

Review Article

Insulin Resistance in Osteoarthritis: Similar Mechanisms to Type 2 Diabetes Mellitus

Elena V Tchetina ¹, Galina A Markova,¹ and Eugeniya P Sharapova²

¹*Immunology & Molecular Biology Laboratory, Nasonova Research Institute of Rheumatology, Moscow 115522, Russia*

²*Osteoarthritis Laboratory, Nasonova Research Institute of Rheumatology, Moscow 115522, Russia*

Correspondence should be addressed to Elena V Tchetina; etchetina@mail.ru

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Osteoarthritis (OA) and type 2 diabetes mellitus (T2D) are two of the most widespread chronic diseases. OA and T2D have common epidemiologic traits, are considered heterogenic multifactorial pathologies that develop through the interaction of genetic and environmental factors, and have common risk factors. In addition, both of these diseases often manifest in a single patient. Despite differences in clinical manifestations, both diseases are characterized by disturbances in cellular metabolism and by an insulin-resistant state primarily associated with the production and utilization of energy. However, currently, the primary cause of OA development and progression is not clear. In addition, although OA is manifested as a joint disease, evidence has accumulated that it affects the whole body. As pathological insulin resistance is viewed as a driving force of T2D development, now, we present evidence that the molecular and cellular metabolic disturbances associated with OA are linked to an insulin-resistant state similar to T2D. Moreover, the alterations in cellular energy requirements associated with insulin resistance could affect many metabolic changes in the body that eventually result in pathology and could serve as a unified mechanism that also functions in many metabolic diseases. However, these issues have not been comprehensively described. Therefore, here, we discuss the basic molecular mechanisms underlying the pathological processes associated with the development of insulin resistance; the major inducers, regulators, and metabolic consequences of insulin resistance; and instruments for controlling insulin resistance as a new approach to therapy.

1. Introduction

Osteoarthritis (OA) and diabetes mellitus (DM) represent the most prevalent chronic disorders. OA is prevalent primarily in women and is considered a major reason for disability and economic losses as almost half of adult OA patients are limited in their everyday activities and are restricted in their working capacity. Diabetes is associated with an increased incidence of death and serious side effects, such as stroke, chronic kidney disease, and amputation of the lower extremities [1].

OA is presently viewed as a chronic, systemic disease with slow progression and involves whole joint destruction that mainly includes degradation of hyaline articular cartilage, ligaments, menisci in the knee, hypertrophic changes

of the joint capsule as well as subchondral bone remodelling, osteophyte formation, and synovial inflammation. It is accompanied by pain, dysfunction of the lower extremities, and joint deformities [2].

DM is a metabolic disorder characterized by insulin resistance (IR) and pancreatic β -cell dysfunction as a consequence of prolonged hyperglycaemia in the body [3]. Traditionally, DM is considered to have two forms. Type 1 diabetes mellitus (T1D) is a chronic immune-mediated disease characterized by the selective loss of insulin-producing- β -cells in the pancreatic islets in genetically susceptible individuals resulting in severe insulin deficiency. T1D is often observed in children and may have an autoimmune and postinfection aetiology. Type 2 diabetes mellitus (T2D) is more common in adults. T2D involves IR at

the cellular level and a relative deficiency in insulin secretion. T2D development involves a progressive disturbance in glucose tolerance, which is associated with initial hyperplasia of the pancreatic islets followed by the proliferation of Langerhans β -cells. These processes are accompanied by an inflammatory response that induces the accumulation of the extracellular matrix (fibrosis) and the apoptotic death of islet β -cells, resulting in the progressive failure of β -cells to produce sufficient levels of insulin, followed by a decrease in insulin-stimulated glucose uptake and long-lasting hyperglycaemia. The concomitant development of osmotic and oxidative stresses produces lesions in the kidney, nervous system, and other tissues, which negatively affect the patient's lifespan [4].

OA and DM are common diseases that are significantly associated. Epidemiological studies report that the overall risk of OA in patients with DM is 1.46 while that of DM in patients with OA is 1.41 [5]. The prevalence of OA among patients with DM and that of DM in patients with OA was 29.5% and 14.4%, respectively [5]. Additionally, T2D independently predicts the risk of total joint replacement [6] and is considered a specific risk factor for OA. T2D related hyperglycaemia and IR result in loss of insulin receptor sensitivity, which might reduce chondrocyte survival and differentiation capacity, thereby accelerating OA progression [7]. Manifestations of OA are commonly observed in patients with T1D who lack the ability to produce insulin [8]. For example, radiological glenohumeral OA was observed in 35% T1D patients [9]. This is also associated with hyperglycaemia, which occurs in patients with both types of DM, and DM is known to negatively affect connective tissues [10].

Studies have reported a stronger association between TD and knee OA in women; however, no such association was observed in women aged <30 years [11], implicating the role of oestrogen in the pathogenesis of both conditions. Animal studies have shown that ovariectomy increased the risk of T2D [12]; in contrast, a few studies have shown that 17 β -estradiol (ovarian oestrogen) improves insulin sensitivity and protects insulin production in the diabetic state [13]. Mechanistically, glucose-stimulated insulin secretion requires an increase in the intracellular calcium concentration that is facilitated by oestrogen. This process involves an increase in the adenosine triphosphate (ATP):adenosine diphosphate (ADP) ratio in pancreatic β -cells, closure of ATP-sensitive potassium channels, depolarization of cell membrane, and opening of L-type voltage-dependent calcium channels [14].

Furthermore, OA and T2D are significantly associated with obesity. For example, a positive association is observed between both overweight and obesity and the risk of hand, hip, and knee OA. The strength of this association increases in a dose-dependent manner with increasing body mass index (BMI). For example, incidence rates of knee OA were 5-fold higher in patients with grade II obese than in individuals at a normal weight [15]. Recent studies have reported a significant association between BMI and T2D (relative risk = 6.88) [16]. These effects are attributable to the release of increased quantities of proinflammatory mediators, such as tumour necrosis factor (TNF) α , interleukin (IL)-6, and

IL-1 from adipocytes in individuals with obesity. These agents are known to trigger systemic low-grade inflammation in patients with both OA and TD [17].

Therefore, T2D and OA are both associated with patient age, low-grade inflammation, and obesity and result in disturbances in cellular metabolism; these disorders might have common pathophysiological mechanisms [1]. These similarities suggest that common approaches could be applied to ameliorate the metabolic imbalance associated with these diseases. Therefore, the aim of the study was to demonstrate that OA is not a disease solely of a single joint but like T2D is a pathology of the whole body, which results from insulin resistance-related traits that develop before the onset of the disease. Besides, we suggest that common mechanisms in the development of metabolic disturbances related to both pathologies might be associated with an imbalance in energy provision. These issues will be discussed in the present review.

2. Methodology

A comprehensive search of peer-reviewed literature published in the past two decades was undertaken. The search was performed in Medline/PubMed using the following keywords: (“osteoarthritis” OR “articular cartilage” OR “type 2 diabetes mellitus”) AND (“inflammation” OR “obesity” OR “oxidative phosphorylation” OR “AMPK” OR “insulin resistance” OR “fibrosis” OR “ATP” OR “exercise” OR “pain”). Exclusion criteria were as follows: studies not centred on the outcome of interest; studies only with abstract or with no available data; studies where a full description of research design was not provided or available; papers not written in English; all grey literature; duplication publications; educational materials and research publications intended for health care professionals to help them give verbal recommendations to patients; theses, forecasts, and journals not accessible online; and PowerPoint presentations, blogs, or websites. In addition, the reference lists of appropriate studies were also examined to identify additional relevant studies. Both authors independently screened all titles and abstracts for eligible studies and extracted data independently. Full texts for all relevant papers were collected and reevaluated by both authors for appropriateness. All papers satisfying the criteria and approved by both authors were included. No search restrictions were imposed.

3. Results and Discussion

3.1. Insulin Is a Universal Regulator of Cellular Metabolism. Insulin controls the expression and activity of more than 150 genes in many organs and tissues by regulating their transcription, messenger ribonucleic acid (mRNA) stability, and translation [18]. Mechanistically, the binding of insulin to the insulin receptor results in the autophosphorylation of its tyrosine residues. Receptor activation leads to the phosphorylation of tyrosine residues on adaptor proteins, members of the insulin receptor substrate family (IRS), which promote signalling to downstream targets [19].

Insulin is primarily involved in the maintenance of physiological blood glucose levels which depends on a complex interaction between the insulin responsiveness of skeletal muscle, liver, and adipose tissue and glucose-stimulated insulin secretion by pancreatic β -cells [20]. For example, in adipose cells, glucose transporters (GLUTs) are located intracellularly in the absence of insulin. Insulin stimulates the phosphorylation of tyrosine residues on IRS-1 and allows insulin receptors to bind to GLUT, subsequently inducing the recruitment of GLUT from the extracellular pool to the plasma membrane and exposing functional glucose transporters to the extracellular compartment containing glucose [21].

In pancreatic β -cells, respiration and the rate of adenosine triphosphate (ATP) production in oxidative phosphorylation (OXPHOS) are highly dependent on glucose availability due to the inability of β -cells to metabolize glucose by aerobic glycolysis, as β -cells are not capable of converting pyruvate to lactate [22]. Therefore, when the glucose concentration in circulation increases after a meal, the increased ATP/ADP (adenosine diphosphate) ratio in β -cells initiates the closing of ATP-sensitive K-channels, thus depolarizing the plasma membrane and activating voltage-gated L-type Ca^{2+} channels to stimulate Ca^{2+} -dependent exocytosis of insulin-containing secretory granules [23]. As in the majority of other cell types, cellular energy requirements are regulated by respiration rates and ATP availability; the increase in circulating insulin concentrations initiates biosynthetic activity in different tissues such as skeletal muscle, liver, white adipose tissue, and brain for the utilization of increased nutrient supply followed by restoration of the blood glucose concentration to its original level [24].

Insulin is also of primary importance for articular cartilage. Human chondrocytes express the functional insulin receptor, which is more abundant in healthy cells relative to OA. Insulin is capable of inducing anabolic responses that promote the synthesis of type II collagen and proteoglycan, inhibit expression of the collagenases, matrix metalloproteinase- (MMP-) 1 and 13, and a disintegrin and metalloproteinase with thrombospondin motifs (ADAMTS), and counteract the effects related to IL-1 β [25]. In animal and human studies, insulin treatment reversed cartilage loss, which was associated with fracture healing in diabetic mice [26] and ameliorated cartilage degradation [27].

3.2. Mechanisms of Insulin Resistance Development. The IR state involves induction of the inhibitory serine phosphorylation of IRS-1, preventing the involvement of IRS-1 in insulin receptor signalling and thus inhibiting insulin action [19]. In an insulin-resistant state, the ability of insulin to stimulate glucose uptake via insulin-dependent GLUTs is impaired, requiring higher than normal concentrations of extracellular insulin to maintain normal circulating glucose levels. Initially, higher insulin concentrations are maintained by β -cell overstimulation. Persistent insulin activation induces hyperphosphorylation of the serine/threonine

(Ser/Thr) residues on the IRS. This hyperphosphorylation results in reduced insulin sensitivity of the insulin receptor, inhibition of insulin signalling, or the uncoupling of IRS-1 and its degradation [28].

3.3. Physiological Insulin Resistance. Physiological insulin resistance in healthy cells represents a survival strategy and resolves after cessation of the abnormal condition. The survival of every organism depends on the capacity to withstand starvation and develop a strong acute immune response to pathogens or tissue injury [29]. For this purpose, the energy obtained from nutrients should be transferred for expenditure during a strong immune response. The inflammatory response involves proinflammatory cytokine production, which is an energy-consuming process [30]. Proinflammatory cells rely on aerobic glycolysis to convert pyruvate to lactate, and in an alternative route, reoxidising the NADH produced by glycolysis. Simultaneously, these cells downregulate oxidative mitochondrial metabolism, which has an anti-inflammatory effect. This downregulation is accompanied by changes in the tricarboxylic acid (TCA) cycle function from a catabolic function that generates ATP into a partially anabolic function, as citrate withdrawal from the cycle is used for fatty acid synthesis [30].

As adipocytes and hepatocytes are located adjacent to blood vessels carrying immune cells (macrophages), they would be the first cells to interact with the immune system. Therefore, the activation of the inflammatory response would transiently block major anabolic signalling pathways such as the insulin pathway in tissues other than immune cells by activating serine kinases such as c-Jun N-terminal kinase (JNK), I κ B kinase (IKK) β , extracellular-signal-regulated kinase (ERK), ribosomal protein S6 kinase (S6k), mechanistic target of rapamycin (mTOR), protein kinase C (PKC), and glycogen synthase kinase (GSK)3 β and inducing inhibitory serine phosphorylation of IRS-1 [31]. Therefore, inflammation serves as a driver of physiological IR, which is associated with transient redirection of energy for immune system activation.

3.4. Pathological Insulin Resistance. The insulin-resistant state is an early and key feature of T2D that develops 10–20 years before the onset of the disease [11]. Recently, IR traits were also described in OA patients [32]. For example, an increase in serum insulin concentration was found in 82% of patients with OA, while high insulin concentrations (10–500 nM) were shown to be capable of increasing articular cartilage degradation [33].

3.4.1. Overnutrition. Pathological insulin resistance in OA patients might result from overnutrition (Figure 1). According to the obesity-related model of IR development, increased oxidation of fatty acids (FAs) produces high levels of intracellular acetyl-CoA and citrate, which are capable of inhibiting two enzymes involved in glucose utilization, namely, pyruvate dehydrogenase and phosphofructokinase, a key rate-limiting enzyme in glycolysis. Subsequent

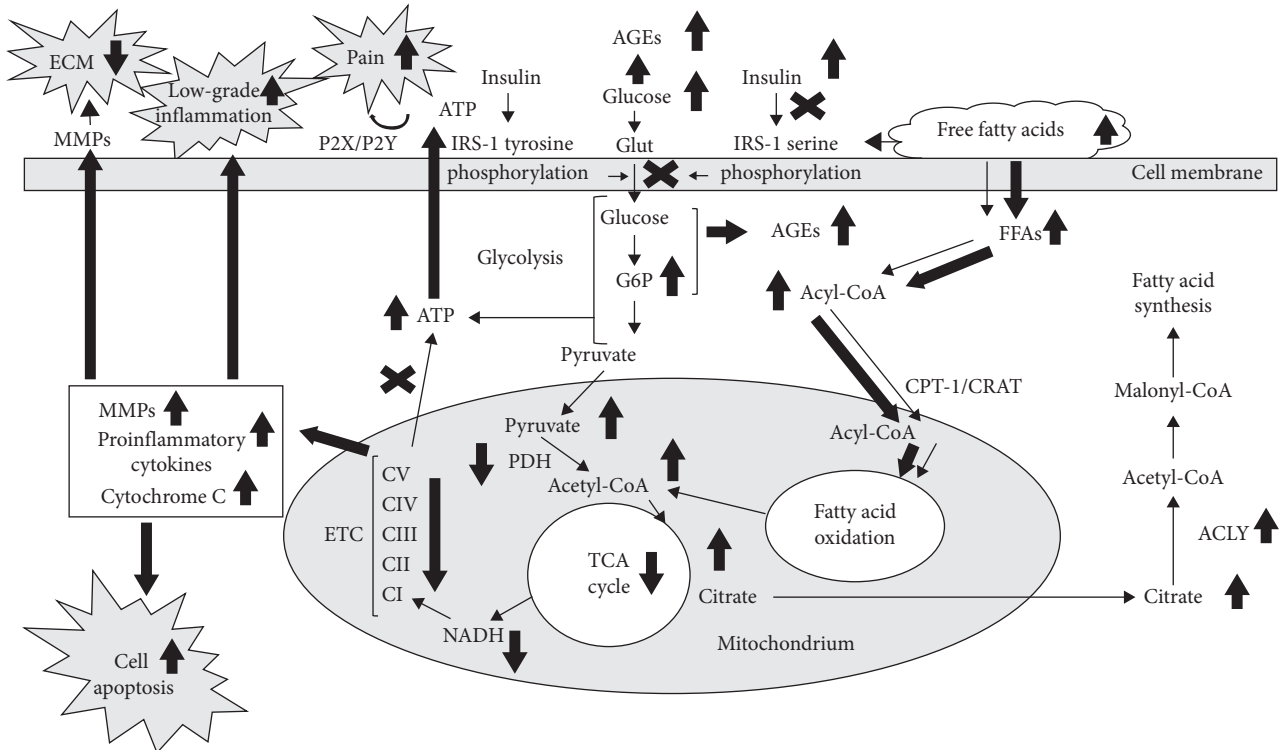


FIGURE 1: Metabolic changes in healthy and osteoarthritic chondrocytes associated with insulin resistance. Thin arrows, healthy chondrocyte. Bold arrows, insulin resistance-related changes in osteoarthritic chondrocyte. ECM, extracellular matrix; MMPs-matrix metalloproteinases; P2X/P2Y, purinergic receptors; IRS, insulin receptor substrate; ATP, adenosine triphosphate; PDH, pyruvate dehydrogenase; ETC, electron transport chain; C, Complex; NADH, nicotinamide adenine dinucleotide reduced; G6P, glucose-6-phosphate; AGEs, advanced glycation endproducts; TCA, tricarboxylic acid; FFAs, free fatty acids; CPT, carnitine:palmitoyl-CoA transferase; CRAT, carnitine acyltransferase; ACLY, ATP-citrate lyase.

accumulation of glucose-6-phosphate would inhibit hexokinase II activity, resulting in an increase in intracellular glucose concentrations and decreased glucose uptake [34].

It was also shown that fatty acids could affect the GLUT4 transporter directly while elevated plasma FA concentrations abolished insulin-stimulated IRS-associated phosphoinositide 3- (PI3-) kinase activity by phosphorylation of serine/threonine sites on insulin receptor substrates, which fail to associate with PI3-kinase and thus decreased activation of glucose transport. Therefore, the accumulation of intracellular fatty-acyl-CoAs or other FA metabolites leads to insulin signalling defects and IR [35].

A sustained high citrate concentration associated with IR could also result from the low expression of ATP-citrate lyase (ACLY), which was observed in the pancreatic β -cells of obese patients [36]. ACLY is a cytosolic protein that catalyses the formation of acetyl-CoA and oxaloacetate from citrate and coenzyme A in the presence of ATP and thus regulates the switch between glucose and fatty acid metabolism.

The increase in glycolytic flux can be associated with a switch in the expression of liver phosphofructokinase-2/fructose-2,6-bisphosphatase (PFKFB) isoforms from PFKFB1 to PFKFB3 and results in the accumulation of fructose-2,6-bisphosphate [30]. Indeed, PFKFB3 disruption in mice ameliorated high-fat diet-induced adipose tissue inflammation but exacerbated systemic IR, whereas the

overexpression of PFKFB3 in adipose tissue improved systemic insulin sensitivity [37].

Obesity due to overnutrition likewise affects the severity of joint degradation, risk of OA progression, and joint surgery requirements [38]. At the same time, significant weight loss was required to achieve symptomatic and structural improvements in humans, while moderate weight loss protected subjects at high risk of knee OA [39]. The downregulation of PFKFB3 expression and increased activity of citrate synthase were also observed in human OA cartilage and in $\text{TNF}\alpha$ - or $\text{IL-1}\beta$ -treated chondrocytes [40].

Adipokines, primarily resistin, are shown to be involved in the development of IR in patients with DM and OA [41]. Resistin is a 12 kDa cysteine-rich polypeptide hormone secreted by macrophages and adipocytes. It suppresses insulin-induced glucose uptake [42], promotes the proliferation, migration, and activation of human endothelial cells, increases expression of vascular endothelial growth factor receptors and MMPs, and activates ERK1/2 and p38 signalling pathways [43]. Recent studies have reported that insulin function regulation is initiated in hypothalamic nuclei, which control energy homeostasis [44], and resistin reduces insulin sensitivity and signalling in the hypothalamic nuclei, as well as in adipose tissue, liver, and skeletal muscles by increasing serine phosphorylation of insulin receptor substrate-1 (IRS1) [41].

Resistin was also associated with radiographic knee OA, promoted OA progression, was positively correlated with OA severity and leptin levels, and induced production of MMPs and proinflammatory cytokines in human chondrocytes [45]. Interestingly, resistin was not associated with cartilage volume loss and even promoted cartilage proteoglycan synthesis [46]. Therefore, the adverse effects of resistin in patients with OA might be attributable to increased expression of proinflammatory cytokines, as well as the production of tissue degrading enzymes and down-regulation of synthesis of structural proteins, which are formed in the pericellular matrix of the articular cartilage [47].

3.4.2. Inflammation. Increased nutrient concentrations associated with long-term overnutrition can induce inflammation in all tissues involved in energy homeostasis, such as fat, muscle, liver, islets, and nutrient-transporting blood vessels [29]. Free fatty acids (FFAs) are considered to be important proinflammatory agents that can activate the innate immune system. For example, FFAs can either bind directly to Toll-like receptors (TLRs) or induce the TLR dimerization required for signalling [48]. Adaptive immunity can also be involved in IR development, promoting the release of autoantigens in concert with alarmins and further activating the immune system or stimulating obesity-induced inflammation due to the accumulation of T cells in adipose tissue. This mechanism is supported by the elevated circulating levels of acute-phase proteins, cytokines, and chemokines observed in T2D and OA patients. In addition, proinflammatory cytokines such as TNF α are capable of activating various signal transduction pathways that inhibit insulin action and promote hyperglycaemia [49].

Obesity also triggers a local low-grade subacute inflammatory response due to the infiltration of proinflammatory M1-polarized macrophages in OA synovium prior to cartilage degradation [50]. These findings are supported by the observation that, in early OA, 266 genes were differentially expressed between inflamed synovium and healthy synovial tissue [51]. Furthermore, increased concentrations of IL-1 α , IL-18, and TNF α were associated with OA severity, while baseline IL-18 levels predicted OA progression [32]. Besides, patients with knee OA had increased the levels of IL-6 released from the infrapatellar fat pad relative to subcutaneous adipose tissue while individuals with a higher baseline body mass index and increased serum IL-6 levels were more likely to be diagnosed with radiographic knee OA after a 5-year follow-up [52].

Therefore, pathological IR results from disturbances of cellular metabolic processes, primarily triggered by overnutrition, and is associated with chronic low-grade inflammation.

3.4.3. Hyperglycaemia. In many cells and tissues, primarily articular chondrocytes, glucose serves as a major energy-producing substrate. During pathological IR conditions (Figure 1), the above-described inhibition of the glycolytic pathway produces an increase in intracellular glucose and

glucose-6-phosphate concentrations, potentially resulting in reduced insulin-stimulated glucose transport activity in target tissues [25]. Subsequent transient upregulation of glycolysis increases intracellular ATP concentrations as indicated by functional inhibition of K(ATP) channels, which sense intracellular ATP/ADP levels and are impaired in OA chondrocytes [53].

Glucose and glucose-6-phosphate accumulation can result in the formation of advanced glycation end products (AGEs), which are formed by the nonenzymatic glycation of reducing sugars such as glucose, with amino groups of proteins, lipids, and nucleic acids [54]. Glycation is a concentration-dependent process that is enhanced in diabetes and aging and was observed in OA [55]. AGE-related inhibition of ATP production and AMP-activated protein kinase (AMPK) phosphorylation [56] further point to high ATP concentrations in OA chondrocytes. In addition, AGEs can induce transcription factor nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) and stress kinase ERK1/2 expression through the activation of the recognition receptor RAGE, followed by NOD-like receptor (NLR) family member NLRP3 inflammasome formation, and IL-1 receptor activation and thereby systemic inflammation [57].

3.4.4. Mitochondrial Dysfunction. Disturbances in mitochondrial FA oxidation associated with pathological IR might result from carnitine insufficiency and/or carnitine acetyltransferase (CRAT) deficiency (Figure 1). Carnitine is responsible for mitochondrial transportation and oxidation of long-chain FAs and also serves as an acceptor of acyl groups, which enables the export of excess of acylcarnitines from mitochondria. Carnitine requirements are increased during sustained metabolic stress, while decreased carnitine availability is associated with IR states such as old age, diabetes, and diet-induced obesity [58]. In human knee OA, L-carnitine supplementation improved the patient's clinical status, demonstrating significant reductions in serum malondialdehyde, total cholesterol, and low-density lipoprotein cholesterol concentrations compared to those at baseline. This was accompanied by a significant decrease in serum IL-1 β and MMP-1 levels [59].

Another possibility regarding the development of IR-related mitochondrial dysfunction might involve insufficient cellular antioxidant activity. The mitochondrial respiratory chain is the main source of superoxide radicals in many cell types, including pancreatic β -cells [60]. 0.15% of total oxygen used in OXPHOS is converted to superoxide anion, which is transformed to hydrogen peroxide by superoxide dismutase (SOD) [61]. Superoxide production is increased in β -cells in rodents with T2D, in cultured β -cells under hyperglycaemic conditions or lipid excess in obese humans, and is associated with attenuation of the respiratory chain [62]. However, SOD activity was reduced in T2D patients [63], in human OA chondrocytes and articular cartilage [64]. Reduced activities of Complexes I, II, and III of the mitochondrial respiratory chain and mitochondrial depolarization in human OA articular chondrocytes in culture relative

to normal chondrocytes were also noted, while the inhibition of Complexes III and V were capable of stimulating proinflammatory cytokine and matrix metalloproteinase production in these cells [65].

Overall, the attenuation of mitochondrial function during IR development is linked to the reduction of mitochondrial oxidative enzyme and electron transport chain activity and hence decreased ATP synthesis.

3.4.5. Insulin Resistance-Related Pain. IR might also be associated with pain manifestations involving central sensitization [66]. It has been shown that, in addition to peripheral tissues, the central nervous system is also involved in the regulation of tissue-specific insulin sensitivity, as insulin receptors were found throughout the brain [67]. The origin of insulin in the brain is mostly peripheral, as insulin crosses the blood brain barrier by insulin receptor-mediated active transport [68]. Studies in diabetic rats have demonstrated that nerve injury is capable of downregulating the expression of insulin receptors in skeletal muscle innervated by the injured nerve [69] while acute pain has been reported to reduce insulin sensitivity due to decreased glucose uptake in the body [70].

Another mechanism of IR involvement in the increased pain sensitivity in OA and T2D might be associated with excessive amounts of cellular ATP, a key pain mediator capable of activating purinergic receptors P2X and P2Y, which are involved in both neuropathic diabetic and rheumatic pain sensitivity and hyperalgesia in arthritic joints [71] (Figure 1). For example, extracellular ATP was detected in most nonexcitatory cells, including chondrocytes, presumably due to nonlytic ATP release by conductance effluxes, specific transporters, constitutive secretory pathways, or regulated exocytosis [72]. This is supported by the observation that increased ATP levels were present in the knee synovial fluid of OA patients and were related to pain intensity [73].

3.5. AMPK Is a Major Regulator of Insulin Resistance. AMPK is a serine/threonine kinase that regulates metabolic pathways responsible for cellular energy generation and is involved in the control of whole-body energy balance by responding to the hormonal and nutrient signals in the central nervous system and peripheral tissues that govern nutrient uptake and energy expenditure [30]. AMPK is activated by adenosine monophosphate (AMP) and antagonized by increased concentrations of ATP, thus controlling changes in the AMP/ATP ratio. AMPK stimulated glucose uptake and suppressed glycogen synthesis in skeletal muscle [74]. AMPK activation inhibits anabolic pathways and upregulates catabolic pathways.

The major AMPK-regulated pathway, which reduces the risk of obesity and IR, stimulates fatty acid (FA) oxidation and inhibits FA synthesis by phosphorylating and downregulating acetyl-CoA carboxylase, a rate-limiting reaction in the conversion of acetyl-CoA to malonyl-CoA. In addition, AMPK stimulates malonyl-CoA decarboxylase, which catalyses malonyl-CoA degradation. A decrease in

malonyl-CoA prevents the inhibition of carnitine:palmitoyl-CoA transferase-1 (CPT-1), which is responsible for long-chain fatty acyl-CoA transport into mitochondria and fatty acid synthase controlling FA biosynthesis [75]. Chronic AMPK activation has been shown to stimulate mitochondrial biogenesis in skeletal muscle mediated by an increase in the concentration of proliferator-activated receptor γ coactivator- (PGC-) 1α , which is downregulated in T2D and OA [76, 77]. AMPK can increase insulin sensitivity either by suppressing the mechanistic target of rapamycin (mTOR) directly or by activating its inhibitor tuberous sclerosis complex (TSC), resulting in decreased activity of the TSC-mTOR-S6K1 pathway [78].

3.5.1. AMPK as an Indicator of the Efficacy of ATP Turnover. Effective oxidative capacity is determined by energy expenditure, and therefore, the mitochondrial respiratory dysfunction associated with elevated FFA levels reflects decreased ATP turnover [79]. For example, the treatment of primary myoblasts with palmitate was associated with a decreased rate of de novo protein synthesis, a major ATP consumer. Generally, FFAs reduce the overall absolute rate of mitochondrial ATP supply, which may be a consequence of lower ATP demand. Therefore, depressed mitochondrial respiration followed by decreased mitochondrial ATP synthesis might not involve intrinsic defects but could be an adaptive response to altered energy demand [80]. In addition, the treatment of cultured bovine articular chondrocytes with high doses of exogenous ATP induced an increase in extracellular inorganic phosphate accumulation, a by-product of ATP degradation. This effect was accompanied by the upregulation of MMP-13 and proinflammatory mediators, preventing extracellular matrix (ECM) formation [81].

3.5.2. AMPK Activity and Insulin Resistance. High insulin concentrations inhibit AMPK [82]. Animal studies using transgenic mice that expressed the inactive form of the AMPK α 2 subunit in skeletal muscle developed impaired whole-body glucose tolerance and IR in skeletal muscle, specifically when fed a high-fat diet [83]. These results were supported by genetic studies in the Japanese population, which demonstrated that polymorphisms in the gene encoding the AMPK α 2 subunit were linked to IR and T2D [84]. The decrease in AMPK activity associated with the IR state has been also shown in the adipose tissue of 75% of severely obese patients undergoing bariatric surgery. This association was accompanied by an upregulation of proinflammatory genes, including cytokines and chemokines, and the decreased expression of PGC- 1α , enzymes related to the β -oxidation of FA and the citric acid cycle in visceral adipose tissue [85]. Skeletal muscle AMPK expression and activity were also decreased in T2D patients [86].

Phosphorylated AMPK α is constitutively present in normal articular chondrocytes and cartilage, while OA chondrocytes are deficient in AMPK activity [87]. Decreased AMPK activity enhances the pro-catabolic responses of chondrocytes to the proinflammatory cytokines IL- 1β and

TNF α [88]. Alternatively, the upregulation of AMPK was associated with decreased collagen degradation activity and increased the expression of type II collagen accompanied by the upregulation of TCA-related genes and the down-regulation of proinflammatory cytokines and metalloproteinases [89]. In animal studies, pharmacological AMPK activation limited knee OA development and progression [90].

Therefore, AMPK is a primary regulator of cellular energy metabolism and IR state, while the acquisition of AMPK expression levels similar to that in healthy subjects could serve as an indicator of treatment efficacy.

3.6. Putative Role of Chronic Inflammation and Fibrosis in Energy Balance Improvement. Physiological inflammation represents a short-term adaptive response as a component of tissue repair and involves the integration of numerous signals in different cells and organs [29]. For example, the activation of NF κ B observed in pregnancy is mediated by limited hyperglycaemia that induces β -cell proliferation and increase in the pancreas mass pointing to the increased requirements for insulin [91].

The prolonged, low-grade or chronic inflammation observed in many metabolic diseases including OA and T2D is generally not considered beneficial. Chronic inflammation is associated with the proliferation of immune cells, abnormal production of cytokines, and activation of proinflammatory signalling pathways [31]. Chronic inflammation was observed in T2D patients, while increases in interleukin-(IL-) 1 β , IL-6, and C-reactive protein (CRP) levels were considered predictors of the disease [92]. Differences in cytokine/chemokine levels were also found between normal and OA serum samples [93]. Increased expression of IL-1 β and TNF α was observed in the peripheral blood of OA patients relative to that of healthy controls [94]. In addition, knee cartilage volume was negatively associated with the concentrations of the circulating cytokines IL-6 and TNF α and CRP [95].

Furthermore, chronic inflammation is associated with tissue fibrosis, which is characterized by fibroblast proliferation and excessive ECM deposition [96]. For example, high glucose levels stimulated TLR4 and resulted in NF- κ B activation followed by upregulation of growth factors such as transforming growth factor (TGF) β and connective tissue growth factor (CTGF), which was accompanied by ECM accumulation in tendons in T2D patients [97]. These effects were associated with the enhanced expression of MMP-1, MMP-2, and MMP-9, reflecting the increase in ECM degradation under hyperglycaemic conditions [98]. In human OA, inflammation and fibrosis also occur together with ECM destruction, as early changes prior to radiographic alterations were related to synovial inflammation, indicated by thickening of the lining layer and an increase in the proliferation of lining cells in sites of chondral defects [99] and by profibrotic changes in infrapatellar fat pad cells [100].

However, protein synthesis and cell proliferation are highly ATP-dependent energy-consuming processes [101]. The reduced AMPK expression in OA articular

chondrocytes [88, 89] and in the cells of IR tissues in T2D [86] suggests increased levels of intracellular ATP. Additionally, the chronic systemic inflammation associated with profibrotic changes in tissues is accompanied by increased AMPK expression in peripheral blood, indicating additional energy requirements [89]. These effects might induce an energy appeal reaction, a process that involves the redirection of energy-rich fuels from energy-rich compartments, such as IR tissues in T2D and OA articular chondrocytes, as has been indicated by studies of the activated immune system [102].

Therefore, as catabolic pathways are responsible for energy provision [103], we may speculate that the redirection of energy fuels to support immune cell functions and/or fibrotic tissue synthesis might improve the energy balance of the whole body, which is disturbed by the disease.

3.7. Factors Associated with Control of Insulin Resistance in Osteoarthritis. OA is considered an incurable disease; therefore, all current treatments are symptomatic in nature and are primarily focused on managing the secondary manifestations of IR-induced profound disturbances of cellular energy metabolism such as inflammation, articular cartilage destruction, and pain. Notably, OA is treated with paracetamol, nonsteroidal anti-inflammatory drugs (NSAIDs), corticosteroid injections, and tramadol based on Guidelines [104]. Novel treatments such as capsaicin (a transient receptor potential cation channel subfamily V member 1 (TPRV1) agonist) and galcanezumab (for calcitonin gene-related peptide (CGRP) blockade) target pain, whereas biological agents such as lutikizumab (for IL-1 α/β blockade) or etanercept (for TNF blockade) target inflammation [105]. However, some of these agents are associated with prodiabetic effects. For example, glucosamine (the most widely used OA medication in postmenopausal women) is known to cause glucose intolerance and IR and inhibits insulin production by pancreatic β -cells, resulting in apoptosis and deranged glucose metabolism [106]. Therefore, research is focused on the development of newer approaches to target pathways affected by the OA.

3.7.1. Cellular and Subcellular Particle-Based Therapies. Cell therapies utilizing stem, progenitor, primary, or stem cell derivatives (including embryonic stem cells, induced pluripotent stem cells, or mesenchymal stem cells (MSCs)) constitute a promising therapeutic approach to OA. These cells replace injured cells or tissues to stimulate the self-healing of endogenous tissues through nutritive effects [107]. However, studies have shown that MSCs promote carcinogenesis and maldifferentiation of cells into types that may cause disease exacerbation [108]. Therefore, recently extracellular vesicles (exosomes) that are actively secreted by MSCs are considered important contributors for OA amelioration [109]. Exosomes are nanosized membranous vesicles (30–120 nm in diameter), which are released by many cell types [110] and are involved in the intercellular transfer of bioactive molecules such as proteins, lipids, mRNAs, or microRNAs. Exosomes respond to cell injury

and participate in the regulation of cell proliferation and differentiation [111] and may contribute to cartilage repair and regeneration [112]. MSC exosomes were shown to alleviate pain, inflammation, and matrix degradation via activation of the ERK, protein kinase B (Akt), and AMPK pathways in an animal model of temporomandibular OA [113]. Exosomes have also shown efficacy as potential therapeutic agents for T1D in animal studies [114]. The absence of genomic content and the consequent lack of replication capacity reduce the malignant potential of exosomes [115]. However, the widespread application of exosomes is limited by the lack of adequate knowledge regarding the exact molecular mechanisms through which they participate in cartilage repair [109]. For example, some studies have reported that chondrocytes showed decreased anabolic activity in response to MSC exosomes unless they were capable of expressing miR-140-5p [116]. Additionally, exosomes from different cell types are highly heterogeneous and show varying survival and healing capacities secondary to old age or unfavourable health conditions in donors [117].

The recent introduction of high-throughput sequencing has resulted in the identification of endogenous circular RNAs (circRNAs) derived from precursor mRNA in different tissues [118]. CircRNAs represent endogenous noncoding RNAs characterized by covalently closed cyclic structures with nonpolyadenylated tails, which function as translational regulators, RNA-binding sponges, and microRNA sponges [119]. They are widely expressed, highly stable, evolutionary conserved elements among species that show tissue-specific and developmental stage-specific expression patterns [120]. For example, research has shown that some circRNAs were aberrantly expressed in articular cartilage of patients with OA and underwent upregulation in chondrocytes stimulated by IL-1 and TNF α [121]. Overexpression of some circRNAs was associated with the upregulation of ECM degrading enzymes and inhibition of matrix proteins such as type II collagen and aggrecan [122]. Dysregulation of circRNA was also observed in patients with DM and was primarily associated with regulation of insulin secretion by β -cells as well as vascular complications including diabetic nephropathy, neuropathy, and cardiomyopathy [118]. CircRNAs are also involved in the regulation of miRNA activity by the formation of miRNA sponges that interact with RNA-binding proteins (RBPs) to regulate gene transcription and protein translation [123]. Notably, miRNAs represent small noncoding RNAs capable of posttranslational regulation of target mRNAs transcription via binding the corresponding miRNA response elements (MREs) [124]. Some circRNAs contain MREs that compete for miRNA-binding sites (miRNA sponges) and result in functional sequestration of miRNAs [125]. Therefore, circRNAs are considered useful diagnostic and predictive biomarkers of some disorders, and inhibition of their activity might promote ECM recovery and slow down the process of joint degradation [123]. It is noteworthy that although many circRNAs are described in humans, only a few are known to produce biological effects [126]. Other studies investigating circRNA silencing by specific siRNAs have reported a lack of stability against extracellular and intracellular nucleases and a number of nonspecific effects [127].

3.7.2. Therapeutic Potential of Mimetic Molecules. Recent *in vitro* and animal studies have suggested that OA could be ameliorated using mimetics of known molecules affected by the disease. For example, lubricin mimetic, an extracellular matrix component structural analogue, attenuated OA progression [128] whereas PG545, a heparan sulphate mimetic in cultured human OA chondrocytes, inhibited the effect of heparanase that is known to stimulate catabolic MMP13 and ADAMTS and downregulate aggrecan and type II collagen synthesis [129]. Animal studies have proved that the glucose tolerance factor (GTF), a dietary agent extracted from yeast, functions as a putative insulin mimetic to lower blood glucose and lipid concentrations. It is also known to participate in phosphorylation of IRS1, Akt, and mitogen-activated protein kinase (MAPK) independent of insulin receptor phosphorylation [130]. However, these effects are questionable because GTF has not been purified, and its chemical structure remains unclear. Cobalt chloride, a hypoxia-mimetic agent has been used to confirm the role of glucose metabolism in articular chondrocytes. These studies have shown that the energy metabolism of chondrocytes undergoes adaptations under hypoxic conditions; however, reduced GLUT1 activity impairs OA cartilage repair [131]. Synthetic SOD mimetics such as nitroxides (tempol) and compound M40403 efficiently decreased hydroxyl radical production and tissue inflammation and injury and also reduced neutrophil infiltration at the site of inflammation in addition to protecting against cell death and loss of mitochondrial content in articular cartilage after mechanical overload [132].

Furthermore, studies on mimetic molecules might unravel some molecular mechanisms of the disease. For example, the analysis of SOD mimetics has shown upregulation of superoxide production only during normal articular chondrocyte loading, whereas chronic overloading resulted in more complex changes involving altered lipid metabolism [133]. This finding might be attributed to the fact that anabolic responses of the articular cartilage to normal loads depend on mitochondrial electron transport and, therefore, the mitochondrial function can control chondrocyte inflammatory response [134] evidenced by attenuation of such a response following the inhibition of electron transport by rotenone (mitochondrial Complex I inhibitor) or by the addition of antioxidants [135]. In contrast, severe impact significantly increases oxidant production, and prevention of cell death could be achieved only by antioxidant supplementation or inhibition of mitochondria (a source of reactive oxygen species [ROS]), by rotenone [136]. Therefore, repetitive mechanical overload *in vitro* was shown to cause a significant decrease in mitochondrial SOD concentrations despite increased oxidative damage, which is often observed in chondrocytes from severely arthritic knees [137].

3.7.3. Repurposing of Antidiabetic Agents for the Treatment of Osteoarthritis. Human and animal studies have shown that nearly all antidiabetic drugs are effective in treating OA. For example, antidiabetic treatment with insulin or pioglitazone

effectively prevented severe experimental OA in rodents with diabetes compared to those without diabetes in a diabetic rodent model [138]. Some human studies have shown protective effects of antidiabetic medications in knee OA; these drugs were shown to reduce knee OA progression [139]. Additionally, antidiabetic drug repaglinide used in combination with oral NSAIDs is known to alter the pharmacokinetics of the latter in patients with OA [140]. In vitro studies have shown that exenatide, a specific glucagon-like peptide agonist, prevents degradation of type II collagen and aggrecan in articular cartilage by inhibiting MMPs and ADAMTS. It was also shown to reduce oxidative stress and inhibit p38-dependent activation of nuclear factor kappa-light-chain-enhancer of activated B cells (NFkB) [141]. Saxagliptin, a novel dipeptidyl peptidase IV (DPP-4) inhibitor, suppresses the degradation of type II collagen in primary human chondrocytes by decreasing MMP, ADAMTS, and ROS production and increasing glutathione levels mediated by p38/NFkB pathway [142]. Metformin, the most commonly used antidiabetic drug, reduced inflammation, decreased the number of T helper 17 (Th17) cells, and increased the number of regulatory T (Treg) cells in a mouse model of collagen-induced arthritis [143]. Therefore, it was reasonable to conclude that antidiabetic medications attenuate the deleterious effects of DM in patients with OA, and this action halts disease progression [144].

Notably, studies have proved that metformin also effectively treats pancreatic cancer [145], colorectal cancer [146], all types of breast cancer [147], and benign prostate hyperplasia [148]. Moreover, metformin inhibits hepatitis C virus replication in Huh7 cells [149] and counteracts the effects of *Plasmodium* (the parasite causing malaria) [150] as well as other intracellular pathogens such as *Mycobacterium tuberculosis* and *Legionella pneumoniae* [151, 152]. Metformin might also be useful in patients with coronavirus (COVID19) disease, particularly in those with obesity and diabetes [153]. The broad specificity of antidiabetic drugs such as metformin is attributable to their capacity to inhibit hepatic gluconeogenesis and lower blood glucose concentration, improve insulin sensitivity in muscle, and lower plasma concentrations of free fatty acids [154]. The mechanism of action of metformin includes activation of AMPK expression [155] and inhibition of mTOR, which is upregulated in OA [156] either directly by AMPK stimulation or in AMPK-independent process mediated by upregulation of the regulated in development and DNA damage responses 1 (REDD1) [157]. It also affects the mitochondrial oxidative phosphorylation system by targeting Complex I of the mitochondrial respiratory chain [158]. Therefore, the efficacy of antidiabetic medications for the treatment of various disorders including OA could be attributed to their ability to target specific mitochondrial components such as OXPHOS complexes, oxygen consumption, mitochondrial replication, and biogenesis as well as enzymatic activity [158] primarily, via a metformin-mediated decrease in ATP production by the inhibition of the mitochondrial electron transport Complex I [159] resulting in improved insulin sensitivity.

3.7.4. Calorie Restriction for Amelioration and Prevention of Osteoarthritis. OA is associated with obesity and diabetes. Moreover, the maintenance of healthy body weight is essential for the prevention and amelioration of several chronic diseases [160]. Therefore, calorie restriction (CR) could serve as a useful therapeutic alternative in patients with OA. CR represents a nongenetic and non-pharmacological intervention to improve patients' life- and healthspan, primarily by focusing on weight loss [161]. Dietary restrictions mainly act by inhibiting key nutrient sensing and proinflammatory pathways, activating molecular pathways stimulating proteostasis, and enabling genome stability, stress resistance, and stem cell function [162]. The most important result of weight loss is a significant reduction in IR [163] because visceral obesity shows a strong inverse correlation with insulin sensitivity [164]. For example, studies performed in patients with T2D have shown that although the effect of CR on fasting glucose concentrations was variable, fasting insulin concentrations were reduced by 10–40% after 6–12 weeks of CR [165]. Animal studies (in Hartley guinea pigs) have also shown that CR with a regular chow diet reduced knee OA and decreased local and systemic inflammation [166]. However, lifelong CR was not associated with changes in OA incidence of joint synovitis in aged mice, indicating that the reduction of calorie intake is not sufficient to prevent age-related OA [167]. This observation is supported by the results of numerous studies that prove that CR is more effective when combined with physical activity [161].

3.7.5. Activation of Energy Expenditure by Physical Training. Muscle tissue is the first tissue to become insulin-resistant; therefore, changes in plasma glucose and insulin are primarily determined by the muscle IR state rather than insulin insensitivity in adipose or hepatic tissues [168]. Exercise has been demonstrated to activate the mobilization of progenitor cells from bone marrow, neural, and cardiac tissues, which are involved in the recovery of corresponding organs [169]. Therefore, exercise is considered an important physiological regulator of energy utilization as it is capable of increasing the energy metabolism of the whole body by up to 20-fold [170]. The mitochondrial biogenesis induced by exercise also occurs in tissues other than skeletal muscle, including the brain, adipose tissue, and kidney, providing evidence that exercise increases metabolic demand in these tissues and/or stimulates interorgan crosstalk [169].

The high-intensity exercise involves increased glucose oxidation followed by the upregulation of oxidative phosphorylation and anaerobic glycolysis. These processes occur independently of insulin, as the concentration of insulin is very low during exercise [171]. Indeed, insulin and exercise can independently increase glucose uptake as contraction stimulates GLUT-4 translocation through different molecular mechanisms. In muscle cell plasma membranes, the GLUT-4 concentration is regulated by the relative efficacy of exocytosis, which is increased by insulin, and endocytosis, which is reduced by insulin. Muscle contraction increases the rate of exocytosis, while AMPK activation decreases the

rate of endocytosis. In addition, the existence of two putative intracellular pools of GLUT-4 was suggested, one of which is recruited by insulin while the other is activated by contractions [172].

During prolonged exercise, FAs start to be used as an energy source, promoting lipogenesis and increasing OXPHOS and protein synthesis associated with increased mitochondrial biogenesis [173]. These effects improve insulin sensitivity [174]. However, in the absence of physical activity, the TCA cycle remained transcriptionally inactivated and was inhibited by ROS and insufficient levels of intermediates [175].

Elevated insulin-independent glucose uptake during exercise was reversed at 2-3 h after exercise. However, increased whole-body insulin sensitivity can persist up to 24–48 h [176]. After acute exercise, insulin-resistant humans and rats can reach glucose uptake values similar to healthy but unexercised controls [177]. In OA animal studies, exercise improved glucose tolerance and inhibited the expression of proinflammatory cytokines while upregulating the expression of the anti-inflammatory cytokine IL-10 [178]. In patients with knee OA, increases in intra-articular and perisynovial concentrations of anti-inflammatory IL-10 accompanied by decreased levels of TNF receptor 1 and increased physical function were also observed in response to exercise [179].

Recent studies have shown that AMPK activators can serve as exercise mimetics [180]. However, both direct and indirect AMPK activators can target one or several cellular pathways, while IR can be caused by numerous mechanisms. Therefore, so far, only exercise can alter homeostasis in all affected cell types [169] and improve insulin sensitivity, while any pharmacological strategy aiming to mimic exercise would need to induce increased energy expenditure similar to that in insulin-sensitive tissues [181].

Therefore, physical training targeting the activation of energy expenditure in the insulin-resistant tissues is viewed as a treatment option for OA and T2D patients.

4. Conclusions

Insulin sensitivity is a metabolic tool for discriminating between nutritional challenges and body defence against pathogenic agents and is regulated by reprogramming cellular energy metabolism in both peripheral tissues and immune cells. Therefore, the metabolic changes associated with acute and chronic inflammation might play an adaptive role in restoring the energy homeostasis of the whole body. Insulin resistance is observed in a wide range of different metabolic diseases, including T2D and OA, indicating the inability of the body to overcome energy-related disturbances. Hence, reprogramming energy metabolism could be considered a unifying concept to better understand the pathophysiological mechanisms of chronic inflammatory metabolic diseases such as T2D and OA and to provide new approaches for treatment. As a crucial regulator of energy metabolism at both the cellular and whole-body levels, AMPK controls an integrated signalling network responsible for metabolic adaptation. In addition, AMPK can serve as an

indicator of cellular energy imbalance as it is regulated by the AMP/ATP ratio. The energy gridlock indicated by the downregulation of AMPK in insulin-resistant tissues might represent a new therapeutic target in the treatment of T2D and OA. As presently available AMPK activator mimetics fail to resolve tissue energy congestion, exercise could be considered an important physiological regulator of energy utilization, increasing the energy expenditure of the whole body, including AMPK upregulation in different tissues. Therefore, long-term physical training accompanied by calorie restriction and antidiabetic agent treatment could be viewed as a current therapeutic solution for the management of metabolic disorders such as OA.

Disclosure

This article does not contain any studies with animals or human subjects performed by any of the authors.

Conflicts of Interest

All authors declared that they have no conflicts of interest.

References

- [1] K. Louati, C. Vidal, F. Berenbaum, and J. Sellam, "Association between diabetes mellitus and osteoarthritis: systematic literature review and meta-analysis," *RMD Open*, vol. 1, Article ID e000077, 2015.
- [2] R. F. Loeser, S. R. Goldring, C. R. Scanzello, and M. B. Goldring, "Osteoarthritis: a disease of the joint as an organ," *Arthritis & Rheumatism*, vol. 64, no. 6, pp. 1697–1707, 2012.
- [3] American Diabetes Association, "Classification and Diagnosis of Diabetes: Standards of Medical Care in Diabetes—2018," *Diabetes Care*, vol. 41, pp. S13–S27, 2018.
- [4] I. Hameed, S. R. Masoodi, S. A. Mir, M. Nabi, K. Ghazanfar, and B. A. Ganai, "Type 2 diabetes mellitus: from a metabolic disorder to an inflammatory condition," *World Journal of Diabetes*, vol. 6, no. 4, pp. 598–612, 2015.
- [5] J. Sellam and F. Berenbaum, "Is osteoarthritis a metabolic disease?" *Joint Bone Spine*, vol. 80, no. 6, pp. 568–573, 2013.
- [6] G. Schett, A. Kleyer, C. Perricone et al., "Diabetes is an independent predictor for severe osteoarthritis: results from a longitudinal cohort study," *Diabetes Care*, vol. 36, pp. 403–409, 2013.
- [7] R. Malaguarnera, A. Sacco, C. Voci, G. Pandini, R. Vigneri, and A. Belfiore, "Proinsulin binds with high affinity the insulin receptor isoform A and predominantly activates the mitogenic pathway," *Endocrinology*, vol. 153, no. 5, pp. 2152–2163, 2012.
- [8] W. Marik, S. F. Nemeč, Š. Zbýň et al., "Changes in cartilage and tendon composition of patients with type I diabetes mellitus," *Investigative Radiology*, vol. 51, no. 4, pp. 266–272, 2016.
- [9] N. G. Juel, J. I. Brox, J. C. Hellund, E. Merckoll, K. B. Holte, and T. J. Berg, "Radiological glenohumeral osteoarthritis in long-term type 1 diabetes. Prevalence and reliability of three classification systems. The Dialong shoulder study," *Skeletal Radiology*, vol. 47, no. 9, pp. 1245–1251, 2018.
- [10] B. W. Vogt, E. D. Schleicher, and O. H. Wieland, "ε-amino-lysine-bound glucose in human tissues obtained at autopsy:

- increase in diabetes mellitus," *Diabetes*, vol. 31, no. 12, pp. 1123–1127, 1982.
- [11] N. Dubey, D. Ningrum, R. Dubey et al., "Correlation between diabetes mellitus and knee osteoarthritis: a dry-to-wet lab approach," *International Journal of Molecular Sciences*, vol. 19, no. 10, p. 3021, 2018.
- [12] E. Riant, A. Waget, H. Cogo, J.-F. Arnal, R. Burcelin, and P. Gourdy, "Estrogens protect against high-fat diet-induced insulin resistance and glucose intolerance in mice," *Endocrinology*, vol. 150, no. 5, pp. 2109–2117, 2009.
- [13] F. Mauvais-Jarvis, D. J. Clegg, and A. L. Hevener, "The role of estrogens in control of energy balance and glucose homeostasis," *Endocrine Reviews*, vol. 34, no. 3, pp. 309–338, 2013.
- [14] P. Rorsman and E. Renström, "Insulin granule dynamics in pancreatic beta cells," *Diabetologia*, vol. 46, no. 8, pp. 1029–1045, 2003.
- [15] C. Reyes, K. M. Leyland, G. Peat, C. Cooper, N. K. Arden, and D. Prieto-Alhambra, "Association between overweight and obesity and risk of clinically diagnosed knee, hip, and hand osteoarthritis: a population-based cohort study," *Arthritis & Rheumatology*, vol. 68, no. 8, pp. 1869–1875, 2016.
- [16] V. Bellou, L. Belbasis, I. Tzoulaki, and E. Evangelou, "Risk factors for type 2 diabetes mellitus: an exposure-wide umbrella review of meta-analyses," *PLoS One*, vol. 13, no. 3, Article ID e0194127, 2018.
- [17] S. Schwarz, I. Mrosewski, S. Silawal, and G. Schulze-Tanzil, "The interrelation of osteoarthritis and diabetes mellitus: considering the potential role of interleukin-10 and in vitro models for further analysis," *Inflammation Research*, vol. 67, no. 4, pp. 285–300, 2018.
- [18] C. Mounier and B. I. Posner, "Transcriptional regulation by insulin: from the receptor to the gene," *Canadian Journal of Physiology and Pharmacology*, vol. 84, no. 7, pp. 713–724, 2006.
- [19] S. Schenk, M. Saberi, and J. M. Olefsky, "Insulin sensitivity: modulation by nutrients and inflammation," *Journal of Clinical Investigation*, vol. 118, no. 9, pp. 2992–3002, 2008.
- [20] B. B. Lowell and G. I. Shulman, "Mitochondrial dysfunction and type 2 diabetes," *Science*, vol. 307, no. 5708, pp. 384–387, 2005.
- [21] S. W. Cushman and L. J. Wardzala, "Potential mechanism of insulin action on glucose transport in the isolated rat adipose cell. Apparent translocation of intracellular transport systems to the plasma membrane," *The Journal of Biological Chemistry*, vol. 255, no. 255, pp. 4758–4762, 1980.
- [22] F. M. Ashcroft and P. Rorsman, "Diabetes mellitus and the β cell: the last ten years," *Cell*, vol. 148, no. 6, pp. 1160–1171, 2012.
- [23] M. Soty, M. Visa, S. Soriano, M. del Carmen Carmona, Áa Nadal, and A. Novials, "Involvement of ATP-sensitive potassium (KATP) channels in the loss of beta-cell function induced by human islet amyloid polypeptide," *Journal of Biological Chemistry*, vol. 286, no. 47, pp. 40857–40866, 2011.
- [24] P. Rorsman, M. Braun, and Q. Zhang, "Regulation of calcium in pancreatic α - and β -cells in health and disease," *Cell Calcium*, vol. 51, no. 3–4, pp. 300–308, 2012.
- [25] S. C. Rosa, A. T. Rufino, F. Judas, C. Tenreiro, M. C. Lopes, and A. F. Mendes, "Expression and function of the insulin receptor in normal and osteoarthritic human chondrocytes: modulation of anabolic gene expression, glucose transport and GLUT-1 content by insulin," *Osteoarthritis and Cartilage*, vol. 19, no. 6, pp. 719–727, 2011.
- [26] R. A. Kayal, J. Alblowi, E. McKenzie et al., "Diabetes causes the accelerated loss of cartilage during fracture repair which is reversed by insulin treatment," *Bone*, vol. 44, pp. 357–363, 2009.
- [27] E. V. Tchetina, J. Antoniou, M. Tanzer, D. J. Zukor, and A. R. Poole, "Transforming growth factor- β_2 suppresses collagen cleavage in cultured human osteoarthritic cartilage, reduces expression of genes associated with chondrocyte hypertrophy and degradation, and increases prostaglandin E2 production," *The American Journal of Pathology*, vol. 168, no. 1, pp. 131–140, 2006.
- [28] K. D. Coppins and M. F. White, "Regulation of insulin sensitivity by serine/threonine phosphorylation of insulin receptor substrate proteins IRS1 and IRS2," *Diabetologia*, vol. 55, no. 10, pp. 2565–2582, 2012.
- [29] M. Y. Donath, Éa Dalmas, N. S. Sauter, and M. Böni-Schnetzler, "Inflammation in obesity and diabetes: islet dysfunction and therapeutic opportunity," *Cell Metabolism*, vol. 17, no. 6, pp. 860–872, 2013.
- [30] L. A. J. O'Neill and D. G. Hardie, "Metabolism of inflammation limited by AMPK and pseudo-starvation," *Nature*, vol. 493, no. 7432, pp. 346–355, 2013.
- [31] G. S. Hotamisligil, "Inflammation and metabolic disorders," *Nature*, vol. 444, no. 7121, pp. 860–867, 2006.
- [32] F. Berenbaum, T. M. Griffin, and R. Liu-Bryan, "Review: metabolic regulation of inflammation in osteoarthritis," *Arthritis & Rheumatology*, vol. 69, no. 1, pp. 9–21, 2017.
- [33] M. Ribeiro, P. López de Figueroa, F. J. Blanco, A. F. Mendes, and B. Caramés, "Insulin decreases autophagy and leads to cartilage degradation," *Osteoarthritis and Cartilage*, vol. 24, no. 4, pp. 731–739, 2016.
- [34] P. Ebeling, H. A. Koistinen, and V. A. Koivisto, "Insulin-independent glucose transport regulates insulin sensitivity," *FEBS Letters*, vol. 436, no. 3, pp. 301–303, 1998.
- [35] G. I. Shulman, "Cellular mechanisms of insulin resistance," *Journal of Clinical Investigation*, vol. 106, no. 2, pp. 171–176, 2000.
- [36] M. Zhong, Y. Wu, W. Ou, L. Huang, and L. Yang, "Identification of key genes involved in type 2 diabetic islet dysfunction: a bioinformatics study," *Bioscience Reports*, vol. 39, no. 5, 2019.
- [37] Y. Huo, X. Guo, H. Li et al., "Disruption of inducible 6-phosphofructo-2-kinase ameliorates diet-induced adiposity but exacerbates systemic insulin resistance and adipose tissue inflammatory response," *Journal of Biological Chemistry*, vol. 285, no. 6, pp. 3713–3721, 2010.
- [38] M. Duclos, "Osteoarthritis, obesity and type 2 diabetes: the weight of waist circumference," *Annals of Physical and Rehabilitation Medicine*, vol. 59, no. 3, pp. 157–160, 2016.
- [39] A. J. Teichtahl, A. E. Wluka, S. K. Tanamas et al., "Weight change and change in tibial cartilage volume and symptoms in obese adults," *Annals of the Rheumatic Diseases*, vol. 74, no. 6, pp. 1024–1029, 2015.
- [40] J. Qu, D. Lu, H. Guo, W. Miao, G. Wu, and M. Zhou, "PFKFB3 modulates glycolytic metabolism and alleviates endoplasmic reticulum stress in human osteoarthritis cartilage," *Clinical and Experimental Pharmacology and Physiology*, vol. 43, no. 3, pp. 312–318, 2016.
- [41] M. J. Saeedi Borujeni, E. Esfandiary, G. Taheripak, P. Codoñer-Franch, E. Alonso-Iglesias, and H. Mirzaei, "Molecular aspects of diabetes mellitus: resistin, microRNA, and exosome," *Journal of Cellular Biochemistry*, vol. 119, no. 2, pp. 1257–1272, 2018.

- [42] C. W. Zhao, Y. H. Gao, W. X. Song et al., "An update on the emerging role of resistin on the pathogenesis of osteoarthritis," *Mediators Inflamm*, vol. 2019, Article ID 1532164, 2019.
- [43] H. Mu, R. Ohashi, S. Yan et al., "Adipokine resistin promotes in vitro angiogenesis of human endothelial cells," *Cardiovascular Research*, vol. 70, no. 1, pp. 146–157, 2006.
- [44] M. A. Burmeister, J. E. Ayala, H. Smouse et al., "The hypothalamic glucagon-like peptide 1 receptor is sufficient but not necessary for the regulation of energy balance and glucose homeostasis in mice," *Diabetes*, vol. 66, no. 2, pp. 372–384, 2017.
- [45] S. Zheng, J. Xu, S. Xu et al., "Association between circulating adipokines, radiographic changes, and knee cartilage volume in patients with knee osteoarthritis," *Scandinavian Journal of Rheumatology*, vol. 45, no. 3, pp. 224–229, 2016.
- [46] J. Martel-Pelletier, J.-P. Raynaud, M. Dorais, F. Abram, and J.-P. Pelletier, "The levels of the adipokines adiponin and leptin are associated with knee osteoarthritis progression as assessed by MRI and incidence of total knee replacement in symptomatic osteoarthritis patients: a post hoc analysis," *Rheumatology*, vol. 55, no. 4, pp. 680–688, 2016.
- [47] J. H. Lee, T. Ort, K. Ma et al., "Resistin is elevated following traumatic joint injury and causes matrix degradation and release of inflammatory cytokines from articular cartilage in vitro," *Osteoarthritis and Cartilage*, vol. 17, no. 5, pp. 613–620, 2009.
- [48] S. W. Wong, M.-J. Kwon, A. M. K. Choi, H.-P. Kim, K. Nakahira, and D. H. Hwang, "Fatty acids modulate Toll-like Receptor 4 activation through regulation of receptor dimerization and recruitment into lipid rafts in a reactive oxygen species-dependent manner," *Journal of Biological Chemistry*, vol. 284, no. 40, pp. 27384–27392, 2009.
- [49] C. Herder, E. J. Brunner, W. Rathmann et al., "Elevated levels of the anti-inflammatory interleukin-1 receptor antagonist precede the onset of type 2 diabetes: the Whitehall II study," *Diabetes Care*, vol. 32, no. 3, pp. 421–423, 2009.
- [50] A. R. Sun, S. K. Panchal, T. Friis et al., "Obesity-associated metabolic syndrome spontaneously induces infiltration of pro-inflammatory macrophage in synovium and promotes osteoarthritis," *PLoS One*, vol. 12, no. 3, Article ID e0183693, 2017.
- [51] C. R. Scanzello, B. McKeon, B. H. Swaim et al., "Synovial inflammation in patients undergoing arthroscopic meniscectomy: molecular characterization and relationship to symptoms," *Arthritis & Rheumatism*, vol. 63, no. 2, Article ID 391e400, 2011.
- [52] E. Distel, T. Cadoudal, S. Durant, A. Poignard, X. Chevalier, and C. Benelli, "The infrapatellar fat pad in knee osteoarthritis: an important source of interleukin-6 and its soluble receptor," *Arthritis & Rheumatism*, vol. 60, no. 11, pp. 3374–3377, 2009.
- [53] A. T. Rufino, S. C. Rosa, F. Judas, A. Mobasher, M. C. Lopes, and A. F. Mendes, "Expression and function of K(ATP) channels in normal and osteoarthritic human chondrocytes: possible role in glucose sensing," *Journal of Cellular Biochemistry*, vol. 114, no. 8, pp. 1879–1889, 2013.
- [54] J. DeGroot, N. Verzijl, M. J. G. Wenting-Van Wijk et al., "Accumulation of advanced glycation end products as a molecular mechanism for aging as a risk factor in osteoarthritis," *Arthritis & Rheumatism*, vol. 50, no. 4, pp. 1207–1215, 2004.
- [55] M. Lotfy, J. Adegate, H. Kalasz, J. Singh, and E. Adegate, "Chronic complications of diabetes mellitus: a mini review," *Current Diabetes Reviews*, vol. 13, no. 13, pp. 3–10, 2017.
- [56] H. Huang, Z.-J. Wang, H.-B. Zhang et al., "The function of PPAR γ /AMPK/SIRT-1 pathway in inflammatory response of human articular chondrocytes stimulated by advanced glycation end products," *Biological and Pharmaceutical Bulletin*, vol. 42, no. 8, pp. 1303–1309, 2019.
- [57] A. Bierhaus and P. P. Nawroth, "Multiple levels of regulation determine the role of the receptor for AGE (RAGE) as common soil in inflammation, immune responses and diabetes mellitus and its complications," *Diabetologia*, vol. 52, no. 11, pp. 2251–2263, 2009.
- [58] D. M. Muoio, R. C. Noland, J.-P. Kovalik et al., "Muscle-specific deletion of carnitine acetyltransferase compromises glucose tolerance and metabolic flexibility," *Cell Metabolism*, vol. 15, no. 5, pp. 764–777, 2012.
- [59] A. Malek Mahdavi, R. Mahdavi, and S. Kolahi, "Effects of l-carnitine supplementation on serum inflammatory factors and matrix metalloproteinase enzymes in females with knee osteoarthritis: a randomized, double-blind, placebo-controlled pilot study," *Journal of the American College of Nutrition*, vol. 35, no. 7, pp. 597–603, 2016.
- [60] P. Ježek, T. Olejár, K. Smolková et al., "Antioxidant and regulatory role of mitochondrial uncoupling protein UCP2 in pancreatic beta-cells," *Physiological Research*, vol. 63, pp. S73–S91, 2014.
- [61] M. C. Sádaba, I. Martín-Estal, J. E. Puche, and I. Castilla-Cortázar, "Insulin-like growth factor 1 (IGF-1) therapy: mitochondrial dysfunction and diseases," *Biochimica et Biophysica Acta (BBA)—Molecular Basis of Disease*, vol. 1862, no. 7, pp. 1267–1278, 2016.
- [62] J. M. Moreno-Navarrete, G. Blasco, G. Xifra et al., "Obesity is associated with gene expression and imaging markers of iron accumulation in skeletal muscle," *The Journal of Clinical Endocrinology & Metabolism*, vol. 101, no. 3, pp. 1282–1289, 2016.
- [63] Y. Panahi, N. Khalili, E. Sahebi et al., "Antioxidant effects of curcuminoids in patients with type 2 diabetes mellitus: a randomized controlled trial," *Inflammopharmacology*, vol. 25, no. 1, pp. 25–31, 2017.
- [64] R. S. Lane, Y. Fu, S. Matsuzaki, M. Kinter, K. M. Humphries, and T. M. Griffin, "Mitochondrial respiration and redox coupling in articular chondrocytes," *Arthritis Research & Therapy*, vol. 17, p. 54, 2015.
- [65] M. J. López-Armada, B. Caramés, M. A. Martín et al., "Mitochondrial activity is modulated by TNF α and IL-1 β in normal human chondrocyte cells," *Osteoarthritis and Cartilage*, vol. 14, no. 10, pp. 1011–1022, 2006.
- [66] D. Ziegler, "Current concepts in the management of diabetic polyneuropathy," *Current Diabetes Reviews*, vol. 7, no. 3, pp. 208–220, 2011.
- [67] E. T. Parlevliet, C. P. Coomans, P. C. N. Rensen, and J. A. Romijn, "The brain modulates insulin sensitivity in multiple tissues," *Frontiers of Hormone Research*, vol. 42, pp. 50–58, 2014.
- [68] K. Gerozisis, "Brain insulin: regulation, mechanisms of action and functions," *Cellular and Molecular Neurobiology*, vol. 23, no. 1, pp. 1–25, 2003.
- [69] X. Zhai, C. Sun, P. Rong et al., "A correlative relationship between chronic pain and insulin resistance in Zucker fatty rats: role of downregulation of insulin receptors," *The Journal of Pain*, vol. 17, no. 4, pp. 404–413, 2016.
- [70] J. Greisen, C. B. Juhl, T. Grøfte, H. Vilstrup, T. S. Jensen, and O. Schmitz, "Acute pain induces insulin resistance in humans," *Anesthesiology*, vol. 95, no. 3, pp. 578–584, 2001.
- [71] G. Burnstock, "Purinergic signalling: therapeutic developments," *Frontiers in Pharmacology*, vol. 8, p. 661, 2017.

- [72] E. R. Lazarowski, R. C. Boucher, and T. K. Harden, "Mechanisms of release of nucleotides and integration of their action as P2X- and P2Y-receptor activating molecules," *Molecular Pharmacology*, vol. 64, no. 4, pp. 785–795, 2003.
- [73] N. Kumahashi, K. Naitou, H. Nishi et al., "Correlation of changes in pain intensity with synovial fluid adenosine triphosphate levels after treatment of patients with osteoarthritis of the knee with high-molecular-weight hyaluronic acid," *The Knee*, vol. 18, no. 3, pp. 160–164, 2011.
- [74] L. Miyamoto, T. Toyoda, T. Hayashi et al., "Effect of acute activation of 5'-AMP-activated protein kinase on glycogen regulation in isolated rat skeletal muscle," *Journal of Applied Physiology*, vol. 102, no. 3, pp. 1007–1013, 2007.
- [75] A. K. Saha, A. J. Schwarsin, R. Roduit et al., "Activation of malonyl-CoA decarboxylase in rat skeletal muscle by contraction and the AMP-activated protein kinase activator 5-Aminoimidazole-4-carboxamide-1- β -d-ribofuranoside," *Journal of Biological Chemistry*, vol. 275, no. 32, pp. 24279–24283, 2000.
- [76] Y. Wang, X. Zhao, M. Lotz, R. Terkeltaub, and R. Liu-Bryan, "Mitochondrial biogenesis is impaired in osteoarthritis chondrocytes but reversible via peroxisome proliferator-activated receptor γ coactivator 1 α ," *Arthritis & Rheumatology*, vol. 67, no. 8, pp. 2141–2153, 2015.
- [77] H. Zong, J. M. Ren, L. H. Young et al., "AMP kinase is required for mitochondrial biogenesis in skeletal muscle in response to chronic energy deprivation," *Proceedings of the National Academy of Sciences*, vol. 99, no. 25, pp. 15983–15987, 2002.
- [78] D. M. Gwinn, D. B. Shackelford, D. F. Egan et al., "AMPK phosphorylation of raptor mediates a metabolic checkpoint," *Molecular Cell*, vol. 30, no. 2, pp. 214–226, 2008.
- [79] B. B. Kahn, T. Alquier, D. Carling, and D. G. Hardie, "AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism," *Cell Metabolism*, vol. 1, no. 1, pp. 15–25, 2005.
- [80] C. Affourtit, "Mitochondrial involvement in skeletal muscle insulin resistance: a case of imbalanced bioenergetics," *Biochimica et Biophysica Acta (BBA)—Bioenergetics*, vol. 1857, no. 10, pp. 1678–1693, 2016.
- [81] J. Usprech, G. Chu, R. Giardini-Rosa, K. Martin, and S. D. Waldman, "The therapeutic potential of exogenous adenosine triphosphate (ATP) for cartilage tissue engineering," *Cartilage*, vol. 3, no. 4, pp. 364–373, 2012.
- [82] S. Horman, D. Vertommen, R. Heath et al., "Insulin antagonizes ischemia-induced Thr172Phosphorylation of AMP-activated protein kinase α -subunits in heart via hierarchical phosphorylation of ser485/491," *Journal of Biological Chemistry*, vol. 281, no. 9, pp. 5335–5340, 2006.
- [83] N. Fujii, R. C. Ho, Y. Manabe et al., "Ablation of AMP-activated protein kinase 2 activity exacerbates insulin resistance induced by high-fat feeding of mice," *Diabetes*, vol. 57, no. 11, pp. 2958–2966, 2008.
- [84] M. Horikoshi, K. Hara, J. Ohashi et al., "A polymorphism in the AMPK 2 subunit gene is associated with insulin resistance and type 2 diabetes in the Japanese population," *Diabetes*, vol. 55, no. 4, pp. 919–923, 2006.
- [85] M. Qatanani, Y. Tan, R. Dobrin et al., "Inverse regulation of inflammation and mitochondrial function in adipose tissue defines extreme insulin sensitivity in morbidly obese patients," *Diabetes*, vol. 62, no. 3, pp. 855–863, 2013.
- [86] G. K. Bandyopadhyay, J. G. Yu, J. Ofrecio, and J. M. Olefsky, "Increased malonyl-CoA levels in muscle from obese and type 2 diabetic subjects lead to decreased fatty acid oxidation and increased lipogenesis; thiazolidinedione treatment reverses these defects," *Diabetes*, vol. 55, no. 8, pp. 2277–2285, 2006.
- [87] R. K. June, R. Liu-Bryan, F. Long, and T. M. Griffin, "Emerging role of metabolic signaling in synovial joint remodeling and osteoarthritis," *Journal of Orthopaedic Research*, vol. 34, no. 12, pp. 2048–2058, 2016.
- [88] R. Terkeltaub, B. Yang, M. Lotz, and R. Liu-Bryan, "Chondrocyte AMP-activated protein kinase activity suppresses matrix degradation responses to proinflammatory cytokines interleukin-1 β and tumor necrosis factor α ," *Arthritis & Rheumatism*, vol. 63, no. 7, pp. 1928–1937, 2011.
- [89] E. V. Tchetina, G. A. Markova, A. R. Poole et al., "Deferoxamine suppresses collagen cleavage, protease, cytokine, COL10A1 expression and upregulates AMPK and Krebs cycle genes in human osteoarthritic cartilage," *International Journal of Rheumatology*, vol. 2016, no. 14, Article ID 6432867, 2016.
- [90] L.-Y. Chen, Y. Wang, R. Terkeltaub, and R. Liu-Bryan, "Activation of AMPK-SIRT3 signaling is chondroprotective by preserving mitochondrial DNA integrity and function," *Osteoarthritis and Cartilage*, vol. 26, no. 11, pp. 1539–1550, 2018.
- [91] D. M. Schumann, K. Maedler, I. Franklin et al., "The Fas pathway is involved in pancreatic beta cell secretory function," *Proceedings of the National Academy of Sciences*, vol. 104, no. 8, pp. 2861–2866, 2007.
- [92] A. D. Pradhan, J. E. Manson, N. Rifai, J. E. Buring, and P. M. Ridker, "C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus," *JAMA*, vol. 286, no. 3, pp. 327–334, 2001.
- [93] B. J. Heard, M. J. Fritzler, J. P. Wiley et al., "Intraarticular and systemic inflammatory profiles may identify patients with osteoarthritis," *The Journal of Rheumatology*, vol. 40, no. 8, pp. 1379–1387, 2013.
- [94] M. Attur, I. Belitskaya-Levy, C. Oh et al., "Increased interleukin-1 β gene expression in peripheral blood leukocytes is associated with increased pain and predicts risk for progression of symptomatic knee osteoarthritis," *Arthritis & Rheumatism*, vol. 63, no. 7, Article ID 1908e17, 2011.
- [95] X. Jin, J. R. Beguerie, W. Zhang et al., "Circulating C reactive protein in osteoarthritis: a systematic review and meta-analysis," *Annals of the Rheumatic Diseases*, vol. 74, no. 4, pp. 703–710, 2015.
- [96] D. F. G. Remst, E. N. Blaney Davidson, and P. M. van der Kraan, "Unravelling osteoarthritis-related synovial fibrosis: a step closer to solving joint stiffness," *Rheumatology*, vol. 54, no. 11, pp. 1954–1963, 2015.
- [97] J. Ma, S. J. Chadban, C. Y. Zhao et al., "TLR4 activation promotes podocyte injury and interstitial fibrosis in diabetic nephropathy," *PLoS One*, vol. 9, no. 5, Article ID e97985, 2014.
- [98] W. C. Tsai, F. C. Liang, J. W. Cheng et al., "High glucose concentration up-regulates the expression of matrix metalloproteinase-9 and -13 in tendon cells," *BMC Musculoskeletal Disorders*, vol. 14, no. 1, p. 255, 2013.
- [99] R. Ene, R. D. Sinescu, P. Ene, M. M. Cirstoiu, and F. C. Cirstoiu, "Synovial inflammation in patients with different stages of knee osteoarthritis," *Romanian journal of Morphology and Embryology*, vol. 56, no. 56, pp. 169–173, 2015.
- [100] F. Eymard, A. Pigenet, D. Citadelle et al., "Knee and hip intra-articular adipose tissues (IAATs) compared with autologous subcutaneous adipose tissue: a specific phenotype

- for a central player in osteoarthritis," *Annals of the Rheumatic Diseases*, vol. 76, no. 6, pp. 1142–1148, 2017.
- [101] C. M. Spies, R. H. Straub, and F. Buttgerit, "Energy metabolism and rheumatic diseases: from cell to organism," *Arthritis Research & Therapy*, vol. 14, no. 3, p. 216, 2012.
- [102] R. H. Straub, M. Cutolo, F. Buttgerit, and G. Pongratz, "Review: energy regulation and neuroendocrine-immune control in chronic inflammatory diseases," *Journal of Internal Medicine*, vol. 267, no. 6, pp. 543–560, 2010.
- [103] R. H. Straub, "Concepts of evolutionary medicine and energy regulation contribute to the etiology of systemic chronic inflammatory diseases," *Brain, Behavior, and Immunity*, vol. 25, no. 1, pp. 1–5, 2011.
- [104] A. E. Nelson, K. D. Allen, Y. M. Golightly, A. P. Goode, and J. M. Jordan, "A systematic review of recommendations and guidelines for the management of osteoarthritis: the chronic osteoarthritis management initiative of the U.S. bone and joint initiative," *Seminars in Arthritis and Rheumatism*, vol. 43, pp. 701–712, 2014.
- [105] M. Kloppenburg and F. Berenbaum, "Osteoarthritis year in review 2019: epidemiology and therapy," *Osteoarthritis Cartilage*, vol. 28, no. 3, pp. 242–248, 2020.
- [106] L. Kang, C.-H. Chen, M.-H. Wu, J.-K. Chang, F.-M. Chang, and J.-T. Cheng, "17 β -Estradiol protects against glucosamine-induced pancreatic β -cell dysfunction," *Menopause*, vol. 21, no. 11, pp. 1239–1248, 2014.
- [107] E. Buzhor, L. Leshansky, J. Blumenthal et al., "Cell-based therapy approaches: the hope for incurable diseases," *Regenerative Medicine*, vol. 9, no. 5, pp. 649–672, 2014.
- [108] M. Packer, "The alchemist's nightmare," *Circulation*, vol. 137, no. 19, pp. 2068–2073, 2018.
- [109] E. Mianehsaz, H. R. Mirzaei, M. Mahjoubin-Tehran et al., "Mesenchymal stem cell-derived exosomes: a new therapeutic approach to osteoarthritis?" *Stem Cell Research & Therapy*, vol. 10, p. 340, 2019.
- [110] H. Mirzaei, A. Sahebkar, M. R. Jaafari, M. Goodarzi, and H. R. Mirzaei, "Diagnostic and therapeutic potential of exosomes in cancer: the beginning of a new tale?" *Journal of Cellular Physiology*, vol. 14, p. 25739, 2016.
- [111] W. S. Toh, R. C. Lai, J. H. P. Hui, and S. K. Lim, "MSC exosome as a cell-free MSC therapy for cartilage regeneration: implications for osteoarthritis treatment," *Seminars in Cell and Developmental Biology*, vol. 67, no. 67, pp. 56–64, 2017.
- [112] S. Zhang, W. C. Chu, R. C. Lai, S. K. Lim, J. H. P. Hui, and W. S. Toh, "Exosomes derived from human embryonic mesenchymal stem cells promote osteochondral regeneration," *Osteoarthritis and Cartilage*, vol. 24, no. 12, pp. 2135–2140, 2016.
- [113] S. Zhang, K. Y. W. Teo, S. J. Chuah, S. K. Lai, S. K. Lim, and W. S. Toh, "MSC exosomes alleviate temporomandibular joint osteoarthritis by attenuating inflammation and restoring matrix homeostasis," *Biomaterials*, vol. 200, pp. 35–47, 2019.
- [114] J. Mohammadi Ayenehdeh, B. Niknam, S. Rasouli et al., "Immunomodulatory and protective effects of adipose tissue-derived mesenchymal stem cells in an allograft islet composite transplantation for experimental autoimmune type 1 diabetes," *Immunology Letters*, vol. 188, pp. 21–31, 2017.
- [115] M. J. Saeedi Borujeni, E. Esfandiary, A. Baradaran et al., "Molecular aspects of pancreatic β -cell dysfunction: oxidative stress, microRNA, and long noncoding RNA," *Journal of Cellular Physiology*, vol. 234, no. 6, pp. 8411–8425, 2019.
- [116] S.-C. Tao, T. Yuan, Y.-L. Zhang, W.-J. Yin, S.-C. Guo, and C.-Q. Zhang, "Exosomes derived from miR-140-5p-over-expressing human synovial mesenchymal stem cells enhance cartilage tissue regeneration and prevent osteoarthritis of the knee in a rat model," *Theranostics*, vol. 7, no. 1, pp. 180–195, 2017.
- [117] N. Heldring, I. Mäger, M. J. A. Wood, K. Le Blanc, and S. E. L. Andaloussi, "Therapeutic potential of multipotent mesenchymal stromal cells and their extracellular vesicles," *Human Gene Therapy*, vol. 26, no. 8, pp. 506–517, 2015.
- [118] M. Zaiou, "circRNAs signature as potential diagnostic and prognostic biomarker for diabetes mellitus and related cardiovascular complications," *Cells*, vol. 9, no. 3, p. 659, 2020.
- [119] K. Abbaszadeh-Goudarzi, S. Radbakhsh, M. H. Pourhanifeh et al., "Circular RNA and diabetes: epigenetic regulator with diagnostic role," *Current Molecular Medicine*, vol. 20, 2020.
- [120] J. Salzman, C. Gawad, P. L. Wang, N. Lacayo, and P. O. Brown, "Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types," *PLoS One*, vol. 7, no. 2, Article ID e30733, 2012.
- [121] Q. Liu, X. Zhang, X. Hu et al., "Circular RNA related to the chondrocyte ECM regulates MMP13 expression by functioning as a MiR-136 "sponge" in human cartilage degradation," *Scientific Reports*, vol. 6, no. 1, p. 22572, 2016.
- [122] Q. Liu, X. Zhang, X. Hu et al., "Emerging roles of circRNA related to the mechanical stress in human cartilage degradation of osteoarthritis," *Molecular Therapy—Nucleic Acids*, vol. 7, pp. 223–230, 2017.
- [123] C.-X. Yu and S. Sun, "An emerging role for circular RNAs in osteoarthritis," *Yonsei Medical Journal*, vol. 59, no. 3, pp. 349–355, 2018.
- [124] B. Liu, J. Li, and M. J. Cairns, "Identifying miRNAs, targets and functions," *Briefings in Bioinformatics*, vol. 15, no. 1, pp. 1–19, 2014.
- [125] T. B. Hansen, T. I. Jensen, B. H. Clausen et al., "Natural RNA circles function as efficient microRNA sponges," *Nature*, vol. 495, no. 7441, pp. 384–388, 2013.
- [126] L.-L. Chen, "The biogenesis and emerging roles of circular RNAs," *Nature Reviews Molecular Cell Biology*, vol. 17, no. 4, pp. 205–211, 2016.
- [127] P. Barata, A. K. Sood, and D. S. Hong, "RNA-targeted therapeutics in cancer clinical trials: current status and future directions," *Cancer Treatment Reviews*, vol. 50, pp. 35–47, 2016.
- [128] D. Nemirov, Y. Nakagawa, Z. Sun et al., "Effect of lubricin mimetics on the inhibition of osteoarthritis in a rat anterior cruciate ligament transection model," *The American Journal of Sports Medicine*, vol. 48, no. 3, pp. 624–634, 2020.
- [129] G. Gibor, N. Ilan, S. Journo et al., "Heparanase is expressed in adult human osteoarthritic cartilage and drives catabolic responses in primary chondrocytes," *Osteoarthritis and Cartilage*, vol. 26, no. 8, pp. 1110–1117, 2018.
- [130] S. Weksler-Zangen, T. Mizrahi, I. Raz, and N. Mirsky, "Glucose tolerance factor extracted from yeast: oral insulin-mimetic and insulin-potentiating agent: in vivo and in vitro studies," *British Journal of Nutrition*, vol. 108, no. 5, pp. 875–882, 2012.
- [131] S. Peansukmanee, A. Vaughan-Thomas, S. D. Carter et al., "Effects of hypoxia on glucose transport in primary equine chondrocytes in vitro and evidence of reduced GLUT1 gene expression in pathologic cartilage in vivo," *Journal of Orthopaedic Research*, vol. 27, no. 4, pp. 529–535, 2009.
- [132] V. Afonso, R. Champy, D. Mitrovic, P. Collin, and A. Lomri, "Reactive oxygen species and superoxide dismutases: role in

- joint diseases," *Joint Bone Spine*, vol. 74, no. 4, pp. 324–329, 2007.
- [133] M. Coleman, M. Brouillette, N. Andresen, R. Oberley-Deegan, and J. Martin, "Differential effects of superoxide dismutase mimetics after mechanical overload of articular cartilage," *Antioxidants*, vol. 6, no. 4, p. 98, 2017.
- [134] R. Terkeltaub, K. Johnson, A. Murphy, and S. Ghosh, "Invited review: the mitochondrion in osteoarthritis," *Mitochondrion*, vol. 1, no. 4, pp. 301–319, 2002.
- [135] K. J. Wolff, P. S. Ramakrishnan, M. J. Brouillette et al., "Mechanical stress and ATP synthesis are coupled by mitochondrial oxidants in articular cartilage," *Journal of Orthopaedic Research*, vol. 31, no. 2, pp. 191–196, 2013.
- [136] W. Goodwin, D. McCabe, E. Sauter et al., "Rotenone prevents impact-induced chondrocyte death," *Journal of Orthopaedic Research*, vol. 28, pp. 1057–1063, 2010.
- [137] C. Gavriilidis, S. Miwa, T. von Zglinicki, R. W. Taylor, and D. A. Young, "Mitochondrial dysfunction in osteoarthritis is associated with down-regulation of superoxide dismutase 2," *Arthritis & Rheumatism*, vol. 65, no. 2, pp. 378–387, 2013.
- [138] A. Courties and J. Sellam, "Osteoarthritis and type 2 diabetes mellitus: what are the links?" *Diabetes Research and Clinical Practice*, vol. 122, pp. 198–206, 2016.
- [139] Y. Y. Leung, J. C. Allen, L. W. Ang, J. M. Yuan, and W. P. Koh, "Diabetes mellitus and the risk of total knee replacement among Chinese in Singapore, the Singapore Chinese Health Study," *Scientific Reports*, vol. 7, p. 40671, 2017.
- [140] D. Perry, M. R. Kolber, C. Korownyk et al., "Top studies relevant to primary care practice," *Canadian Family Physician*, vol. 64, no. 64, pp. 280–285, 2018.
- [141] C. Tong, H. Liang, X. Liu et al., "The protective effects of exenatide against AGEs-induced articular matrix degradation in human primary chondrocytes," *American Journal of Translational Research*, vol. 11, no. 11, pp. 2081–2089, 2019.
- [142] N. Hu, X. Gong, S. Yin et al., "Saxagliptin suppresses degradation of type II collagen and aggrecan in primary human chondrocytes: a therapeutic implication in osteoarthritis," *Artificial Cells, Nanomedicine, and Biotechnology*, vol. 47, no. 1, pp. 3239–3245, 2019.
- [143] H. J. Son, J. Lee, S. Y. Lee et al., "Metformin attenuates experimental autoimmune arthritis through reciprocal regulation of Th17/Treg balance and osteoclastogenesis," *Mediators of Inflammation*, vol. 2014, Article ID 973986, 13 pages, 2014.
- [144] I. V. Shirinsky and V. S. Shirinsky, "Effects of medication-treated diabetes on incidence and progression of knee osteoarthritis: a longitudinal analysis of the osteoarthritis initiative data," *Rheumatology International*, vol. 37, no. 6, pp. 983–991, 2017.
- [145] W. Yue, C. S. Yang, R. S. DiPaola, and X.-L. Tan, "Repurposing of metformin and aspirin by targeting AMPK-mTOR and inflammation for pancreatic cancer prevention and treatment," *Cancer Prevention Research*, vol. 7, no. 4, pp. 388–397, 2014.
- [146] F. V. N. Din, A. Valanciute, V. P. Houde et al., "Aspirin inhibits mTOR signaling, activates AMP-activated protein kinase, and induces autophagy in colorectal cancer cells," *Gastroenterology*, vol. 142, no. 7, pp. 1504–1515, 2012.
- [147] E. Varghese, S. Samuel, M. Abotaleb, S. Cheema, R. Mamtani, and D. Büsselberg, "The 'Yin and Yang' of natural compounds in anticancer therapy of triple-negative breast cancers," *Cancers*, vol. 10, no. 10, p. 346, 2018.
- [148] G. Koren, G. Nordon, K. Radinsky, and V. Shalev, "Identification of repurposable drugs with beneficial effects on glucose control in type 2 diabetes using machine learning," *Pharmacology Research & Perspectives*, vol. 7, Article ID e00529, 2019.
- [149] K. Nakashima, K. Takeuchi, K. Chihara, H. Hotta, and K. Sada, "Inhibition of hepatitis C virus replication through adenosine monophosphate-activated protein kinase-dependent and -independent pathways," *Microbiology and Immunology*, vol. 55, no. 11, pp. 774–782, 2011.
- [150] I. M. Vera, M. T. Grilo Ruivo, L. F. Lemos Rocha et al., "Targeting liver stage malaria with metformin," *JCI Insight*, vol. 4, Article ID e127441, 2019.
- [151] A. Singhal, L. Jie, P. Kumar et al., "Metformin as adjunct antituberculosis therapy," *Science Translational Medicine*, vol. 6, no. 263, Article ID 263ra159, 2014.
- [152] C. Kajiwara, Y. Kusaka, S. Kimura et al., "Metformin mediates protection against *Legionella pneumonia* through activation of AMPK and mitochondrial reactive oxygen species," *The Journal of Immunology*, vol. 200, no. 2, pp. 623–631, 2018.
- [153] L. Luzi and M. G. Radaelli, "Influenza and obesity: its odd relationship and the lessons for COVID-19 pandemic," *Acta Diabetologica*, vol. 5, no. 6, 2020.
- [154] A. K. Madiraju, Y. Qiu, R. J. Perry et al., "Metformin inhibits gluconeogenesis via a redox-dependent mechanism in vivo," *Nature Medicine*, vol. 24, no. 9, pp. 1384–1394, 2018.
- [155] Y. K. Choi and K.-G. Park, "Metabolic roles of AMPK and metformin in cancer cells," *Molecules and Cells*, vol. 36, no. 4, pp. 279–287, 2013.
- [156] E. V. Tchetina, A. R. Poole, E. M. Zaitseva et al., "Differences in Mammalian target of rapamycin gene expression in the peripheral blood and articular cartilages of osteoarthritic patients and disease activity," *Arthritis*, vol. 2013, Article ID 461486, 14 pages, 2013.
- [157] I. Ben Sahra, C. Regazzetti, G. Robert et al., "Metformin, independent of AMPK, induces mTOR inhibition and cell-cycle arrest through REDD1," *Cancer Research*, vol. 71, no. 13, pp. 4366–4372, 2011.
- [158] S. Aminzadeh-Gohari, D. D. Weber, S. Vidali, L. Catalano, B. Kofler, and R. G. Feichtinger, "From old to new—repurposing drugs to target mitochondrial energy metabolism in cancer," *Seminars in Cell & Developmental Biology*, vol. 98, pp. 211–223, 2020.
- [159] E. Fontaine, "Metformin-induced mitochondrial complex I inhibition: facts, uncertainties, and consequences," *Frontiers in Endocrinology*, vol. 9, p. 753, 2018.
- [160] L. Fontana and F. B. Hu, "Optimal body weight for health and longevity: bridging basic, clinical, and population research," *Aging Cell*, vol. 13, no. 3, pp. 391–400, 2014.
- [161] J. Most, V. Tosti, L. M. Redman, and L. Fontana, "Calorie restriction in humans: an update," *Ageing Research Reviews*, vol. 39, pp. 36–45, 2017.
- [162] L. Fontana and L. Partridge, "Promoting health and longevity through diet: from model organisms to humans," *Cell*, vol. 161, no. 1, pp. 106–118, 2015.
- [163] R. F. Hamman, R. R. Wing, S. L. Edelstein et al., "Effect of weight loss with lifestyle intervention on risk of diabetes," *Diabetes Care*, vol. 29, no. 9, pp. 2102–2107, 2006.
- [164] M.-J. Lee, Y. Wu, and S. K. Fried, "Adipose tissue heterogeneity: implication of depot differences in adipose tissue for obesity complications," *Molecular Aspects of Medicine*, vol. 34, no. 1, pp. 1–11, 2013.

- [165] A. R. Barnosky, K. K. Hoddy, T. G. Unterman, and K. A. Varady, "Intermittent fasting vs daily calorie restriction for type 2 diabetes prevention: a review of human findings," *Translational Research*, vol. 164, no. 4, pp. 302–311, 2014.
- [166] L. B. Radakovich, A. J. Marolf, L. A. Culver, and K. S. Santangelo, "Calorie restriction with regular chow, but not a high-fat diet, delays onset of spontaneous osteoarthritis in the Hartley Guinea pig model," *Arthritis Research & Therapy*, vol. 21, p. 145, 2019.
- [167] J. N. McNeill, C. L. Wu, K. N. Rabey, D. Schmitt, and F. Guilak, "Life-long caloric restriction does not alter the severity of age-related osteoarthritis," *Age*, vol. 36, p. 9669, 2014.
- [168] T. Pearson, J. A. D. Wattis, J. R. King, I. A. MacDonald, and D. J. Mazzatti, "The effects of insulin resistance on individual tissues: an application of a mathematical model of metabolism in humans," *Bulletin of Mathematical Biology*, vol. 78, no. 6, pp. 1189–1217, 2016.
- [169] P. D. Neufer, M. M. Bamman, D. M. Muoio et al., "Understanding the cellular and molecular mechanisms of physical activity-induced health benefits," *Cell Metabolism*, vol. 22, no. 1, pp. 4–11, 2015.
- [170] B. Kiens, "Skeletal muscle lipid metabolism in exercise and insulin resistance," *Physiological Reviews*, vol. 86, no. 1, pp. 205–243, 2006.
- [171] L. J. Goodyear and B. B. Kahn, "Exercise, glucose transport, and insulin sensitivity," *Annual Review of Medicine*, vol. 49, no. 49, pp. 235–261, 1998.
- [172] E. A. Richter and M. Hargreaves, "Exercise, GLUT4, and skeletal muscle glucose uptake," *Physiological Reviews*, vol. 93, no. 3, pp. 993–1017, 2013.
- [173] F. G. S. Toledo and B. H. Goodpaster, "The role of weight loss and exercise in correcting skeletal muscle mitochondrial abnormalities in obesity, diabetes and aging," *Molecular and Cellular Endocrinology*, vol. 379, no. 1-2, pp. 30–34, 2013.
- [174] P. M. Coen, C. J. Tanner, N. L. Helbling et al., "Clinical trial demonstrates exercise following bariatric surgery improves insulin sensitivity," *Journal of Clinical Investigation*, vol. 125, no. 1, pp. 248–257, 2015.
- [175] T. R. Koves, J. R. Ussher, R. C. Noland et al., "Mitochondrial overload and incomplete fatty acid oxidation contribute to skeletal muscle insulin resistance," *Cell Metabolism*, vol. 7, no. 1, pp. 45–56, 2008.
- [176] E. B. Arias, J. Kim, K. Funai, and G. D. Cartee, "Prior exercise increases phosphorylation of Akt substrate of 160 kDa (AS160) in rat skeletal muscle," *American Journal of Physiology-Endocrinology and Metabolism*, vol. 292, no. 4, pp. E1191–E1200, 2007.
- [177] C. M. Castorena, E. B. Arias, N. Sharma, and G. D. Cartee, "Postexercise improvement in insulin-stimulated glucose uptake occurs concomitant with greater AS160 phosphorylation in muscle from normal and insulin-resistant rats," *Diabetes*, vol. 63, no. 7, pp. 2297–2308, 2014.
- [178] M. Ferretti, R. Gassner, Z. Wang et al., "Biomechanical signals suppress proinflammatory responses in cartilage: early events in experimental antigen-induced arthritis," *The Journal of Immunology*, vol. 177, no. 12, pp. 8757–8766, 2006.
- [179] I. C. Helmark, U. R. Mikkelsen, J. Børglum et al., "Exercise increases interleukin-10 levels both intraarticularly and perisynovially in patients with knee osteoarthritis: a randomized controlled trial," *Arthritis Research & Therapy*, vol. 12, no. 4, p. R126, 2010.
- [180] V. A. Narkar, M. Downes, R. T. Yu et al., "AMPK and PPAR δ agonists are exercise mimetics," *Cell*, vol. 134, no. 3, pp. 405–415, 2008.
- [181] B. H. Goodpaster and L. M. Sparks, "Metabolic flexibility in health and disease," *Cell Metabolism*, vol. 25, no. 5, pp. 1027–1036, 2017.