* and docking studies, compounds **3** and **8** stand out as promising anti-glioma agents.

**Key words:** Apoptosis, Akt activity, cancer, molecular docking, thiadiazole

## ÖZ

**Amaç:** Akt, antikanser ilaç keşfinde ve gelişiminde önemli bir hedef olarak düşünülmektedir ve bu nedenle Akt enzimini hedefleyen yeni potent antikanser ajanların keşfi için yoğun çaba harcanmıştır.

**Gereç ve Yöntemler:** Tiyadiazollerin antikanser ilaç keşfindeki önemine bağlı olarak, bu çalışmada sekiz adet 1,3,4-tiyadiazol türevinin C6 sıçan glioma ve A549 insan akciğer adenokarsinoma hücre dizileri üzerindeki sitotoksik etkileri MTT deneyi kullanılarak araştırılmıştır. En etkili antikanser ajanların apoptoz, kaspaz-3 aktivasyonu, mitokondriyal membran potansiyeli, hücre döngüsü arresti üzerindeki etkileri BD FACSAria (I) akış sitometrisi ile belirlenmiştir. Akt aktivite, C6 ve A549 hücre dizilerinde ELISA kolorimetrik yöntem kullanılarak ölçülmüştür. **3** ve **8** no'lu bileşiklerin Akt enziminin (PDB kod: 3OW4) aktif bölgesindeki olası bağlanma biçimlerini araştırmak için Schrödinger's Maestro moleküler modelleme programı kullanılmıştır.

**Bulgular:** *N*-(4-Klorofenil)-2-[(5-((4-nitrofenil)amino)-1,3,4-tiyadiazol-2-il)tiyo]asetamit (**3**) ve *N*-(6-nitrobenzotiyazol-2-il)-2-[(5-((4-nitrofenil)amino)-1,3,4-tiyadiazol-2-il)tiyo]asetamit (**8**), C6 hücre dizisinde apoptozu ve hücre döngüsü tutuklanmasını Akt aktivitesi inhibisyonu aracılığıyla

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(sırasıyla %92.36 ve %86.52) indüklemişlerdir. Bu bileşiklerin docking sonuçları, gözlemlenen Akt inhibitör aktiviteden  $\pi$ - $\pi$  etkileşimleri, hidrojen bağları ve tuz köprüsü oluşumunun sorumlu olduğunu belirtmektedir.

**Sonuç:** *In vitro* ve docking çalışmalarına göre, **3** ve **8** no'lu bileşikler umut vadeden anti glioma ajanlar olarak dikkat çekmektedirler.

**Anahtar kelimeler:** Apoptoz, Akt aktivite, kanser, moleküler docking, tiyadiazol

## INTRODUCTION

Targeted cancer therapies, which are used to inhibit tumor growth, progression, and metastasis by interfering with specific molecular targets, have emerged as a promising therapeutic approach for the management of cancer.<sup>1</sup>

Akt, also known as protein kinase B, is overexpressed or activated in a variety of human cancers, including gliomas and lung, breast, ovarian, gastric, and pancreatic cancers.<sup>1,2</sup> Inhibition of Akt signaling results in induction of apoptosis and inhibition of tumor growth and therefore Akt has attracted a great deal of attention as a promising target for anticancer drug discovery and development.<sup>1-6</sup>

Thiadiazole has been studied extensively for more than one hundred years due to its outstanding therapeutic applications. The sulfur atom of the thiadiazole ring imparts improved liposolubility and the mesoionic nature of thiadiazoles also allows these compounds to cross cellular membranes and interact with biological targets with distinct affinities. 1,3,4-Thiadiazoles display a wide spectrum of biological activities including anticancer, antimicrobial, antiviral, antiepileptic, antidiabetic, analgesic, and anti-inflammatory activities.<sup>7-14</sup> In particular, recent studies have pointed out the significance of the 1,3,4-thiadiazole scaffold in the field of current cancer research. Thiadiazole-based anticancer agents exert potent antitumor activity against a variety of human cancer cell lines through the inhibition of diverse molecular targets including histone deacetylase, Abl tyrosine kinase, focal adhesion kinase, Akt, and tubulin polymerization.<sup>7-22</sup>

Prompted by the aforementioned findings, herein we focused on the *in vitro* antiproliferative effects of a series of 1,3,4-thiadiazoles on A549 human lung adenocarcinoma and C6 rat glioma cell lines. Further *in vitro* and *in silico* studies were also carried out to determine the mechanism of antitumor action of the most potent anticancer agents in this series.

## MATERIALS AND METHODS

### Chemistry

5-(4-Nitrophenyl)amino-1,3,4-thiadiazole-2(3*H*)-thione was synthesized via the ring closure reaction of 4-(4-nitrophenyl) thiosemicarbazide with carbon disulfide in the presence of potassium hydroxide. Finally, the reaction of 5-(4-nitrophenyl) amino-1,3,4-thiadiazole-2(3*H*)-thione with *N*-(alkyl/aryl)-2-chloroacetamide/4-(chloroacetyl)morpholine in the presence of potassium carbonate afforded compounds **1-8**. The synthetic procedure and the spectral data of compounds **1-8** were reported previously by our research group.<sup>23</sup> The chemical structures of the test compounds are given in Table 1.

### Biochemistry

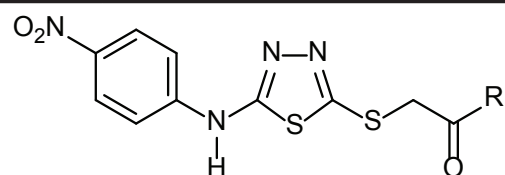
#### Cell culture and drug treatment

C6 rat glioma and NIH/3T3 mouse embryonic fibroblast cells were incubated in Dulbecco's Modified Eagle's Medium (Sigma, Deisenhofen, Germany) supplemented with 10% fetal calf serum (Gibco, Paisley, Scotland). A549 human lung adenocarcinoma cells were incubated in 90% RPMI supplemented with 10% fetal bovine serum (Gibco, Paisley, Scotland). All media were supplemented with 100 IU/mL penicillin-streptomycin (Gibco, Paisley, Scotland) and the cells were incubated at 37°C in a humidified atmosphere of 95% air and 5% CO<sub>2</sub>. Exponentially growing cells were plated at 2×10<sup>4</sup> cells/mL into 96-well microtiter tissue culture plates (Nunc, Denmark) and incubated for 24 h before the addition of the drugs (the optimum cell number for cytotoxicity assays was determined in preliminary experiments). The stock solutions of the compounds were prepared in dimethyl sulfoxide (DMSO; Sigma Aldrich, Poole, UK) and further dilutions were made with fresh culture medium (the concentration of DMSO in the final culture medium was <0.1%, which had no effect on cell viability).

#### MTT assay

The level of cellular 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) (Sigma) reduction was quantified as previously described in the literature<sup>24,25</sup> with small modifications.<sup>26</sup> Compounds **1-8** were investigated for their anticancer activity against A549 human lung adenocarcinoma and C6 rat glioma cell lines. NIH/3T3 mouse embryonic fibroblast cells were used to evaluate the selectivity of the compounds.

**Table 1.** The chemical structures of compounds 1-8



Compound	R
1	Diethylamino
2	(3-Chlorophenyl)amino
3	(4-Chlorophenyl)amino
4	(4-Nitrophenyl)amino
5	(1,3-Benzodioxol-5-ylmethyl)amino
6	Morpholin-4-yl
7	(Benzothiazol-2-yl)amino
8	(6-Nitrobenzothiazol-2-yl)amino

After 24 h of preincubation, compounds **1-8** and cisplatin (positive control) were added to give a final concentration in the range 3.9-500  $\mu\text{g}/\text{mL}$  and the cells were incubated for 24 h. This concentration range was chosen based on our previous studies.<sup>26</sup> At the end of this period, MTT was added to a final concentration of 0.5 mg/mL and the cells were incubated for 4 h at 37°C. After the medium was removed, the formazan crystals formed by MTT metabolism were solubilized by addition of 200  $\mu\text{L}$  of DMSO to each well and absorbance was read at 540 nm with a microtiter plate spectrophotometer (Bio-Tek plate reader, Winooski, VT, USA). Every concentration was repeated in three wells. The half maximal inhibitory concentration ( $\text{IC}_{50}$ ) values were defined as the drug concentrations that reduced absorbance to 50% of control values.

#### *Flow cytometric analyses of apoptosis*

After the cells were incubated with compounds **1, 2, 3, 4, 5, 8**, and cisplatin at  $\text{IC}_{50}$  concentrations, phosphatidylserine externalization, which indicates early apoptosis, was measured by Annexin V-propidium iodide (PI) (BD Pharmingen, San Jose, CA, USA) on a BD FACSAria flow cytometer for 24 h. The Annexin V staining protocol was applied according to the manufacturer's instructions (BD Pharmingen, San Jose, CA, USA). The cells were then briefly washed with cold phosphate buffer saline (PBS) and suspended in a binding buffer at a concentration of  $1 \times 10^6$  cells/mL. Then 100  $\mu\text{L}$  of this solution containing  $1 \times 10^5$  cells was transferred to a 5 mL test tube. After addition of 5  $\mu\text{L}$  of Annexin-V and PI, the cells were incubated for 15 min at room temperature (RT) in the dark. Then 400  $\mu\text{L}$  of 1X binding buffer was added to each tube and the cells were processed for data acquisition, and analyzed on a Becton Dickinson FACSAria flow cytometer using BD FACSDiva software version 6.1.1 (BD Biosciences, San Jose, CA, USA).<sup>26</sup>

#### *Flow cytometric analyses of caspase-3*

After C6 cells were incubated with compounds **1, 2, 3, 4, 5, 8**, and cisplatin at  $\text{IC}_{50}$  concentrations for 24 h, the caspase-3 activity measurement protocol was applied according to the manufacturer's instructions (BD Pharmingen, San Jose, CA, USA). In brief, the cells were washed with cold PBS 1X cells and incubated with 0.5 mL of perm lyse solution for 30 min at RT in the dark. The pellets were washed twice with 0.5 mL of perm wash buffer. The cells were resuspended in 100  $\mu\text{L}$  of perm wash buffer and 10  $\mu\text{L}$  of caspase-3 antibody was added over 20 min at RT in the dark. At least 10,000 cells were counted for each sample and the cells were analyzed by BD FACSAria flow cytometry using BD FACSDiva software version 6.1.1 (BD Biosciences, San Jose, CA, USA).

#### *Analysis of mitochondrial membrane potential (MMP) by flow cytometry*

The cells were seeded in six-well plates at a density of  $10^5$  cells/mL, and the  $\text{IC}_{50}$  doses of compounds **1, 2, 3, 4, 5, 8**, and cisplatin were added to cells. The cells were incubated in 5%  $\text{CO}_2$  air-conditioned atmosphere at 37°C. After 48 h of incubation, the cells were trypsinized, washed with PBS, and centrifuged at  $400 \times g$  for 5 min. 5,5',6,6'-Tetrachloro-1,1',3,3'-tetraethylbenz

imidazolylcarbocyanine iodide (JC-1) dye solution (1X assay buffer + JC-1 stock solution) was added to the cells. The stock solution was prepared by dissolving DMSO. Then the samples were incubated at 37°C for 10-15 min. After incubation, the cells were washed twice with an assay buffer and analyzed by BD FACSAria flow cytometry using BD FACSDiva software version 6.1.1 (BD Biosciences, San Jose, CA, USA). The cells showing mitochondrial membrane potential disruption were determined as a percentage of all cells.<sup>26</sup>

#### *Cell cycle analysis*

After C6 and A549 cells were incubated with  $\text{IC}_{50}$  concentrations of the compounds for 24 h, the cell cycle analysis measurement protocol was applied according to the manufacturer's instructions (BD Biosciences, San Jose, CA, USA). The cells were briefly suspended in citrate buffer. The cells were then centrifuged at  $400 \times g$  for 5 min at RT. The supernatant was decanted and 250  $\mu\text{L}$  of solution A was added to the pellet and kept at RT for 10 min. Then 200  $\mu\text{L}$  of solution B was added and the resulting mixture was gently mixed and kept at RT for 10 min. Then 200  $\mu\text{L}$  of solution C was added. After being gently mixed, it was kept in the dark at 4°C for 10 min and then analyzed on a BD FACSAria flow cytometer using BD Bioscience's ModFit software.<sup>27</sup>

#### *Inhibition of Akt enzyme*

After 10,000 cells/well were incubated with compounds **1, 2, 3, 4, 5, 8**, and cisplatin at  $\text{IC}_{50}$  concentrations for 24 h, the in-cell ELISA colorimetric Akt activity protocol was applied according to the manufacturer's instructions (Thermo Fisher Scientific, Rockford, IL, USA). Briefly, the medium was removed and 100  $\mu\text{L}$  of 4% formaldehyde was added to each well. The plate was incubated in a fume hood at RT for 15 min. Formaldehyde was removed and the plate was washed twice with 100  $\mu\text{L}$ /well of 1X TBS. The 1X TBS was removed and 100  $\mu\text{L}$ /well of 1X permeabilization buffer was added, followed by incubation for 15 min at RT. Permeabilization buffer was removed and the plate was washed once with 100  $\mu\text{L}$ /well of 1X TBS. The 1X TBS was removed and 100  $\mu\text{L}$ /well quenching solution was added, followed by incubation at RT for 20 min. The quenching solution was removed and the plate was washed once with 100  $\mu\text{L}$ /well of 1X TBS. The 1X TBS was removed and 100  $\mu\text{L}$ /well of blocking buffer was added, followed by incubation at RT for 30 min. After the blocking buffer was removed, 50  $\mu\text{L}$ /well of primary antibody was added. A plate sealer was applied and incubation was conducted overnight at 4°C. The primary antibody solution was removed and the plate was washed three times with 100  $\mu\text{L}$ /well of 1X wash buffer. After the wash buffer was removed, 100  $\mu\text{L}$ /well of diluted horseradish peroxidase conjugate was added, followed by incubation for 30 min at RT. The wash buffer was removed and 100  $\mu\text{L}$ /well of 3,3',5,5'-tetramethylbenzidine substrate was added. Then the plate was incubated at RT, protected from light. Next, 100  $\mu\text{L}$ /well of 3,3',5,5'-tetramethylbenzidine stop solution was added and the absorbance was measured at 450 nm within 30 min of stopping the reaction. The experiment was performed in triplicate wells. The values of blank wells were subtracted from

each well of treated and control cells. Percent Akt activity was defined as the relative absorbance of treated versus untreated control cells.

#### Statistical analysis

Statistical Package for the Social Sciences (SPSS) (Chicago, IL, USA) for Windows 15.0 was used for statistical analysis. The data were expressed as mean  $\pm$  standard deviation. Comparisons were performed by one-way analysis of variance for normally distributed continuous variables and post-hoc analyses of group differences were expressed by the Tukey test. Probability values less than 0.05 ( $p < 0.05$ ) were regarded as significant.

#### Molecular docking studies

Compounds **3** and **8** were docked to the active site of Akt enzyme. Ligands were set to the physiological pH (pH= 7.4) at the protonation step and the crystal structure of Akt enzyme was retrieved from the Protein Data Bank server (PDB code: 3OW4). The structures of compounds **3** and **8** were submitted to the protein preparation module of Schrödinger's Maestro molecular modeling package (Schrödinger Release 2016-2: Schrödinger, LLC, New York, NY, USA). In molecular docking simulations, Glide/XP docking protocols were applied for prediction of the topologies of compounds **3** and **8** in the active site of the target structure.<sup>28</sup>

## RESULTS AND DISCUSSION

The MTT assay was carried out to determine the anticancer effects of the compounds on A549 human lung adenocarcinoma and C6 rat glioma cell lines (Table 2).

Compounds **3** and **4** were more effective on the C6 cell line than cisplatin ( $IC_{50} = 24.33 \pm 0.58 \mu\text{g/mL}$ ). Compounds **3** and **4** showed antiproliferative effects on the C6 cell line with  $IC_{50}$  values of  $22.00 \pm 3.00 \mu\text{g/mL}$  and  $18.50 \pm 4.95 \mu\text{g/mL}$ , respectively. This outcome clearly indicated that *p*-chloro and *p*-nitro substituents significantly enhanced anticancer activity against the C6 cell line. Compounds **1**, **2**, **5**, and **8** exhibited notable cytotoxic activity against the C6 cell line with  $IC_{50}$  values of  $50.66 \pm 12.50$ ,

$42.33 \pm 2.52$ ,  $46.67 \pm 2.89$ , and  $42.67 \pm 2.08 \mu\text{g/mL}$ , respectively. These results showed the importance of the alkyl and aryl groups attached to the acetamido moiety for anticancer activity against C6 cells.

Compound **3** was found to be the most promising anticancer agent against the A549 cell line with an  $IC_{50}$  value of  $21.00 \pm 1.15 \mu\text{g/mL}$  when compared with cisplatin ( $IC_{50} = 13.50 \pm 2.12 \mu\text{g/mL}$ ). Compounds **1**, **5**, and **8** also showed anticancer activity against the A549 cell line with  $IC_{50}$  values of  $46.33 \pm 2.31$ ,  $42.67 \pm 2.52$ , and  $41.33 \pm 1.15 \mu\text{g/mL}$ , respectively. Interestingly, compound **4**, the most potent anticancer agent against the C6 cell line, did not show any inhibitory activity against the A549 cell line ( $IC_{50} > 500 \mu\text{g/mL}$ ). This outcome indicated that *p*-nitro substituent significantly decreased anticancer activity against the A549 cell line.

Toxicity to host cells is an important characteristic to assess the safety of drug candidates early in the drug discovery process. In order to evaluate whether the compounds were toxic or nontoxic to healthy cells, the cytotoxic effects of compounds **1-8** on NIH/3T3 mouse embryonic fibroblast cells were investigated using the MTT assay (Table 2). Generally, the most potent anticancer agents in this series showed low cytotoxicity against the NIH/3T3 cell line with  $IC_{50}$  values higher than their effective  $IC_{50}$  values.

After the 24 h incubation period, the apoptotic effects of compounds **1**, **2**, **3**, **4**, **5**, and **8** were analyzed based on Annexin V-PI binding capacities in flow cytometry. Following flow cytometric analyses, the early and late apoptotic effects of compounds **1**, **2**, **3**, **4**, **5**, and **8** (for  $IC_{50}$  doses) on the C6 cell line were determined as 25.7%, 23.8%, 22.7%, 13.6%, 17.4%, and 10.0%, respectively (Table 3, Figure 1). On the other hand, the early and late apoptotic effects of compounds **1**, **3**, **5**, and **8** (for  $IC_{50}$  doses) on the A549 cell line were very low (1.5%, 0.3%, 0.2%, and 2.3%, respectively). However, their necrotic cell percentages were very high (60.2%, 25.5%, 81.1%, and 54.0%,

**Table 3. Percentages of typical quadrant analysis of Annexin V FITC/PI flow cytometry of C6 cells treated with compounds 1, 2, 3, 4, 5, 8, and cisplatin**

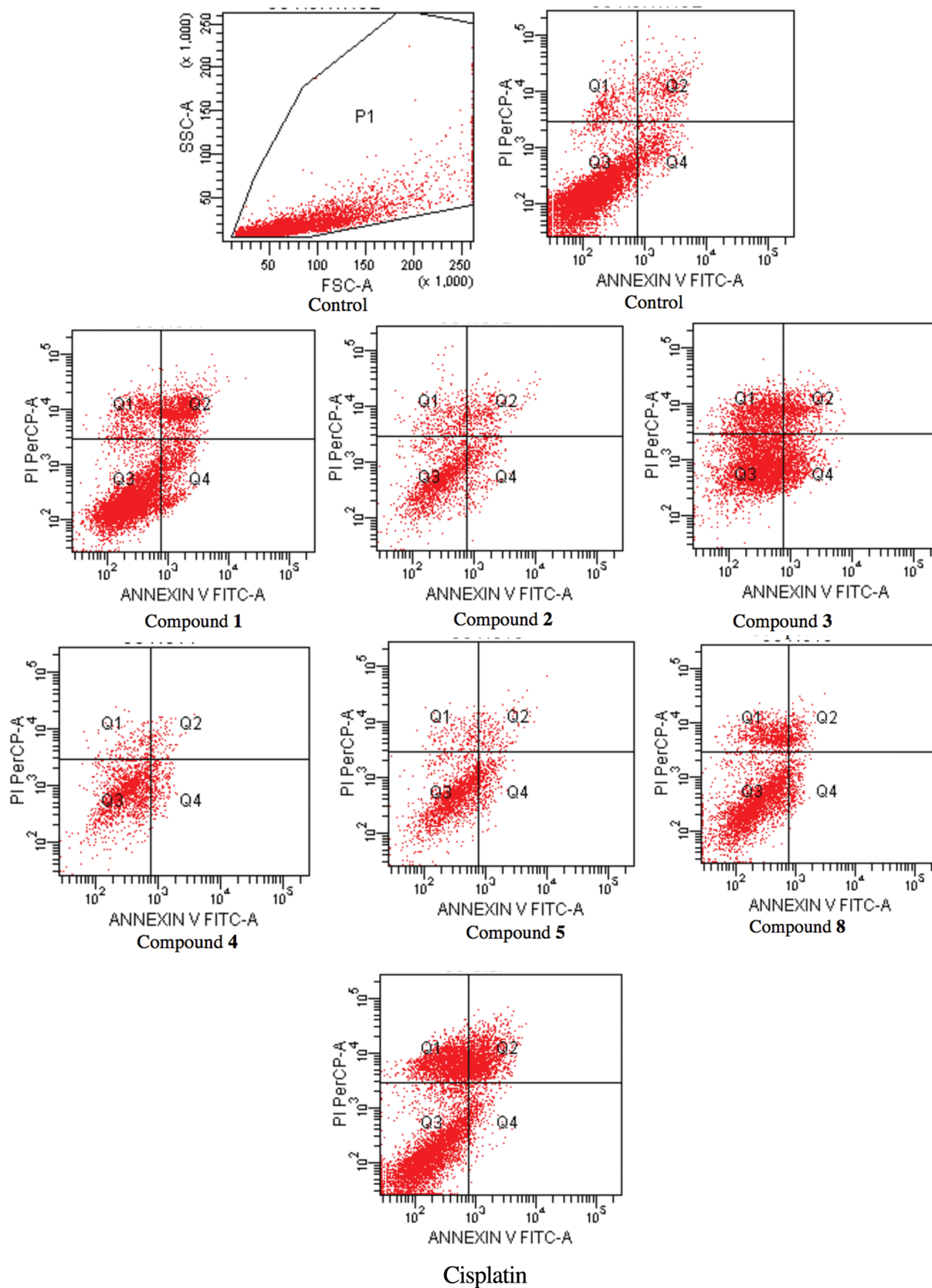
Groups	Early apoptotic cells %	Late apoptotic cells %	Viable cells %	Necrotic cells %
Control (untreated)	5.2	4.3	87.0	3.1
Compound 1 treated cells	9.7	16.0	66.1	8.2
Compound 2 treated cells	12.2	11.6	63.5	12.8
Compound 3 treated cells	12.8	9.9	21.8	21.8
Compound 4 treated cells	9.6	4.0	78.7	7.6
Compound 5 treated cells	11.1	6.3	74.6	8.0
Compound 8 treated cells	4.7	5.3	71.7	18.3
Cisplatin treated cells	3.4	15.4	55.5	25.7

C6 cells were cultured for 24 h in medium with compounds **1**, **2**, **3**, **4**, **5**, **8**, and cisplatin at  $IC_{50}$  values. At least 10,000 cells were analyzed per sample and quadrant analysis was performed; PI: Propidium iodide

**Table 2.  $IC_{50}$  values of the compounds against A549, C6, and NIH/3T3 cells for 24 h**

Compound	C6 cell line	A549 cell line	NIH/3T3 cell line
1	$50.66 \pm 12.50$	$46.33 \pm 2.31$	>500
2	$42.33 \pm 2.52$	$160.00 \pm 34.64$	$175.00 \pm 35.35$
3	$22.00 \pm 3.00$	$21.00 \pm 1.15$	$91.67 \pm 7.64$
4	$18.50 \pm 4.95$	>500	$275.00 \pm 35.36$
5	$46.67 \pm 2.89$	$42.67 \pm 2.52$	$450.00 \pm 70.71$
6	$135.00 \pm 21.21$	$91.67 \pm 2.89$	>500
7	$76.67 \pm 2.89$	$88.33 \pm 7.64$	$480.00 \pm 26.46$
8	$42.67 \pm 2.08$	$41.33 \pm 1.15$	$125.00 \pm 35.36$
Cisplatin	$24.33 \pm 0.58$	$13.50 \pm 2.12$	nt

nt: Not tested; Values are given as mean  $\pm$  standard deviation



**Figure 1.** Typical quadrant analysis of Annexin V-FITC/PI flow cytometry of C6 cells treated with compounds 1, 2, 3, 4, 5, 8, and cisplatin. At least 10,000 cells were analyzed per sample and quadrant analysis was performed. The portion (%) of cell number is shown in each quadrant. Q1, necrotic cells; Q2, late apoptotic cells; Q3, viable cells; Q4, early apoptotic cells. C6 cells were cultured for 24 h in medium with  $IC_{50}$  concentrations of the compounds, cisplatin, and untreated control cells

respectively) (Table 4, Figure 2). According to these findings, compounds **1**, **2**, and **3** (25.7%, 23.8%, and 22.7%) showed more apoptotic activity than cisplatin (18.8%) against C6 cells. On the other hand, compounds **1**, **3**, **5**, and **8** caused necrotic cell death in A549 cells.

Due to the key role of caspase-3 activation in the initiation of cellular events during the early apoptotic process,<sup>29</sup> the effects of compounds **1**, **2**, **3**, **4**, **5**, and **8** on caspase-3 activation in the C6 cell line were determined. Caspase-3 positive cell percentages of compounds **1**, **3**, and cisplatin (for IC<sub>50</sub> doses) were determined as 18.6%, 49.7%, and 14.9%, respectively (Table 5, Figure 3). On the other hand, caspase-3 negative cell percentages of these compounds and cisplatin (for IC<sub>50</sub> doses) were determined as 80.4%, 49.5%, and 85.0%, respectively. These findings indicated that compound **3** was the most effective compound on caspase-3 activation in the C6 cell line.

In order to investigate the effects of compounds **1**, **2**, **3**, **4**, **5**, and **8** on the MMP of C6 and A549 cells, the cells were incubated with IC<sub>50</sub> concentrations of these compounds for 24

h. Compounds **3** and **8** caused higher disturbance to MMP than cisplatin in both cell line. Mitochondrial membrane depolarized cell percentages of compounds **3**, **8** and cisplatin (for IC<sub>50</sub> doses) were determined as 27.6, 27.9, and 16.9 in the C6 cell line (Table 6, Figure 4), while mitochondrial membrane depolarized cell percentages of these compounds and cisplatin (for IC<sub>50</sub> doses) were determined as 44.7, 28.2, and 24.1, respectively, in the A549 cell line (Table 6, Figure 5).

Due to the importance of cell cycle checkpoints for the progression of cell proliferation,<sup>30</sup> the compounds were analyzed for their effects on the cell cycle in A549 and C6 cells (Table 7, Figure 6). Compounds **2**, **3**, **4**, **5**, and **8** induced G1/S phase arrest in C6 cells. Among them, compound **3** caused more

**Table 4.** Percentages of typical quadrant analysis of Annexin V FITC/PI flow cytometry of A549 cells treated with compounds **1**, **3**, **5**, **8**, and cisplatin

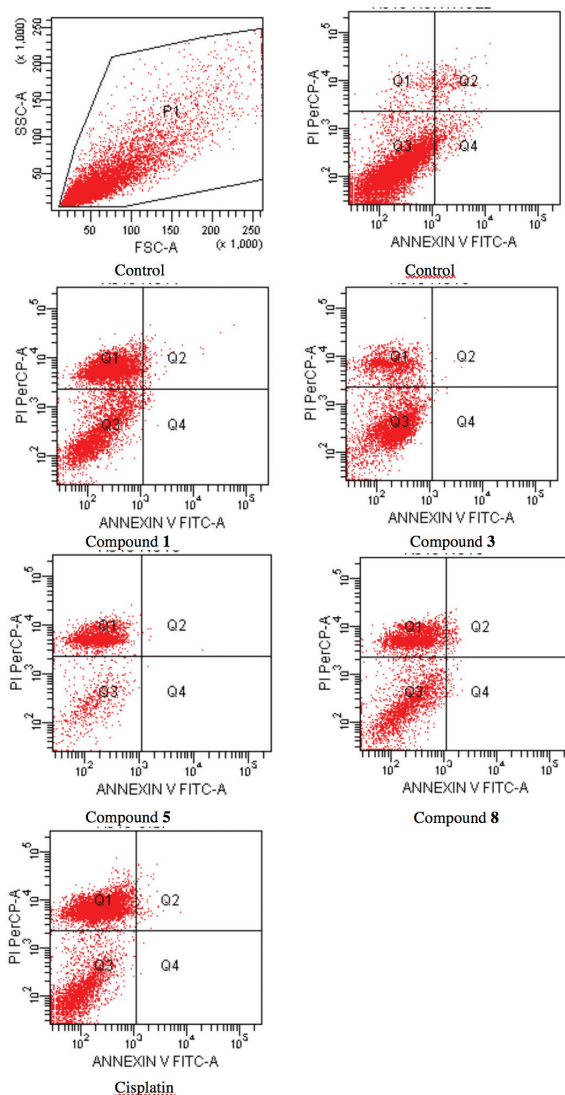
Groups	Early apoptotic cells %	Late apoptotic cells %	Viable cells %	Necrotic cells %
Control (untreated)	3.6	2.5	91.7	2.2
Compound <b>1</b> treated cells	0.3	1.2	38.2	60.2
Compound <b>3</b> treated cells	0.1	0.2	74.2	25.5
Compound <b>5</b> treated cells	0.0	0.2	18.7	81.1
Compound <b>8</b> treated cells	0.7	1.6	43.6	54.0
Cisplatin treated cells	0.0	0.8	30.5	68.7

A549 cells were cultured for 24 h in medium with compounds **1**, **3**, **5**, **8**, and cisplatin at IC<sub>50</sub> values. At least 10,000 cells were analyzed per sample and quadrant analysis was performed; PI: Propidium iodide

**Table 5.** Percentages of quadrant analysis of active caspase-3 phycoerythrin staining by flow cytometry of C6 cells treated with IC<sub>50</sub> concentrations of compounds **1**, **2**, **3**, **4**, **5**, **8**, and cisplatin

Groups	Caspase-3 (-) cells %	Caspase-3 (+) cells %
Control (untreated)	96.3	3.1
Compound <b>1</b> treated cells	80.4	18.6
Compound <b>2</b> treated cells	88.6	11.1
Compound <b>3</b> treated cells	49.5	49.7
Compound <b>4</b> treated cells	87.7	12.7
Compound <b>5</b> treated cells	89.4	10.1
Compound <b>8</b> treated cells	97.4	2.0
Cisplatin treated cells	85.0	14.9

C6 cells were cultured for 24 h in medium with compounds **1**, **2**, **3**, **4**, **5**, **8**, and cisplatin at IC<sub>50</sub> concentrations. At least 10,000 cells were analyzed per sample and quadrant analysis was performed



**Figure 2.** Typical quadrant analysis of Annexin V-FITC/PI flow cytometry of A549 cells treated with compounds **1**, **3**, **5**, **8**, and cisplatin. At least 10,000 cells were analyzed per sample and quadrant analysis was performed. The portion (%) of cell number is shown in each quadrant. Q1, necrotic cells; Q2, late apoptotic cells; Q3, viable cells; Q4, early apoptotic cells. A549 cells were cultured for 24 h in medium with IC<sub>50</sub> concentrations of the compounds, cisplatin, and untreated control cells

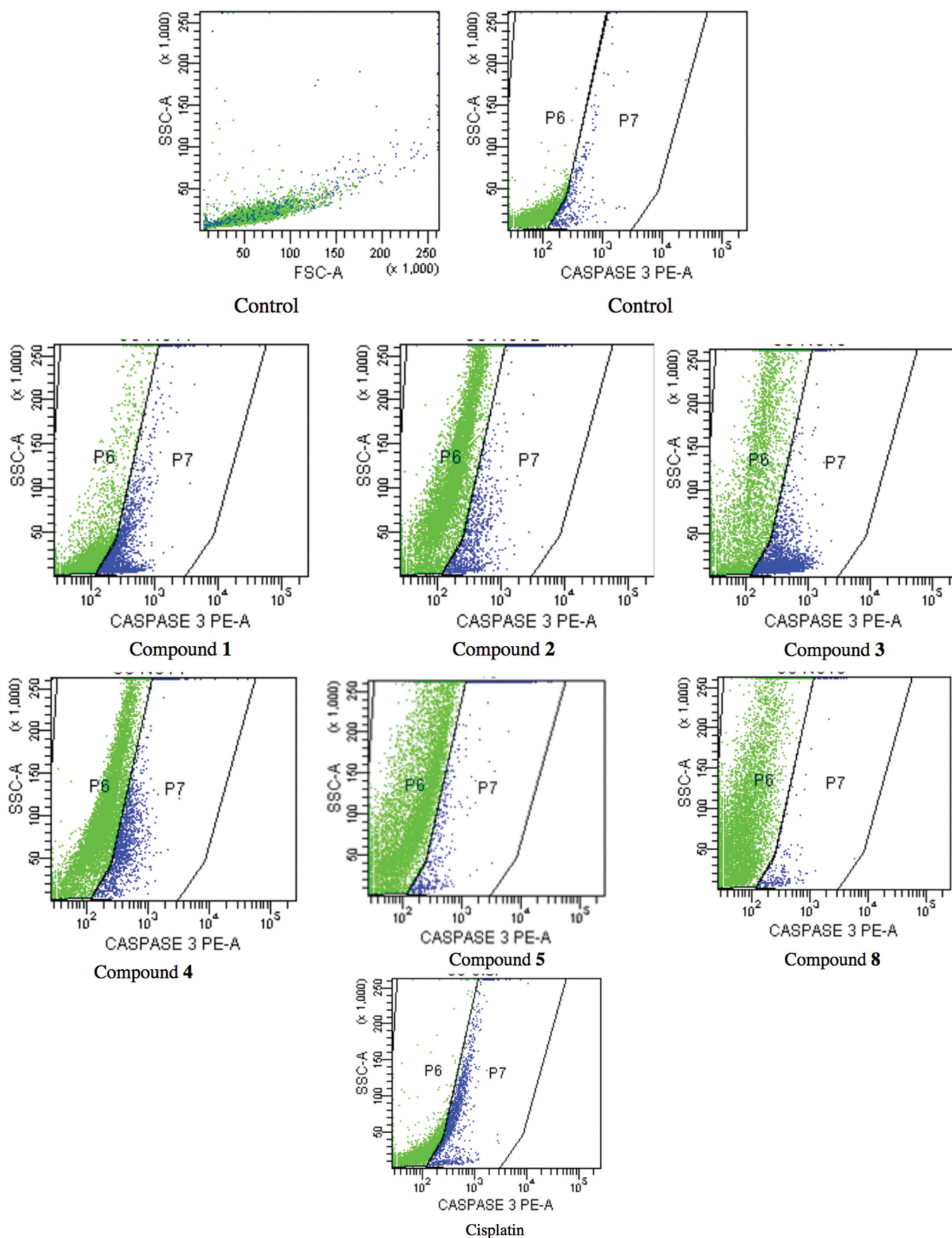
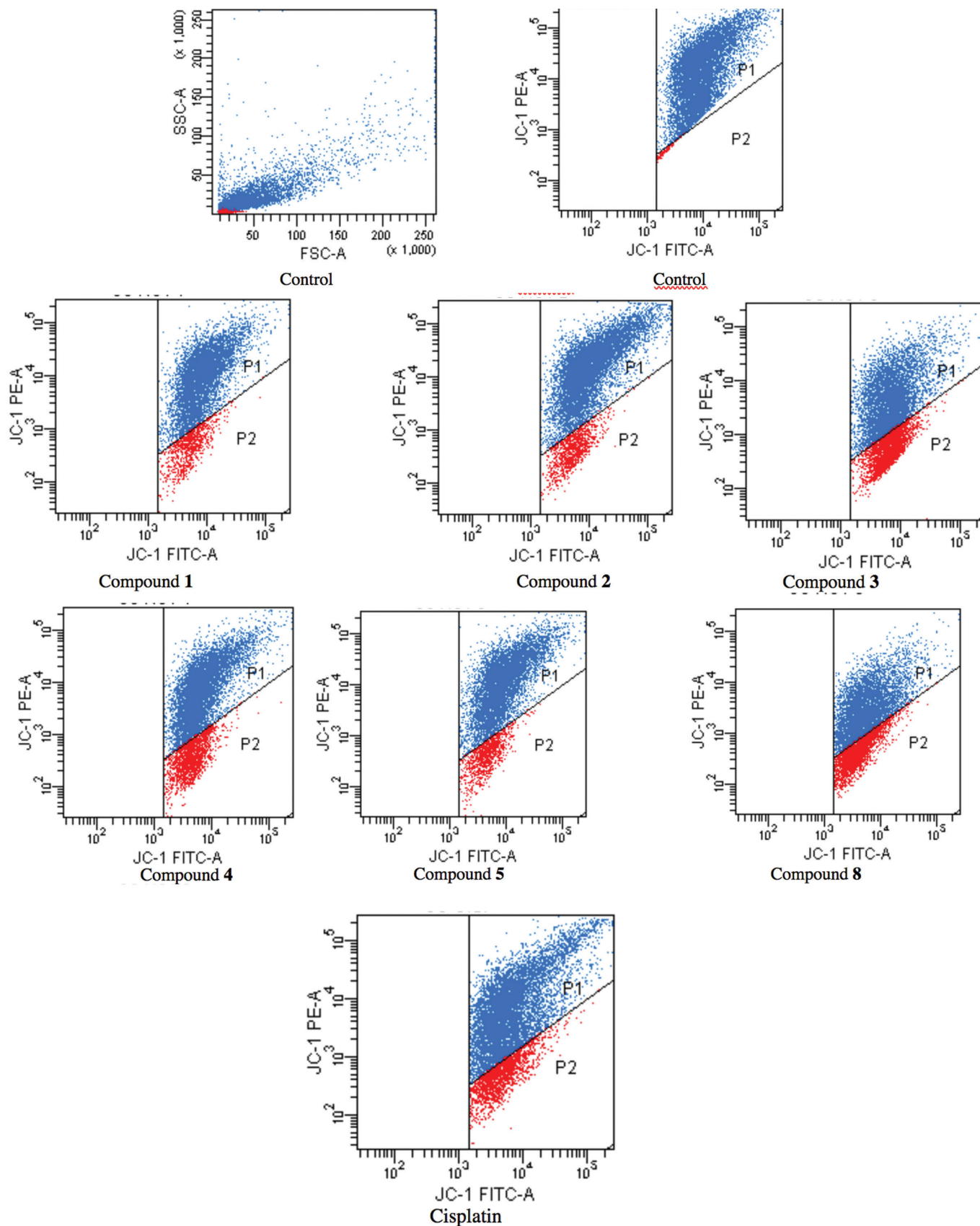
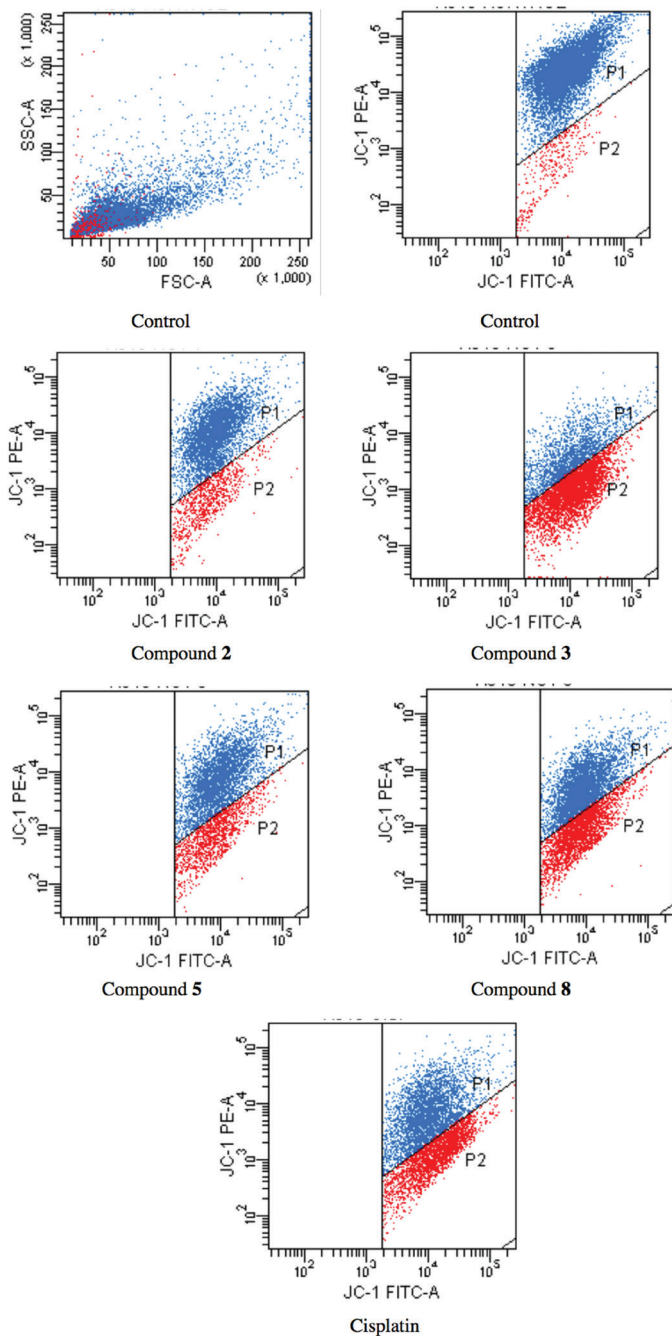


Figure 3. Caspase 3 activity of C6 cells treated with IC<sub>50</sub> concentrations of compounds 1, 3, 5, 8, and cisplatin



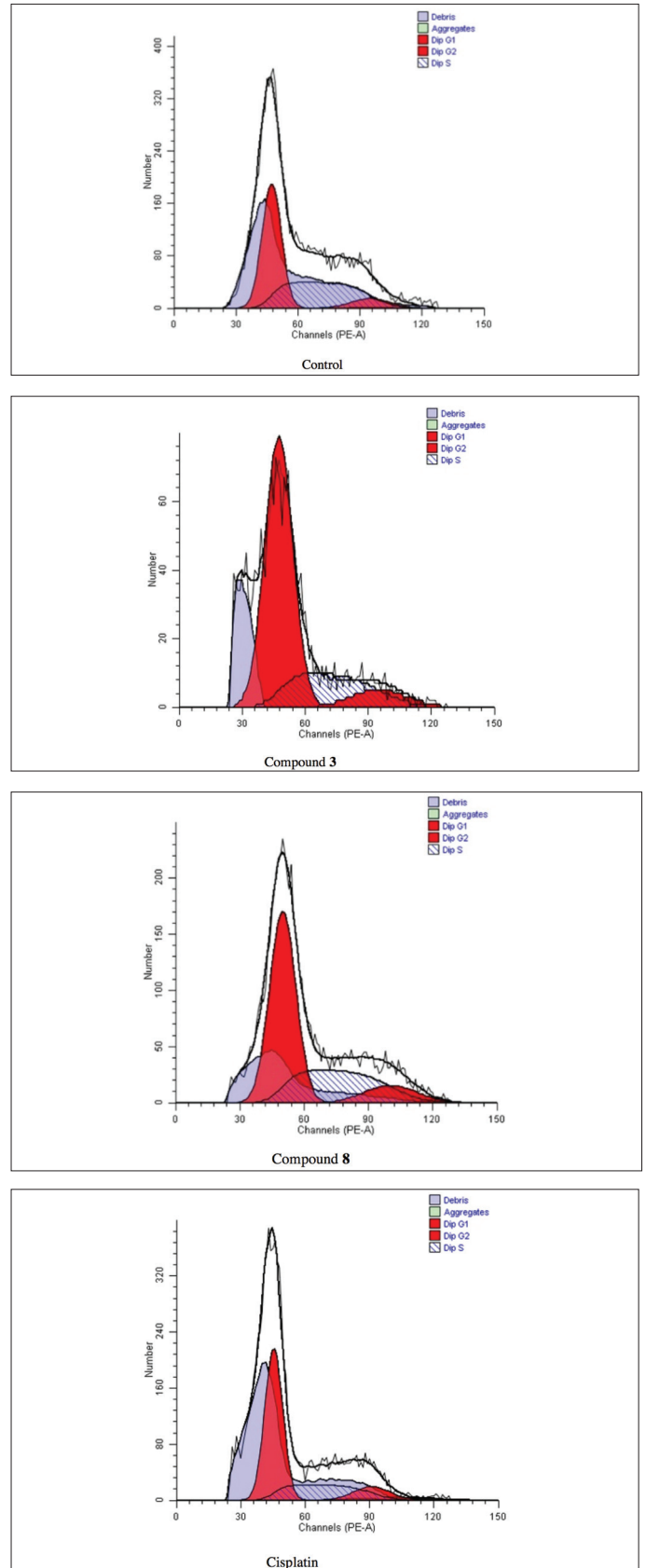
**Figure 4.** The reduction in the MMP in the C6 cell line by the compounds. The cells treated or untreated with IC<sub>50</sub> doses of the compounds for 24 h were stained with mitochondrial-selective JC-1 dye and analyzed by flow cytometry. P1: mitochondrial membrane polarized cells, P2: mitochondrial membrane depolarized cells



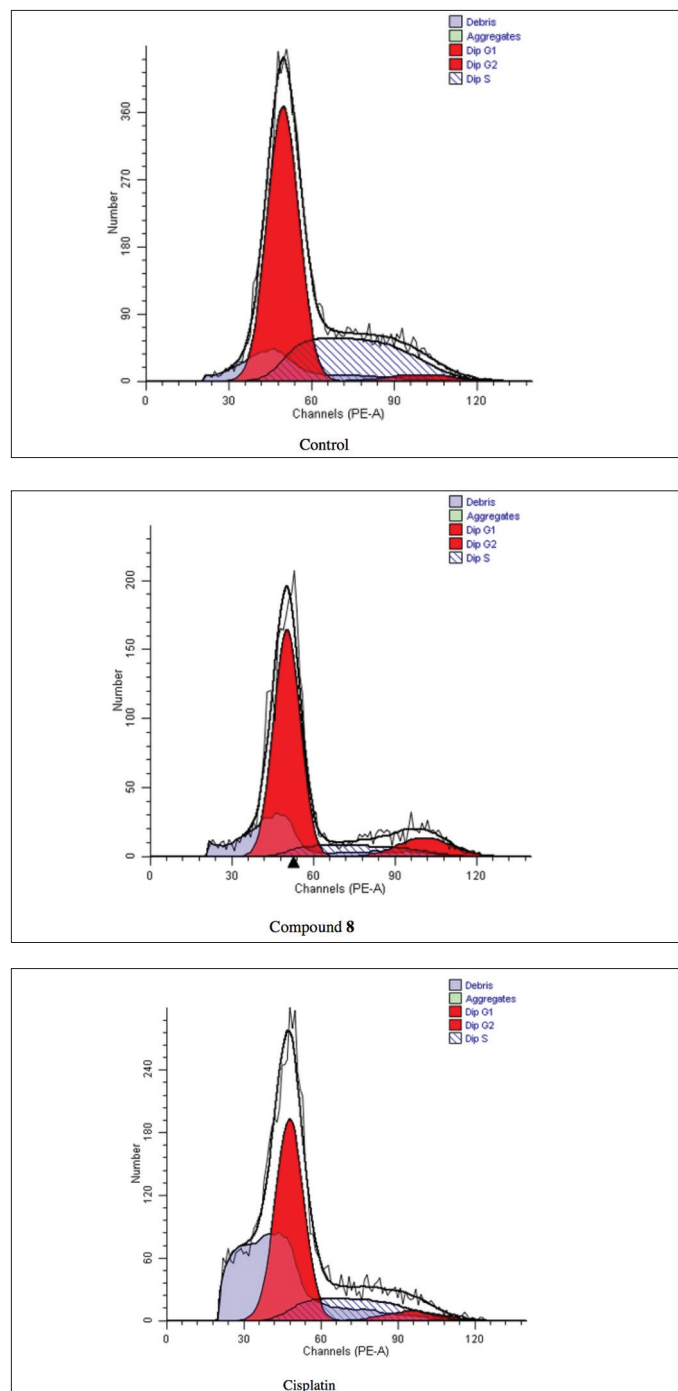


**Figure 5.** The reduction in the MMP in the A549 cell line by the compounds. The cells treated or untreated with  $IC_{50}$  doses of the compounds for 24 h were stained with the mitochondrial-selective JC-1 dye and analyzed by flow cytometry. P1: mitochondrial membrane polarized cells, P2: mitochondrial membrane depolarized cells

G1/S phase arrest (67.21%) than cisplatin (62.57%). Compounds 1, 2, 3, 4, 5, and 8 arrested the G2/M cell cycle in C6 cells. On the other hand, compounds 1, 5, and 8 caused G2/M cell cycle arrest in A549 cells (Table 7, Figure 7). Only compound 8 caused G1/S cell cycle phase arrest in A549 cells. The effects of compound 8 on G1/S arrest were more significant in A549 cells than in C6 cells.



**Figure 6.** Cell cycle distribution of C6 cells treated with  $IC_{50}$  concentrations of the compounds for 24 h. At least 10,000 cells were analyzed per sample. Blue color shows debris, green color shows aggregates, and red color shows cells in G1 and G2 phases



**Figure 7.** Cell cycle distribution of A549 cells treated with  $IC_{50}$  concentration of compound **8** and cisplatin for 24 h. At least 10,000 cells were analyzed per sample. Blue color shows debris, green color shows aggregates, and red color shows cells in G1 and G2 phases

As a consequence of the pivotal role of Akt in regulating diverse cellular functions including cell growth, proliferation, and survival,<sup>1-6</sup> the most potent anticancer agents were investigated for their inhibitory effects on Akt activity (Table 8). Compounds **3** ( $92.36 \pm 0.70\%$  and  $91.22 \pm 0.16\%$  for C6 and A549 cells, respectively) and **8** ( $86.52 \pm 0.37\%$  and  $70.48 \pm 13.28\%$  for C6 and A549 cells, respectively) were the most potent Akt inhibitors in this series compared to control cells ( $p < 0.001$ ).

**Table 6.** Reduction in the MMP in C6 and A549 cells by  $IC_{50}$  concentrations of the compounds and cisplatin

Groups	Mitochondrial membrane polarized (P1) cells (%)		Mitochondrial membrane depolarized cells (P2) (%)	
	C6 cells	A549 cells	C6 cells	A549 cells
Control (untreated)	96.0	92.5	1.3	3.3
Compound <b>1</b> treated cells	61.7	9.0	45.4	8.3
Compound <b>2</b> treated cells	87.2	-	10.8	-
Compound <b>3</b> treated cells	58.1	19.9	27.6	44.7
Compound <b>4</b> treated cells	78.5	-	20.1	-
Compound <b>5</b> treated cells	75.5	37.1	11.6	12.9
Compound <b>8</b> treated cells	63.0	44.8	27.9	28.2
Cisplatin treated cells	77.0	38.8	16.9	24.1

P1: Mitochondrial membrane polarized cells, P2: Mitochondrial membrane depolarized cells; Cells treated or untreated with  $IC_{50}$  concentrations of the compounds and cisplatin for 24 h were stained with mitochondrial-selective JC-1 dye and analyzed by flow cytometry

**Table 7.** C6 and A549 cell percentages on G1/S and G2/M cell cycle phases

Compounds	A549 cell line		C6 cell line	
	G1/S phase	G2/M phase	G1/S phase	G2/M phase
Control	62.9	2.85	49.99	7.69
Compound <b>1</b> treated cells	58.05	5.51	49.26	10.91
Compound <b>2</b> treated cells	-	-	51.13	8.42
Compound <b>3</b> treated cells	-	-	67.21	8.59
Compound <b>4</b> treated cells	-	-	56.99	11.48
Compound <b>5</b> treated cells	61.84	4.64	57.71	7.96
Compound <b>8</b> treated cells	73.86	11.87	57.53	10.35
Cisplatin	68.22	6.27	62.57	11.21

Molecular docking simulations were performed to elucidate the possible binding modes of compounds **3** and **8** in the active site of Akt enzyme (PDB code: 3OW4). The docking results of compounds **3** and **8** indicated that  $\pi$ - $\pi$  interactions, H bonds, and salt-bridge formation were responsible for the observed affinity (Figure 8). The nitrophenyl and chlorophenyl groups and acetamido moiety of compound **3** formed  $\pi$ - $\pi$  interactions and H-bonds with Ala230, Lys179, and Asp292 residues, respectively. However, the nitrophenylamino group and acetamido moiety of compound **8** presented  $\pi$ - $\pi$  interactions, H-bonds, and salt-bridge formation with Asp439, Glu234, Arg4, and Lys158 and 276, Phe442, and Ser7 residues, respectively. Nitro substitution on the benzothiazole ring was engaged in  $\pi$ - $\pi$  interactions with Phe161. Docking scores were  $-5.55$  kcal/mol

Table 8. Akt inhibitory effects of compounds 1, 2, 3, 4, 5, 8, and cisplatin

Compound	Inhibition %	
	C6 cell line	A549 cell line
1	38.68±0.75*	43.39±5.86*
2	66.37±2.47**	-
3	92.36±0.70***	91.22±0.16***
4	43.30±32.03*	-
5	68.84±20.82**	66.57±8.75**
8	86.52±0.37***	70.48±13.28***
Cisplatin	23.64±5.09*	54.18±28.50**

Significant differences versus control values, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ . Values are given as mean  $\pm$  standard deviation

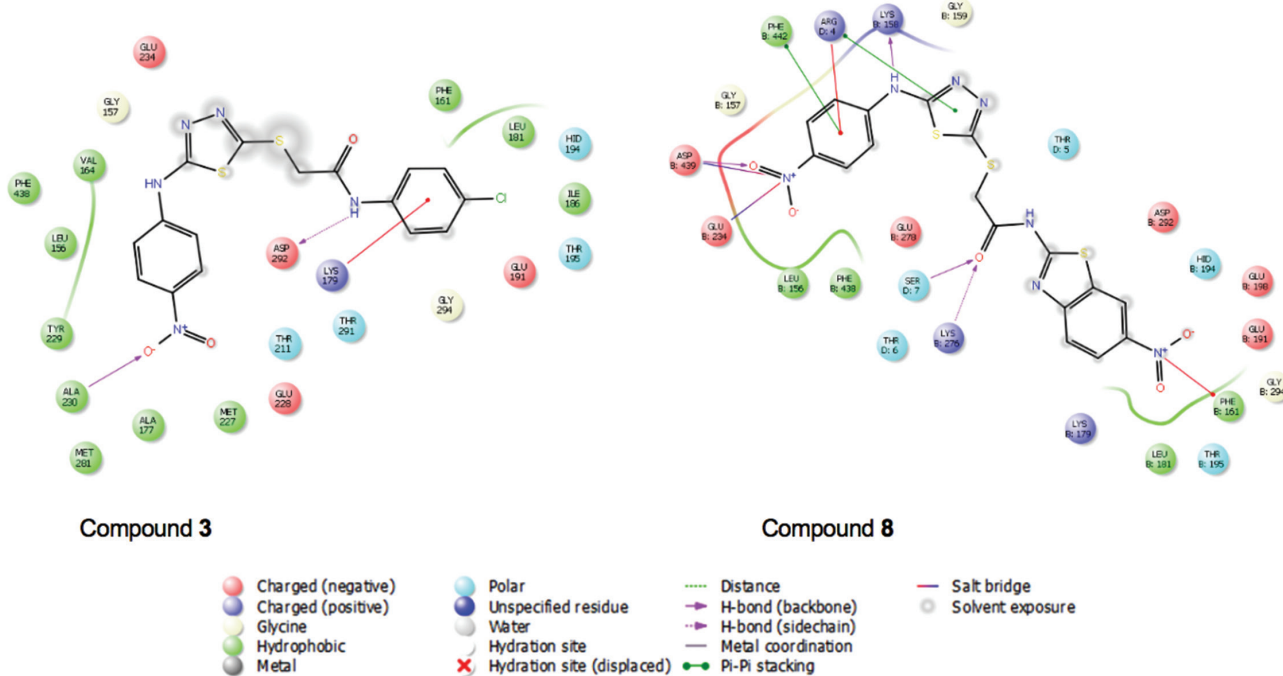
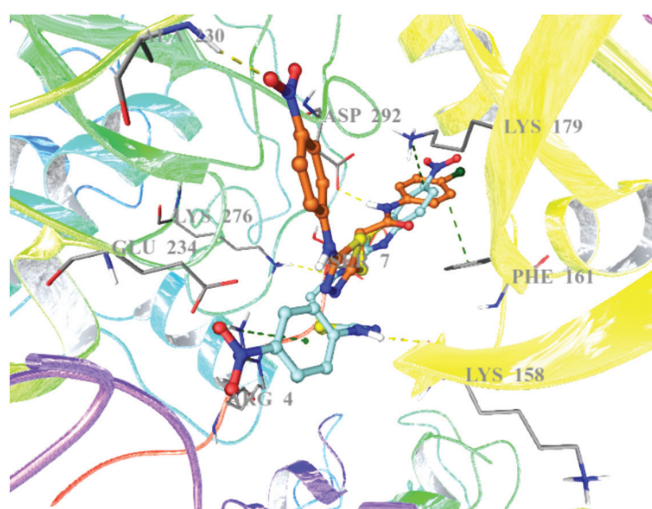


Figure 8. Docking positions and interactions of compounds 3 and 8 in the active site of Akt enzyme, respectively (Ligand custom carbons are colored orange for compound 3 and turquoise for compound 8)

for compound **3** and -5.33 kcal/mol for compound **8** in the active site of Akt enzyme.

## CONCLUSIONS

In the current work, *in vitro* and *in silico* studies were carried out to determine the mechanism of antitumor action of thiadiazole-based anticancer agents. Compounds **3** and **8** induced apoptosis and cell cycle arrest in G1/S and G2/M phases in the C6 cell line through inhibition of Akt activity. Docking studies also confirmed that compounds **3** and **8** demonstrated high affinity to the active site of Akt enzyme by forming  $\pi$ - $\pi$  interactions, hydrogen bonds, and salt-bridge formation with appropriate residues. According to *in vitro* and docking studies, compounds **3** and **8** stand out as promising antiglioma agents for further *in vitro* and *in vivo* studies.

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