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## Rewiring regulatory T cells for tumour killing

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## Abstract

In mouse models of cancer, the inhibition of a set of regulatory proteins improves checkpointblockade therapy by causing regulatory T cells to produce the cytokine interferon- $\gamma$ .

Immune surveillance is the first line of the body's defence against the uncontrolled proliferation of tumour cells, as leukocytes such as natural killer (NK) cells and cytotoxic CD8<sup>+</sup> T lymphocytes (CTL) efficiently kill neoplastic cells. But tumour cells can explore immunosuppressive networks and evade anti-tumour immunity<sup>1</sup>. As an endogenous control mechanism over autoimmunity that can nevertheless also result in impaired anti-tumour immunity, forkhead-box P3 (FOXP3)<sup>+</sup> CD25<sup>+</sup> regulatory T (T<sub>REG</sub>) cells suppress the effector function of NK cells, CTLs and antigen-presenting cells (APCs) through multiple pathways<sup>2</sup> (Fig. 1). In light of their immune suppressor activities<sup>2–4</sup>, and given that high numbers of T<sub>REG</sub> cells in the tumour microenvironment are associated with poor prognosis, T<sub>REG</sub> cells are an attractive target for anticancer therapy.

In humans,  $T_{REG}$ -cell deficiency caused by mutations in the  $T_{REG}$ -cell master transcription regulator FOXP3 results in IPEX (immune dysregulation, polyendocrinopathy, enteropathy, X-linked) syndrome — a severe and systemic autoimmune disorder. The dramatic effects of systemic  $T_{REG}$ -cell deficiency suggest that the complete removal of  $T_{REG}$  cells is not a feasible approach for cancer immunotherapy. In a study published in *Nature*, Thorsten Mempel and colleagues now report that the disruption of the T-cell receptor (TCR) signalosome complex CARMA1–BCL10–MALT1 (CBM) in  $T_{REG}$  cells causes a reversion in the cell's phenotype, from immune suppression to immune activation, leading to the production of interferon (IFN)- $\gamma$  in the tumour microenvironment and enhancing local antitumour activity<sup>5</sup>.

Mempel and colleagues identified a role for the CBM signalosome in  $T_{REG}$  cells by crossing  $Foxp3^{YFP-Cre}$  mice to *Carma1* flox/flox mice, which produced offspring with  $T_{REG}$ -cell-specific *Carma1* deficiency. In these mice, genetic deletion of *Carma1* in  $T_{REG}$  cells did not affect the expression of FOXP3 or other markers of  $T_{REG}$  cells, but triggered the expression of the immune-effector transcription factors T-box transcription factor TBX21 (T-bet) and RAR-related orphan receptor- $\gamma t$  (ROR- $\gamma t$ ), as well as the production of the pro-inflammatory cytokines tumour necrosis factor (TNF), interleukin (IL)-17 and IFN- $\gamma$ . Importantly, the monoallelic deletion of *Carma1* in  $T_{REG}$  cells also resulted in tumour

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control while avoiding systemic autoimmunity in mice bearing either poorly immunogenic D4M.3A melanoma or immunogenic MC38 adenocarcinoma. In these models, IFN- $\gamma$  production by T<sub>REG</sub> cells was critical for controlling tumour progression in a process restricted to the tumour microenvironment.

To explore the mechanism underlying the anti-tumour effect of *Carma1<sup>-/-</sup>* T<sub>REG</sub> cells, Mempel and colleagues explored the effects of the acute deletion of *Carma1* in the tamoxifen-induced *Foxp3*<sup>Cre–ERT2</sup> *Carma1*<sup>flox/flox</sup> model. In agreement with prior data, tumour growth was arrested in these animals only after tamoxifen administration. Notably, macrophages from *Foxp3*<sup>Cre–ERT2</sup> *Carma1*<sup>flox/flox</sup> tumour-bearing animals expressed higher levels of major histocompatibility complex (MHC) class II when compared with *Foxp3*<sup>Cre–ERT2</sup> *Carma1*<sup>+/+</sup> control mice. Concurrently, tumour cells expressed higher levels of MHC-I, making them more sensitive to CTL-dependent killing. However, tumour cells also expressed higher levels of programmed death-ligand 1 (PD-L1, also known as B7-H1), which may have contained the anti-tumour immune response. This was found to be the case as, while *Foxp3*<sup>Cre–ERT2</sup> *Carma1*<sup>+/+</sup> animals responded moderately to PD-1 blockade, *Foxp3*<sup>Cre–ERT2</sup> *Carma1*<sup>flox/flox</sup> mice manifested significantly enhanced tumour regression in response to the checkpoint inhibitor.

The lack of pharmacological inhibitors for CARMA1 prevents the non-genetic interrogation of the function of this factor in  $T_{REG}$  cells, but inhibitors exist for MALT1, an enzymatic component of the CBM complex. Administration of two MALT1 inhibitors (mepazine and MI-2) in the D4M.3A tumour model triggered a phenotype that approximates that of conditional deletion of *Carma1* in  $T_{REG}$  cells, including an increase in the frequency of T cells producing IFN- $\gamma$  and TNF, and the elevated expression of MHC-I and PD-L1 on cancer cells. Furthermore, when D4M.3A melanoma-bearing male mice were treated with mepazine and anti-PD-1, the combination therapy yielded synergistic anti-tumour activity. Hence, the authors conclude that the inhibition of the CARMA1–BCL10–MALT1 signalosome induces an inflammatory phenotype in  $T_{REG}$  cells and generates potent antitumour immunity (Fig. 2).

Human  $T_{REG}$  cells can express inflammatory and effector cytokines, including IL-8, IL-17 and IFN- $\gamma$  in the tumour microenvironment and in inflammatory conditions<sup>6–8</sup>. Thus, that Mempel and colleagues observed IFN- $\gamma^+$   $T_{REG}$  cells in the mouse tumour microenvironment is not surprising. However, that CARMA1 can dramatically alter the functional properties of FOXP3<sup>+</sup> cells to enable IFN- $\gamma$  expression in  $T_{REG}$  cells is an important observation. Although the specific deletion of neuropilin-1 results in IFN- $\gamma$ expression in  $T_{REG}$  cells<sup>9</sup>, whether the CBM signalosome and neuropilin-1 are connected in regulating the  $T_{REG}$  phenotype is unknown.

As  $T_{REG}$  cells are regulated by the strength of the TCR signal<sup>10</sup>, the CBM signalosome may quantitatively and qualitatively alter TCR signalling in  $T_{REG}$  cells. This potential mechanistic relationship would partially explain why  $T_{REG}$  cells express IFN- $\gamma$  in the tumour and not in lymph nodes. In addition, it is known that the CBM signalosome can activate several downstream pathways, including nuclear factor- $\kappa$ B (NF- $\kappa$ B), activator protein 1 and mammalian target of rapamycin (mTOR). Mempel and co-authors provided

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evidence that NF- $\kappa$ B has no role in their models. Given that the integrity of T<sub>REG</sub> cells is particularly regulated by the balance of mTORC1 and mTORC2 (ref.<sup>11</sup>), that the phenotypic plasticity of T<sub>REG</sub> cells can be regulated by the crosstalk of metabolic pathways, and that IFN- $\gamma$  production is linked to the metabolic state of T cells<sup>12</sup>, it is sound to investigate the mechanistic association between CBM signalosome components, mTOR and IFN- $\gamma$ expression in T<sub>REG</sub> cells. The authors show that CARMA1 deficiency induces the expression of T-bet and ROR- $\gamma$ t; it is therefore tempting to speculate whether CARMA1– CBM has a role in the interplay between the three key T-cell transcription factors FOXP3, Tbet and ROR- $\gamma$ t in T<sub>REG</sub> cells, and whether this interplay may determine the functional fate of T<sub>REG</sub> cells.

The observation of an increase in MHC-I expression in macrophages, and of PD-L1 expression on tumour cells under stimulation by IFN- $\gamma^+$  T<sub>REG</sub> cells, offers a mechanism for the therapeutic efficacy of the phenotypically reverted T<sub>REG</sub> cells and would explain why PD-1 blockade concomitant with *Carma1*-deletion leads to the strong rejection of tumours. However, it is unclear whether IFN- $\gamma$  from IFN- $\gamma^+$  T<sub>REG</sub> cells also led to the induction of PD-L1 expression on macrophages or myeloid dendritic cells (mDCs). PD-L1<sup>+</sup> antigenpresenting cells are the primary targets of PD-L1 and PD-1 blockade therapy<sup>13–15</sup>, and several studies have documented that PD-L1 is expressed on macrophages and mDCs in human tumour-draining lymph nodes and in the tumour microenvironments. Ultimately, multiple environmental, genetic and metabolic determinants may shape the outcome of PD-1 and PD-L1 blockade therapy in different experimental and clinical settings<sup>16</sup>. A clear understanding of the IFN- $\gamma^+$  T<sub>REG</sub> cell's interacting partners may help to develop new mechanistically informed therapeutic strategies for cancer patients.

## References

1. Zou W Nat. Rev. Cancer 5, 263–274 (2005). [PubMed: 15776005]

2. Zou W Nat. Rev. Immunol 6, 295–307 (2006). [PubMed: 16557261]

3. Curiel TJ et al. Nat. Med 10, 942–949 (2004). [PubMed: 15322536]

4. Zou L et al. Cancer Res. 64, 8451–8455 (2004). [PubMed: 15548717]

5. Di Pilato M et al. Nature 570, 112–116 (2019). [PubMed: 31092922]

6. Kryczek I et al. J. Immunol 186, 4388–4395 (2011). [PubMed: 21357259]

7. Duhen T et al. Blood 119, 4430–4440 (2012). [PubMed: 22438251]

8. Kryczek I et al. Oncoimmunology 5, e1105430 (2016). [PubMed: 27622054]

9. Overacre-Delgoffe AE et al. Cell 169, 1130–1141 (2017). [PubMed: 28552348]

10. Sakaguchi S Nat. Immunol 6, 345–352 (2005). [PubMed: 15785760]

11. Zeng H et al. Nature 499, 485–490 (2013). [PubMed: 23812589]

12. Chang CH et al. Cell 153, 1239–1251 (2013). [PubMed: 23746840]

13. Curiel TJ et al. Nat. Med 9, 562–567 (2003). [PubMed: 12704383]

14. Lin H et al. J. Clin. Invest 128, 805–815 (2018). [PubMed: 29337305]

15. Tang H et al. J. Clin. Invest 128, 580–588 (2018). [PubMed: 29337303]

16. Zou W, Wolchok JD & Chen L Sci. Transl. Med 8, 328rv324 (2016).

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### Fig. 1 |. Effect of $T_{\mbox{REG}}$ cells on immune cells.

 $T_{REG}$  cells are developed in the thymus (central  $T_{REG}$  cells;  $cT_{REG}$ ) or generated and expanded outside the thymus (peripheral  $T_{REG}$  cells;  $pT_{REG}$ ); can directly target antigenpresenting cells, NK cells and effector T cells; and mediate immunosuppression via distinct molecular mechanisms.

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#### Fig. 2 |. Effect of the cBm signalosome on the phenotype of $T_{\mbox{REG}}$ cells.

**a**, Expression levels of the CBM-complex scaffold protein CARMA1 are associated with different levels of IFN- $\gamma$  production in T<sub>REG</sub> cells following in vitro stimulation and in vivo tumour challenge. LN, lymph node. **b**, The genetic or pharmacological loss of CBM signalling results in the increased production of IFN- $\gamma$  in tumour-associated T<sub>REG</sub> cells. IFN- $\gamma$  production by T<sub>REG</sub> cells may affect MHC-I, MHC-II and PD-L1 on tumour cells and on antigen-presenting cells.  $\uparrow$ , increased expression; ?, unknown effects.

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