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Systemic exposure to *Ginkgo biloba* extract in male F344/NCrl rats: Relevance to humans

Suramya Waidyanatha^{1,*}, Esra Mutlu¹, Seth Gibbs², Billie Stiffler², Jon Andre², Brian Burbach², Cynthia V. Rider¹

¹Division of the National Toxicology Program, National Institute of Environmental Health Sciences, Research Triangle Park, NC, USA;

²Battelle, Columbus, OH, USA

Abstract

Ginkgo biloba extract (GBE) is a popular botanical dietary supplement used worldwide and the safety of use is a public health concern. While GBE is a complex mixture, the terpene trilactones and flavonol glycosides are believed to elicit the pharmacological and/or toxicological effects of GBE. In a National Toxicology Program (NTP) 2-year rodent bioassay with GBE, hepatotoxicity was observed in rodents (100 mg/kg in rats, 200 mg/kg in mice). Subsequently, questions arose about whether or not the GBE used in NTP studies was representative of other GBE products and how rodent doses are related to human doses. To address these, we generated systemic exposure data for terpene trilactones in male rats following oral administration of 30, 100, and 300 mg/kg GBE test article from the 2-year bioassay. Dose-normalized C_{max} and AUC_{∞} for terpene trilactones from the current study were within 5-fold of published rodent studies using a standardized GBE preparation. Comparison of our rat systemic exposure data at 100 mg/kg GBE to published human data following ingestion of 240 mg GBE-containing product showed that the rat/human exposure multiple was 3 to 22, for terpene trilactones. These data demonstrate the relevance of NTP rodent toxicity data to humans.

Keywords

Botanical dietary supplements; *Ginkgo biloba* extract; Terpene trilactones; Flavonols; Flavonol glycosides; Systemic exposure

Introduction

The botanical dietary supplement industry is a multibillion-dollar industry worldwide with sales surpassing 8 billion dollars in the U.S. in 2017 (Smith et al., 2018). According to a National Health Information Survey conducted in 2012, approximately 18% of adults in the United States are using these supplements (Clark et al., 2015). Currently, there are over

*Correspondence: Suramya Waidyanatha, Division of the National Toxicology Program, National Institute of Environmental Health Sciences, P.O. Box 12233, Drop K2-07, Research Triangle Park, North Carolina 27709, waidyanathas@niehs.nih.gov.

Declaration of Interest

The authors report no conflicts of interest

26,000 supplements in this category in the United States marketplace, according to the Dietary Supplement Label Database (<https://dsld.nlm.nih.gov/dsld/>). Because of widespread use and limited availability of safety data, these products represent an important public health concern.

Ginkgo biloba leaf extract (GBE) is a popular botanical dietary supplement used worldwide for its purported cognitive and cardiovascular benefits (Brinkley et al., 2010; DeKosky et al., 2008; Snitz et al., 2009). Other reported beneficial activities of GBE include antioxidative (Goh and Barlow, 2002), antitumor (Agha et al., 2001) and anti-inflammatory (Lee et al., 2011) actions. Approximately 3 million people in the U.S. use dietary supplements containing GBE with a recommended daily dose of 120 to 240 mg (DeFeudis, 2003). Some reports indicate that GBE-containing products are generally well-tolerated in humans (Diamond et al., 2000; Liebermann et al., 2017).

GBE is a complex mixture containing different classes of constituents such as flavonol glycosides, terpene trilactones, proanthocyanidines, ginkgolic acids, biflavones, polyflavones and ginkgotoxins (Singh et al., 2008; Leistner and Drewke, 2009). The purported pharmacological properties of GBE are believed to be due to flavonol glycosides and terpene trilactones (Ding et al., 2009), although unlike terpene trilactones which are unique to *Ginkgo biloba*, flavonol glycosides are ubiquitous in the plant kingdom. It is accepted that GBE should contain 22–27% flavonol glycosides, 5–7% terpene trilactones and 5 ppm ginkgolic acids (van Beek, 2002). Standardized GBEs in the marketplace should conform to these established levels, including the unofficial industry gold standard, EGb761®, produced by Dr. Wilmar Schwabe Pharmaceuticals (Karlsruhe, Germany). However, a wide range of levels have been reported in GBE products for terpene trilactones (27–358% of standardized) and flavonol glycosides (86–418% of standardized) (Fransen et al., 2010). This variability poses a challenge for selecting a representative GBE product for efficacy and/or toxicity testing.

Absorption, distribution, metabolism and excretion (ADME) studies of GBE have focused mainly on the two purported active classes, terpene trilactones and flavonol glycosides. There is a significant amount of work characterizing the ADME properties of terpene trilactones (e.g., bilobalide, ginkgolides A, B, C, and J). Terpene trilactones are well-distributed in rodents and are highly bioavailable following ingestion in both rodents and humans with short plasma elimination half-lives (rodents, 1–3 hours; humans, 2–11 hours) (Biber and Koach, 1999; Biber 2003; Cheng et al., 2013; Drago et al., 2002; Fourtillian et al., 1995; Kressmann et al., 2002; Mauri et al., 2001, 2006; Rossi et al., 2009; Zheng et al., 2015; Zheng et al., 2016; Woelkart et al., 2010; Ude et al., 2011; summarized in Ude et al., 2013). Data on the metabolism of terpene trilactones are limited. The work by Wang et al. (2008) demonstrated that terpene trilactones are metabolized by cleavage of the lactone ring and/or hydroxylation of the terpene backbone. Flavonol glycosides on the other hand are less studied, likely due to their existence as a variety of glycosides and poor bioavailability (Liu and Hu, 2007; Pietta et al., 1995, 1997). These properties may have led to low systemic concentrations following ingestion, making measurement of compounds in this class analytically challenging. Flavonol glycosides are studied as corresponding aglycone, flavonol, (e.g., quercetin, kaempferol, and isorhamnetin) following hydrolysis of

corresponding glycosides. Reported plasma elimination half-lives, depending on the flavonol, were 2–4 hours in humans (Ding et al., 2006; Wójcicki et al., 1995) and 19–24 hours in rodents (Rangel-Ordóñez et al., 2010; Zheng et al., 2015). It has been suggested that neither the flavonol glycosides nor the aglycones, but other downstream metabolites are responsible for pharmacological effects of GBE (summarized in Ude et al., 2013).

There have been a handful of preclinical studies evaluating the toxicity of GBE in rodents. Following oral exposure of rats to 4, 20, and 100 mg/kg EGb761®, no treatment-related effects were reported, although details of the study are not available (Woerdenbag and De Smet, 2000). In another study, following oral exposure of rats and mice to a GBE preparation (details not provided), with doses ranging from 100 to 1600 mg/kg, there was no evidence of treatment-related effects (Salvador, 1995). The lack of toxicity reported in these studies from the literature contrasts with observed toxicity in the 90-day and 2-year GBE studies conducted by the National Toxicology Program (NTP). In the NTP 2-year rodent bioassay, GBE demonstrated clear hepatotoxicity in rats (100, 300, and 1000 mg/kg GBE) and mice (200, 600, and 2000 mg/kg GBE) of both sexes following gavage administration (NTP, 2013; Rider et al., 2014). The GBE preparation used in these studies contained 31.2% flavonol glycosides, 15.4% terpene trilactones, and 10.5 ppm ginkgolic acid (NTP, 2013). During the public reporting of the NTP GBE studies, questions arose regarding the relevance of the findings to human exposure. These questions centered around whether or not: 1) the test article was relevant to other GBE products on the market and 2) the doses used in the rodent bioassays were relevant to human exposure levels.

Data from well-designed ADME studies could help in understanding how different constituent levels among different GBE products might impact observed effects and allow for comparison of animal doses to human exposure levels. Ultimately, these data will aid in understanding the human health risk posed by GBE. As such, a study in rats was designed to generate systemic exposure data to aid in bridging this gap. The rat strain, route of exposure, and the GBE used were the same as those used in the NTP 2-year bioassay (NTP, 2013; Rider et al., 2014). The doses selected were 30, 100, and 300 mg/kg. The two highest doses were the same as those used in the 2-year bioassay; a lower dose was also included to generate data closer to the human recommended daily dose.

Materials and methods

Chemicals and reagents:

GBE (CASRN 90045-36-6; lot # 020703) used in the current study (and NTP 2-year bioassay) was obtained from Shanghai Xing Ling Science and Technology Pharmaceutical Company, Limited (Shanghai, China). The GBE lot was characterized as described elsewhere (Rider et al., 2014; NTP, 2013). The lot contains ~31% flavonol glycosides (16.7% quercetin, 12.2% kaempferol, and 2.4% isorhamnetin), ~15% terpene lactones (6.9% bilobalide, 3.7% ginkgolide A, 1.6% ginkgolide B, 3.1% ginkgolide C), and 10.5 ppm ginkgolic acids. Standards of ginkgolide A, ginkgolide C, isorhamnetin, and quercetin were obtained from Sigma-Aldrich (St. Louis, MO). Standards of bilobalide, ginkgolide B, ginkgolide J, and kaempferol hydrate were obtained from TCI America (Portland, OR). Hesperetin and limonene (used as internal standards) were also obtained from Sigma-

Aldrich (St. Louis, MO). All other chemicals and reagents were procured from commercial sources.

Animals:

Male F344/NCrl rats were obtained from Charles River Kingston (Stone Ridge, NY). The animals were quarantined for at least 7 days before they were randomized using Provantis software (Instem, Stone, UK) into dosing groups and used in the study. Animals had *ad libitum* access to certified, irradiated NTP 2000 wafer feed (Ziegler Bros, Inc., Gardners, PA) and city (West Jefferson, OH) tap water. Feed was used within 120 days of milling. Environmental conditions included: room temperature 69 to 75 °F (22 ± 2 °C), relative humidity 35 to 65% and a 12-h light/dark cycle. Animals were 7–8 weeks old at the time of dosing.

Animal studies were conducted at Battelle (Columbus, OH) and approved by the Institutional Animal Care and Use Committee. Animals were housed in facilities that are fully accredited by the Association for Assessment and Accreditation of Laboratory Animal Care International. Animal procedures were in accordance with the “Guide for the Care and Use of Laboratory Animals” (National Research Council, 2011).

Study design and plasma collection:

Oral dose formulations of GBE were prepared in corn oil at 6, 20, and 60 mg/mL and stored at ambient temperature. Formulations were analyzed by liquid chromatography-mass spectrometry (LC-MS) using a Shimadzu LC-20AD LC (Columbia, MD) coupled to an AB Sciex tandem triple quadrupole mass spectrometer (Framingham, MA) and a Phenomenex Prodigy ODS 3 column (250 mm \times 4.6 mm, 5 μ m) (Torrance, CA). Mobile phases A (water:methanol, 9:1, with 0.1% formic acid) and B (methanol with 0.1% formic acid) were run at an initial condition of 15% B (5 min) and then changed linearly to 95% B in 10 min. The turbospray ion source was set at 400 °C in negative ion mode. Analytes quantified were bilobalide and quercetin representing terpene trilactone and flavonol classes, respectively. Transitions monitored were m/z 325 \rightarrow 163 and m/z 301 \rightarrow 151 for bilobalide and quercetin, respectively. All formulations were within 10% of target concentration except for quercetin at the lowest concentration where it was within 20% of target concentration. Prior to study initiation, formulation stability was confirmed for up to 42 days when stored in sealed plastic bottles at approximately 5 °C and at least 3 hours under simulated animal room conditions (within 10% of the day 0 value).

A single dose of GBE was administered by intragastric gavage at 30, 100, and 300 mg/kg ($n = 18$ animals per dose group) in a volume of 5 mL/kg using a syringe equipped with a ball-tipped 16-gauge gavage needle. Following dose administration, blood was collected using K₃EDTA as an anticoagulant from 3 animals per time point at target times of 0 (predose), 5, 10, 15, 30, 60, 90, 120, 240, 480, 720 and 1440 minutes from all dose groups. Two blood samples were obtained from each rat; the first sample (~ 500 μ L) was obtained via retro-orbital plexus under CO₂/O₂ anesthesia after which the animal was returned to its home cage pending a second blood collection. Terminal blood collection was via cardiac puncture following euthanasia with CO₂. The blood sampling times were separated by at least 2 hours

for each rat. For non-terminal collections the total volume collected did not exceed 2% of body weight. The actual times for blood collection were recorded at each time point. Plasma was isolated from blood within 1 hour of collection. All samples were stored at -70°C within 2 hours of collection.

Quantitation of terpene trilactones and flavonols:

An LC-MS/MS method based on literature methods was developed to quantify a suite of GBE constituents in plasma (Guan et al., 2014; Zheng et al., 2016). Plasma was hydrolyzed to convert flavonol glycosides to corresponding aglycones. The analytes quantified were bilobalide, ginkgolides A, B, C, and J, quercetin, kaempferol, and isorhamnetin (Figure 1).

Two stock solutions of combined terpene trilactone and flavonol standards were prepared in acetonitrile:[methanol:water (7:3)] (1:1), except isorhamnetin where acetonitrile:water (1:1) was used. The weight of kaempferol hydrate was adjusted to the anhydrous weight prior to preparing solutions. Standards in the working range (20 to 20,000 ng/mL depending on the analyte) were prepared by diluting alternate stock solutions in methanol:water (7:3). Stock solutions of internal standards (hesperetin and limonene) were prepared in methanol:water (7:3) and diluted in water to prepare a combined working internal standard solution at 500 ng/mL with respect to each analyte.

Plasma calibration standards (ginkgolide A, 4.3 to 400 ng/mL; ginkgolides B, C, 1 to 200 ng/mL; ginkgolide J, 10 to 1000 ng/mL; bilobalide, 5 to 1000 ng/mL; quercetin, 3 to 100 ng/mL, kaempferol, 40 to 500 ng/mL; isorhamnetin, 5 to 50 ng/mL) were prepared by adding analytes to plasma using working standard solutions.

For the determination of terpene trilactones and flavonols mentioned above, 100 μL plasma aliquots were transferred to microcentrifuge tubes and 20 μL of internal standard solution were added. To each tube 100 μL 4M hydrochloric acid/20 mM ascorbic acid were added, and the samples were digested at 80°C for 30 minutes. Following digestion, samples were extracted with 1 mL of ethyl acetate, supernatants were collected and evaporated to dryness under a stream of nitrogen, and residues were reconstituted in 100 μL methanol.

Methanol extracts were analyzed by LC-MS/MS using a Waters Acquity QSM (Milford, MA) LC coupled to a XEVO TQ-S mass spectrometer. Chromatography was performed on a Waters Acquity BEH C18 column (100×2.1 mm, $1.7 \mu\text{m}$) (Milford, MA). Mobile phases A (0.1% aqueous formic acid) and B (0.1% formic acid in acetonitrile) were run with initial condition of 5% B for 0.5 minutes followed by 5 to 70% B over 4.5 min with a 2 min hold at a flow rate of 0.4 mL/min. The column temperature was maintained at 40°C . The electrospray ion source was operated in negative mode. Due to the large number of analytes to be quantified, samples were injected twice and monitored in 2 groups as given in Table 1; corresponding ion source temperatures were 450°C and 550°C and ion spray voltages were -3000 or -4500 V. The transitions monitored, corresponding retention times, and the respective internal standard used are given in Table 1.

A linear regression with $1/X$ weighing was used to relate LC-MS/MS peak area response ratio of analyte to internal standard and analyte concentration. The concentration of each

analyte was calculated using its response ratio, the regression equation, and dilution when applicable. The concentration of analytes was expressed as ng/mL of plasma. The method was linear ($r = 0.97$ for all analytes except kaempferol where it was 0.87) with limits of quantitation (LOQ) 10 ng/mL and limits of detection (LOD) 5 ng/mL for all analytes except kaempferol where LOQ and LOD was 40 ng/mL. Average accuracy determined as relative error (RE) was $\pm 16\%$ and precision determined as relative standard deviation (RSD) in general was 20% for terpene trilactones. Of the flavanols, average RE and RSD for quercetin was $\pm 16\%$ and 23% , respectively. For kaempferol and isorhamnetin RE was $\pm 16\%$, but RSD was higher with values 33% .

Toxicokinetic analysis:

All concentrations above LODs were used. Individual animal data were evaluated for aberrant concentrations and time points. The actual blood collection times were within 5% from nominal time for timepoints ≤ 4 h and within 15 min for timepoints greater than 4 h and hence, the nominal collection times were used for TK analysis. All concentrations were evaluated to identify outliers by performing Q-tests. Based on these assessments, no values were eliminated from TK analysis.

Group mean plasma concentration versus time curves were analyzed using non-compartmental analysis (NCA) with Phoenix WinNonlin software (Version 7.0, Certara L.P., Princeton, NJ). When using the NCA models, no weighting factors were used to obtain the initial estimates. Also, the software algorithm was allowed to select the points used to calculate slope of the terminal linear phase based on determining an optimal coefficient of determination.

Results

Terpene trilactones:

All terpene trilactones were detected in plasma from rats following oral administration of GBE. Bilobalide, ginkgolide B, and ginkgolide C were quantifiable in all samples except for the 1440 minute time point for the 30 and 100 mg/kg groups. Ginkgolide A was quantifiable in all samples through 720 minutes. For ginkgolide J, all samples from the 30 mg/kg group were below LOD; for the 100 mg/kg, levels were quantifiable from 30 to 240 minutes and for 300 mg/kg from 5 to 480 minutes.

Plasma concentration-versus-time plots are given in Figure 2 for the 300 mg/kg group as an example. Toxicokinetic data for all dose groups are presented in Table 2. For all terpene trilactones, C_{max} and AUC_{∞} increased with the dose but with an apparent less than dose-proportional increase (Table 2). The rank order for C_{max} and AUC_{∞} was bilobalide $>$ ginkgolide A $>$ ginkgolide B $>$ ginkgolide C $>$ ginkgolide J which reflects their abundance in the GBE preparation used in the study (NTP, 2013). Plasma elimination half-lives estimated were similar for terpene trilactones with values ranging from 127 to 230 minutes. The plasma elimination half-life for ginkgolide J could not be estimated since the terminal phase could not be defined due to insufficient timepoints above the LOD for the analyte.

Flavonols:

All three flavonols were detected in plasma from rats. Quercetin was quantifiable in all samples from the study. Isorhamnetin was also quantifiable in all samples from the study, however, the analytical method was less precise for isorhamnetin than for quercetin (see methods). Kaempferol was not detected in plasma from 30 and 100 mg/kg dose groups; in the 300 mg/kg dose group only 4 time points had quantifiable levels of kaempferol. This was unexpected given that the GBE preparation used had kaempferol levels similar to quercetin and higher than isorhamnetin levels (16.71% quercetin, 12.20% kaempferol, and 2.37% isorhamnetin) and may reflect the lower analytical method sensitivity for kaempferol compared to the other 2 analytes (LOD: quercetin, 1 ng/mL; kaempferol, 40 ng/mL; isorhamnetin, 5 ng/mL). For these reasons, kaempferol data are not reported.

Plasma concentration-versus-time plots for quercetin and isorhamnetin are given in Figure 3 for the 300 mg/kg group as an example. Toxicokinetic data are presented in Table 2. For 300 mg/kg quercetin and 100 and 300 mg/kg isorhamnetin, plasma elimination half-life and AUC_{∞} could not be estimated because the elimination phase could not be well defined due to insufficient timepoints above the LOD for those analytes. For both analytes, C_{max} and AUC_{∞} increased with dose (AUC_{last} was used in the absence of AUC_{∞}) (Table 2). At 300 mg/kg dose, the rank order for C_{max} and AUC_{∞} was quercetin > isorhamnetin, which is the same as their respective abundance within the GBE preparation used in the study (NTP, 2013).

Plasma elimination half-lives for quercetin were ~35 and 9.5 h, respectively, following a single administration of 30 and 100 mg/kg, suggesting a decrease in half-life with increasing dose. Plasma elimination half-life for isorhamnetin at 30 mg/kg was ~55 h and was similar to that of quercetin at a similar dose.

Discussion

There are numerous GBE preparations including tinctures, capsules, and tablets, which can contain GBE alone or in combination with other botanical ingredients in the marketplace. Despite the availability of specifications for standardized GBE (22–27% flavonol glycosides, 5–7% terpene trilactones and 5 ppm ginkgolic acids), the composition varies significantly between different GBE products (Fransen et al., 2010). The variation can arise from numerous sources including differences in raw plant material, storage, extraction procedures, as well as both intentional (e.g., drugs) or economically-motivated (e.g., alternate plant species or less expensive material) adulteration (Shipkowski et al., 2018). The complex chemistry of GBE combined with variability of products in the marketplace have made selecting a representative GBE product for safety testing challenging. In the current investigation we 1) designed a study to evaluate the systemic exposure to GBE constituents following dosing with the GBE preparation used in the NTP 2-year bioassay and compared the data to values from the literature from a study using a standardized GBE preparation, and 2) compared the NTP 2-year bioassay doses to human exposure levels using systemic exposure parameters in our rodent study and human data from the literature to determine the relationship between the NTP 2-year doses and recommended doses in people.

In F344/NCrl rats, terpene trilactones were absorbed rapidly following exposure of rats to a single dose of GBE, with both C_{max} and AUC_{∞} of GBE constituents increasing with dose. Interestingly, systemic exposure parameters for terpene trilactones reflected the abundance of these constituents in the GBE preparation suggesting disposition of these constituents in the rat was similar. Plasma elimination half-lives estimated for terpene trilactones in the current study (~ 2 to 4 hours) were similar to those reported for rats previously (~ 2 to 7 hours) (Zheng et al., 2016; Biber and Koch, 1999; summarized in Ude et al., 2013). The analytical method did not perform as well for the flavonols. The accuracy and precision were lower for kaempferol and hence, data for this analyte were not reported. Based on the data generated for quercetin and isorhamnetin, systemic exposure of flavonols also tracked with the abundance of the flavonols in the GBE preparation fairly well, with quercetin showing higher values than isorhamnetin. The reported plasma elimination half-lives (19–24 h) following a single administration of 600 mg/kg GBE in rats (Rangel-Ordóñez et al., 2010) were in the same range as reported in the current study (9.5 – 55 h).

The systemic exposure data obtained from the current study for terpene trilactones were compared, after normalizing to the dose of GBE administered, with two published studies where male rats were administered a single oral dose of standardized GBE preparations (Biber, 2003; Zheng et al., 2016) (Table 3). In this comparison, the dose-normalized C_{max} was 3-fold for bilobalide, 2-fold for ginkgolide A and were similar for ginkgolide B between studies. Similarly, dose-normalized AUC_{∞} was 5-fold for bilobalide, 3.5-fold for ginkgolide A, and 2-fold for ginkgolide B between studies (Table 3). These findings demonstrate the similarities between the GBE preparations used between studies.

Next, the systemic exposure data obtained from our rat study were compared with data available in the literature following exposure of humans to a clinically-relevant dose of standardized GBE, EGB761®. A single clinical dose of 240 mg in a 70 kg-person translates to 3.4 mg GBE/kg/day. When human and rat doses were compared, based on mg/kg dose, rat doses of 30 and 300 mg/kg/day were 9- and 88-fold higher than the relevant human dose of 3.4 mg/kg/day, respectively (Table 4). To account for different metabolic rates of the two species, the mg/kg/day doses were converted to $mg/m^2/day$, using a K_m value of 6 for rats and 37 for humans (Reagan-Shaw et al., 2008). Based on the $mg/m^2/day$ dose, a rat dose of 100 mg/kg was only 5 times higher than the clinically relevant daily dose of 240 mg (3.4 mg/kg/day) (Table 4). A similar comparison was made between rats and humans using AUC_{∞} data available in humans for terpene trilactones (Biber, 2003), which takes into account species differences in ADME. For example, comparison of rat data to human data indicates that a 30 mg/kg dose in rats is equivalent to a typical 240 mg/day dose in humans based on ginkgolide B, but with ginkgolide A and bilobalide, a 30 mg/kg dose in rats is 3- and 10-fold higher, respectively than the 240 mg/day dose in humans. When a similar comparison was made using rat data for 100 mg/kg dose, the rat/human exposure multiple ranged from 3 to 22, depending on the terpene trilactone used for comparison. The estimated rat/human exposure multiple was 3 to 8 when data previously reported by Biber and Koch (1999) in Sprague Dawley rats using EGB761® was used for this comparison, suggesting minimal difference in the 2 types of GBE preparations used (Table 4). Preliminary work that has compared 12 GBE constituents including terpene trilactones, ginkgolic acids, ginkgotoxin and flavonols to hepatotoxicity markers showed that the terpene trilactones

correlate with increased liver weight and cholesterol in rats, as well as associated changes in gene expression (unpublished data). Since the observed hepatotoxicity could be due to a combination of the terpene trilactones, it may be more relevant to use total levels of bilobalide, ginkgolide A, ginkgolide B, and ginkgolide C to estimate the totality of exposure because the disposition of individual terpene trilactones in the class may vary between animals and humans. However, it should be noted that other GBE constituents, namely ginkgolic acids have also been shown to elicit cytotoxicity in liver cells *in vitro* (Qing-Qing, 2018) and hepatotoxicity in rats at doses of 100 or 900 mg/kg ginkgolic acid extract (Qian et al., 2017).

Taken collectively, these data demonstrate that systemic exposure to constituents from the GBE preparation used in our studies were similar to those of standardized GBE preparations. In addition, systemic exposure to GBE-specific terpene trilactones, following a 100 mg GBE/kg/day dose in rats, where liver effects were observed, was similar to a single clinically-relevant dose of 240 mg (3.4 mg/kg/day) in humans, demonstrating the relevance of NTP rodent toxicity data to humans. The question of whether the mechanism of toxicity and observed effects in animal studies are relevant to humans requires further consideration by the risk assessment community.

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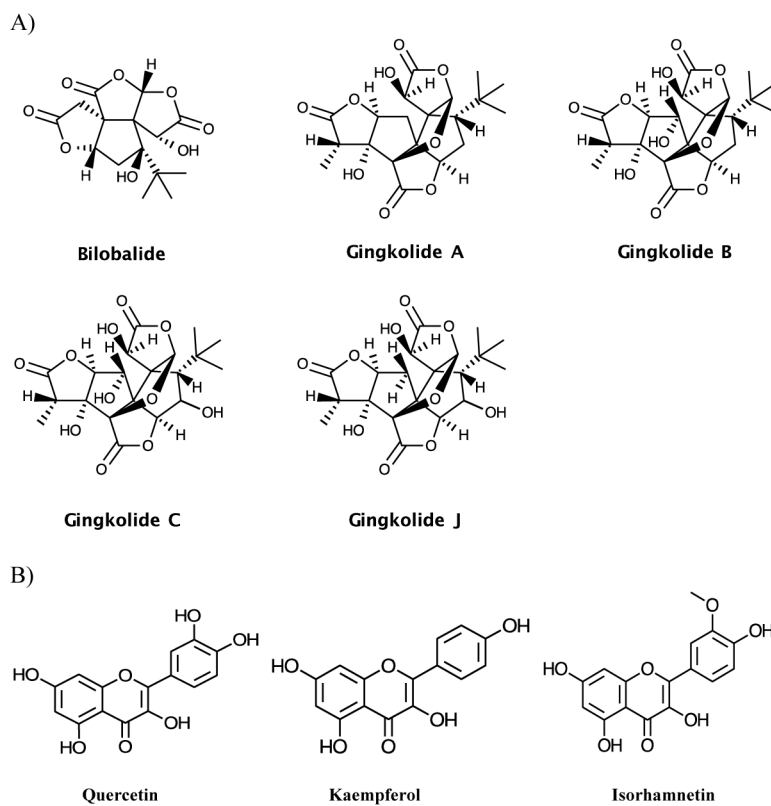
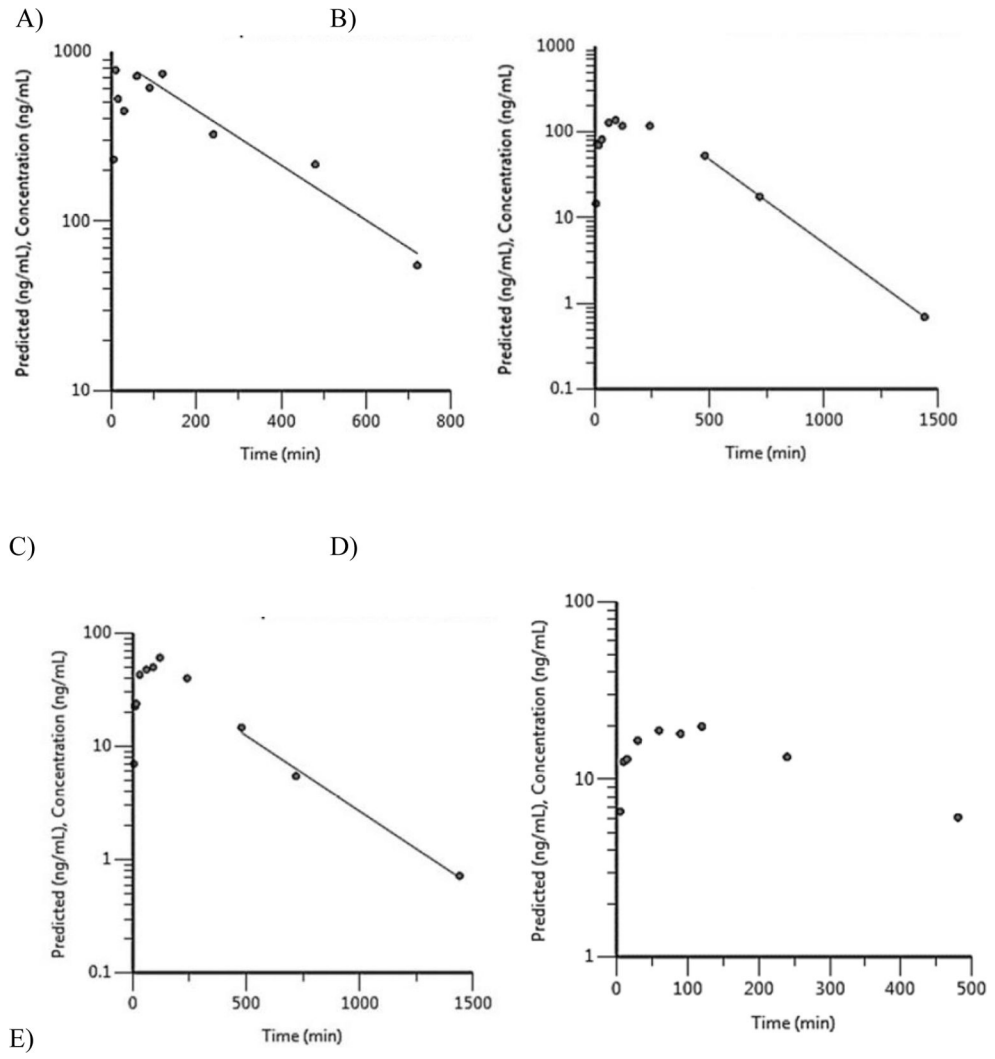


Figure 1.
Constituents of *Ginkgo biloba* extract quantified A) terpene trilactones B) flavonols



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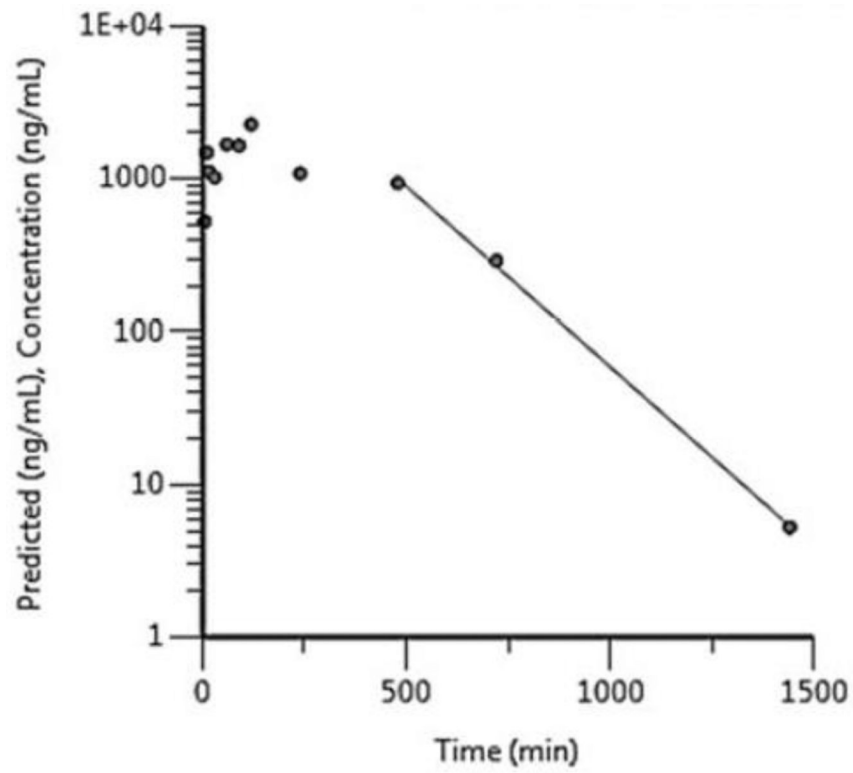


Figure 2. Plasma concentration versus time profiles for terpene trilactones following a single gavage administration of 300 mg/kg *Ginkgo biloba* extract in male F344/NCrl rats. A) ginkgolide A, B) ginkgolide B, C) ginkgolide C, D) ginkgolide J, and E) bilobalide.

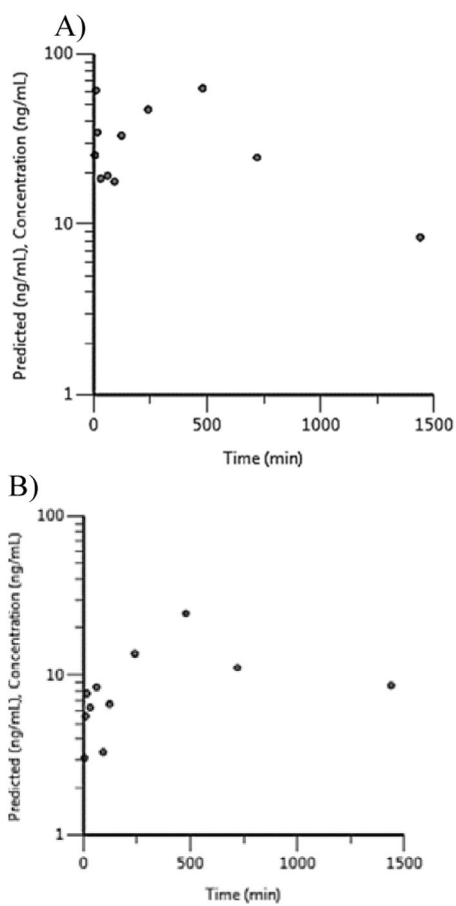


Figure 3. Plasma concentration versus time profiles for flavonols following a single gavage administration of 300 mg/kg *Ginkgo biloba* extract in male F344/NCrl rats. A) quercetin B) isorhamnetin

Table 1.

Transitions monitored for the quantitation of *Ginkgo biloba* extract constituents and corresponding retention times

	Analyte	Abbreviation	Retention Time (min)	Transition (m/z)
Group 1 ^a	Ginkgolide A	GLA	4.27	407 → 229
	Ginkgolide C	GLC	3.66	439 → 383
	Bilobalide	BLL	3.69	325 → 163
	Quercetin	QCT	4.73	301 → 151
	Limonene	LIM	5.20	469 → 229
	Hesperetin	HES	4.74	301 → 164
Group 2	Ginkgolide B	GLB	4.45	423 → 367
	Ginkgolide J	GLJ	3.63	423 → 349
	Kaempferol	KMF	4.69	285 → 93
	Isorhamnetin	ISR	4.75	315 → 300
	Limonene	LIM	5.20	469 → 229
	Hesperetin	HES	4.74	301 → 164

^aEach sample was injected twice and ions monitored in each injection are given as groups 1 or 2.

Table 2.

Plasma toxicokinetic parameters for selected constituents following a single gavage administration of *Ginkgo biloba* extract in male F344/NCrl rats^a

Analyte	Dose (mg/kg)	T _{max_obs} ^b (min)	C _{max_obs} (ng/mL)	Half-life (min)	AUC _{last} (min*ng/mL)	AUC _∞ (min*ng/mL)
BLL	30	15	326	230	132000	148000
	100	120	714	162	301000	319000
	300	120	2280	127	880000	881000
GLA	30	15	127	230	36900	41600
	100	60	277	156	95100	99100
	300	10	781	186	233000	251000
GLB	30	90	23.2	197	9020	9820
	100	60	65.2	169	24400	25800
	300	90	138	154	62600	62700
GLC	30	240	9.1	ND ^c	3790	ND
	100	60	38.3	134	11500	11800
	300	120	60.7	226	22500	22700
GLJ ^d	100	60	9.9	ND	2000	ND
	300	120	19.9	ND	6350	ND
ISR	30	15	5.92	3320	6710	25700
	100	720	6.81	ND	8430	ND
	300	480	24.6	ND	18000	ND
QCT	30	120	8.15	2120	4780	12400
	100	10	13.2	570	13500	17100
	300	480	63.4	ND	43400	ND

^aEstimated using non-compartmental analysis

^bT_{max_obs}, observed to reach maximum concentration; C_{max_obs}, observed maximum concentration; AUC_{last}, area under the concentration versus time curve to last timepoint; AUC_∞, area under the concentration versus time curve to infinity; BLL, bilobalide; GLA, ginkgolide A; GLB, ginkgolide B; GLC, ginkgolide C; GLJ, ginkgolide J

^cND, not determined. Elimination phase could not be defined and therefore half-life and AUC_∞ could not be estimated

^dConcentrations for 30 mg/kg were below the detection limit of the assay (5 ng/mL)

Table 3.

Comparison of dose-normalized systemic exposure parameters for selected terpene trilactones in rats using different *Ginkgo biloba* extract preparations

Species	Dose (mg/kg)	Dose-normalized C _{max} (ng/mL)/(mg/kg)			Dose-normalized AUC _∞ (ng.min/mL)/(mg/kg)		
		BLL ^a	GLA	GLB	BLL	GLA	GLB
Rat ^b	30	10.9	4.2	0.77	4933	1387	327
	100	7.1	2.8	0.65	3190	991	258
	300	7.6	2.6	0.46	2937	837	209
Rat ^c	30	5.3	2.3	1.3	994	441	286
	55	5.0	2.7	1.3	968	425	277
	100	4.0	3.0	1.0	1199	467	221
Rat ^d	25	4.1	2.1	0.8	922	399	179

^aBLL, bilobalide; GLA, ginkgolide A; GLB, ginkgolide B

^bData from the current study. GBE used was Shanghai Xing Ling Science and Technology Pharmaceutical Company, Ltd. (lot # GBE-50-001003) (NTP, 2013)

^cData from Biber and Koch (1999). GBE used was EGB761[®]. Dose-normalized values shown are after adjusting for comparable units for C_{max} and AUC_∞ to the current study

^dData from Zheng et al., 2016. GBE used was standardized Ginaton. Dose-normalized values shown are after adjusting for comparable units to the current study

Table 4.

Comparison of exposure multiples between rats^a and humans^b for selected terpene trilactones following ingestion of *Ginkgo biloba* extract

Rat doses		Exposure multiple (rat/human) ^c				
mg GBE/kg	mg GBE/m ² ^d	mg GBE/kg	mg GBE/m ²	AUC		
				BLL ^e	GLA	GLB
30	180	9	1.4	10	3	1
100	600	30	5	22	8	3
300	1800	88	14	59	20	7

^aData used was from the current study

^bData from Biber (2003) following ingestion of 240 mg EGB761® in adults was used. Assuming a 70 kg body weight, this corresponds to 3.4 mg/kg b.wt or 126 mg/m², using a K_m value of 37 for humans (Reagan-Shaw et al., 2008). AUC values reported in this study for BLL, GLA, and GLB, respectively, were: 247.1, 211.1, and 140.69 h*ng/mL (or 14826, 12666 and 8441 min*ng/mL)

^cRat/human exposure multiple was estimated 3 ways: mg/kg, mg/m² and AUC

^dDose in mg/m² was estimated by multiplying mg/kg by a K_m value of 6 for rats (Reagan-Shaw et al., 2008)

^eBLL, bilobalide; GLA, ginkgolide A; GLB, ginkgolide B