

## Neuroendocrine, epigenetic, and intergenerational effects of general anesthetics

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**Author contributions:** The authors conducted literature review and analysis, drafted and critically revised the manuscript, and gave final approval.

**Supported by** National Institutes of Health, No. R01NS091542; National Natural Science Foundation of China, No. 81771149, No. U1704165.

**Conflict-of-interest statement:** No potential conflicts of interest.

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**Manuscript source:** Invited manuscript

**Received:** December 24, 2019

**Peer-review started:** December 24, 2019

**First decision:** February 20, 2020

**Revised:** March 18, 2020

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### Abstract

The progress of modern medicine would be impossible without the use of general anesthetics (GAs). Despite advancements in refining anesthesia approaches, the effects of GAs are not fully reversible upon GA withdrawal. Neurocognitive deficiencies attributed to GA exposure may persist in neonates or endure for weeks to years in the elderly. Human studies on the mechanisms of the long-term adverse effects of GAs are needed to improve the safety of general anesthesia but they are hampered not only by ethical limitations specific to human research, but also by a lack of specific biological markers that can be used in human studies to safely and objectively study such effects. The latter can primarily be attributed to an insufficient understanding of the full range of the biological effects induced by GAs and the molecular mechanisms mediating such effects even in rodents, which are far more extensively studied than any other species. Our most recent experimental findings in rodents suggest that GAs may adversely affect many more people than is currently anticipated. Specifically, we have shown that anesthesia with the commonly used GA sevoflurane induces in exposed animals not only neuroendocrine abnormalities (somatic effects), but also epigenetic reprogramming of germ cells (germ cell effects). The latter may pass the neurobehavioral effects of parental sevoflurane exposure to the offspring, who may be affected even at levels of anesthesia that are not harmful to the exposed parents. The large number of patients who require general anesthesia, the even larger number of their future unexposed offspring whose health may be affected, and a growing number of neurodevelopmental disorders of unknown etiology underscore the translational importance of investigating the intergenerational effects of GAs. In this mini review, we discuss emerging

**Accepted:** March 25, 2020  
**Article in press:** March 25, 2020  
**Published online:** May 19, 2020

**P-Reviewer:** Hosak L, Ishizawa K, Shiina A

**S-Editor:** Dou Y

**L-Editor:** A

**E-Editor:** Liu JH



experimental findings on neuroendocrine, epigenetic, and intergenerational effects of GAs.

**Key words:** Brain; General anesthetic; Sevoflurane; Corticosterone; Cortisol; Histone acetylation; Deoxyribonucleic acid methylation; Intergenerational effects; Gamma aminobutyric acid

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**Core tip:** The GABAergic general anesthetics may act as stressors and endocrine disruptors in neonates and young adults. They may induce two distinct types of long-term adverse effects: Neuroendocrine effects (the somatic effects) and epigenetic reprogramming of germ cells (the germ cell effects). The latter may pass neurobehavioral abnormalities to male offspring. Compared to the somatic cells, the germ cells may be more sensitive to the deleterious effects of general anesthetics, raising the possibility that the offspring may be affected even when levels of anesthesia are not harmful to the exposed parents. Further rigorous experimental testing of all these possibilities is required.

**Citation:** Martynyuk AE, Ju LS, Morey TE, Zhang JQ. Neuroendocrine, epigenetic, and intergenerational effects of general anesthetics. *World J Psychiatr* 2020; 10(5): 81-94

**URL:** <https://www.wjgnet.com/2220-3206/full/v10/i5/81.htm>

**DOI:** <https://dx.doi.org/10.5498/wjp.v10.i5.81>

## INTRODUCTION

The number of surgeries performed globally has rapidly increased from 226.4 million in 2004 to 312.9 million in 2012, according to 2016 World Health Organization estimates<sup>[1]</sup>. Most of these surgeries and many non-surgical procedures require general anesthesia, which can be viewed as a state of pharmacologically induced “reversible brain coma”<sup>[2]</sup>. Despite complete reversibility of the primary effect of general anesthetics (GAs), *i.e.*, induction of the general anesthesia state, many studies in humans and almost all studies in laboratory animals provide evidence that GAs may leave persistent footprints of their brief presence in the body, *i.e.*, anesthetic exposure may lead to long-lasting functional abnormalities<sup>[3-7]</sup>. Investigations of anesthesia-induced abnormalities are currently restricted primarily to evaluating neurocognitive function in the exposed subjects<sup>[8-12]</sup>, although it is biologically plausible that GAs may affect other functions/systems because their actions are not limited to neuronal effects involved in the mediation of a general anesthesia state. Still, as a result of these studies, the long-term adverse effects of GAs are an increasingly recognized health concern, especially in the very young and elderly<sup>[3-7]</sup>. The adverse effects of GAs during the perinatal period are of special concern<sup>[13-17]</sup> because mounting evidence indicates that at this stage of life the central nervous system and other body systems are highly susceptible to reprogramming by environmental factors/stressors<sup>[18-21]</sup>. Such environmental factors may include GAs, given the multiple molecular targets known to mediate their actions in the brain and throughout the body<sup>[22-25]</sup>. In support of this contention are reports of learning disabilities, long-term memory impairment, and attention-deficit hyperactivity disorders in patients who had anesthesia early in life<sup>[3,5,11,26,27]</sup>. Although several recent studies have not found negative neurocognitive consequences of relatively short ( $\leq 1$  h) anesthesia exposures in children<sup>[6,7,28]</sup>, both clinical and laboratory studies, including the most recent clinical assessments<sup>[29]</sup>, agree that prolonged or repeated exposures to GAs that are frequently required for very sick children may result in significant neurocognitive abnormalities later in life<sup>[30]</sup>. Furthermore, a recent report of the effects on brain development of relatively short anesthesia exposure for cesarean delivery<sup>[31]</sup> further confirms that the current understanding of this phenomenon remains in an early stage.

Because of a widely accepted dogma that the brain is most susceptible to the deleterious effects of environmental stressors at the extreme of ages, investigations of the long-term adverse effects of GAs in young adults are scarce even in animal models. Several studies have assessed the effects of GAs in young adult rats,

primarily using rats of this age as comparisons to other age groups<sup>[4,32-34]</sup>. Aside from the fact that these studies found long-term effects of isoflurane in young adult rats, different isoflurane concentrations and exposure regimens make it difficult to compare the effects across these studies. Clearly, further research is needed to elucidate the full range of long-term effects of GAs in young adults. Importantly, the germ cells, which pass the genetic and epigenetic information from parents to offspring, can be susceptible to epigenetic reprogramming by environmental factors throughout the lifespan<sup>[35-39]</sup>.

Laboratory and clinical studies provide evidence that alcohol, stress, endocrine disruptors, obesity, and even physical exercise may affect embryonic development and the phenotype of the offspring<sup>[40-44]</sup>. GAs share many molecular mechanisms of action with alcohol<sup>[45-49]</sup> and may act as endocrine disruptors and environmental stressors in animal models and humans. The spectrum of molecular actions of GAs and susceptibility of germ cells to epigenetic reprogramming by environmental factors across the lifespan support the possibility that the offspring may be affected by parental exposure to GAs regardless of the parental age at the time of exposure to GAs. In this mini review, we discuss emerging experimental findings on the neuroendocrine, epigenetic, and intergenerational effects of GAs.

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## NEUROENDOCRINE, EPIGENETIC, AND SECOND-GENERATION EFFECTS OF GENERAL ANESTHETICS

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Research studies on the epigenetic multigenerational effects of environmental factors, such as alcohol, stress, endocrine disruptors, and others, have changed our thinking about the susceptibility of somatic and germ cells to alterations by environmental factors and the persistence of such alterations not only across the lifespan, but also through generations<sup>[36-38,40,42]</sup>. The potential of GAs to induce similar epigenetic effects, including reprogramming of the germ cell epigenome, and by extension intergenerational effects is supported by the notion that GAs share many molecular mechanisms of action with alcohol and may act as endocrine disruptors and environmental stressors in animal models and humans<sup>[45-49]</sup>.

Endocrine disruptors can broadly be defined as agents that interfere with the functioning of the endocrine system. In support of the neuroendocrine effects of GAs are reports of significant rises in cortisol levels in pediatric patients after surgery or after anesthesia without surgery in healthy children<sup>[50,51]</sup>. Also, measurements of salivary cortisol levels in response to different levels of sedation in healthy children found a more than threefold increase in cortisol values, with the highest cortisol levels during the recovery phase<sup>[52]</sup>. Adult patients who received isoflurane-based tracheal general anesthesia compared to those who received bupivacaine-based epidural anesthesia had more than two times higher plasma levels of cortisol at the end of surgery, which was also more than four times higher compared to baseline levels in the same patients<sup>[53]</sup>. For a recent comprehensive review on cortisol levels associated with anesthesia/surgery see<sup>[54]</sup>. Importantly, our findings in rodents support the notion that GABAergic anesthetics act *via* specific molecular mechanisms to induce stress-like responses, rather than that GA-caused increases in glucocorticoid levels are the result of the systemic stress response because of uncontrolled physiological parameters during anesthesia. Thus, in neonatal rats, the gamma aminobutyric acid (GABA) type A receptor (GABA<sub>A</sub>R) antagonist bicuculline at a low dose (0.01 mg/kg) or aromatase inhibitor formestane, administered prior to anesthesia with sevoflurane, prevented the sevoflurane-increased corticosterone secretion without an obvious effect on the sedation depth induced by the anesthetic (unpublished observations).

The inhibitory control of the corticotropin-releasing hormone-secreting hypothalamic paraventricular neurons by GABA<sub>A</sub>R-mediated signaling and the positive modulation of this signaling by neuroactive steroids is one of the fundamental mechanisms of downregulating the stress response<sup>[55,56]</sup>. Due to relatively high and low expressions of the Cl<sup>-</sup> transporters Na<sup>+</sup>-K<sup>+</sup>-2Cl<sup>-</sup> (NKCC1) and K<sup>+</sup>-2Cl<sup>-</sup> (KCC2), respectively, immature neurons have elevated intracellular concentrations of Cl<sup>-</sup>, the main charge carriers through GABA<sub>A</sub>R channels<sup>[57-61]</sup>, a major substrate for the otherwise inhibitory effects of GABAergic anesthetics<sup>[22-25]</sup>. Activation of GABA<sub>A</sub>Rs in immature neurons causes Cl<sup>-</sup> efflux, membrane depolarization, activation of the voltage-gated Ca<sup>++</sup> channels, and relief of the Mg<sup>++</sup>-block of Ca<sup>++</sup> permeable N-methyl-D-aspartate receptors<sup>[58-62]</sup>. The GABA-initiated Ca<sup>++</sup> influxes regulate a wide spectrum of developmental processes from gene expression to synapse formation<sup>[61,62]</sup>. During the second postnatal week, GABA<sub>A</sub>R-mediated signaling in the brain undergoes a fundamental transition from predominantly stimulating/excitatory to inhibitory, which is caused by a concomitant developmental downregulation of NKCC1 and,

most importantly, upregulation of neuronal-specific KCC2<sup>[61,62]</sup>. It is plausible that in the neonatal brain such GABA<sub>A</sub>R inhibitory signaling-mediated control of the stress response system is weakened or GABAergic anesthetics may even stimulate the stress response through positive modulation of depolarizing/excitatory GABA<sub>A</sub>R signaling at this age.

Consistent with stressor-like effects of GABAergic anesthetics, we found that a single exposure of neonatal rats to the GABAergic GAs sevoflurane or propofol was sufficient to cause multifold increases in corticosterone secretion and electroencephalography-detectable seizures at the time of anesthesia<sup>[63-67]</sup>. The anesthetic-caused increases in excitatory GABA<sub>A</sub>R signaling and corticosterone levels may be required for neonatal GABAergic anesthetic-induced seizures to occur<sup>[68,69]</sup>. Importantly, a single exposure to sevoflurane or propofol early in life induced neuroendocrine abnormalities<sup>[64,66,70-73]</sup> similar to those induced by repeated, but not single, maternal separations, a widely used rodent model of developmental effects of early-life stress in humans<sup>[74-78]</sup>. The GABAergic anesthetic-induced long-term neuroendocrine abnormalities, which were more robust in males, included increased anxiety-like behavior and exacerbated corticosterone responses to stress<sup>[64-67]</sup>. In addition, the rats, neonatally exposed to anesthesia, had elevated *Crh* mRNA levels in the hypothalamus, as well as up- and downregulated hypothalamic and hippocampal mRNA levels of *Nkcc1* and *Kcc2*, respectively<sup>[66,67,73]</sup>. Notably, delays in the GABA<sub>A</sub>R signaling transition to inhibitory have been linked in animal models and humans to a number of cognitive neuropsychiatric disorders, such as schizophrenia, autism spectrum disorder (ASD), and Rett syndrome<sup>[79-85]</sup>. For example, in Rett syndrome, a severe form of ASD, the methyl CpG binding protein 2 deficiency-induced KCC2 downregulation may play an important causal role<sup>[84,85]</sup>.

When used at a low dose, bumetanide, a loop diuretic, is the most selective of the available inhibitors of NKCC1 activity<sup>[86,87]</sup>. Our lab and others have demonstrated that pretreatment of neonatal rats with bumetanide prior to anesthetic exposure ameliorated many of the acute and lasting developmental effects of GABAergic anesthetics, including: (1) Seizures; (2) Downregulated *Kcc2* levels; (3) Elevated levels of *Crh* mRNA; (4) Exacerbated corticosterone responses to acute stress; and (5) Behavioral abnormalities<sup>[63-66,88]</sup>. Bumetanide's ameliorating effects suggest that anesthetic-exacerbated GABA<sub>A</sub>R-mediated stimulation/excitation in the neonatal rodent brain is an initial step in anesthetic-induced developmental abnormalities. Importantly, bumetanide exhibits promising therapeutic effects against ASD, schizophrenia, and Fragile X syndrome in animal models and humans<sup>[89-93]</sup>, suggesting that these diseases and the GABAergic anesthetic-induced abnormalities may share similar mediating mechanisms. Our current understanding of rodent and human ontogeny supports the possibility that similar GA-sensitive mechanisms operate in rodents and humans early in life. Based on the intensity of synaptogenesis, a 1-wk-old rat can be compared to a 2- to 3-year-old human<sup>[94-98]</sup>. In animals, neonatal anesthetic exposure<sup>[63,65,72,99,100]</sup> and early-life stress<sup>[101-103]</sup> have profound long-term effects on synaptic morphology and function, suggesting that humans are vulnerable well into the postnatal period. Similar to the rodent brain during the first 2 postnatal weeks, the human brain is more excitable during the first year of life than at any other time, with seizures occurring in 3.5 per 1000 live births<sup>[104-106]</sup>. Human neonatal seizures are resistant to GABAergic antiepileptic drugs (AEDs) because of depolarizing/excitatory GABA<sub>A</sub>R signaling at this age<sup>[107,108]</sup>. Many human neonatal seizures can be detected only through electroencephalographies because they are not accompanied by clinical signs such as convulsions<sup>[109]</sup>, which helps explain why epileptic seizures are not routinely reported in anesthetized human infants. In neonatal seizures with clinical manifestations, GABA<sub>A</sub>R-enhancing AEDs depress convulsions but may exacerbate electrographic cortical seizure activity (electroclinical uncoupling)<sup>[110]</sup>. The NKCC1 inhibitor bumetanide, administered alone or in combination with GABAergic AEDs, may alleviate neonatal seizures in rodents and humans<sup>[111,112]</sup>. In humans, the KCC2 protein levels at birth are only about 20% of adult levels and significantly increase during the first postnatal year<sup>[86]</sup>. This late KCC2 increase during brain development may make KCC2 a highly susceptible molecular target for modulation by environmental factors, including GAs.

The heightened corticosterone responses to acute stress months after exposure to the GABAergic anesthetics early in life suggest that stressors in post-anesthesia life can further exacerbate developmental abnormalities, initially programmed by GABAergic anesthetics. This concept is supported by our findings that adult rats exposed neonatally for a relatively short time to anesthesia with etomidate or sevoflurane followed by a subsequent single episode of maternal separation exhibited developmental abnormalities significantly greater than those exposed to only one of the two interventions<sup>[66,73,113]</sup>. In further support of the idea that the long-term adverse outcomes of early-life anesthesia may result from a combination of the effects of GAs

at the time of anesthesia and the effects of “post-anesthesia” environmental factors, several laboratories have demonstrated that the adverse developmental effects of neonatal anesthesia in rodents may be alleviated, not only by pharmacological interventions before exposure to anesthesia, but also by post-weaning housing of the exposed animals in an enriched environment<sup>[114-116]</sup>. Environmental factors may alleviate or exacerbate the effects of neonatal anesthesia. In other words, two subjects exposed to the same anesthesia regimen may have different long-term outcomes based on post-anesthesia life experiences. In support of this possibility, Zhang *et al*<sup>[117]</sup> reported that rats neonatally exposed to sevoflurane and then housed from the time of weaning in isolation in enrichment-deprived environments exhibited reduced levels of brain-derived neurotrophic factor (BDNF), synaptic protein markers, and survival of new granule cells in the hippocampus, as well as behavioral abnormalities. On the other hand, rats that were exposed to the same regimen of anesthesia with sevoflurane and housed in groups in enriched environments had the same outcomes as their counterparts who were not exposed to sevoflurane. These findings further support the possibility that GABAergic anesthetics administered during early life can be considered environmental stressors that predispose the exposed subjects to stress vulnerability later in life. Identifying environmental factors that predispose humans to abnormal stress reactivity later in life and the mechanisms underlying their effects is of important clinical and basic neuroscience concern because dysregulated stress response systems have been linked to the pathophysiology of several neuropsychiatric disorders<sup>[118]</sup>.

Environmental enrichment, which may alleviate the neurodevelopmental effects of early-life exposure to GAs in rodents, is not an issue for the majority of human patients because this is typical for most children. If similar mechanisms operate in humans, then brain development in healthy human patients who experience normal stress levels may be minimally affected, if at all, after exposure to general anesthesia early in life. However, most children who require general anesthesia during the early postnatal period inevitably experience a variety of stressors during life post-anesthesia exposure (*e.g.*, diseases, pain, hunger, psychological stress). Such patients may be at risk of developing early-life, anesthetic-programmed neuroendocrine and neurocognitive abnormalities. The exacerbating effects of environmental stressors on long-term adverse outcomes of early-life exposure to GAs may be why several recent studies have not found negative neurocognitive consequences of relatively short ( $\leq 1$  h) anesthesia exposures in healthy children<sup>[6,7,28]</sup>, while studies in very sick children have<sup>[29]</sup>. It will be important to take this factor into consideration when planning new clinical studies. Further investigation of the interaction of adverse effects of early-life exposure to GAs and post-anesthesia stressors is important because it may identify not only the most vulnerable patients, but also those who are at diminished risk and would not benefit from delaying needed anesthesia-required interventions.

Histone acetylation and DNA methylation are important epigenetic mechanisms whereby environmental factors, in particular stress, affect brain development and function<sup>[119-121]</sup>. Histone acetylation facilitates gene transcription by enabling chromatin relaxation, while histone deacetylation results in stronger histone interaction with DNA, more compact chromatin structure, and repression of gene transcription. Histone acetylation is regulated by adding and removing acetyl groups to the N-terminal of histone tails by acetyltransferases and histone deacetylases, respectively<sup>[122-126]</sup>. Jia *et al*<sup>[99]</sup> found that repeated exposure of neonatal rats to sevoflurane led to increased levels of histone deacetylases 3 and 8 and reduced levels of acetylated histones H3 and H4 in the hippocampus. The sevoflurane-exposed rats had lower hippocampal density of dendritic spines and synaptic protein markers and exhibited impaired hippocampus function-based behavior. These effects of sevoflurane were alleviated by treatments with the histone deacetylase inhibitor sodium butyrate, suggesting a potential role of histone acetylation as one of the mediators of the developmental effects of GAs. The role of histone acetylation in GA-induced abnormalities in neonatal rodents has also been reported by other laboratories<sup>[127,128]</sup>. Furthermore, histone acetylation may be involved in the mediation of learning and memory dysfunction in offspring after pregnant rats exposed to sevoflurane, isoflurane, or propofol<sup>[129-131]</sup>.

Ju *et al*<sup>[100]</sup> have found that repeated exposure of neonatal rats to sevoflurane resulted in increased expression of hippocampal DNA methyltransferases 3A/B (DNMT3A/B), but not DNA methyltransferase 1 (DNMT1). These enzymes catalyze DNA methylation at the 5' position of cytosine residues adjacent to guanines (CpG sites), typically leading to long-term transcriptional repression. The activity of DNMT1 is responsible for maintenance of the remaining 5mC marks during cell divisions because of DNMT1 selectivity to hemi-methylated DNA, while DNMT3A/B are *de novo* DNMTs, which are induced by internal and external (environmental) stimuli<sup>[132,133]</sup>. In addition to increased expression of DNMT3A/B, the rats neonatally

exposed to sevoflurane had downregulated methyl CpG binding protein 2, hypermethylated the *Bdnf* gene, downregulated BDNF levels, and exhibited behavioral deficiencies<sup>[100]</sup>. The ability of sevoflurane to induce these abnormalities was significantly diminished in rats that were pretreated with a non-selective DNMT inhibitor, 5-aza-2'-deoxycytidin, prior to sevoflurane exposure<sup>[100]</sup>. On the other hand, Wu *et al.*<sup>[134]</sup> found that exposure of neonatal rats to isoflurane, another inhaled anesthetic that has positive modulation of GABA<sub>A</sub>R activity like sevoflurane, induced a significant increase in the expression of hippocampal DNMT1. More experimental studies are needed before we can draw conclusions on the specific epigenetic mechanisms involved in the mediation of the adverse effects of a given GA or a class of GAs.

It has been established in a wide range of species, including humans, that environmental factors, in particular stress, acting *via* epigenetic mechanisms may affect not only the exposed subjects, but also future generations<sup>[135-139]</sup>. In contrast to studies of famine or war survivors, in which large groups of people in relatively compact living areas within a specific time period were affected<sup>[135,137,140]</sup>, it is not trivial to link someone's neurodevelopmental abnormalities to his/her parent's relatively short exposure to GAs. Still, the emerging laboratory and clinical data demonstrate that the adverse effects of GAs may include epigenetic modifications, not only in the somatic cells of the exposed subjects, but also in their germ cell epigenome. Of most direct support of the possibility of epigenetic germ cell effects of general anesthesia in humans, Donkin *et al.*<sup>[141]</sup> found that the DNA methylation status of 1509 genes in the sperm of male patients was changed 1 wk after bariatric surgery. The findings of epigenetic changes in spermatozoa just 1 wk after anesthesia/surgery suggest that the anesthesia/surgery may be an important cause of such changes and that even the mature human sperm is susceptible to epigenetic reprogramming by environmental factors. The latter is supported by the presence of DNMTs in mature human sperm<sup>[142]</sup>. Importantly, of the 1509 genes altered at 1 wk after the anesthesia/surgery, 1004 genes remained altered 1 year later<sup>[141]</sup>. Alarming, several small pilot clinical assessments found that anesthesia care providers may have altered female/male offspring ratios, also suggesting that persistent exposure to traces of the escaped GAs from scavenging in operating rooms might affect germ cells, and, hence, the next generation(s)<sup>[143-147]</sup>.

To test whether neonatal exposure to anesthesia with sevoflurane can affect not only the exposed animals, but also their future offspring, we exposed male and female postnatal day (P) 5 rats (generation F0) to 6 h anesthesia with 2.1% sevoflurane<sup>[67]</sup>. On P90, the exposed and control rats were used as breeders to produce the second generation of rats (generation F1). We have found that adult offspring of parents who were neonatally exposed to sevoflurane exhibited neurobehavioral abnormalities. Irrespective of whether sires, dams, or both parents were exposed to neonatal sevoflurane, only F1 males, but not F1 females, were affected. F1 males exhibited reduced *Kcc2* Cl<sup>-</sup> exporter expression and behavioral abnormalities. Bisulfate sequencing revealed CpG dinucleotide hypermethylation in the *Kcc2* promoter in the F0 sperm and ovary and in the hypothalamus and hippocampus of F1 males<sup>[67]</sup>. The correlation of impaired hippocampal *Kcc2* expression and hippocampus-dependent behavior in F1 males points to the involvement of epigenetic *Kcc2* modulation in the mediation of the intergenerational effects of sevoflurane<sup>[67]</sup>. Because DNA methylation is often associated with transcriptional repression, these data suggest that methylation of the *Kcc2* promoter in F0 gametes may contribute to sevoflurane-induced intergenerational impairment in *Kcc2* expression. The role of DNA methylation in the intergenerational effects of neonatal sevoflurane exposure in rats was also reported by Chastain-Potts *et al.*<sup>[148]</sup>. Surprisingly, despite similar, but not identical, neurobehavioral abnormalities in the exposed parents and their male offspring, including downregulation of the *Kcc2* expression, the male offspring, in contrast to their exposed parents, exhibited corticosterone responses to stress that were the same as the corticosterone responses to stress in the male offspring of control parents<sup>[67]</sup>. These findings, taken together with the effects of GABAergic anesthetics in the exposed animals, suggest that GA-induced modulation of GABA<sub>A</sub>R signaling through reduced expression of *Kcc2* may be required but not sufficient to induce exacerbated responses to stress in adulthood.

Importantly, stress-like effects of GABAergic anesthetics may not be limited to the early postnatal period. Thus, exposure of young adult rats to sevoflurane on 3 alternating days starting on P56 resulted in similar increases in serum levels of corticosterone in male and female rats 1 h after the last exposure to the anesthetic<sup>[149]</sup>. However, long term, the exposed female rats were the same as the controls; they were not affected. On the other hand, contrary to the currently generally accepted view and our initial hypothesis that adult rats are resilient to the long-term adverse effects of GAs, the exposed young adult male rats developed neuroendocrine and behavioral

abnormalities. More than 3 months after exposure to sevoflurane, they exhibited not only an exacerbated hypothalamic pituitary adrenal axis response to stress, but their serum levels of luteinizing and testosterone hormones were also significantly increased, as was their expression of the hypothalamic gonadotropin-releasing hormone gene. The hypothalamic pituitary testicular axis functioning in the exposed adult male rats was also altered at the level of expressions of the hypothalamic aromatase and estrogen receptor  $\alpha$  and  $\beta$  genes. The expressions of the aromatase and estrogen receptor  $\alpha$  genes were significantly increased, while the expression of the estrogen receptor  $\beta$  gene was slightly, but significantly, decreased<sup>[149]</sup>. These findings demonstrate that anesthesia with sevoflurane alters (disrupts) not only the functioning of the hypothalamic pituitary adrenal axis, but also the hypothalamic pituitary testicular axis functioning<sup>[53]</sup>. We have previously demonstrated the role of estradiol in the acute adverse effects of sevoflurane in neonatal rats<sup>[69]</sup>.

Similar to the effects of neonatal exposure to sevoflurane, sevoflurane administered to young adult male rats induced significant impairment in expressions of the hypothalamic and hippocampal *Kcc2* genes. Similar to adult rats that were neonatally exposed to sevoflurane, male and female rats exposed to sevoflurane in young adulthood had hypermethylated *Kcc2* gene in spermatozoa and ovarian tissue, respectively<sup>[149]</sup>. Interestingly, we have analyzed the same CpG sites in the *Kcc2* promoter region in the germ cells of rats that were exposed to sevoflurane neonatally or in young adulthood and found similar changes in their methylation regardless of the age of sevoflurane exposure<sup>[67,149]</sup>. These findings suggest that sevoflurane may induce germ cell effects *via* similar mediating mechanisms when administered to rats over a wide range of ages, from neonates to young adults. The similarities between the effects of neonatal and young adult exposure to sevoflurane were also evident in the offspring of the exposed parents. Thus, the *Kcc2* gene was hypermethylated and exhibited reduced expression in the hypothalamus and hippocampus of the F1 male, but not female, offspring of the exposed parents. These changes in the *Kcc2* gene in the hypothalamus and hippocampus of adult F1 male offspring were accompanied by behavioral deficiencies in the elevated plus maze and prepulse inhibition (PPI) of acoustic startle response tests, but their corticosterone responses to stress were not different from the controls<sup>[67,149]</sup>. The sevoflurane-exposed young adult male and female rats had similar acute increases in serum corticosterone levels and changes in DNA methylation status of the *Kcc2* gene in spermatozoa and ovarian tissue and passed neurobehavioral abnormalities to their male offspring, despite the finding that the exposed dams lacked the somatic effects<sup>[149]</sup>. These findings allow us to hypothesize that sevoflurane-induced corticosterone secretion at the time of anesthesia is involved in the anesthetic-induced germ cell effects and by extension the intergenerational effects. Also, the findings that the exposed but long-term physiologically unaffected dams, similar to the exposed and affected sires, pass deleterious effects of sevoflurane to their unexposed male offspring suggest that compared to the somatic cells, the germ cells are more sensitive to the deleterious effects of sevoflurane. This raises the possibility that male offspring may be affected even when the anesthesia level/duration is insufficient to induce significant abnormalities in their exposed parents. Future studies will be needed to test these hypotheses that may have important translational applicability.

We have tested the role of the neuron-specific *Kcc2* gene as a mediator of the germ cell and intergenerational effects of sevoflurane only because we have extensive background data supporting its involvement in the mediation of the somatic effects of neonatal and young adult exposure to sevoflurane in the exposed animals<sup>[64-67,149]</sup>. The findings that the *Kcc2* gene was affected in the parental somatic (brain) cells and germ cells, two effects that may not necessarily have similar mediating mechanisms, was surprising, on the one hand, but on the other hand suggested that the *Kcc2* gene could be one of many genes involved in passing on the intergenerational effects of the anesthetic. Indeed, using genome-wide reduced representation bisulfite sequencing, we found more than 2000 differentially methylated DNA regions in the sperm of adult rats neonatally exposed to sevoflurane compared to unexposed controls (unpublished observations). Future detailed investigation of various genes that are involved in the mediation of the intergenerational effects of sevoflurane and other GAs may help to identify a full spectrum of the biological intergenerational effects of GAs and may lead to new, unexpected parental GA-induced phenotypes in offspring. It is likely that the intergenerational effects of exposure to GAs in young adulthood are not limited to sevoflurane, as Tang *et al.*<sup>[150]</sup> have shown behavioral abnormalities in the offspring of mice that were exposed to enflurane at 11 wk of age.

The weakness of most rodent behavioral paradigms is that they are difficult to directly replicate in humans. The results of our studies demonstrate that the PPI of startle was impaired in F0 rats exposed to sevoflurane as neonates or in young adulthood, as well as in their F1 male offspring<sup>[64-67,149]</sup>. Also, male and female rats,

exposed to sevoflurane neonatally, and male rats exposed to sevoflurane in young adulthood, exhibited heightened corticosterone responses to stress long term<sup>[64-67,149]</sup>. In humans, PPI can be measured safely *via* changes in the eyeblink reflex using nearly identical parameters as in rodents<sup>[151-155]</sup>, while heightened stress-related cortisol levels can be readily measured in saliva<sup>[156,157]</sup>. If GAs induce similar effects in humans, the PPI of startle and saliva levels of cortisol may be used as objective, translatable, and easily and safely measurable biological markers of the adverse effects of GAs in humans. Identification of such a biomarker(s) will facilitate investigation of the underlying mechanisms of the adverse effects of GAs in humans to guide development of safer anesthetic approaches. Of particular relevance, one of the effective, tested therapeutic agents to alleviate the developmental effects of GABAergic anesthetics in rodent models, the NKCC1 inhibitor bumetanide<sup>[63-66,88]</sup>, is approved for the treatment of various pediatric conditions and exhibits promising therapeutic effects in human studies of neurodevelopmental disorders<sup>[89-93]</sup>.

Considering relatively well-studied molecular targets for GA actions and emerging evidence of similarities between the adverse outcomes of exposure to GABAergic anesthetics and psychiatric disorders, understanding the molecular mechanisms of the adverse effects of GAs may help to elucidate the mechanistic basis and etiology of complex neurodevelopmental disorders.

## CONCLUSION

The results of recent studies raise many intriguing questions related to the types of adverse effects of GABAergic GAs and their underlying mechanisms, the answers to which may have important translational applicability for establishing safer general anesthesia, in particular, and for better understanding of the nature of neuropsychiatric disorders, in general. They suggest that GABAergic GAs, in particular sevoflurane, may act as stressors and endocrine disruptors in neonates and young adults. These stress-like effects of GABAergic GAs may be involved in the mediation of two distinct types of long-term adverse effects of GAs in the exposed rodents: neuroendocrine effects (the somatic effects) and epigenetic reprogramming of their germ cells (the germ cell effects). The latter may pass neurobehavioral abnormalities to male offspring. The intergenerational effects of sevoflurane are similar, but not identical, when administered to neonatal and young adult rats, suggesting that similar mediating mechanisms are involved over a wide range of ages at the time of anesthesia. The initial data suggest that compared to the somatic cells, the germ cells are more sensitive to the deleterious effects of sevoflurane, raising the possibility that the offspring may be affected even when levels of anesthesia are not harmful to the exposed parents. The long-term adverse effects of GAs in the exposed young adult male rats suggest that current views on the window of vulnerability to the adverse effects of GAs in rodents (up to the first 2 postnatal weeks)<sup>[158,159]</sup>, and, hence, the United States Food and Drug Administration recommendations to avoid GAs in children younger than 3<sup>[160]</sup>, may need to be reconsidered to include more advanced ages. All of these possibilities may have important translational applicability if confirmed; further rigorous experimental testing is required.

## REFERENCES

- 1 Weiser TG, Haynes AB, Molina G, Lipsitz SR, Esquivel MM, Uribe-Leitz T, Fu R, Azad T, Chao TE, Berry WR, Gawande AA. Size and distribution of the global volume of surgery in 2012. *Bull World Health Organ* 2016; **94**: 201-209F [PMID: 26966331 DOI: 10.2471/BLT.15.159293]
- 2 Brown EN, Lydic R, Schiff ND. General anesthesia, sleep, and coma. *N Engl J Med* 2010; **363**: 2638-2650 [PMID: 21190458 DOI: 10.1056/NEJMr0808281]
- 3 Vutskits L, Xie Z. Lasting impact of general anaesthesia on the brain: mechanisms and relevance. *Nat Rev Neurosci* 2016; **17**: 705-717 [PMID: 27752068 DOI: 10.1038/nrn.2016.128]
- 4 Crosby C, Culley DJ, Baxter MG, Yukhananov R, Crosby G. Spatial memory performance 2 weeks after general anesthesia in adult rats. *Anesth Analg* 2005; **101**: 1389-1392 [PMID: 16243999 DOI: 10.1213/01.ANE.0000180835.72669.AD]
- 5 Hu D, Flick RP, Zaccariello MJ, Colligan RC, Katusic SK, Schroeder DR, Hanson AC, Buenvenida SL, Gleich SJ, Wilder RT, Sprung J, Warner DO. Association between Exposure of Young Children to Procedures Requiring General Anesthesia and Learning and Behavioral Outcomes in a Population-based Birth Cohort. *Anesthesiology* 2017; **127**: 227-240 [PMID: 28609302 DOI: 10.1097/ALN.0000000000001735]
- 6 Sun LS, Li G, Miller TL, Salorio C, Byrne MW, Bellinger DC, Ing C, Park R, Radcliffe J, Hays SR, DiMaggio CJ, Cooper TJ, Rauh V, Maxwell LG, Youn A, McGowan FX. Association Between a Single General Anesthesia Exposure Before Age 36 Months and Neurocognitive Outcomes in Later Childhood. *JAMA* 2016; **315**: 2312-2320 [PMID: 27272582 DOI: 10.1001/jama.2016.6967]
- 7 Davidson AJ, Disma N, de Graaff JC, Withington DE, Dorris L, Bell G, Stargatt R, Bellinger DC,



- Schuster T, Arnup SJ, Hardy P, Hunt RW, Takagi MJ, Giribaldi G, Hartmann PL, Salvo I, Morton NS, von Ungern Sternberg BS, Locatelli BG, Wilton N, Lynn A, Thomas JJ, Polaner D, Bagshaw O, Szmuk P, Absalom AR, Frawley G, Berde C, Ormond GD, Marmor J, McCann ME; GAS consortium. Neurodevelopmental outcome at 2 years of age after general anaesthesia and awake-regional anaesthesia in infancy (GAS): an international multicentre, randomised controlled trial. *Lancet* 2016; **387**: 239-250 [PMID: 26507180 DOI: 10.1016/S0140-6736(15)00608-X]
- 8 **Graham MR**, Brownell M, Chateau DG, Dragan RD, Burchill C, Fransoo RR. Neurodevelopmental Assessment in Kindergarten in Children Exposed to General Anesthesia before the Age of 4 Years: A Retrospective Matched Cohort Study. *Anesthesiology* 2016; **125**: 667-677 [PMID: 27655179 DOI: 10.1097/ALN.0000000000001245]
- 9 **Ing CH**, DiMaggio CJ, Malacova E, Whitehouse AJ, Hegarty MK, Feng T, Brady JE, von Ungern-Sternberg BS, Davidson AJ, Wall MM, Wood AJ, Li G, Sun LS. Comparative analysis of outcome measures used in examining neurodevelopmental effects of early childhood anaesthesia exposure. *Anesthesiology* 2014; **120**: 1319-1332 [PMID: 24694922 DOI: 10.1097/ALN.0000000000000248]
- 10 **Filan PM**, Hunt RW, Anderson PJ, Doyle LW, Inder TE. Neurologic outcomes in very preterm infants undergoing surgery. *J Pediatr* 2012; **160**: 409-414 [PMID: 22048043 DOI: 10.1016/j.jpeds.2011.09.009]
- 11 **Sprung J**, Flick RP, Katusic SK, Colligan RC, Barbaresi WJ, Bojanic K, Welch TL, Olson MD, Hanson AC, Schroeder DR, Wilder RT, Warner DO. Attention-deficit/hyperactivity disorder after early exposure to procedures requiring general anesthesia. *Mayo Clin Proc* 2012; **87**: 120-129 [PMID: 22305025 DOI: 10.1016/j.mayocp.2011.11.008]
- 12 **Hansen TG**, Pedersen JK, Henneberg SW, Pedersen DA, Murray JC, Morton NS, Christensen K. Academic performance in adolescence after inguinal hernia repair in infancy: a nationwide cohort study. *Anesthesiology* 2011; **114**: 1076-1085 [PMID: 21368654 DOI: 10.1097/ALN.0b013e31820e77a0]
- 13 **Sun L**. Early childhood general anaesthesia exposure and neurocognitive development. *Br J Anaesth* 2010; **105** Suppl 1: i61-i68 [PMID: 21148656 DOI: 10.1093/bja/aeq302]
- 14 **Ing C**, Sun M, Olfson M, DiMaggio CJ, Sun LS, Wall MM, Li G. Age at Exposure to Surgery and Anesthesia in Children and Association With Mental Disorder Diagnosis. *Anesth Analg* 2017; **125**: 1988-1998 [PMID: 28857799 DOI: 10.1213/ANE.0000000000002423]
- 15 **Block RI**, Thomas JJ, Bayman EO, Choi JY, Kimble KK, Todd MM. Are anesthesia and surgery during infancy associated with altered academic performance during childhood? *Anesthesiology* 2012; **117**: 494-503 [PMID: 22801049 DOI: 10.1097/ALN.0b013e3182644684]
- 16 **DiMaggio C**, Sun LS, Li G. Early childhood exposure to anesthesia and risk of developmental and behavioral disorders in a sibling birth cohort. *Anesth Analg* 2011; **113**: 1143-1151 [PMID: 21415431 DOI: 10.1213/ANE.0b013e3182147f42]
- 17 **DiMaggio C**, Sun LS, Kakavouli A, Byrne MW, Li G. A retrospective cohort study of the association of anesthesia and hernia repair surgery with behavioral and developmental disorders in young children. *J Neurosurg Anesthesiol* 2009; **21**: 286-291 [PMID: 19955889 DOI: 10.1097/ANA.0b013e3181a71f11]
- 18 **Heindel JJ**, Balbus J, Birnbaum L, Brune-Drisse MN, Grandjean P, Gray K, Landrigan PJ, Sly PD, Suk W, Cory Slechta D, Thompson C, Hanson M. Developmental Origins of Health and Disease: Integrating Environmental Influences. *Endocrinology* 2015; **156**: 3416-3421 [PMID: 26241070 DOI: 10.1210/EN.2015-1394]
- 19 **Barker DJ**. The origins of the developmental origins theory. *J Intern Med* 2007; **261**: 412-417 [PMID: 17444880 DOI: 10.1111/j.1365-2796.2007.01809.x]
- 20 **Gluckman PD**, Hanson MA, Mitchell MD. Developmental origins of health and disease: reducing the burden of chronic disease in the next generation. *Genome Med* 2010; **2**: 14 [PMID: 20236494 DOI: 10.1186/gm135]
- 21 **Baird J**, Jacob C, Barker M, Fall CH, Hanson M, Harvey NC, Inskip HM, Kumaran K, Cooper C. Developmental Origins of Health and Disease: A Lifecourse Approach to the Prevention of Non-Communicable Diseases. *Healthcare (Basel)* 2017; **5** [PMID: 28282852 DOI: 10.3390/healthcare5010014]
- 22 **Antkowiak B**, Rudolph U. New insights in the systemic and molecular underpinnings of general anesthetic actions mediated by  $\gamma$ -aminobutyric acid A receptors. *Curr Opin Anaesthesiol* 2016; **29**: 447-453 [PMID: 27168087 DOI: 10.1097/ACO.0000000000000358]
- 23 **Kotani N**, Akaike N. The effects of volatile anesthetics on synaptic and extrasynaptic GABA-induced neurotransmission. *Brain Res Bull* 2013; **93**: 69-79 [PMID: 22925739 DOI: 10.1016/j.brainresbull.2012.08.001]
- 24 **Khan KS**, Hayes I, Buggy DJ. Pharmacology of anaesthetic agents II: inhalation anaesthetic agents. *Continuing Education in Anaesthesia Critical Care & Pain* 2014; **14**: 106-111 [DOI: 10.1093/bjaceaccp/mkt038]
- 25 **Brohan J**, Goudra BG. The Role of GABA Receptor Agonists in Anesthesia and Sedation. *CNS Drugs* 2017; **31**: 845-856 [PMID: 29039138 DOI: 10.1007/s40263-017-0463-7]
- 26 **Wilder RT**, Flick RP, Sprung J, Katusic SK, Barbaresi WJ, Mickelson C, Gleich SJ, Schroeder DR, Weaver AL, Warner DO. Early exposure to anesthesia and learning disabilities in a population-based birth cohort. *Anesthesiology* 2009; **110**: 796-804 [PMID: 19293700 DOI: 10.1097/01.anes.0000344728.34332.5d]
- 27 **Kalkman CJ**, Peelen L, Moons KG, Veenhuizen M, Bruens M, Sinnema G, de Jong TP. Behavior and development in children and age at the time of first anesthetic exposure. *Anesthesiology* 2009; **110**: 805-812 [PMID: 19293699 DOI: 10.1097/ALN.0b013e31819c7124]
- 28 **McCann ME**, de Graaff JC, Dorris L, Disma N, Withington D, Bell G, Grobler A, Staggart R, Hunt RW, Sheppard SJ, Marmor J, Giribaldi G, Bellinger DC, Hartmann PL, Hardy P, Frawley G, Izzo F, von Ungern Sternberg BS, Lynn A, Wilton N, Mueller M, Polaner DM, Absalom AR, Szmuk P, Morton N, Berde C, Soriano S, Davidson AJ; GAS Consortium. Neurodevelopmental outcome at 5 years of age after general anaesthesia or awake-regional anaesthesia in infancy (GAS): an international, multicentre, randomised, controlled equivalence trial. *Lancet* 2019; **393**: 664-677 [PMID: 30782342 DOI: 10.1016/S0140-6736(18)32485-1]
- 29 **Banerjee P**, Rossi MG, Anghelescu DL, Liu W, Breazeale AM, Reddick WE, Glass JO, Phillips NS, Jacola LM, Sabin ND, Inaba H, Srivastava D, Robison LL, Pui CH, Hudson MM, Krull KR. Association Between Anesthesia Exposure and Neurocognitive and Neuroimaging Outcomes in Long-term Survivors of Childhood Acute Lymphoblastic Leukemia. *JAMA Oncol* 2019 [PMID: 31219514 DOI: 10.1001/jamaoncol.2019.1094]
- 30 **Lin EP**, Lee JR, Lee CS, Deng M, Loepke AW. Do anesthetics harm the developing human brain? An integrative analysis of animal and human studies. *Neurotoxicol Teratol* 2017; **60**: 117-128 [PMID: 28111111 DOI: 10.1016/j.ntt.2017.05.001]

- 27793659 DOI: [10.1016/j.ntt.2016.10.008](https://doi.org/10.1016/j.ntt.2016.10.008)]
- 31 **Deoni SC**, Adams SH, Li X, Badger TM, Pivik RT, Glasier CM, Ramakrishnaiah RH, Rowell AC, Ou X. Cesarean Delivery Impacts Infant Brain Development. *AJNR Am J Neuroradiol* 2019; **40**: 169-177 [PMID: [30467219](https://pubmed.ncbi.nlm.nih.gov/30467219/) DOI: [10.3174/ajnr.A5887](https://doi.org/10.3174/ajnr.A5887)]
  - 32 **Culley DJ**, Baxter M, Yukhananov R, Crosby G. The memory effects of general anesthesia persist for weeks in young and aged rats. *Anesth Analg* 2003; **96**: 1004-1009, table of contents [PMID: [12651650](https://pubmed.ncbi.nlm.nih.gov/12651650/) DOI: [10.1213/01.ane.0000052712.67573.12](https://doi.org/10.1213/01.ane.0000052712.67573.12)]
  - 33 **Stratmann G**, Sall JW, May LD, Bell JS, Magnusson KR, Rau V, Visrodia KH, Alvi RS, Ku B, Lee MT, Dai R. Isoflurane differentially affects neurogenesis and long-term neurocognitive function in 60-day-old and 7-day-old rats. *Anesthesiology* 2009; **110**: 834-848 [PMID: [19293705](https://pubmed.ncbi.nlm.nih.gov/19293705/) DOI: [10.1097/ALN.0b013e31819c463d](https://doi.org/10.1097/ALN.0b013e31819c463d)]
  - 34 **Zhu C**, Gao J, Karlsson N, Li Q, Zhang Y, Huang Z, Li H, Kuhn HG, Blomgren K. Isoflurane anesthesia induced persistent, progressive memory impairment, caused a loss of neural stem cells, and reduced neurogenesis in young, but not adult, rodents. *J Cereb Blood Flow Metab* 2010; **30**: 1017-1030 [PMID: [20068576](https://pubmed.ncbi.nlm.nih.gov/20068576/) DOI: [10.1038/jcbfm.2009.274](https://doi.org/10.1038/jcbfm.2009.274)]
  - 35 **Ly L**, Chan D, Aarabi M, Landry M, Behan NA, MacFarlane AJ, Trasler J. Intergenerational impact of paternal lifetime exposures to both folic acid deficiency and supplementation on reproductive outcomes and imprinted gene methylation. *Mol Hum Reprod* 2017; **23**: 461-477 [PMID: [28535307](https://pubmed.ncbi.nlm.nih.gov/28535307/) DOI: [10.1093/molehr/gax029](https://doi.org/10.1093/molehr/gax029)]
  - 36 **Rompala GR**, Homanics GE. Intergenerational Effects of Alcohol: A Review of Paternal Preconception Ethanol Exposure Studies and Epigenetic Mechanisms in the Male Germline. *Alcohol Clin Exp Res* 2019; **43**: 1032-1045 [PMID: [30908630](https://pubmed.ncbi.nlm.nih.gov/30908630/) DOI: [10.1111/acer.14029](https://doi.org/10.1111/acer.14029)]
  - 37 **Rodgers AB**, Morgan CP, Bronson SL, Revello S, Bale TL. Paternal stress exposure alters sperm microRNA content and reprograms offspring HPA stress axis regulation. *J Neurosci* 2013; **33**: 9003-9012 [PMID: [23699511](https://pubmed.ncbi.nlm.nih.gov/23699511/) DOI: [10.1523/JNEUROSCI.0914-13.2013](https://doi.org/10.1523/JNEUROSCI.0914-13.2013)]
  - 38 **Chan JC**, Nugent BM, Bale TL. Parental Advisory: Maternal and Paternal Stress Can Impact Offspring Neurodevelopment. *Biol Psychiatry* 2018; **83**: 886-894 [PMID: [29198470](https://pubmed.ncbi.nlm.nih.gov/29198470/) DOI: [10.1016/j.biopsych.2017.10.005](https://doi.org/10.1016/j.biopsych.2017.10.005)]
  - 39 **Goldberg LR**, Gould TJ. Multigenerational and transgenerational effects of paternal exposure to drugs of abuse on behavioral and neural function. *Eur J Neurosci* 2019; **50**: 2453-2466 [PMID: [29949212](https://pubmed.ncbi.nlm.nih.gov/29949212/) DOI: [10.1111/ejn.14060](https://doi.org/10.1111/ejn.14060)]
  - 40 **Chastain LG**, Sarkar DK. Alcohol effects on the epigenome in the germline: Role in the inheritance of alcohol-related pathology. *Alcohol* 2017; **60**: 53-66 [PMID: [28431793](https://pubmed.ncbi.nlm.nih.gov/28431793/) DOI: [10.1016/j.alcohol.2016.12.007](https://doi.org/10.1016/j.alcohol.2016.12.007)]
  - 41 **Rattan S**, Flaws JA. The epigenetic impacts of endocrine disruptors on female reproduction across generations†. *Biol Reprod* 2019; **101**: 635-644 [PMID: [31077281](https://pubmed.ncbi.nlm.nih.gov/31077281/) DOI: [10.1093/biolre/iox081](https://doi.org/10.1093/biolre/iox081)]
  - 42 **Gillette R**, Son MJ, Ton L, Gore AC, Crews D. Passing experiences on to future generations: endocrine disruptors and transgenerational inheritance of epimutations in brain and sperm. *Epigenetics* 2018; **13**: 1106-1126 [PMID: [30444163](https://pubmed.ncbi.nlm.nih.gov/30444163/) DOI: [10.1080/15592294.2018.1543506](https://doi.org/10.1080/15592294.2018.1543506)]
  - 43 **Yehuda R**, Daskalakis NP, Lehrner A, Desarnaud F, Bader HN, Makotkine I, Flory JD, Bierer LM, Meaney MJ. Influences of maternal and paternal PTSD on epigenetic regulation of the glucocorticoid receptor gene in Holocaust survivor offspring. *Am J Psychiatry* 2014; **171**: 872-880 [PMID: [24832930](https://pubmed.ncbi.nlm.nih.gov/24832930/) DOI: [10.1176/appi.ajp.2014.13121571](https://doi.org/10.1176/appi.ajp.2014.13121571)]
  - 44 **Pembrey ME**, Bygren LO, Kaati G, Edvinsson S, Northstone K, Sjöström M, Golding J; ALSPAC Study Team. Sex-specific, male-line transgenerational responses in humans. *Eur J Hum Genet* 2006; **14**: 159-166 [PMID: [16391557](https://pubmed.ncbi.nlm.nih.gov/16391557/) DOI: [10.1038/sj.ejhg.5201538](https://doi.org/10.1038/sj.ejhg.5201538)]
  - 45 **Mihic SJ**, Ye Q, Wick MJ, Koltchine VV, Krasowski MD, Finn SE, Mascia MP, Valenzuela CF, Hanson KK, Greenblatt EP, Harris RA, Harrison NL. Sites of alcohol and volatile anaesthetic action on GABA(A) and glycine receptors. *Nature* 1997; **389**: 385-389 [PMID: [9311780](https://pubmed.ncbi.nlm.nih.gov/9311780/) DOI: [10.1038/38738](https://doi.org/10.1038/38738)]
  - 46 **Beckstead MJ**, Phelan R, Mihic SJ. Antagonism of inhalant and volatile anesthetic enhancement of glycine receptor function. *J Biol Chem* 2001; **276**: 24959-24964 [PMID: [11346643](https://pubmed.ncbi.nlm.nih.gov/11346643/) DOI: [10.1074/jbc.M011627200](https://doi.org/10.1074/jbc.M011627200)]
  - 47 **Beckstead MJ**, Phelan R, Trudell JR, Bianchini MJ, Mihic SJ. Anesthetic and ethanol effects on spontaneously opening glycine receptor channels. *J Neurochem* 2002; **82**: 1343-1351 [PMID: [12354281](https://pubmed.ncbi.nlm.nih.gov/12354281/) DOI: [10.1046/j.1471-4159.2002.01086.x](https://doi.org/10.1046/j.1471-4159.2002.01086.x)]
  - 48 **Rees DC**, Knisely JS, Breen TJ, Balster RL. Toluene, halothane, 1,1,1-trichloroethane and oxazepam produce ethanol-like discriminative stimulus effects in mice. *J Pharmacol Exp Ther* 1987; **243**: 931-937 [PMID: [3694538](https://pubmed.ncbi.nlm.nih.gov/3694538/)]
  - 49 **Bowen SE**, Balster RL. Desflurane, enflurane, isoflurane and ether produce ethanol-like discriminative stimulus effects in mice. *Pharmacol Biochem Behav* 1997; **57**: 191-198 [PMID: [9164572](https://pubmed.ncbi.nlm.nih.gov/9164572/) DOI: [10.1016/s0091-3057\(96\)00308-5](https://doi.org/10.1016/s0091-3057(96)00308-5)]
  - 50 **Khilnani P**, Munoz R, Salem M, Gelb C, Todres ID, Chernow B. Hormonal responses to surgical stress in children. *J Pediatr Surg* 1993; **28**: 1-4 [PMID: [8429460](https://pubmed.ncbi.nlm.nih.gov/8429460/) DOI: [10.1016/s0022-3468\(05\)80343-4](https://doi.org/10.1016/s0022-3468(05)80343-4)]
  - 51 **Rains PC**, Rampersad N, De Lima J, Murrell D, Kinchington D, Lee JW, Maguire AM, Donaghue KC. Cortisol response to general anaesthesia for medical imaging in children. *Clin Endocrinol (Oxf)* 2009; **71**: 834-839 [PMID: [19508604](https://pubmed.ncbi.nlm.nih.gov/19508604/) DOI: [10.1111/j.1365-2265.2009.03591.x](https://doi.org/10.1111/j.1365-2265.2009.03591.x)]
  - 52 **Hsu AA**, von Elten K, Chan D, Flynn T, Walker K, Barnhill J, Naun C, Pedersen AM, Ponaman M, Fredericks GJ, Crudo DF, Pinsker JE. Characterization of the cortisol stress response to sedation and anesthesia in children. *J Clin Endocrinol Metab* 2012; **97**: E1830-E1835 [PMID: [22855336](https://pubmed.ncbi.nlm.nih.gov/22855336/) DOI: [10.1210/jc.2012-1499](https://doi.org/10.1210/jc.2012-1499)]
  - 53 **Aggo AT**, Fyeface-Ogan S, Mato CN. The differential impact of two anesthetic techniques on cortisol levels in Nigerian surgical patients. *Niger J Clin Pract* 2012; **15**: 68-74 [PMID: [22437094](https://pubmed.ncbi.nlm.nih.gov/22437094/) DOI: [10.4103/1119-3077.94102](https://doi.org/10.4103/1119-3077.94102)]
  - 54 **Prete A**, Yan Q, Al-Tarrah K, Akturk HK, Prokop LJ, Alahdab F, Foster MA, Lord JM, Karavitaki N, Wass JA, Murad MH, Arlt W, Bancos I. The cortisol stress response induced by surgery: A systematic review and meta-analysis. *Clin Endocrinol (Oxf)* 2018; **89**: 554-567 [PMID: [30047158](https://pubmed.ncbi.nlm.nih.gov/30047158/) DOI: [10.1111/cen.13820](https://doi.org/10.1111/cen.13820)]
  - 55 **Mody I**, Maguire J. The reciprocal regulation of stress hormones and GABA(A) receptors. *Front Cell Neurosci* 2011; **6**: 4 [PMID: [22319473](https://pubmed.ncbi.nlm.nih.gov/22319473/) DOI: [10.3389/fncel.2012.00004](https://doi.org/10.3389/fncel.2012.00004)]
  - 56 **Kakizawa K**, Watanabe M, Mutoh H, Okawa Y, Yamashita M, Yanagawa Y, Itoi K, Suda T, Oki Y, Fukuda A. A novel GABA-mediated corticotropin-releasing hormone secretory mechanism in the median

- eminence. *Sci Adv* 2016; **2**: e1501723 [PMID: 27540587 DOI: 10.1126/sciadv.1501723]
- 57 **Salmon CK**, Pribrag H, Gizowski C, Farmer WT, Cameron S, Jones EV, Mahadevan V, Bourque CW, Stellwagen D, Woodin MA, Murai KK. Depolarizing GABA Transmission Restrains Activity-Dependent Glutamatergic Synapse Formation in the Developing Hippocampal Circuit. *Front Cell Neurosci* 2020; **14**: 36 [PMID: 32161521 DOI: 10.3389/fncel.2020.00036]
- 58 **Ben-Ari Y**. The GABA excitatory/inhibitory developmental sequence: a personal journey. *Neuroscience* 2014; **279**: 187-219 [PMID: 25168736 DOI: 10.1016/j.neuroscience.2014.08.001]
- 59 **Khazipov R**, Valeeva G, Khalilov I. Depolarizing GABA and developmental epilepsies. *CNS Neurosci Ther* 2015; **21**: 83-91 [PMID: 25438879 DOI: 10.1111/cns.12353]
- 60 **Ito S**. GABA and glycine in the developing brain. *J Physiol Sci* 2016; **66**: 375-379 [PMID: 26951057 DOI: 10.1007/s12576-016-0442-7]
- 61 **Farrant M**, Kaila K. The cellular, molecular and ionic basis of GABA(A) receptor signalling. *Prog Brain Res* 2007; **160**: 59-87 [PMID: 17499109 DOI: 10.1016/S0079-6123(06)60005-8]
- 62 **Dehorter N**, Vinay L, Hammond C, Ben-Ari Y. Timing of developmental sequences in different brain structures: physiological and pathological implications. *Eur J Neurosci* 2012; **35**: 1846-1856 [PMID: 22708595 DOI: 10.1111/j.1460-9568.2012.08152.x]
- 63 **Edwards DA**, Shah HP, Cao W, Gravenstein N, Seubert CN, Martynyuk AE. Bumetanide alleviates epileptogenic and neurotoxic effects of sevoflurane in neonatal rat brain. *Anesthesiology* 2010; **112**: 567-575 [PMID: 20124973 DOI: 10.1097/ALN.0b013e3181cf9138]
- 64 **Tan S**, Xu C, Zhu W, Willis J, Seubert CN, Gravenstein N, Summers C, Martynyuk AE. Endocrine and neurobehavioral abnormalities induced by propofol administered to neonatal rats. *Anesthesiology* 2014; **121**: 1010-1017 [PMID: 24992523 DOI: 10.1097/ALN.0000000000000366]
- 65 **Xu C**, Tan S, Zhang J, Seubert CN, Gravenstein N, Summers C, Vasilopoulos T, Martynyuk AE. Anesthesia with sevoflurane in neonatal rats: Developmental neuroendocrine abnormalities and alleviating effects of the corticosteroid and Cl(-) importer antagonists. *Psychoneuroendocrinology* 2015; **60**: 173-181 [PMID: 26150359 DOI: 10.1016/j.psyneuen.2015.06.016]
- 66 **Ju LS**, Yang JJ, Gravenstein N, Seubert CN, Morey TE, Summers C, Vasilopoulos T, Yang JJ, Martynyuk AE. Role of environmental stressors in determining the developmental outcome of neonatal anesthesia. *Psychoneuroendocrinology* 2017; **81**: 96-104 [PMID: 28433802 DOI: 10.1016/j.psyneuen.2017.04.001]
- 67 **Ju LS**, Yang JJ, Morey TE, Gravenstein N, Seubert CN, Resnick JL, Zhang JQ, Martynyuk AE. Role of epigenetic mechanisms in transmitting the effects of neonatal sevoflurane exposure to the next generation of male, but not female, rats. *Br J Anaesth* 2018; **121**: 406-416 [PMID: 30032879 DOI: 10.1016/j.bja.2018.04.034]
- 68 **Willis J**, Zhu W, Perez-Downes J, Tan S, Xu C, Seubert C, Gravenstein N, Martynyuk A. Propofol-induced electroencephalographic seizures in neonatal rats: the role of corticosteroids and  $\gamma$ -aminobutyric acid type A receptor-mediated excitation. *Anesth Analg* 2015; **120**: 433-439 [PMID: 25390279 DOI: 10.1213/ANE.0000000000000529]
- 69 **Zhang J**, Xu C, Puentes DL, Seubert CN, Gravenstein N, Martynyuk AE. Role of Steroids in Hyperexcitatory Adverse and Anesthetic Effects of Sevoflurane in Neonatal Rats. *Neuroendocrinology* 2016; **103**: 440-451 [PMID: 26159049 DOI: 10.1159/000437267]
- 70 **Cao W**, Pavlinec C, Gravenstein N, Seubert CN, Martynyuk AE. Roles of aldosterone and oxytocin in abnormalities caused by sevoflurane anesthesia in neonatal rats. *Anesthesiology* 2012; **117**: 791-800 [PMID: 22854980 DOI: 10.1097/ALN.0b013e318266c62d]
- 71 **Seubert CN**, Zhu W, Pavlinec C, Gravenstein N, Martynyuk AE. Developmental effects of neonatal isoflurane and sevoflurane exposure in rats. *Anesthesiology* 2013; **119**: 358-364 [PMID: 23619170 DOI: 10.1097/ALN.0b013e318291c04e]
- 72 **Xu C**, Seubert CN, Gravenstein N, Martynyuk AE. Propofol, but not etomidate, increases corticosterone levels and induces long-term alteration in hippocampal synaptic activity in neonatal rats. *Neurosci Lett* 2016; **618**: 1-5 [PMID: 26923669 DOI: 10.1016/j.neulet.2016.02.045]
- 73 **Yang J**, Ju L, Jia M, Zhang H, Sun X, Ji M, Yang J, Martynyuk AE. Subsequent maternal separation exacerbates neurobehavioral abnormalities in rats neonatally exposed to sevoflurane anesthesia. *Neurosci Lett* 2017; **661**: 137-142 [PMID: 28982596 DOI: 10.1016/j.neulet.2017.09.063]
- 74 **Majcher-Masłanka I**, Solarz A, Chocyk A. Maternal separation disturbs postnatal development of the medial prefrontal cortex and affects the number of neurons and glial cells in adolescent rats. *Neuroscience* 2019; **423**: 131-147 [PMID: 31705889 DOI: 10.1016/j.neuroscience.2019.10.033]
- 75 **Heydari A**, Esmailpour K, Sheibani V. Maternal separation impairs long term-potential in CA3-CA1 synapses in adolescent female rats. *Behav Brain Res* 2019; **376**: 112239 [PMID: 31526768 DOI: 10.1016/j.bbr.2019.112239]
- 76 **Brunson KL**, Kramár E, Lin B, Chen Y, Colgin LL, Yanagihara TK, Lynch G, Baram TZ. Mechanisms of late-onset cognitive decline after early-life stress. *J Neurosci* 2005; **25**: 9328-9338 [PMID: 16221841 DOI: 10.1523/JNEUROSCI.2281-05.2005]
- 77 **Furukawa M**, Tsukahara T, Tomita K, Iwai H, Sonomura T, Miyawaki S, Sato T. Neonatal maternal separation delays the GABA excitatory-to-inhibitory functional switch by inhibiting KCC2 expression. *Biochem Biophys Res Commun* 2017; **493**: 1243-1249 [PMID: 28962859 DOI: 10.1016/j.bbrc.2017.09.143]
- 78 **Patchev VK**, Montkowski A, Rouskova D, Koranyi L, Holsboer F, Almeida OF. Neonatal treatment of rats with the neuroactive steroid tetrahydrocorticosterone (THDOC) abolishes the behavioral and neuroendocrine consequences of adverse early life events. *J Clin Invest* 1997; **99**: 962-966 [PMID: 9062354 DOI: 10.1172/JCI119261]
- 79 **Merner ND**, Chandler MR, Bourassa C, Liang B, Khanna AR, Dion P, Rouleau GA, Kahle KT. Regulatory domain or CpG site variation in SLC12A5, encoding the chloride transporter KCC2, in human autism and schizophrenia. *Front Cell Neurosci* 2015; **9**: 386 [PMID: 26528127 DOI: 10.3389/fncel.2015.00386]
- 80 **Ben-Ari Y**, Khalilov I, Kahle KT, Cherubini E. The GABA excitatory/inhibitory shift in brain maturation and neurological disorders. *Neuroscientist* 2012; **18**: 467-486 [PMID: 22547529 DOI: 10.1177/1073858412438697]
- 81 **Merner ND**, Mercado A, Khanna AR, Hodgkinson A, Bruat V, Awadalla P, Gamba G, Rouleau GA, Kahle KT. Gain-of-function missense variant in SLC12A2, encoding the bumetanide-sensitive NKCC1 cotransporter, identified in human schizophrenia. *J Psychiatr Res* 2016; **77**: 22-26 [PMID: 26955005 DOI: 10.1016/j.jpsychires.2016.02.016]
- 82 **Genç F**, Kara M, Ünal Y, Uygur Küçükseymen E, Biçer Gömçeli Y, Kaynar T, Tosun K, Kutlu G.

- Methylation of cation-chloride cotransporters NKCC1 and KCC2 in patients with juvenile myoclonic epilepsy. *Neurol Sci* 2019; **40**: 1007-1013 [PMID: 30759289 DOI: 10.1007/s10072-019-03743-4]
- 83 **Han P**, Welsh CT, Smith MT, Schmidt RE, Carroll SL. Complex Patterns of GABAergic Neuronal Deficiency and Type 2 Potassium-Chloride Cotransporter Immaturity in Human Focal Cortical Dysplasia. *J Neuropathol Exp Neurol* 2019; **78**: 365-372 [PMID: 30856249 DOI: 10.1093/jnen/nlz009]
- 84 **Tang X**, Kim J, Zhou L, Wengert E, Zhang L, Wu Z, Carroumeu C, Muotri AR, Marchetto MC, Gage FH, Chen G. KCC2 rescues functional deficits in human neurons derived from patients with Rett syndrome. *Proc Natl Acad Sci USA* 2016; **113**: 751-756 [PMID: 26733678 DOI: 10.1073/pnas.1524013113]
- 85 **Tang X**, Drotar J, Li K, Clairmont CD, Brumm AS, Sullins AJ, Wu H, Liu XS, Wang J, Gray NS, Sur M, Jaenisch R. Pharmacological enhancement of *KCC2* gene expression exerts therapeutic effects on human Rett syndrome neurons and *Mecp2* mutant mice. *Sci Transl Med* 2019; **11** [PMID: 31366578 DOI: 10.1126/scitranslmed.aau0164]
- 86 **Dzhala VI**, Talos DM, Sdrulla DA, Brumback AC, Mathews GC, Benke TA, Delpire E, Jensen FE, Staley KJ. NKCC1 transporter facilitates seizures in the developing brain. *Nat Med* 2005; **11**: 1205-1213 [PMID: 16227993 DOI: 10.1038/nm1301]
- 87 **Ben-Ari Y**. NKCC1 Chloride Importer Antagonists Attenuate Many Neurological and Psychiatric Disorders. *Trends Neurosci* 2017; **40**: 536-554 [PMID: 28818303 DOI: 10.1016/j.tins.2017.07.001]
- 88 **Liu G**, Zhu T, Zhang A, Li F, Qian W, Qian B. Heightened stress response and cognitive impairment after repeated neonatal sevoflurane exposures might be linked to excessive GABAAR-mediated depolarization. *J Anesth* 2016; **30**: 834-841 [PMID: 27435414 DOI: 10.1007/s00540-016-2215-0]
- 89 **Du L**, Shan L, Wang B, Li H, Xu Z, Staal WG, Jia F. A Pilot Study on the Combination of Applied Behavior Analysis and Bumetanide Treatment for Children with Autism. *J Child Adolesc Psychopharmacol* 2015; **25**: 585-588 [PMID: 26258842 DOI: 10.1089/cap.2015.0045]
- 90 **Lemonnier E**, Robin G, Degrez C, Tyzio R, Grandgeorge M, Ben-Ari Y. Treating Fragile X syndrome with the diuretic bumetanide: a case report. *Acta Paediatr* 2013; **102**: e288-e290 [PMID: 23647528 DOI: 10.1111/apa.12235]
- 91 **Lemonnier E**, Lazartigues A, Ben-Ari Y. Treating Schizophrenia With the Diuretic Bumetanide: A Case Report. *Clin Neuropharmacol* 2016; **39**: 115-117 [PMID: 26966887 DOI: 10.1097/WNF.0000000000000136]
- 92 **Lemonnier E**, Villeneuve N, Sonie S, Serret S, Rosier A, Roue M, Brosset P, Viellard M, Bernoux D, Rondeau S, Thummler S, Ravel D, Ben-Ari Y. Effects of bumetanide on neurobehavioral function in children and adolescents with autism spectrum disorders. *Transl Psychiatry* 2017; **7**: e1056 [PMID: 28291262 DOI: 10.1038/tp.2017.10]
- 93 **Hadjikhani N**, Åsberg Johnels J, Lassalle A, Zürcher NR, Hippolyte L, Gillberg C, Lemonnier E, Ben-Ari Y. Bumetanide for autism: more eye contact, less amygdala activation. *Sci Rep* 2018; **8**: 3602 [PMID: 29483603 DOI: 10.1038/s41598-018-21958-x]
- 94 **Semple BD**, Blomgren K, Gimlin K, Ferriero DM, Noble-Haesslein LJ. Brain development in rodents and humans: Identifying benchmarks of maturation and vulnerability to injury across species. *Prog Neurobiol* 2013; **106-107**: 1-16 [PMID: 23583307 DOI: 10.1016/j.pneurobio.2013.04.001]
- 95 **Schachtele SJ**, Losh J, Dailey ME, Green SH. Spine formation and maturation in the developing rat auditory cortex. *J Comp Neurol* 2011; **519**: 3327-3345 [PMID: 21800311 DOI: 10.1002/cne.22728]
- 96 **Lenroot RK**, Giedd JN. Brain development in children and adolescents: insights from anatomical magnetic resonance imaging. *Neurosci Biobehav Rev* 2006; **30**: 718-729 [PMID: 16887188 DOI: 10.1016/j.neubiorev.2006.06.001]
- 97 **Petanjek Z**, Judaš M, Šimic G, Rasin MR, Uylings HB, Rakic P, Kostovic I. Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proc Natl Acad Sci USA* 2011; **108**: 13281-13286 [PMID: 21788513 DOI: 10.1073/pnas.1105108108]
- 98 **Liu X**, Somel M, Tang L, Yan Z, Jiang X, Guo S, Yuan Y, He L, Oleksiak A, Zhang Y, Li N, Hu Y, Chen W, Qiu Z, Pääbo S, Khaitovich P. Extension of cortical synaptic development distinguishes humans from chimpanzees and macaques. *Genome Res* 2012; **22**: 611-622 [PMID: 22300767 DOI: 10.1101/gr.127324.111]
- 99 **Jia M**, Liu WX, Yang JJ, Xu N, Xie ZM, Ju LS, Ji MH, Martynyuk AE, Yang JJ. Role of histone acetylation in long-term neurobehavioral effects of neonatal Exposure to sevoflurane in rats. *Neurobiol Dis* 2016; **91**: 209-220 [PMID: 27001149 DOI: 10.1016/j.nbd.2016.03.017]
- 100 **Ju LS**, Jia M, Sun J, Sun XR, Zhang H, Ji MH, Yang JJ, Wang ZY. Hypermethylation of Hippocampal Synaptic Plasticity-Related genes is Involved in Neonatal Sevoflurane Exposure-Induced Cognitive Impairments in Rats. *Neurotox Res* 2016; **29**: 243-255 [PMID: 26678494 DOI: 10.1007/s12640-015-9585-1]
- 101 **Bock J**, Braun K. The impact of perinatal stress on the functional maturation of prefronto-cortical synaptic circuits: implications for the pathophysiology of ADHD? *Prog Brain Res* 2011; **189**: 155-169 [PMID: 21489388 DOI: 10.1016/B978-0-444-53884-0.00023-3]
- 102 **Ohta KI**, Suzuki S, Warita K, Kaji T, Kusaka T, Miki T. Prolonged maternal separation attenuates BDNF-ERK signaling correlated with spine formation in the hippocampus during early brain development. *J Neurochem* 2017; **141**: 179-194 [PMID: 28178750 DOI: 10.1111/jnc.13977]
- 103 **Danielewicz J**, Hess G. Early life stress alters synaptic modification range in the rat lateral amygdala. *Behav Brain Res* 2014; **265**: 32-37 [PMID: 24556204 DOI: 10.1016/j.bbr.2014.02.012]
- 104 **Lanska MJ**, Lanska DJ, Baumann RJ, Kryscio RJ. A population-based study of neonatal seizures in Fayette County, Kentucky. *Neurology* 1995; **45**: 724-732 [PMID: 7723962 DOI: 10.1212/wnl.45.4.724]
- 105 **van Rooij LG**, Hellström-Westas L, de Vries LS. Treatment of neonatal seizures. *Semin Fetal Neonatal Med* 2013; **18**: 209-215 [PMID: 23402893 DOI: 10.1016/j.siny.2013.01.001]
- 106 **Volpe J**. Neurology of the newborn. 5th ed. Philadelphia, PA: Saunders, 2008
- 107 **Painter MJ**, Scher MS, Stein AD, Armatti S, Wang Z, Gardiner JC, Paneth N, Minnich B, Alvin J. Phenobarbital compared with phenytoin for the treatment of neonatal seizures. *N Engl J Med* 1999; **341**: 485-489 [PMID: 10441604 DOI: 10.1056/NEJM199908123410704]
- 108 **Boylan GB**, Rennie JM, Chorley G, Pressler RM, Fox GF, Farrer K, Morton M, Binnie CD. Second-line anticonvulsant treatment of neonatal seizures: a video-EEG monitoring study. *Neurology* 2004; **62**: 486-488 [PMID: 14872039 DOI: 10.1212/01.wnl.0000106944.59990.e6]
- 109 **Boylan GB**, Stevenson NJ, Vanhatalo S. Monitoring neonatal seizures. *Semin Fetal Neonatal Med* 2013; **18**: 202-208 [PMID: 23707519 DOI: 10.1016/j.siny.2013.04.004]
- 110 **Connell J**, Oozeer R, de Vries L, Dubowitz LM, Dubowitz V. Clinical and EEG response to anticonvulsants in neonatal seizures. *Arch Dis Child* 1989; **64**: 459-464 [PMID: 2730114 DOI: 10.1136/adc.1989.064059.459]

- 10.1136/adc.64.4\_spec\_no.459]
- 111 **Puskarjov M**, Kahle KT, Ruusuvoori E, Kaila K. Pharmacotherapeutic targeting of cation-chloride cotransporters in neonatal seizures. *Epilepsia* 2014; **55**: 806-818 [PMID: 24802699 DOI: 10.1111/epi.12620]
- 112 **Kahle KT**, Barnett SM, Sassower KC, Staley KJ. Decreased seizure activity in a human neonate treated with bumetanide, an inhibitor of the Na(+)-K(+)-2Cl(-) cotransporter NKCC1. *J Child Neurol* 2009; **24**: 572-576 [PMID: 19406757 DOI: 10.1177/0883073809333526]
- 113 **Yang J**, Ju L, Yang C, Xue J, Setlow B, Morey TE, Gravenstein N, Seubert CN, Vasilopoulos T, Martynyuk AE. Effects of combined brief etomidate anesthesia and postnatal stress on amygdala expression of Cl<sup>-</sup> cotransporters and corticotropin-releasing hormone and alcohol intake in adult rats. *Neurosci Lett* 2018; **685**: 83-89 [PMID: 30125644 DOI: 10.1016/j.neulet.2018.08.019]
- 114 **Shih J**, May LD, Gonzalez HE, Lee EW, Alvi RS, Sall JW, Rau V, Bickler PE, Lalchandani GR, Yusupova M, Woodward E, Kang H, Wilk AJ, Carlston CM, Mendoza MV, Guggenheim JN, Schaefer M, Rowe AM, Stratmann G. Delayed environmental enrichment reverses sevoflurane-induced memory impairment in rats. *Anesthesiology* 2012; **116**: 586-602 [PMID: 22354242 DOI: 10.1097/ALN.0b013e318247564d]
- 115 **Zheng H**, Dong Y, Xu Z, Crosby G, Culley DJ, Zhang Y, Xie Z. Sevoflurane anesthesia in pregnant mice induces neurotoxicity in fetal and offspring mice. *Anesthesiology* 2013; **118**: 516-526 [PMID: 23314109 DOI: 10.1097/ALN.0b013e3182834d5d]
- 116 **Chinn GA**, Sasaki Russell JM, Banh ET, Lee SC, Sall JW. Voluntary Exercise Rescues the Spatial Memory Deficit Associated With Early Life Isoflurane Exposure in Male Rats. *Anesth Analg* 2019; **129**: 1365-1373 [PMID: 31517674 DOI: 10.1213/ANE.0000000000004418]
- 117 **Zhang MQ**, Ji MH, Zhao QS, Jia M, Qiu LL, Yang JJ, Peng YG, Yang JJ, Martynyuk AE. Neurobehavioural abnormalities induced by repeated exposure of neonatal rats to sevoflurane can be aggravated by social isolation and enrichment deprivation initiated after exposure to the anaesthetic. *Br J Anaesth* 2015; **115**: 752-760 [PMID: 26475803 DOI: 10.1093/bja/aev339]
- 118 **Atrooz F**, Liu H, Salim S. Stress, psychiatric disorders, molecular targets, and more. *Prog Mol Biol Transl Sci* 2019; **167**: 77-105 [PMID: 31601407 DOI: 10.1016/bs.pmbts.2019.06.006]
- 119 **D'Mello SR**. Regulation of Central Nervous System Development by Class I Histone Deacetylases. *Dev Neurosci* 2019; **41**: 149-165 [PMID: 31982872 DOI: 10.1159/000505535]
- 120 **McClelland S**, Korosi A, Cope J, Ivy A, Baram TZ. Emerging roles of epigenetic mechanisms in the enduring effects of early-life stress and experience on learning and memory. *Neurobiol Learn Mem* 2011; **96**: 79-88 [PMID: 21338703 DOI: 10.1016/j.nlm.2011.02.008]
- 121 **Powell WT**, LaSalle JM. Epigenetic mechanisms in diurnal cycles of metabolism and neurodevelopment. *Hum Mol Genet* 2015; **24**: R1-R9 [PMID: 26105183 DOI: 10.1093/hmg/ddv234]
- 122 **Rudenko A**, Tsai LH. Epigenetic modifications in the nervous system and their impact upon cognitive impairments. *Neuropharmacology* 2014; **80**: 70-82 [PMID: 24495398 DOI: 10.1016/j.neuropharm.2014.01.043]
- 123 **Bannister AJ**, Kouzarides T. Regulation of chromatin by histone modifications. *Cell Res* 2011; **21**: 381-395 [PMID: 21321607 DOI: 10.1038/cr.2011.22]
- 124 **Fischer A**, Sananbenesi F, Mungenast A, Tsai LH. Targeting the correct HDAC(s) to treat cognitive disorders. *Trends Pharmacol Sci* 2010; **31**: 605-617 [PMID: 20980063 DOI: 10.1016/j.tips.2010.09.003]
- 125 **Guan JS**, Haggarty SJ, Giacometti E, Dannenberg JH, Joseph N, Gao J, Nieland TJ, Zhou Y, Wang X, Mazitschek R, Bradner JE, DePinho RA, Jaenisch R, Tsai LH. HDAC2 negatively regulates memory formation and synaptic plasticity. *Nature* 2009; **459**: 55-60 [PMID: 19424149 DOI: 10.1038/nature07925]
- 126 **McQuown SC**, Barrett RM, Matheos DP, Post RJ, Rogge GA, Alenghat T, Mullican SE, Jones S, Rusche JR, Lazar MA, Wood MA. HDAC3 is a critical negative regulator of long-term memory formation. *J Neurosci* 2011; **31**: 764-774 [PMID: 21228185 DOI: 10.1523/JNEUROSCI.5052-10.2011]
- 127 **Dalla Massara L**, Osuru HP, Oklopic A, Milanovic D, Joksimovic SM, Caputo V, DiGrucchio MR, Ori C, Wang G, Todorovic SM, Jevtovic-Todorovic V. General Anesthesia Causes Epigenetic Histone Modulation of c-Fos and Brain-derived Neurotrophic Factor, Target Genes Important for Neuronal Development in the Immature Rat Hippocampus. *Anesthesiology* 2016; **124**: 1311-1327 [PMID: 27028464 DOI: 10.1097/ALN.0000000000001111]
- 128 **Joksimovic SM**, Osuru HP, Oklopic A, Beenhakker MP, Jevtovic-Todorovic V, Todorovic SM. Histone Deacetylase Inhibitor Entinostat (MS-275) Restores Anesthesia-induced Alteration of Inhibitory Synaptic Transmission in the Developing Rat Hippocampus. *Mol Neurobiol* 2018; **55**: 222-228 [PMID: 28840475 DOI: 10.1007/s12035-017-0735-8]
- 129 **Luo F**, Hu Y, Zhao W, Zuo Z, Yu Q, Liu Z, Lin J, Feng Y, Li B, Wu L, Xu L. Maternal Exposure of Rats to Isoflurane during Late Pregnancy Impairs Spatial Learning and Memory in the Offspring by Up-Regulating the Expression of Histone Deacetylase 2. *PLoS One* 2016; **11**: e0160826 [PMID: 27536989 DOI: 10.1371/journal.pone.0160826]
- 130 **Lin J**, Wang S, Feng Y, Zhao W, Zhao W, Luo F, Feng N. Propofol exposure during early gestation impairs learning and memory in rat offspring by inhibiting the acetylation of histone. *J Cell Mol Med* 2018; **22**: 2600-2611 [PMID: 29461008 DOI: 10.1111/jcmm.13524]
- 131 **Wu Z**, Li X, Zhang Y, Tong D, Wang L, Zhao P. Effects of Sevoflurane Exposure During Mid-Pregnancy on Learning and Memory in Offspring Rats: Beneficial Effects of Maternal Exercise. *Front Cell Neurosci* 2018; **12**: 122 [PMID: 29773978 DOI: 10.3389/fncel.2018.00122]
- 132 **Kinney SR**, Pradhan S. Regulation of expression and activity of DNA (cytosine-5) methyltransferases in mammalian cells. *Prog Mol Biol Transl Sci* 2011; **101**: 311-333 [PMID: 21507356 DOI: 10.1016/B978-0-12-387685-0.00009-3]
- 133 **Lyko F**. The DNA methyltransferase family: a versatile toolkit for epigenetic regulation. *Nat Rev Genet* 2018; **19**: 81-92 [PMID: 29033456 DOI: 10.1038/nrg.2017.80]
- 134 **Wu J**, Bie B, Naguib M. Epigenetic Manipulation of Brain-derived Neurotrophic Factor Improves Memory Deficiency Induced by Neonatal Anesthesia in Rats. *Anesthesiology* 2016; **124**: 624-640 [PMID: 26649423 DOI: 10.1097/ALN.0000000000000981]
- 135 **Lehrner A**, Bierer LM, Passarelli V, Pratchett LC, Flory JD, Bader HN, Harris IR, Bedi A, Daskalakis NP, Makotkine I, Yehuda R. Maternal PTSD associates with greater glucocorticoid sensitivity in offspring of Holocaust survivors. *Psychoneuroendocrinology* 2014; **40**: 213-220 [PMID: 24485493 DOI: 10.1016/j.psyneuen.2013.11.019]
- 136 **Khashan AS**, Abel KM, McNamee R, Pedersen MG, Webb RT, Baker PN, Kenny LC, Mortensen PB. Higher risk of offspring schizophrenia following antenatal maternal exposure to severe adverse life events.

- Arch Gen Psychiatry* 2008; **65**: 146-152 [PMID: 18250252 DOI: 10.1001/archgenpsychiatry.2007.20]
- 137 **Pembrey M**, Saffery R, Bygren LO; Network in Epigenetic Epidemiology; Network in Epigenetic Epidemiology. Human transgenerational responses to early-life experience: potential impact on development, health and biomedical research. *J Med Genet* 2014; **51**: 563-572 [PMID: 25062846 DOI: 10.1136/jmedgenet-2014-102577]
- 138 **Bygren LO**, Kaati G, Edvinsson S. Longevity determined by paternal ancestors' nutrition during their slow growth period. *Acta Biotheor* 2001; **49**: 53-59 [PMID: 11368478 DOI: 10.1023/a:1010241825519]
- 139 **Kaati G**, Bygren LO, Edvinsson S. Cardiovascular and diabetes mortality determined by nutrition during parents' and grandparents' slow growth period. *Eur J Hum Genet* 2002; **10**: 682-688 [PMID: 12404098 DOI: 10.1038/sj.ejhg.5200859]
- 140 **van Os J**, Selten JP. Prenatal exposure to maternal stress and subsequent schizophrenia. The May 1940 invasion of The Netherlands. *Br J Psychiatry* 1998; **172**: 324-326 [PMID: 9715334 DOI: 10.1192/bjp.172.4.324]
- 141 **Donkin I**, Versteyhe S, Ingerslev LR, Qian K, Mechta M, Nordkap L, Mortensen B, Appel EV, Jørgensen N, Kristiansen VB, Hansen T, Workman CT, Zierath JR, Barrès R. Obesity and Bariatric Surgery Drive Epigenetic Variation of Spermatozoa in Humans. *Cell Metab* 2016; **23**: 369-378 [PMID: 26669700 DOI: 10.1016/j.cmet.2015.11.004]
- 142 **Marques CJ**, João Pinho M, Carvalho F, Bièche I, Barros A, Sousa M. DNA methylation imprinting marks and DNA methyltransferase expression in human spermatogenic cell stages. *Epigenetics* 2011; **6**: 1354-1361 [PMID: 22048249 DOI: 10.4161/epi.6.11.17993]
- 143 **Wyatt R**, Wilson AM. Children of anaesthetists. *Br Med J* 1973; **1**: 675 [PMID: 4692717 DOI: 10.1136/bmj.1.5854.675-a]
- 144 **Gupta D**, Kaminski E, McKelvey G, Wang H. Firstborn offspring sex ratio is skewed towards female offspring in anesthesia care providers: A questionnaire-based nationwide study from United States. *J Anaesthesiol Clin Pharmacol* 2013; **29**: 221-227 [PMID: 23878446 DOI: 10.4103/0970-9185.111728]
- 145 **Nagella AB**, Ravishankar M, Hemanth Kumar VR. Anaesthesia practice and reproductive outcomes: Facts unveiled. *Indian J Anaesth* 2015; **59**: 706-714 [PMID: 26755835 DOI: 10.4103/0019-5049.170028]
- 146 **Nagella AB**, Ravishankar M, Hemanth Kumar VR. Anaesthesia practice and reproductive outcomes: Facts unveiled. *Indian J Anaesth* 2016; **60**: 225 [PMID: 27053794 DOI: 10.4103/0019-5049.177883]
- 147 **Gupta D**. Firstborn female offsprings are significantly more common among Indian anaesthesiologists as compared to national child sex ratio. *Indian J Anaesth* 2016; **60**: 224 [PMID: 27053793 DOI: 10.4103/0019-5049.177881]
- 148 **Chastain-Potts SE**, Tesic V, Tat QL, Cabrera OH, Quillinan N, Jevtovic-Todorovic V. Sevoflurane Exposure Results in Sex-Specific Transgenerational Upregulation of Target IEGs in the Subiculum. *Mol Neurobiol* 2020; **57**: 11-22 [PMID: 31512116 DOI: 10.1007/s12035-019-01752-0]
- 149 **Ju LS**, Yang JJ, Xu N, Li J, Morey TE, Gravenstein N, Seubert CN, Setlow B, Martynyuk AE. Intergenerational Effects of Sevoflurane in Young Adult Rats. *Anesthesiology* 2019; **131**: 1092-1109 [PMID: 31517640 DOI: 10.1097/ALN.0000000000002920]
- 150 **Tang CK**, Chalton J, Markham JP, Ramanathan S, Turndorf H. Exposure of sires to enflurane affects learning function of murine progeny. *Anesth Analg* 1984; **63**: 729-730 [PMID: 6465557]
- 151 **Swerdlow NR**, Weber M, Qu Y, Light GA, Braff DL. Realistic expectations of prepulse inhibition in translational models for schizophrenia research. *Psychopharmacology (Berl)* 2008; **199**: 331-388 [PMID: 18568339 DOI: 10.1007/s00213-008-1072-4]
- 152 **Greenwood TA**, Light GA, Swerdlow NR, Calkins ME, Green MF, Gur RE, Gur RC, Lazzaroni LC, Nuechterlein KH, Olincy A, Radant AD, Seidman LJ, Siever LJ, Silverman JM, Stone WS, Sugar CA, Tsuang DW, Tsuang MT, Turetsky BI, Freedman R, Braff DL. Gating Deficit Heritability and Correlation With Increased Clinical Severity in Schizophrenia Patients With Positive Family History. *Am J Psychiatry* 2016; **173**: 385-391 [PMID: 26441157 DOI: 10.1176/appi.ajp.2015.15050605]
- 153 **Graham FK**. Presidential Address, 1974. The more or less startling effects of weak prestimulation. *Psychophysiology* 1975; **12**: 238-248 [PMID: 1153628 DOI: 10.1111/j.1469-8986.1975.tb01284.x]
- 154 **Braff DL**, Geyer MA. Sensorimotor gating and schizophrenia. Human and animal model studies. *Arch Gen Psychiatry* 1990; **47**: 181-188 [PMID: 2405807 DOI: 10.1001/archpsyc.1990.01810140081011]
- 155 **Morales-Muñoz I**, Jurado-Barba R, Ponce G, Martínez-Gras I, Jiménez-Arriero MA, Moratti S, Rubio G. Characterizing cannabis-induced psychosis: a study with prepulse inhibition of the startle reflex. *Psychiatry Res* 2014; **220**: 535-540 [PMID: 25175914 DOI: 10.1016/j.psychres.2014.08.010]
- 156 **Tervahartiala K**, Karlsson L, Peltto J, Kortessluoma S, Hyttinen S, Ahtola A, Junntila N, Karlsson H. Toddlers' diurnal cortisol levels affected by out-of-home, center-based childcare and at-home, guardian-supervised childcare: comparison between different caregiving contexts. *Eur Child Adolesc Psychiatry* 2019 [PMID: 31705206 DOI: 10.1007/s00787-019-01432-3]
- 157 **Landram MJ**, Koch AJ, Mayhew JL. Salivary stress hormone response and performance in full competition after linear or undulating periodization training in elite powerlifters. *J Sports Med Phys Fitness* 2020; **60**: 152-159 [PMID: 31663314 DOI: 10.23736/S0022-4707.19.09977-8]
- 158 **Sanders RD**, Hassell J, Davidson AJ, Robertson NJ, Ma D. Impact of anaesthetics and surgery on neurodevelopment: an update. *Br J Anaesth* 2013; **110** Suppl 1: i53-i72 [PMID: 23542078 DOI: 10.1093/bja/aet054]
- 159 **Stratmann G**. Review article: Neurotoxicity of anesthetic drugs in the developing brain. *Anesth Analg* 2011; **113**: 1170-1179 [PMID: 21965351 DOI: 10.1213/ANE.0b013e318232066c]
- 160 **US Food and Drug Administration**. FDA drug safety communication: FDA review results in new warnings about using general anesthetics and sedation drugs in young children and pregnant women. [accessed 2017 Aug 25]. Available from: <https://www.fda.gov/Drugs/DrugSafety/ucm532356.htm>



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