


Serum immunoglobulin G antibody titer to *Fusobacterium nucleatum* is associated with unfavorable outcome after stroke

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Accepted for publication 5 March 2020

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Introduction

A stroke causes damage to the brain due to sudden loss of blood circulation, and its occurrence is broadly categorized into ischemic and hemorrhagic [1]. Although considered to be one of the leading causes of death, a stroke in non-fatal cases can lead to various types of disabilities,

Summary

Stroke can be a cause of death, while in non-fatal cases it is a common cause of various disabilities resulting from associated brain damage. However, whether a specific periodontal pathogen is associated with increased risk of unfavorable outcome after stroke remains unknown. We examined risk factors for unfavorable outcome following stroke occurrence, including serum antibody titers to periodontal pathogens. The enrolled cohort included 534 patients who had experienced an acute stroke, who were divided into favorable ($n = 337$) and unfavorable ($n = 197$) outcome groups according to modified ranking scale (mRS) score determined at 3 months after onset (favorable = score 0 or 1; unfavorable = score 2–6). The associations of risk factors with unfavorable outcome, including serum titers of IgG antibodies to 16 periodontal pathogens, were examined. Logistic regression analysis showed that the initial National Institutes of Health stroke scale score [odds ratio (OR) = 1.24, 95% confidence interval (CI) = 1.18–1.31, $P < 0.001$] and C-reactive protein (OR = 1.29, 95% CI = 1.10–1.51, $P = 0.002$) were independently associated with unfavorable outcome after stroke. Following adjustment with those, detection of the antibody for *Fusobacterium nucleatum* ATCC 10953 in serum remained an independent predictor of unfavorable outcome (OR = 3.12, 95% CI = 1.55–6.29, $P = 0.002$). Determination of the antibody titer to *F. nucleatum* ATCC 10953 in serum may be useful as a predictor of unfavorable outcome after stroke.

Keywords: *Fusobacterium nucleatum*, serum IgG antibody titer, unfavorable outcome after stroke

including physical, cognitive or emotional deficiency, resulting in requirement of partial or complete assistance with performing activities of daily living [2]. Furthermore, related costs, such as home- and hospital-based rehabilitation and care, can place a heavy financial burden on individuals as well as society. Therefore, elucidation of clinical methods

useful for prediction of an unfavorable functional outcome in the early stage following a stroke is important.

Periodontitis, a commonly encountered chronic oral inflammatory disease caused by interactions between host immune responses and periodontal bacteria, is characterized by loss of connective tissue and alveolar bone support, leading to tooth loss [3]. Some cohort studies have shown a relationship of periodontal disease with stroke occurrence. Recently, Lafson *et al.* evaluated the association between periodontal disease and incidence of ischemic and hemorrhagic stroke by performing a meta-analysis of cohort studies, and found that the risk of stroke was significantly increased in individuals with periodontitis, while tooth loss was also shown to be a risk factor [4]. Furthermore, the Atherosclerosis Risk in Communities (ARIC) cohort study of 10 362 stroke-free participants conducted during a 15-year follow-up period showed that periodontal disease was associated with incidence of cardioembolic and atherothrombotic stroke, and that regular dental care may lead to a lower adjusted risk [5].

Recent studies have also examined factors other than tooth loss as indicators of periodontal disease, such as performing measurements of concentrations of serum antibodies to periodontal pathogens, as those findings are considered to more accurately reflect the disease process and have become part of established criteria to identify causative organisms [6]. Recently, serum antibody titers related to a specific periodontal pathogen were revealed to be risk factors for systemic diseases, including ischemic stroke, coronary heart disease, non-alcoholic fatty liver disease and Alzheimer's disease [7-10]. However, it remains unknown whether a serum antibody titer related to a specific periodontal pathogen is associated with increased risk of an unfavorable outcome after stroke. To determine the correlation of specific periodontal pathogens, we examined the relationships between unfavorable outcomes following stroke and associated risk factors, including serum titers of IgG antibodies to several different periodontal pathogens.

Materials and methods

Subjects

We enrolled acute stroke patients, diagnosed as ischemic or hemorrhagic, who had undergone treatment at Hiroshima University Hospital or Suiseikai Kajikawa Hospital from January 2013 to April 2016. The study design was approved by the Ethical Committee of Hiroshima University (Permission no. Epd-614-2) and Suiseikai Kajikawa Hospital (Permission no. 2015-03), and each participant signed an informed consent agreement. All examinations were performed in accordance with relevant guidelines and regulations. Analysis of computed tomography or magnetic resonance imaging results of all patients was performed to determine

a diagnosis of ischemic or hemorrhagic stroke. Patients who were disabled prior to stroke incidence corresponding to a pre-morbid modified Rankin scale (mRS) score ≥ 2 were excluded from analysis, while those with a pre-morbid mRS score of 0 or 1 were included as subjects. Favorable stroke outcome was defined as independence after 3 months, corresponding to an mRS score of 0 and 1, and an unfavorable outcome as an mRS score of 2-6 [11]. The study cohort included a total of 534 patients (mean age = 71.1 years, 57.1% male). The total sample size required for accurate unpaired *t*-test, Mann-Whitney *U*-test, χ^2 test and Fisher's exact test results, obtained using the G*Power software package (version 3.1.9.4; Heinrich-Heine-Universität, Düsseldorf, Germany), was calculated to be 278, 290, 88 and 100 subjects, respectively, with a statistical power of 80%, significance level 5% and effect size 0.3. Baseline clinical characteristic data, including gender, age, smoking status, alcohol drinking status and comorbidities (hypertension, diabetes, hyperlipidemia, atrial fibrillation, peripheral arterial disease, congestive heart failure), were obtained for all enrolled patients. Ischemic stroke subtypes were classified using the Trial of Org 10172 in Acute Stroke Treatment (TOAST) criteria [12] by stroke specialists. Hemorrhagic infarction and trauma-induced hemorrhage were excluded from intracerebral hemorrhage. Stroke severity upon admission was evaluated based on the National Institutes of Health Stroke Scale (NIHSS) score. Hypertension was defined as use of anti-hypertensive medication before admission or confirmed blood pressure of $\geq 140/90$ mmHg at rest when measured 2 weeks after onset. Diabetes mellitus was defined as a glycosylated hemoglobin level $\geq 6.5\%$, fasting blood glucose level ≥ 126 mg/dl or use of anti-diabetes medication. Dyslipidemia was defined as total cholesterol level ≥ 220 mg/dl, low-density lipoprotein cholesterol level ≥ 140 mg/dl, high-density lipoprotein cholesterol level < 40 mg/dl, triglyceride level ≥ 150 mg/dl or use of anti-hyperlipidemia medication. Atrial fibrillation was defined as history of sustained or paroxysmal atrial fibrillation, or atrial fibrillation detection upon arrival or during admission. Diagnosis of ischemic heart disease, peripheral arterial disease and congestive heart failure was determined by attending physicians. Blood samples were collected from all patients within 24 h of admission and C-reactive protein (CRP) concentrations were measured using nephelometry.

Determination of serum immunoglobulin (Ig)G antibody titers to periodontal pathogens

Serum IgG antibody titers to periodontal pathogens were determined using an enzyme-linked immunosorbent assay (ELISA), as previously described in detail [13]. Briefly, serum samples were collected from patients within 3 days after stroke occurrence and stored at -80°C . Sonicated preparations of the following periodontal pathogens were used as bacterial antigens in the present study: *Porphyromonas gingivalis* ATCC33277 (fimA type I), HW24D1 (fimA type

II), 6/26 (fimA type III), W83 (fimA type IV) and HNA99 (fimA type V); *Aggregatibacter actinomycetemcomitans* ATCC29523 (serotype a), Y4 (serotype b) and AUNY67 (serotype c); *Prevotella intermedia* ATCC26511, *P. nigrescens* ATCC33563; *Fusobacterium nucleatum* ATCC25586 (subspecies *nucleatum*) and ATCC 10953 (subspecies *polymorphum*); *Treponema denticola* ATCC35405; *Tannerella forsythensis* ATCC43037; *Campylobacter rectus* ATCC33238; and *Eikenella corrodens* ATCC23834. Serum samples from five healthy subjects were also obtained and pooled, and used for calibration. Bacterial antigen-coated wells were washed with phosphate-buffered saline with Tween (PBST), then serum samples in PBST were added to the wells. After incubation at 4°C overnight, the wells were washed with PBST, then filled with alkaline phosphatase-conjugated goat anti-human IgG (gamma-chain specific; Abcam, Cambridge, MA, USA) in PBST. After another incubation at 37°C for 2 h, the wells were again washed with PBST, then an aliquot of p-nitrophenylphosphate at 1 mg/ml; (Wako Pure Chemical Industries Ltd., Osaka, Japan) in 10% diethanolamine buffer was added to each well as a substrate and incubation was performed at 37°C for 30 min. Optical density at 405 nm was measured using a microplate reader (iMark; Bio-Rad Laboratories Inc., Hercules, CA, USA). Values ≥ 1 were considered to represent more than 2 standard deviations of the mean of the controls and defined as antibody-positive. Absolute serum antibody measurements were used to categorize the samples as positive or negative [8,9].

Statistical analysis

Statistical analysis was performed using the SPSS software package, version 24.0 (SPSS Inc., Chicago, IL, USA). Values are expressed as the mean \pm standard deviation or median (minimum, maximum) for continuous variables, and as

frequencies and percentages for discrete variables. Univariate analysis was performed to evaluate differences between the groups regarding baseline characteristics, risk factors, levels of serum CRP and serum IgG antibody titers to periodontal pathogens. Comparisons between the groups were made using an unpaired *t*-test or Mann-Whitney *U*-test for continuous variables, and Fisher's exact test or a χ^2 test for discrete variables. Subsequently, to assess factors that may be associated with an unfavorable outcome, multivariate logistic regression analysis was performed using variables that showed a trend in univariate analysis ($P < 0.2$) with a forced-entry method. All analyses were two-tailed and a value of $P < 0.05$ was considered to indicate statistical significance.

Results

A total of 664 patients who had experienced an acute stroke were initially registered in this study, 130 of whom with a premorbid mRS score ≥ 2 were excluded from analysis. The stroke subtype of the 534 analyzed patients (mean age = 71.1 years, 57.1% male) was cerebral infarction in 447 (large vessel disease 88, cardioembolic disease 102, small vessel disease 92, other determined cause 20, undetermined cause 145) and cerebral hemorrhage in 87 patients. Subsequently, the 534 patients were categorized into favorable ($n = 337$) and unfavorable ($n = 197$) outcome groups according to the mRS score at 3 months after onset (Fig. 1). The initial clinical characteristics were compared between the groups (Table 1), which revealed that age, stroke subtype, initial NIHSS score and CRP level were significantly higher in the unfavorable group ($P < 0.001$) (Table 2). Additionally, patients with atrial fibrillation and congestive heart failure were

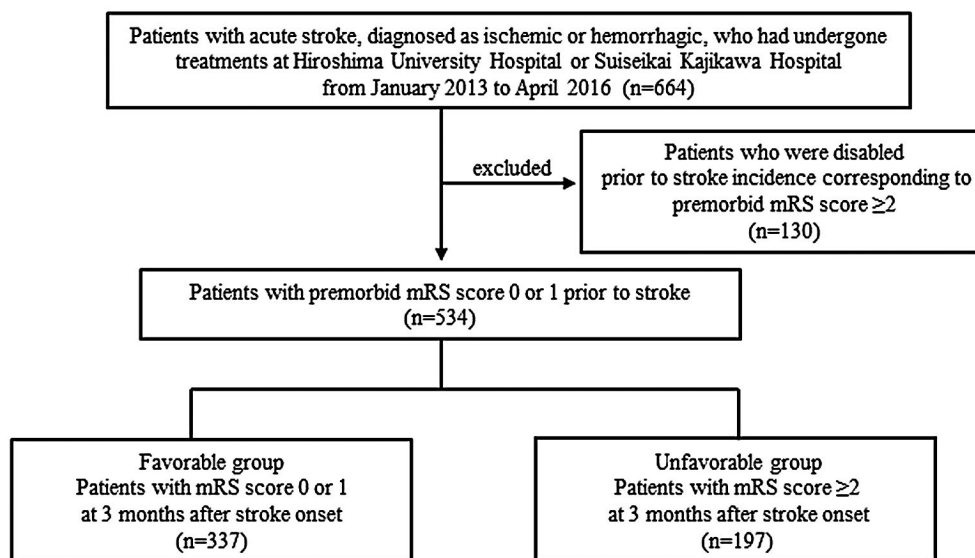


Fig. 1. Flow-chart showing patient selection.

more frequently found in the unfavorable group ($P < 0.001$, $P = 0.01$, respectively), whereas those with dyslipidemia were more frequently found in the favorable group ($P = 0.048$).

Table 1. Baseline clinical characteristics of patients in this study

Clinical parameter $n = 534$	$n = 534$
Age, years	71.1 ± 12.4
Gender, male/female	305/229
BMI, kg/m ²	23.1 ± 4.13
Stroke subtype, ischemia/hemorrhage	447/87
NIHSS score, median (IQR)	3 (1–7.5)
Smoking status, n (%)	
Never	306 (57.3)
Past	100 (18.7)
Current	124 (23.2)
Alcohol status, n (%)	
No	274 (51.3)
Occasionally	109 (20.4)
Daily	128 (24.0)
Hypertension, n (%)	401 (75.1)
Diabetes mellitus, n (%)	129 (24.2)
Dyslipidemia, n (%)	227 (42.5)
Atrial fibrillation, n (%)	93 (17.4)
Ischemic heart disease, n (%)	43 (8.1)
Peripheral arterial disease, n (%)	18 (3.4)
Congestive heart failure, n (%)	21 (3.9)
CRP, mg/l, median (IQR)	0.11 (0.04–0.36)

NIHSS = National Institutes of Health Stroke Scale; IQR = interquartile range; CRP = C-reactive protein; BMI = body mass index.

Next, the frequency of positive findings for serum IgG antibodies to periodontal pathogens were compared between the groups (Table 3). Antibodies to *A. actinomycetemcomitans* AUNY67 (serotype c), *F. nucleatum* ATCC25586 (subspecies *nucleatum*), *F. nucleatum* ATCC10953 (subspecies *polymorphum*) and *C. rectus* ATCC33238 were more frequently detected in the unfavorable compared with the favorable group, while detection of antibodies to the other examined periodontal pathogens was not significantly different between the groups.

Variables with a P -value < 0.20 in univariate analysis were entered into multivariate logistic regression analysis using a forced-entry method (Table 4). Those results showed that initial NIHSS score [odds ratio (OR) = 1.24, 95% confidence interval (CI) = 1.18–1.31; $P < 0.001$] and CRP (OR = 1.29, 95% CI = 1.10–1.51; $P = 0.002$) were independently associated with unfavorable outcome after stroke. We also found that detection of the serum IgG antibody to *F. nucleatum* ATCC 10953 was an independent predictor of unfavorable outcome following stroke (OR = 3.12, 95% CI = 1.55–6.29; $P = 0.002$).

Discussion

Although stroke is a leading cause of death, the rates of mortality of affected individuals have recently been decreasing due to development of advanced medical treatments, which, along with the ageing of society, has resulted in greater numbers who survive such an

Table 2. Comparison of clinical characteristics between favorable and unfavorable groups

Clinical parameter	Favorable ($n = 337$)	Unfavorable ($n = 197$)	P -value
Age, years	69.4 ± 12.2	74.1 ± 12.5	$< 0.001^*$
Gender, male/female	201/136	104/93	0.12
BMI, kg/m ²	23.3 ± 4.1	22.6 ± 9.2	0.03
Stroke subtype, ischemia/hemorrhage	302/35	145/52	$< 0.001^*$
NIHSS score, median (IQR)	2 (1–3)	9.5 (4–18)	$< 0.001^*$
Smoking status, n (%)			0.39
Never	192 (57)	114 (57.9)	
Past	69 (20.5)	31 (15.7)	
Current	75 (22.3)	49 (24.9)	
Alcohol status, n (%)			0.12
No	175 (51.9)	99 (50.3)	
Occasionally	65 (19.3)	44 (22.3)	
Daily	92 (27.3)	36 (18.3)	
Hypertension, n (%)	246 (73.0)	155 (78.7)	0.16
Diabetes mellitus, n (%)	79 (23.4)	50 (25.4)	0.63
Dyslipidemia, n (%)	154 (45.7)	73 (37.1)	0.048*
Atrial fibrillation, n (%)	40 (18.8)	53 (26.9)	$< 0.001^*$
Ischemic heart disease, n (%)	24 (7.1)	19 (9.6)	0.31
Peripheral arterial disease, n (%)	9 (2.7)	9 (4.7)	0.25
Congestive heart failure, n (%)	6 (1.8)	15 (7.6)	0.01*
CRP, mg/l, median (IQR)	0.09 (0.03–0.23)	0.2 (0.06–0.81)	$< 0.001^*$

NIHSS = National Institutes of Health Stroke Scale; IQR = interquartile range; CRP = C-reactive protein; BMI = body mass index.

* $P < 0.05$ (statistically significant).

Table 3. Detection of serum IgG antibody titers to periodontal pathogens in favorable and unfavorable groups after stroke

Periodontal pathogen	Positive for periodontal pathogen, <i>n</i> (%)		<i>P</i> -value
	Favorable	Unfavorable	
	(<i>n</i> = 337)	(<i>n</i> = 197)	
<i>P. gingivalis</i> ATCC33277 (fimA type I)	163 (48.4)	95 (48.2)	0.96
<i>P. gingivalis</i> HW24D1 (fimA type II)	141 (41.8)	90 (45.7)	0.39
<i>P. gingivalis</i> 6/26 (fimA type III)	200 (59.3)	117 (59.4)	0.99
<i>P. gingivalis</i> W83 (fimA type IV)	148 (43.9)	87 (44.1)	0.96
<i>P. gingivalis</i> HNA99 (fimA type V)	191 (56.7)	111 (56.3)	0.94
<i>A. actinomycetemcomitans</i> ATCC29523 (Serotype a)	61 (18.1)	45 (22.8)	0.19
<i>A. actinomycetemcomitans</i> Y4 (serotype b)	100 (29.7)	69 (35.0)	0.20
<i>A. actinomycetemcomitans</i> AUNY67 (Serotype c)	87 (25.8)	67 (34.0)	0.044*
<i>P. intermedia</i> ATCC26511	92 (27.3)	64 (32.5)	0.25
<i>P. nigrescens</i> ATCC33563	55 (16.3)	40 (20.3)	0.24
<i>F. nucleatum</i> ATCC25586	50 (14.8)	64 (32.5)	< 0.001*
<i>F. nucleatum</i> ATCC10953	26 (7.7)	44 (22.3)	< 0.001*
<i>T. denticola</i> ATCC35405	20 (5.9)	18 (9.1)	0.17
<i>T. forsythensis</i> ATCC43037	35 (10.4)	21 (10.7)	0.92
<i>C. rectus</i> ATCC33238	76 (22.6)	61 (31.0)	0.032*
<i>E. corrodens</i> ATCC23834	46 (13.6)	38 (19.3)	0.08

**P* < 0.05 (statistically significant).

occurrence [14]. A study conducted in 2010 estimated that there were 50 million stroke survivors throughout the world at that time who showed various levels of functional disability, with 25–74% requiring some level of assistance or fully dependent on caregivers for activities of daily living [15]. NIHSS score has been shown to be associated with overall functional outcome in patients who have had a stroke [12], while a derivation cohort study conducted in Japan reported that specific NIHSS items, including leg weakness, intra- and extracranial vascular imaging abnormalities, advanced age and female gender, were associated with an unfavorable outcome following a minor ischemic stroke [16]. Additionally, C-reactive protein (CRP), a sensitive inflammatory marker, has been reported to be associated with increased risk of stroke, with higher CRP level shown to be an independent predictor of survival after ischemic stroke and functional outcome after thrombolytic stroke [17,18]. Similar to those previous studies, the present findings revealed that initial NIHSS score and CRP concentration had a dependent association with unfavorable outcome following a stroke in our patients.

Hypertension and smoking status are well-established risk factors for stroke occurrence, with a strong association between those and stroke onset shown in many studies,

Table 4. Multivariate logistic regression analysis to identify predictive factors for unfavorable outcome following stroke

Risk factor	Odds ratio	95% CI	<i>P</i> -value
Age	1.02	1.00–1.05	0.07
Gender	0.93	0.54–1.58	0.08
BMI	0.97	0.91–1.04	0.42
Stroke subtype, ischemia/hemorrhage	1.90	0.97–3.75	0.06
NIHSS score	1.24	1.18–1.31	< 0.001*
Alcohol status	0.85	0.62–1.16	0.31
Hypertension	1.21	0.67–2.17	0.53
Dyslipidemia	0.89	0.53–1.48	0.65
Atrial fibrillation	1.05	0.53–2.09	0.89
Congestive heart failure	2.06	0.57–7.46	0.27
CRP	1.29	1.10–1.51	0.002*
<i>P. gingivalis</i> ATCC33277 (fimA type I)	0.73	0.44–1.20	0.22
<i>A. actinomycetemcomitans</i> ATCC29523 (serotype a)	0.63	0.31–1.29	0.20
<i>A. actinomycetemcomitans</i> Y4 (serotype b)	1.04	0.55–1.99	0.90
<i>A. actinomycetemcomitans</i> AUNY67 (serotype c)	1.21	0.62–2.36	0.57
<i>F. nucleatum</i> ATCC25586	1.63	0.87–3.03	0.13
<i>F. nucleatum</i> ATCC10953	3.12	1.55–6.29	0.002*
<i>T. denticola</i> ATCC35405	1.16	0.46–2.93	0.75
<i>C. rectus</i> ATCC33238	1.45	0.80–2.64	0.22
<i>E. corrodens</i> ATCC23834	0.98	0.47–2.06	0.97

*Based on 20 factors included for analysis with *P*-values < 0.2 shown by univariate Fisher's exact, χ^2 or univariate logistic regression test results. **P* < 0.05 (statistically significant in multivariate logistic regression via forced-entry method). CI = confidence interval; BMI = body mass index.

and also reflected in presented guidelines [19,20]. However, whether these factors are associated with unfavorable outcome after stroke remains unknown. Mahmoud *et al.* examined predictors of clinical outcome after 3 months in ischemic stroke patients and found that cardiovascular disease was associated with an unfavorable outcome, while hypertension status was not different between the unfavorable and favorable outcome groups [21]. Additionally, it has been reported that blood pressure management following an acute ischemic stroke is associated with clinical outcome [22]. Regarding smoking, its impact on unfavorable outcome is controversial. Smoking status in stroke patients was reported to be a predictor of good clinical outcome after intravenous thrombolysis (IVT) [23], although those beneficial effects of smoking after IVT and endovascular treatment were later shown to be mainly related to differences in baseline characteristics and the design of that observational study [24,25]. In the present study, neither hypertension nor smoking status had a significant relationship with unfavorable or favorable outcome after stroke occurrence. It seems that consensus has not

been reached regarding the relationship of unfavorable outcome following a stroke and these factors.

P. gingivalis has been identified as one of the main pathogens responsible for progression of periodontitis [26] and its fimbriae are considered to be an important virulence factor [27]. Furthermore, that bacterium has been classified into six genotypes (types I–V, Ib) based on the nucleotide sequence of the *fimA* gene encoding fimbriae [28]. Some investigators have reported that the risk of systemic disease is associated with an increased serum IgG antibody titer to *P. gingivalis*. Seror *et al.* noted that antibody titers for *P. gingivalis* were associated with occurrence of more severe rheumatoid arthritis in non-smokers in a large cohort of patients with early rheumatoid arthritis [29]. Also, Yamazaki *et al.* found a high frequency of detection of the antibody for *P. gingivalis* Su63 (*fimA* type II), but not for FDC381 (*fimA* type I), in coronary heart disease patients [8]. In addition, Nakahara *et al.* reported a significant correlation between fibrosis progression in non-alcoholic fatty liver disease patients and antibody titers for an *fimA* type IV strain of *P. gingivalis* [9]. In the present study, detection of antibody IgG titers for four different strains of *P. gingivalis* (*fimA* type I–V) was not significantly different between the favorable and unfavorable outcome groups. Therefore, the presence of antibodies for *P. gingivalis*, a major periodontal pathogen associated with incidence of various severe systemic diseases, might not necessarily indicate increased risk of an unfavorable outcome following stroke.

F. nucleatum, a strictly anaerobic Gram-negative rod bacterium normally found in the oral cavity [30], is considered to be a periodontal pathogen because it is frequently isolated from lesions, produces high numbers of tissue irritants and often aggregates with other periodontal pathogens as a bridge between early and late colonizers [30,31]. In the present study, detection of serum IgG antibody titers for *F. nucleatum* ATCC 10953 was independently associated with unfavorable outcome following stroke. Sparks *et al.* examined serum IgG antibody levels of seven periodontal pathogens, including *P. gingivalis* and *A. actinomycetemcomitans*, in patients who eventually converted to Alzheimer's disease (AD), and found that the titers of *F. nucleatum* and *P. intermedia* antibodies were significantly increased at baseline in patients who developed Alzheimer's disease compared with the control group [10]. *F. nucleatum* is able to pass through the blood–brain barrier and has been found to be causative of brain abscesses in some case studies [32,33], while it also has abilities to adhere to and invade host vascular endothelial cells via FadA adhesin molecules, as FadA binds to vascular endothelial–cadherin on the cell surface, which triggers breakdown of endothelial cell-to-cell junctions [34]. Additionally, *F. nucleatum*

has been shown to be associated with portal vein thrombosis in some cases and may have inherent thrombogenic properties [35,36]. Chang *et al.* reported unusual cases of cavernous sinus thrombosis associated with *F. nucleatum* and noted that those patients eventually developed ischemic stroke despite undergoing immediate antibiotic therapy [37]. Following passage through the blood–brain barrier, *F. nucleatum* organisms attack vascular endothelial cells in blood vessels in the brain, which can induce endothelial permeability via loosened cell junctions. Also, *F. nucleatum* is related to an unusual form of thrombosis, which may lead to rebleeding after hemorrhage and insufficient recanalization after infarction, thus promoting an unfavorable prognosis following onset in affected patients.

The virulence of *F. nucleatum* is mediated by endotoxin activity, hemagglutination, co-aggregation and outer membrane proteins, such as FAP-2 and RadD [38,39], while it has also been reported to be induced by death of immune cells [40]. Conversely, this organism causes increased expression of genes associated with host immune responses, such as inflammatory chemokines and cytokines [41]. Some investigators have reported that the virulence and impact of the host immune response differs among *F. nucleatum* strains. Kugan *et al.* examined various strains, including ATCC 25586 and ATCC 10953, with regard to apoptosis, phagocytosis, superoxide generation and proinflammatory cytokine release by neutrophils, and found an *F. nucleatum* strain-specific impact on neutrophil function [42]. Conversely, Bhattacharyya *et al.* noted that expression of mRNA of β -defensin-2 (hBD-2), an anti-microbial peptide, in oral epithelial cells was induced to a greater degree by live cells or cell walls of *F. nucleatum* strain ATCC 25586 compared to those of ATCC 10953, and that differences regarding the amino acid residue of Fad-1 in those strains had effects on regulation of hBD-2 expression [43]. In the present study, antibody titers for *F. nucleatum* ATCC 25586 and ATCC 10953 were significantly greater in the unfavorable group following stroke in univariate analysis, while multivariate analysis revealed that detection of the serum IgG antibody to *F. nucleatum* ATCC 10953 was an independent predictor of an unfavorable outcome. Differences regarding virulence and host immune responses among *F. nucleatum* strains may have varying impacts on unfavorable outcome after stroke.

This study has some limitations. First, we could not accurately evaluate the presence of oral periodontal bacteria in blood, as those organisms are often not detected because an inadequate number of certain bacterial types, such as anaerobic periodontal bacteria, remain alive during blood culture examinations [44]. Secondly, we were not able to evaluate the oral periodontal examination results in a comprehensive manner using

indicators such as probing pocket depth. A third limitation is that IgG antibody titer may reflect either current or previous exposure of the host to periodontal pathogens, although previous reports have noted relationships of serum IgG antibody titers of periodontal pathogens with the severity of periodontitis and deep pockets [45]. Furthermore, periodontal treatments decrease serum IgG antibody titers of periodontal pathogens [45,46]; thus, titer levels seem to reflect the current disease status of individuals with chronic periodontitis. Finally, the present results are limited by use of multiple comparisons. For this study, 20 factors regarding unfavorable outcome after stroke shown to have a *P*-value < 0.20 in univariate analysis were subjected to multivariate logistic regression analysis. Although it is possible that such analysis can be affected by chance when several variable factors are examined together, we found that an unfavorable outcome after stroke was associated with detection of a serum IgG antibody titer to *F. nucleatum* ATCC 10953, as well as NIHSS score and CRP.

In summary, this study is the first to demonstrate that detection of a serum IgG antibody titer to *F. nucleatum* ATCC 10953 is independently associated with unfavorable outcome after stroke. We consider that such detection following stroke occurrence may be useful for predicting patient outcome.

Acknowledgements

We appreciate the expert advice from Tsuyoshi Miyagawa (Center for Integrated Medical Research, Hiroshima University Hospital, Japan). This work was supported by a Grant-in-aid for young scientists (B) (No. 17K17350, 17K17907) and a Grant-in-aid for Scientific Research (C) (No. 15K08615, 17K11839, 18K10746) from the Japanese Ministry of Education, Culture, Sports, and Technology.

Disclosures

The authors have no potential conflicts of interest to declare with respect to authorship and/or publication of this article.

Author contributions

H. N., N. H., K. O. and H. K. conceived the study and designed the protocol. H. N., K. O., T. S., T. O. and M. S. performed the experiments. H. N., N. H., H. S. and S. A. analyzed the data obtained. K. I., M. N., T. N., N. K. and Y. S. collected samples. H. O., H. M., H. K. and H. K. supervised the project. H. N., N. H. and K. O. wrote the manuscript. All authors have reviewed the final version of the manuscript and approved its submission for publishing.

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