

Prion protein glycans reduce intracerebral fibril formation and spongiosis in prion disease

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Posttranslational modifications (PTMs) are common among proteins that aggregate in neurodegenerative disease, yet how PTMs impact the aggregate conformation and disease progression remains unclear. By engineering knockin mice expressing prion protein (PrP) lacking 2 N-linked glycans (*Prnp*^{180Q/196Q}), we provide evidence that glycans reduce spongiform degeneration and hinder plaque formation in prion disease. *Prnp*^{180Q/196Q} mice challenged with 2 subfibrillar, non-plaque-forming prion strains instead developed plaques highly enriched in ADAM10-cleaved PrP and heparan sulfate (HS). Intriguingly, a third strain composed of intact, glycoposphatidylinositol-anchored (GPI-anchored) PrP was relatively unchanged, forming diffuse, HS-deficient deposits in both the *Prnp*^{180Q/196Q} and WT mice, underscoring the pivotal role of the GPI-anchor in driving the aggregate conformation and disease phenotype. Finally, knockin mice expressing triglycosylated PrP (*Prnp*^{187N}) challenged with a plaque-forming prion strain showed a phenotype reversal, with a striking disease acceleration and switch from plaques to predominantly diffuse, subfibrillar deposits. Our findings suggest that the dominance of subfibrillar aggregates in prion disease is due to the replication of GPI-anchored prions, with fibrillar plaques forming from poorly glycosylated, GPI-anchorless prions that interact with extracellular HS. These studies provide insight into how PTMs impact PrP interactions with polyanionic cofactors, and highlight PTMs as a major force driving the prion disease phenotype.

Introduction

Amyloid plaques accumulate in the brain of patients with Alzheimer's disease and certain familial prion diseases, and are often associated with a clinical course that progresses for more than 3 years (1–3). In contrast, in sporadic Creutzfeldt-Jakob disease (sCJD), prion aggregates more commonly form diffuse, synaptic, or plaque-like deposits in the brain and symptoms advance with extraordinary rapidity, with a median of 6 months from clinical onset to terminal disease (4–6). Similar to sCJD, in animal prion diseases, such as chronic wasting disease of cervids, prion aggregates frequently form diffuse or punctate deposits and the clinical phase is brief (7–11). Although the histopathology has been extensively characterized, the molecular mechanisms that drive the prion fold into fibrillar plaques or subfibrillar diffuse deposits are unclear.

Prion aggregates accumulate during the conversion of the cellular prion protein, PrP^C, into a β -sheet rich multimer known as PrP^{Sc} (12, 13). Although the PrP^{Sc} structure has not yet been solved,

several lines of evidence suggest that PrP posttranslational modifications (PTMs), a glycoposphatidylinositol (GPI) anchor and 2 N-linked glycans (14–18), impact the prion conformation (19–21), species barriers (22, 23), and disease progression (24, 25). Previous studies have demonstrated that PrP^{Sc} maintains the sialylated glycans (α 2,6-linked) (17, 26) and that sialylation influences prion conversion and intracellular trafficking (27–31), yet how these bulky, anionic glycans modify prion assembly and the disease phenotype remains unresolved (32). Additionally, while cell and mouse models have revealed that unglycosylated PrP^C can be converted into PrP^{Sc} (24, 33), further study has been hampered by the intracellular retention of unglycosylated PrP^C (34, 35), which normally traffics to the cell surface (36, 37).

To address the question of how glycans impact prion protein aggregation and ultimately disease progression in vivo, here we employed 2 new knockin mouse models expressing cell surface PrP^C with 0 or 3 glycans and 5 complementary approaches. First, using 4 conformationally distinct prion strains, we show that unglycosylated PrP favors extracellular plaque formation and induces severe spongiform degeneration in the brain. Second, using an antibody specific for shed PrP (38), we show that the plaques are highly enriched in ADAM10-cleaved PrP. Third, using mass spectrometry and immunohistochemical labelling, we show that these unglycosylated, ADAM10-cleaved prions tightly bind heparan sulfate (HS) and HS colocalizes to plaques. Fourth, using heparin affinity chromatography, we directly show that unglycosylated

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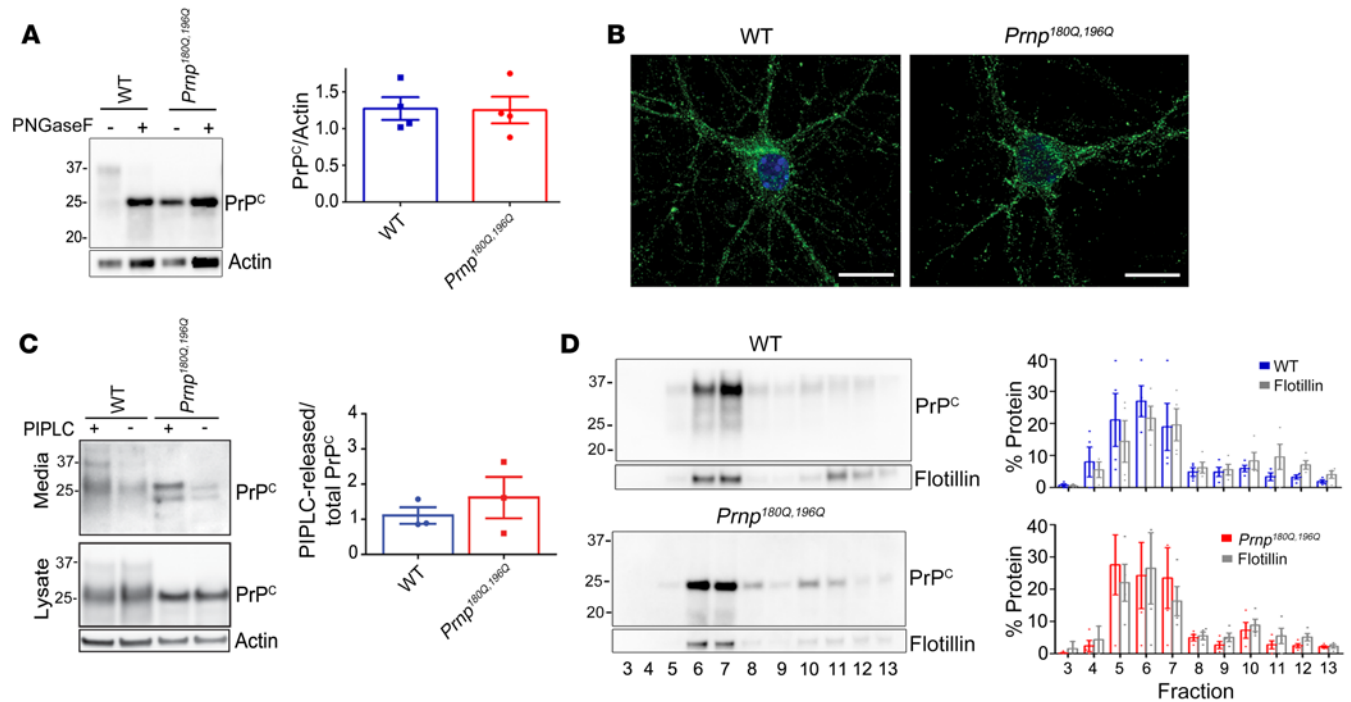


Figure 1. PrP^{180Q/196Q} traffics similarly to WT PrP^C in primary neurons and in mice. (A) Representative Western blot of PNGase-F–treated brain extracts from age-matched *Prnp*^{180Q/196Q} and WT mice reveal similar PrP^C expression levels (quantified in right panel) (100–250 day old mice); $n = 4$ /group. (B) PrP immunocytochemistry shows that unglycosylated PrP^{180Q/196Q} traffics to neuronal processes in primary cortical neurons, as does PrP in WT neurons; $n = 3$ experiments. Scale bars: 10 μm . (C) Representative Western blots of phospholipase C–cleaved (PIPLC–cleaved) PrP^{180Q/196Q} and WT PrP from the surface of cortical neurons show that surface PrP^C levels are similar (media); $n = 3$ experiments. The additional band in the media (~23 kDa) may be a cleaved form of PrP. (D) PrP^{180Q/196Q} and WT PrP^C, together with flotillin, localize to detergent-resistant membranes in the brain; $n = 3$ /group. Unpaired, 2-tailed Student's t test, no significant differences (A and C).

PrP has the highest affinity to heparin and that each additional glycan decreases the binding affinity. Finally, using knockin mice that express PrP having a third N-linked glycan, we show a complete reversal of a typical plaque-forming disease phenotype, as mice instead developed primarily subfibrillar aggregates that lack HS binding and showed a rapid disease progression. Collectively, these studies support the conclusion that PrP glycans can impede fibril formation, potentially through reduced binding of shed, glycosylated PrP to extracellular HS, and suggest that the PTMs on PrP contribute to the profoundly rapid clinical progression observed in the majority of prion diseases.

Results

Prnp^{180Q/196Q} mice generated using the CRISPR-Cas system show normal PrP^C expression and trafficking. Mice expressing unglycosylated PrP^C were generated using a single guide RNA to create 2 point mutations in the endogenous *Prnp* locus, corresponding to asparagine to glutamine substitutions at positions 180 and 196 (mouse PrP numbering) thereby altering the N-glycosylation sequons. Six founder lines were generated, 2 were sequenced, and 1 (line 191) was selected and bred to homozygosity on a C57BL/6 background. *Prnp*^{180Q/196Q} mice developed normally with no clinical signs or histologic lesions observed in the brain ($n = 12$ mice, 150–639 days of age) (Supplemental Figure 1A; supplemental material available online with this article; <https://doi.org/10.1172/JCI131564DS1>). PrP^C was expressed in the brain at levels similar to WT mice, and

the unglycosylated forms showed an equivalent electrophoretic mobility (Figure 1A). To assess the PrP^C distribution in neurons, cortical neurons were isolated from *Prnp*^{180Q/196Q} and WT mice, immunolabelled for PrP, and evaluated by confocal microscopy, revealing an indistinguishable PrP distribution throughout the neuronal cell body and neurites (Figure 1B).

PrP^C is GPI-anchored in the outer leaflet of the cell membrane, yet whether unglycosylated PrP traffics to the cell membrane has been controversial and may depend on the specific amino acids substituted into the N-linked glycan consensus sequence, Asn-X-Ser/Thr (39). Asparagine-to-glutamine substitutions were chosen here due to their structural similarity, differing only by a single methylene. To quantify surface expression in primary cortical neurons isolated from WT and *Prnp*^{180Q/196Q} mice, phosphatidylinositol-specific phospholipase C (PIPLC) hydrolysis was performed on live primary neurons to cleave the GPI-anchor. Similar levels of cleaved PrP were detected in the media, indicating that unglycosylated PrP^{180Q/196Q} localized to the cell surface (Figure 1C). To further confirm that PrP^{180Q/196Q} traffics to the cell surface, PrP-deficient RK13 cells were transfected with *Prnp*^{WT}, *Prnp*^{180Q/196Q}, or *Prnp*^{180A/196A}, a mutant reported to show impaired PrP trafficking (40). Again PrP^{180Q/196Q} was released into the media by PIPLC similar to WT PrP^C, whereas only approximately 40% of PrP^{180A/196A} was released, supporting that PrP with asparagine-to-glutamine substitutions traffics to the cell surface (Supplemental Figure 1B). To assess whether unglycosylated PrP localizes to lipid rafts in

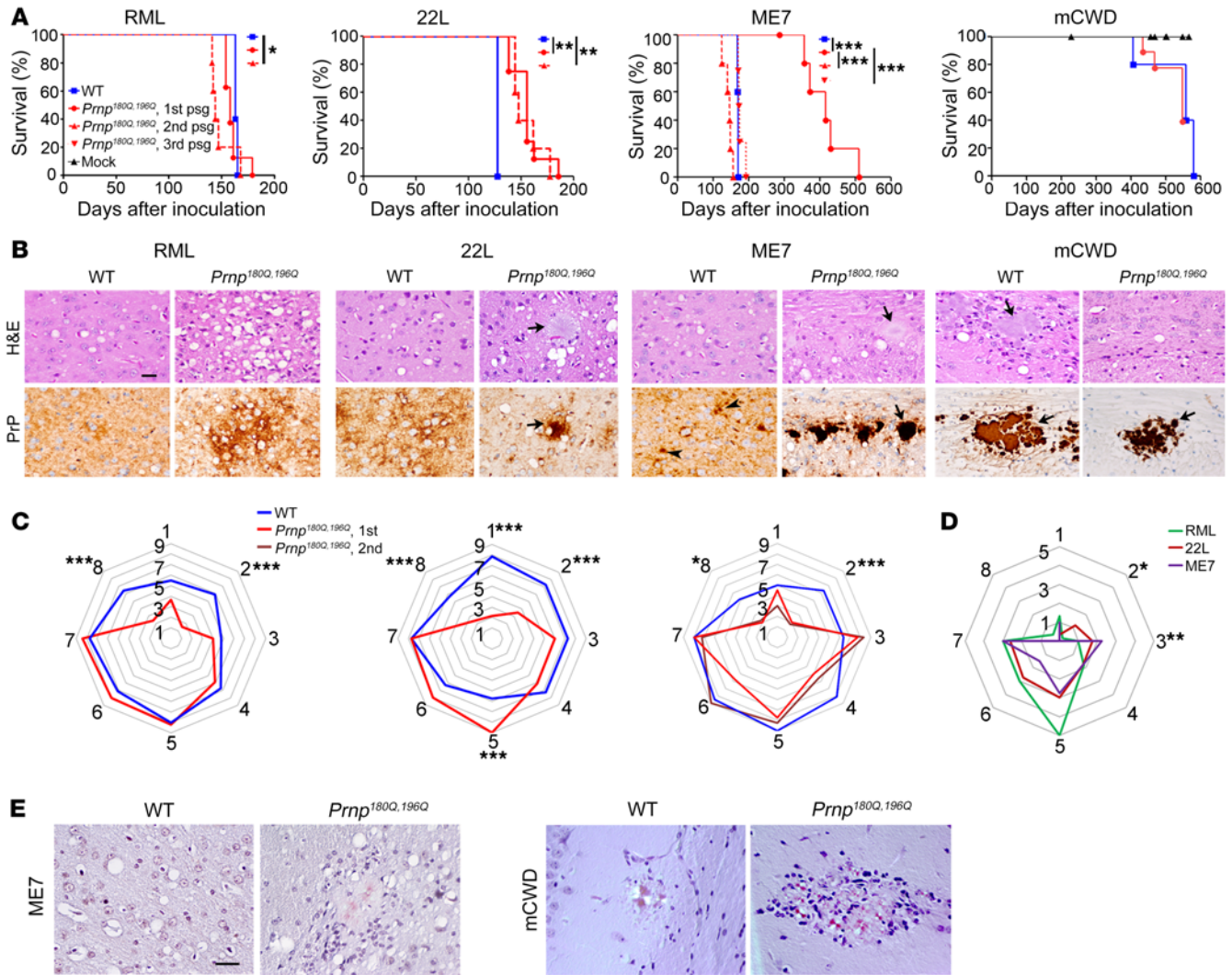


Figure 2. Survival and histopathologic lesions in prion-infected *Prnp*^{180Q/196Q} and WT mice. (A) For mCWD-infected mice, there was no difference in survival times. In contrast, 22L- and ME7-infected *Prnp*^{180Q/196Q} mice showed prolonged survival compared with WT mice. By the second and third passage, ME7-infected *Prnp*^{180Q/196Q} mice showed a 100% attack rate and similar survival times as the WT mice. (B) Brain sections stained with H&E or immunolabelled for PrP revealed severe spongiform degeneration and plaque-like deposits (RML) or plaques (22L, ME7, and mCWD, arrows) in prion-infected *Prnp*^{180Q/196Q} mice. Less spongiform degeneration was noted in the mCWD-infected mice. Scale bar: 50 μ m. (C) Lesion scores of spongiform change, gliosis, and PrP^{Sc} in 8 brain regions differed significantly in RML-, 22L-, and ME7-infected *Prnp*^{180Q/196Q} mice compared with WT mice. Cerebellum was consistently less severely affected in the *Prnp*^{180Q/196Q} mice. 1-dorsal medulla, 2-cerebellum, 3-hypothalamus, 4-medial thalamus, 5-hippocampus, 6-septum, 7-cerebral cortex, and 8-cerebral peduncle. (D) Right panel shows that PrP^{Sc} levels differed in the cerebellum (RML versus 22L; 0.2 ± 0.2 versus 1.2 ± 0.2 , respectively) and hypothalamus (22L versus ME7; 1.8 ± 0.5 versus 2.3 ± 0.7 , respectively) of the *Prnp*^{180Q/196Q} mice. (E) ME7 and mCWD plaques bind Congo red in *Prnp*^{180Q/196Q} brains. Scale bar: 50 μ m. * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$; 1-way ANOVA with Tukey's test (A), 2-way ANOVA with Bonferroni's post hoc test (C and D). RML: $n = 5-8$ mice per group; ME7: $n = 5-7$ mice per group; 22L: $n = 5-8$ mice per group, mCWD: $n = 5-9$ mice per group, mock: $n = 4-8$ mice per group.

vivo, detergent-resistant membranes from WT and *Prnp*^{180Q/196Q} brains were collected following equilibrium density gradient centrifugation, and revealed WT and unglycosylated PrP^C localized in lipid raft fractions together with flotillin (Figure 1D), further confirming surface localization of PrP^{180Q/196Q}. Thus by multiple measures, PrP^{180Q/196Q} traffics to the plasma membrane.

Aged Prnp^{180Q/196Q} mice show no evidence of spontaneous prion disease. To determine whether unglycosylated PrP^C spontaneously aggregates, we histologically and biochemically investigated 12 aged *Prnp*^{180Q/196Q} mice. Brain was immunolabelled for PrP and showed no evidence of prion aggregation (Supplemental Figure 1A). To test for the presence of proteinase K-resistant (PK-resistant) PrP,

WT and PrP^{180Q/196Q} brain extracts were treated with low PK concentrations, which revealed no differences in PrP^C digestion (Supplemental Figure 1C). Last, to test for insoluble PrP, brain homogenates were ultracentrifuged, which revealed that PrP^{180Q/196Q} was highly soluble, similar to WT PrP^C (approximately 92% and 95% soluble, respectively), and differed from a prion-infected brain (approximately 22% soluble) (Supplemental Figure 1D). Thus, *Prnp*^{180Q/196Q} mice show no evidence of aggregated PrP, consistent with their normal lifespan and lack of neurologic disease.

Prnp^{180Q/196Q} mice infected with 4 prion strains develop plaque and plaque-like deposits. We next assessed the susceptibility of the *Prnp*^{180Q/196Q} mice to prion infection. *Prnp*^{180Q/196Q} and WT mice

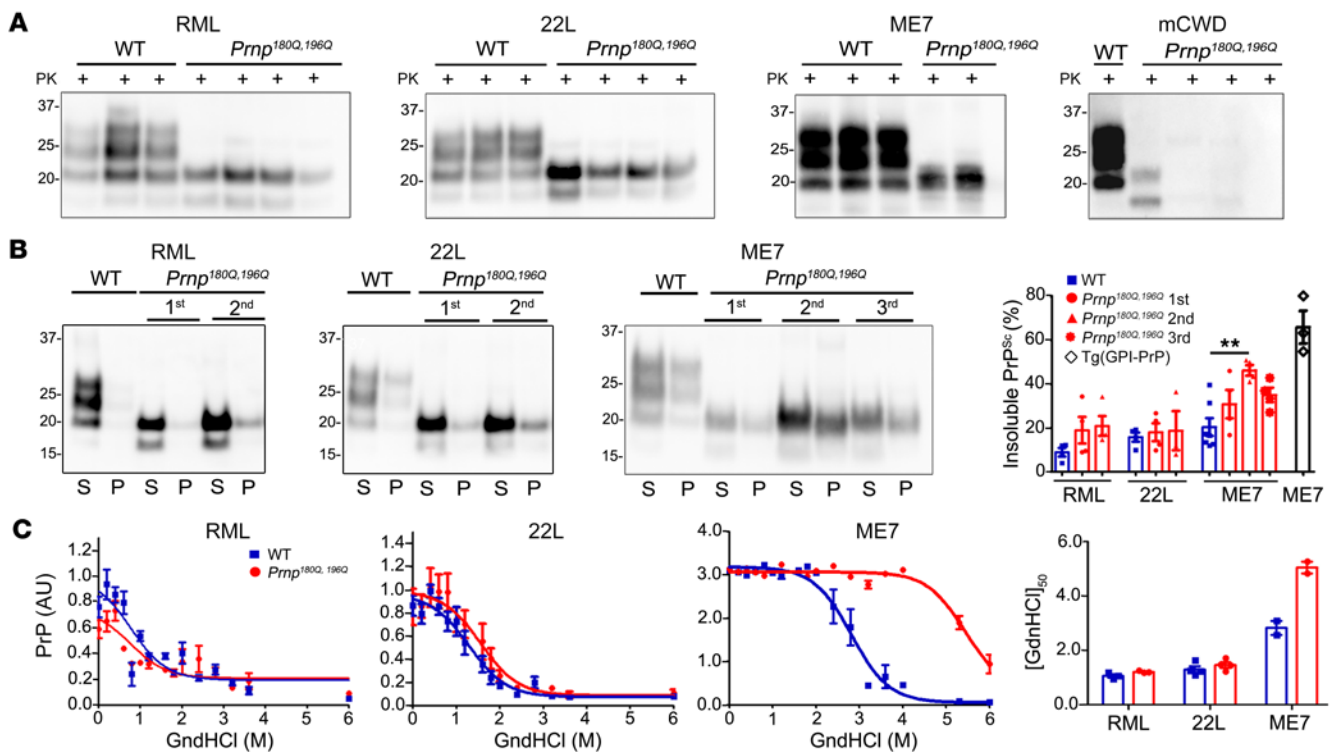


Figure 3. Comparison of PK resistance, solubility, and stability of RML, 22L, ME7, and mCWD prions in *Prnp*^{180Q/196Q} and WT brain samples. (A) Western blots of brain homogenates reveal PK-resistant unglycosylated PrP^{Sc} in RML-, 22L-, ME7-, and mCWD-infected *Prnp*^{180Q/196Q} mice. **(B)** Western blots of soluble (S, supernatant) and insoluble (P, pellet) PrP^{Sc} show similar levels of insoluble PrP^{Sc} in *Prnp*^{180Q/196Q} and WT brains on first passage, but significantly higher levels in ME7-infected *Prnp*^{180Q/196Q} brains on further passage. The GPI-ME7 was included to compare samples with a known highly fibrillar prion; $n = 2$ (ME7, first passage only) and $n = 3$ – 7 /strain (all other strains, including ME7, second and third passage). **(C)** Representative PrP^{Sc} stability curves from single mouse brain samples run in triplicate. The right graph shows the guanidine hydrochloride concentration at which half the PrP^{Sc} remains ($[\text{GdnHCl}]_{50}$) and reveals no significant differences in the stability of RML or 22L, but a significantly more stable ME7 prion after passage in *Prnp*^{180Q/196Q} mice. Plotted (right graph) are the mean and SEM for $n = 2$ (ME7-infected *Prnp*^{180Q/196Q}), $n = 3$ (WT), or $n = 4$ (RML- and 22L-infected *Prnp*^{180Q/196Q}) mice per strain, each run in triplicate. $**P \leq 0.01$, 1-way ANOVA with Tukey's test (**B**).

were inoculated intracerebrally with 4 mouse-adapted prion strains known as RML, 22L, ME7, and mCWD. RML, 22L, and ME7 are subfibrillar strains derived from sheep scrapie and show no fibrils ultrastructurally in situ (41–43), whereas mCWD is a mouse-adapted fibrillar strain derived from deer chronic wasting disease that is characterized by dense plaques composed of bundles of long, extracellular fibrils (44). All RML- and 22L-inoculated *Prnp*^{180Q/196Q} mice developed prion disease. Following inoculation with RML prions, WT and *Prnp*^{180Q/196Q} mice showed no significant difference in survival times on first passage (WT: 164 ± 1 days after inoculation [dpi]; *Prnp*^{180Q/196Q}: 160 ± 3 dpi [mean \pm SEM]), and a modest decrease in survival on second passage in *Prnp*^{180Q/196Q} mice (148 ± 5 dpi) (Figure 2A). Following inoculation with 22L prions, *Prnp*^{180Q/196Q} mice showed a prolonged survival (WT: 127 ± 0 dpi; *Prnp*^{180Q/196Q}: 155 ± 5 dpi), which was unchanged on second passage in *Prnp*^{180Q/196Q} mice (155 ± 6 dpi) (Figure 2A). In contrast, there was a significant transmission barrier following inoculation with ME7 prions, as there was an incomplete attack rate (6/7 mice, 86%) and a highly variable and significantly prolonged incubation period (WT: 169 ± 1 dpi; *Prnp*^{180Q/196Q}: 389 ± 27 dpi). Yet by the second passage, the attack rate was 100% and the incubation period decreased by 60% (142 ± 5 dpi), and by third passage the incubation period was similar to ME7 in WT mice (177 ± 5 dpi), suggestive

of strain adaptation. Similarly, the fibrillar mCWD prions led to an incomplete attack rate on first passage (5/9, 56%), suggestive of a transmission barrier, although the incubation period in WT mice was longer (WT: 569 ± 7 dpi; *Prnp*^{180Q/196Q}: 508 ± 24 dpi) (Figure 2A). Notably, most *Prnp*^{180Q/196Q} mice positive for mCWD prions (4 of 5 positive mice) had low PrP^{Sc} levels in brain and were euthanized due to age ($n = 3$) or concurrent disease ($n = 1$), thus were not likely at terminal disease stages.

A comparison of the histopathologic lesions in RML-, 22L-, and ME7-infected WT and *Prnp*^{180Q/196Q} mice revealed intriguing differences in spongiform degeneration, gliosis, and PrP^{Sc} distribution (Figure 2, B and C). First, the spongiform degeneration was consistently scored as more severe for all 3 strains in the *Prnp*^{180Q/196Q} mice as compared with the WT mice (Figure 2B). Additionally, in the *Prnp*^{180Q/196Q} mice infected with ME7 prions, the hippocampus was consistently markedly atrophied due to severe neuronal loss, with complete loss of the CA1 pyramidal neurons and numerous gemistocytic astrocytes, which persisted upon second passage, whereas WT mice showed a moderate loss of hippocampal neurons (Supplemental Figure 2A). Finally, the cerebellum lacked PrP^{Sc} in all infected *Prnp*^{180Q/196Q} mice, a striking difference from WT mice in which all 3 strains were present in the cerebellum (Figure 2, C and D, region 2). This was not due to a lack

of PrP^C expression, as PrP^{180Q/196Q} was expressed in the cerebellum at levels similar to WT PrP (Supplemental Figure 2B).

PrP^{Sc} morphology also differed from WT mice in that most Prnp^{180Q/196Q} brains showed some plaque-like deposits or dense parenchymal plaques. In WT mice, RML and 22L prions typically form diffuse aggregates, ME7 prions form diffuse and small plaque-like aggregates (15–30 μm), and mCWD prions form large plaques (>100 μm) (Figure 2B). In the Prnp^{180Q/196Q} mouse brain, the RML prion deposits remained mostly diffuse and occasionally formed plaque-like clusters (Figure 2B). In contrast, 22L and ME7 prions formed both diffuse and significantly larger plaques (up to 100 μm for ME7) that were congophilic (Figure 2, B and E; Supplemental Figure 2, C and D). There were also numerous florid ME7 plaques that were surrounded by small vacuoles, similar to those described in variant CJD-infected human brains (45) (Supplemental Figure 2E). Notably, large mCWD plaques were morphologically identical in the WT and Prnp^{180Q/196Q} mice (Figure 2B). Thus the Prnp^{180Q/196Q} mice challenged with the 3 subfibrillar prion strains shared certain newly acquired disease features, including an increase in plaques and plaque-like structures, more severe cortical spongiosis, and a notable lack of prions in the cerebellum.

PrP^{Sc} biochemical properties were altered in Prnp^{180Q/196Q} mice. To assess the differences in the biochemical properties of the unglycosylated prions, we first assessed the presence of PK-resistant PrP^{Sc} in all Prnp^{180Q/196Q} mice. All RML- and 22L-challenged and some ME7- and mCWD-challenged mice showed PK-resistant PrP^{Sc} (Figure 3A). The solubility and stability of RML, 22L, and ME7 PrP^{Sc} were assessed (mCWD prion levels were too low to assess). Notably, the insoluble PrP levels were similar for RML- and 22L-infected WT and Prnp^{180Q/196Q} mice, yet for ME7, we found that further passage in the Prnp^{180Q/196Q} mice was associated with significantly increased levels of insoluble PrP^{Sc} (Figure 3B, second passage).

To next measure the aggregate stability, aliquots of brain homogenate were first denatured with guanidine hydrochloride (GndHCl) ranging from 0 to 6 M, and then diluted and digested with PK. The PrP was measured by ELISA to determine the [GndHCl]_{1/2} concentration at which half the PrP^{Sc} remained. While there were no stability differences for RML and 22L prions in the WT and Prnp^{180Q/196Q} brains, ME7 prions were more stable in the Prnp^{180Q/196Q} brain (Figure 3C). Therefore, the aggregate stability of the 3 unglycosylated prions correlated with plaque formation, with ME7 showing the highest stability.

To further probe the conformation of unglycosylated PrP^{Sc}, we next applied a conformationally sensitive amyloid probe, heptameric formic thiophene acetic acid (h-FTAA), to brain cryosections and performed fluorescence lifetime imaging (FLIM) to measure the decay of emitted light from prion-bound h-FTAA. The decay of light depends on the immediate chemical environment of the h-FTAA, and thus is useful to distinguish differential binding modes of the probe to distinct aggregates. Variations in the decay times have previously been observed for h-FTAA bound to different aggregated morphotypes of amyloid-β (46), as well as prion deposits associated with distinct prion strains (47). Here we found that for every strain compared, the FLIM of prion-bound h-FTAA in the Prnp^{180Q/196Q} mice differed from WT mice. Whereas h-FTAA does not bind RML in WT mice, h-FTAA binds RML in the Prnp^{180Q/196Q} mice (Figure 4). For 22L and ME7 prions, the life-

time distribution was shifted in the Prnp^{180Q/196Q} mice as compared with the WT mice (Figure 4). Since theoretical calculations and solid-state NMR experiments have verified that the binding mode and the optical properties of the oligothiophene are determined by regularly spaced surface charge patterns and highly accessible grooves on the fibril surface (48–50), the observed differences in life time distribution are most likely due to PrP structural variations in Prnp^{180Q/196Q} mice.

ME7 and mCWD prions in Prnp^{180Q/196Q} mice largely consist of ADAM10-cleaved PrP. Unglycosylated PrP showed an increased propensity for fibril formation in a strain-dependent manner. How does the lack of PrP glycans promote fibril and plaque formation? Previously we found that extracellular prion plaques (mCWD strain) were largely, if not entirely, composed of ADAM10-cleaved, GPI-anchorless PrP (Figure 5A) (51), which comprises 10%–15% of PrP^C in the brain (52, 53).

The plaques and plaque-like structures prompted us to investigate the level of ADAM10-cleaved PrP^{Sc} in Prnp^{180Q/196Q} mice. Using antibodies targeting ADAM10-cleaved PrP (sPrP^{G228}) (38) or total PrP (POM1) (54), we found that ADAM10-cleaved PrP levels were very low for RML, yet substantially higher for 22L and ME7, showing a 4-fold and an 18-fold increase as compared with WT mice (Figure 5A). The increase in ADAM10-cleaved PrP^{Sc} was not simply due to an increase in cleaved PrP^C in the Prnp^{180Q/196Q} mice, as the levels were similar to those in the WT mice (Figure 5B).

To next localize the ADAM10-cleaved PrP in the 22L- and ME7-infected brain, we used the sPrP^{G228} and SAF84 antibodies specifically against ADAM10-cleaved PrP and the PrP epitope at residues 166–172 (55, 56), respectively. Remarkably, the ADAM10-cleaved PrP localized to plaques, but not diffuse deposits (Figure 5C and Supplemental Figure 3A). Thus, the relative levels of ADAM10-cleaved PrP^{Sc} in the Prnp^{180Q/196Q} mice correlated with those in WT mice, with ME7 exceeding 22L and RML, yet the absolute levels of cleaved PrP^{Sc} were higher in the Prnp^{180Q/196Q} mice and were localized to extracellular plaques in situ. For mCWD, the prion plaques were composed of ADAM10-cleaved PrP in both the WT and Prnp^{180Q/196Q} mice (Figure 5C).

PrP glycans decrease the heparin-binding affinity. Why does ADAM10-cleaved, unglycosylated PrP have a higher tendency to form fibrils than glycosylated PrP? We previously found high levels of HS bound to ADAM10-cleaved PrP^{Sc} having 0 to 1 glycan (51). We reasoned that unglycosylated PrP may bind with higher affinity to extracellular HS due to a lack in electrostatic repulsion between the anionic glycans and HS. To test this hypothesis, we assessed how PrP glycans impact the binding affinity to heparin, a highly sulfated surrogate of HS, using affinity chromatography and PrP with 0, 1, 2, or 3 glycans from transfected RK13 cells. PrP^C was cleaved from the cell surface by PIPLC hydrolysis and applied to the heparin sepharose column. While PrP^C with 2 to 3 glycans showed a low affinity to heparin, unglycosylated PrP^C showed a high affinity to heparin; the affinity progressively decreased with each additional glycan (Figure 6A). Notably, even within the glycoform mixtures of WT PrP^C (0–2 glycans), the unglycosylated PrP^C showed the highest affinity to heparin (Figure 6B).

To determine whether brain-derived unglycosylated PrP^C also shows a higher affinity to heparin, we next tested the heparin-binding affinity of PrP^C from age-matched WT and Prnp^{180Q/196Q}

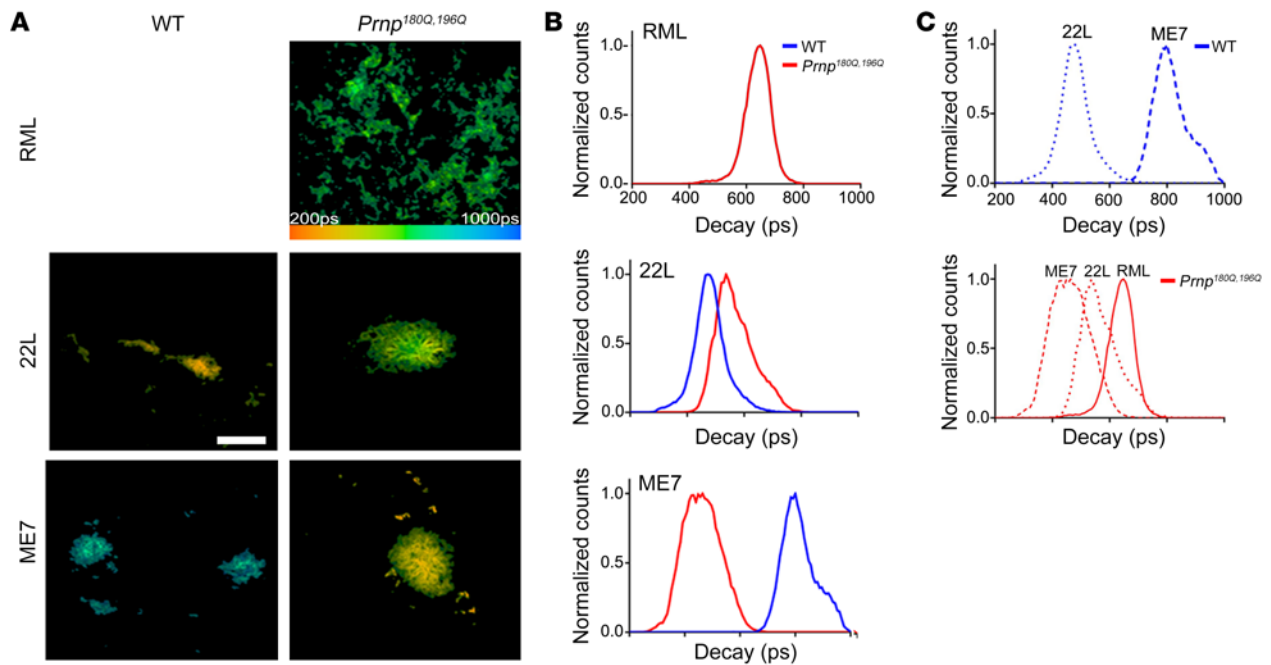


Figure 4. Fluorescence lifetime (FLIM) decay of h-FTAA bound to prion aggregates. (A) Fluorescence lifetime images and (B and C) intensity-weighted mean lifetime (t_i) distributions of h-FTAA-stained RML, 22L, and ME7 prion plaques in *Prnp*^{180Q/196Q} (red) or WT (blue) brain show that the FLIM decay curves in *Prnp*^{180Q/196Q} brain differ from those in WT brain and reveal 3 different aggregate conformers in prion-infected *Prnp*^{180Q/196Q} mice. The decay data were collected with the excitation wavelength set at 490 nm. The color bar represents lifetimes from 200 ps (orange) to 1000 ps (blue) and the images are color coded according to the representative lifetime. Decay curves were collected from 5–10 prion deposits from a minimum of $n = 3$ mice/strain. Scale bar: 20 μ m.

mouse brain homogenates (Figure 6C). Again we found that unglycosylated PrP^C had a higher heparin-binding affinity than glycosylated WT PrP^C (Figure 6, C and D), which also held true for their ADAM10-cleaved counterparts. Interestingly, for both genotypes, ADAM10-cleaved PrP showed a higher heparin-binding affinity than full-length PrP (Figure 6E), suggesting that the GPI anchor with the 3 C-terminal amino acids markedly reduces heparin binding. Collectively, these data suggest that N-linked glycans and the GPI-anchor hinder PrP^C binding to HS.

Unglycosylated, ADAM10-cleaved PrP binds HS in vivo. Sulfated glycosaminoglycans (GAGs) promote fibril assembly in vitro (57, 58) and the sulfated GAG, HS, codeposits with amyloid beta and prion plaques in vivo (59, 60). However, whether HS preferentially binds to unglycosylated prions is unknown. To determine whether the 22L, ME7, and mCWD plaques harbor HS in the *Prnp*^{180Q/196Q} mice, we immunolabelled prion-infected brain sections for HS using the 10E4 antibody, which recognizes an N-sulfated glucosamine residue and does not react with hyaluronan, chondroitin sulfate, or dermatan sulfate. We found that HS clearly localized to prion plaques in *Prnp*^{180Q/196Q} mice, as compared with only a low level of diffuse HS stain in RML-, 22L-, and ME7-infected WT mice (Figure 6F, Supplemental Figure 3B) and RML-infected *Prnp*^{180Q/196Q} mice (Supplemental Figure 3B). Pretreating brain sections with heparin lyases abolished the HS immunolabelling (Supplemental Figure 3C), supporting specificity of the antibody binding.

Mass spectrometry was next used to quantify HS bound to ME7 prions from *Prnp*^{180Q/196Q} and WT brain. ME7 was selected due to the abundance of plaques in the *Prnp*^{180Q/196Q} mice. PrP^{Sc} was purified by solubilization and multiple rounds of ultracentrifuga-

tion (61), levels were quantified against a recombinant PrP standard, and the sample was denatured with sodium hydroxide. HS chains were then depolymerized with a cocktail of heparin lyases, the resulting disaccharides were [¹²C₆] aniline-tagged, and disaccharides were identified by liquid chromatography-mass spectrometry (LC-MS) using [¹³C₆] aniline-tagged disaccharide standards (62, 63). The HS relative to PrP level was calculated for each sample (HS/PrP ratio [μ g/ μ g]). Consistent with the HS immunostaining results, we found significantly higher levels of HS bound to unglycosylated PrP^{Sc} compared with WT PrP^{Sc} (Figure 6G). In addition, the composition of HS bound to ME7 in the WT versus *Prnp*^{180Q/196Q} brain differed, as HS was generally less sulfated in the *Prnp*^{180Q/196Q} brain, with lower levels of N-sulfated and 6-O sulfated HS (Figure 6H). Interestingly, the HS composition in the whole brain lysates from prion-infected WT and *Prnp*^{180Q/196Q} mice was similar. This finding suggests that the observed composition differences in PrP^{Sc}-bound HS were not due to overall differences in HS biosynthesis, but were instead due to PrP^{Sc} selectively binding to specific HS molecules (Figure 6I).

A third glycan on PrP reverses the prion disease phenotype. We reasoned that since unglycosylated, ADAM10-cleaved PrP binds HS and favors plaque formation, the addition of a third glycan may correspondingly decrease plaque formation. To directly test this hypothesis, we developed a new knockin mouse model having a third glycan at position 187 of PrP through a single amino acid mutation (T187N) (mouse numbering). We confirmed the addition of the third glycan and characterized the PrP expression and cell membrane localization in *Prnp*^{187N} mice (Supplemental Figure 4, A and B), which did not show clinical signs or spontaneous dis-

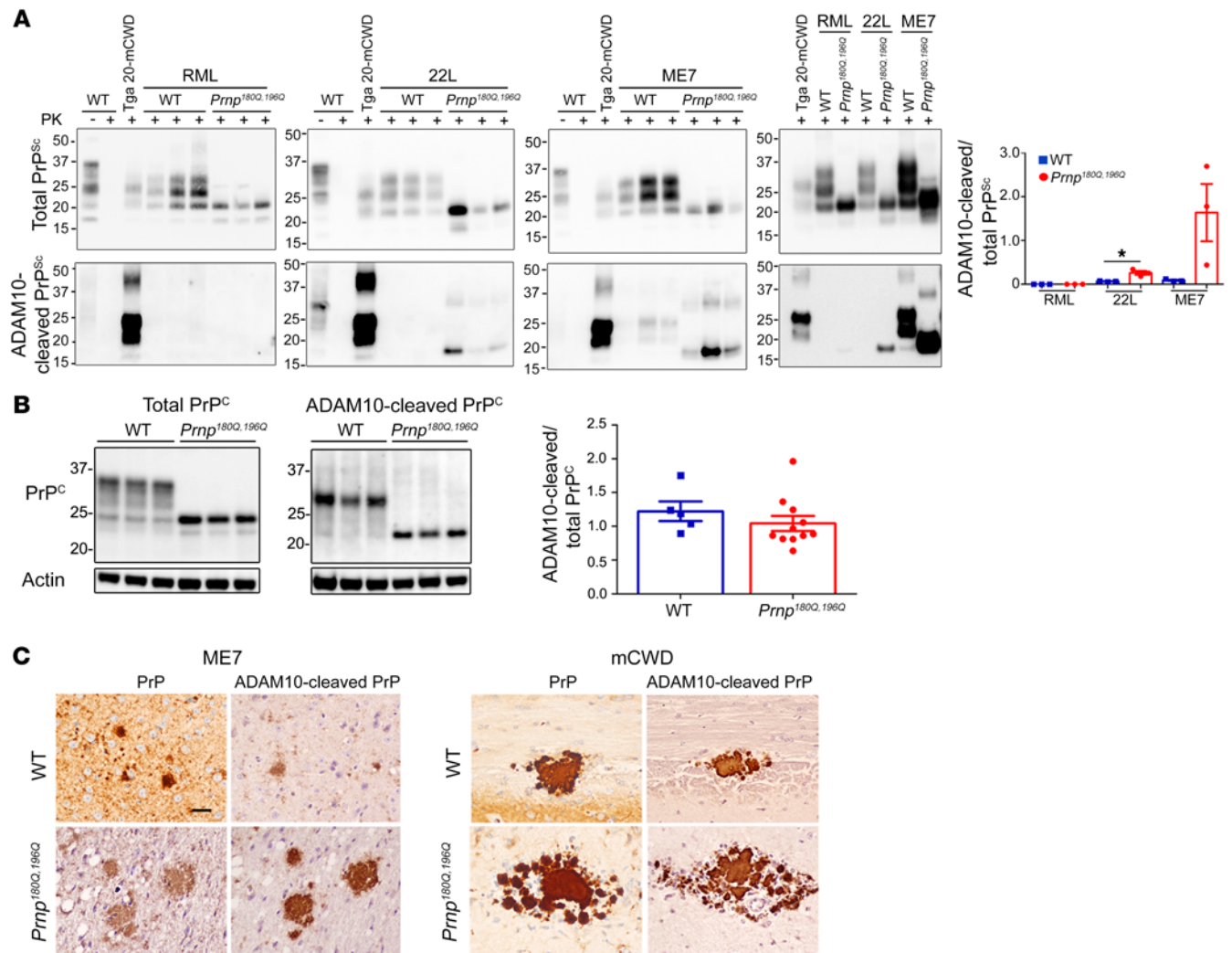


Figure 5. Increased ADAM10-cleaved PrP^{Sc} in prion-infected *Prnp*^{180Q/196Q} mice. (A) Western blots reveal 22L- and ME7-infected *Prnp*^{180Q/196Q} mice harbor higher levels of ADAM10-cleaved PrP^{Sc} in the brain compared with WT mice; $n = 3$ /group. Note that the PK-digested ADAM10-cleaved PrP runs at a lower molecular weight due to lack of the GPI-anchor and terminal 3 amino acids. * $P < 0.05$, unpaired, 2-tailed Student's t test. (B) Western blots show that uninfected *Prnp*^{180Q/196Q} and WT mice have similar levels of ADAM10-cleaved/total PrP^C in the brain; $n = 5$ WT and 11 *Prnp*^{180Q/196Q} mice. (C) Brain sections immunolabeled for PrP with SAF84 (labelled PrP) or with sPrP^{G228} antibody (labelled ADAM10-cleaved PrP) reveal ADAM10-cleaved PrP localizes to plaque-like and plaque deposits in ME7-infected *Prnp*^{180Q/196Q}, mCWD-infected *Prnp*^{180Q/196Q}, and WT brain sections. Note that the diffuse aggregates are not labelled by the sPrP^{G228} antibody. Cortex (ME7-infected WT), thalamus (ME7-infected *Prnp*^{180Q/196Q}), hippocampus (mCWD-infected *Prnp*^{180Q/196Q}), and corpus callosum (WT-infected mCWD) are shown. Scale bar: 50 μm.

ease development. We next challenged mice with a strain known to form long fibrils, mCWD (44). Strikingly, despite an amino acid substitution expected to lead to a transmission barrier and prolong survival, the *Prnp*^{187N} mice developed a profoundly accelerated disease (326 ± 9 dpi), which was approximately 60% of the incubation period of mCWD in WT mice (Figure 7A). Remarkably, on second passage, the incubation period decreased even further to 194 ± 7 dpi, approximately 35% of the WT incubation period (Figure 7A). This disease acceleration was likely due to the additional glycans and not to a difference in the amino acid side chain, as PrP^C-PrP^{Sc} sequence differences typically prolong survival.

To investigate whether the *Prnp*^{187N} mice harbored the large plaques characteristic of mCWD in WT mice, we assessed the histopathological phenotype. We found a notable lack of large plaques

in the *Prnp*^{187N} mice, and instead only diffuse aggregates and rare small plaques in the corpus callosum (Figure 7B). In addition to the rapidly progressive disease and small aggregates, the aggregates no longer bound HS (Figure 7C). Moreover, the prion aggregates were no longer composed of ADAM10-cleaved PrP (Figure 7D), were significantly more soluble (Figure 7E), and showed a similar stability in chaotopes (Figure 7F).

We previously found that mCWD prions do not spread from the tongue into the CNS, thus were nonneuroinvasive, whereas the subfibrillar strains were neuroinvasive (64). To test for neuroinvasion of the new subfibrillar mCWD prions in the *Prnp*^{187N} mice, we inoculated these prions into the tongue. Notably, all *Prnp*^{187N} mice developed prion disease ($5/5$ mice, 312 ± 10 dpi), indicating that the prions had switched from a nonneuroinvasive to a highly

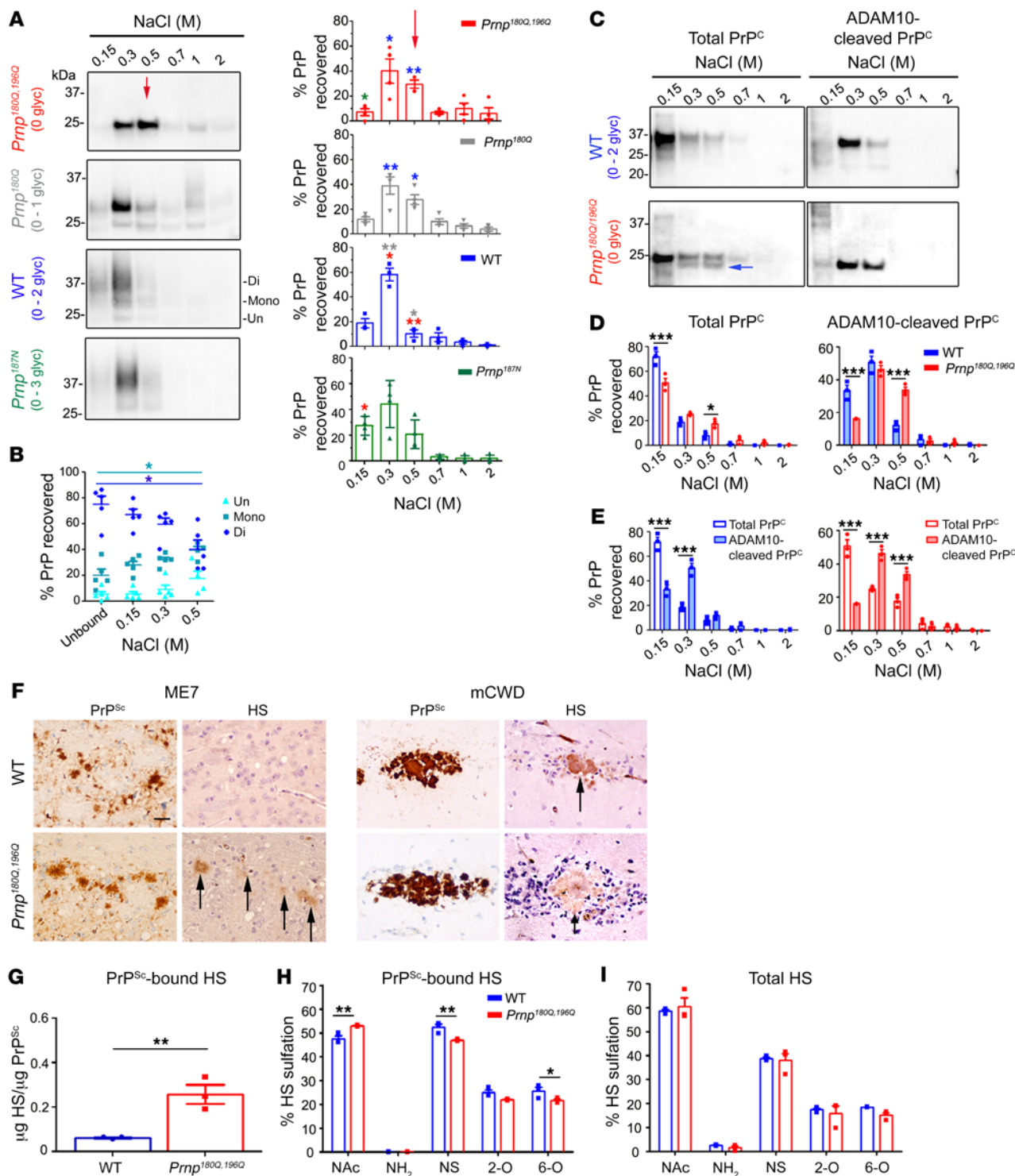


Figure 6. PrP glycans hinder binding to HS. (A) Immunoblots of heparin affinity chromatography experiments assessing variably glycosylated PrP. Relative levels of PrP from each elution are shown in the graphs. Differences in the binding affinity are most notable in the 0.5 M NaCl elution (red arrow). Asterisk color indicates the mutants with significant differences. The triglycosylated PrP isoform level was low in RK13 cells; $n = 3-4$ experiments. (B) Among the WT PrP glycoforms, diglycosylated PrP has a lower heparin affinity than mono- or unglycosylated PrP (unbound is PrP in the flow-through); $n = 5$ experiments, 4 also included in A. (C) Affinity chromatography of the soluble brain fraction reveals that total and ADAM10-cleaved PrP^{180Q/196Q} have significantly higher heparin affinity than the corresponding WT PrP. PrP^{180Q/196Q} shows a second band (blue arrow) that corresponds to ADAM10-cleaved PrP^C. (D) Quantification of PrP shown in C; $n = 3$ /strain. (E) ADAM10-cleaved PrP has a higher heparin-binding affinity than total PrP for both WT PrP and PrP^{180Q/196Q}. (F) Immunolabelling reveals HS colocalizes to ME7 plaques in the Prnp^{180Q/196Q} brain only and to mCWD plaques in both the WT and Prnp^{180Q/196Q} brain; $n = 4$ /strain. Scale bar: 50 μ m. (G) LC-MS reveals approximately 6-fold more HS bound to unglycosylated ME7 PrP^{Sc} than to highly glycosylated ME7 PrP^{Sc} (WT); $n = 3$ /strain. (H) Composition analysis of HS bound to purified PrP^{Sc} (ME7) reveals less N-sulfated (NS) and 6-O sulfated (6-O) HS bound to unglycosylated (Prnp^{180Q/196Q}) as compared with glycosylated (WT) PrP^{Sc}; $n = 3$ /group. (I) The overall HS composition in ME7-infected Prnp^{180Q/196Q} and WT whole-brain lysates are similar; $n = 3$ /group. * $P \leq 0.05$, ** $P \leq 0.01$, and *** $P \leq 0.001$; 2-way ANOVA with Bonferroni's post hoc test (A, D, E, and H). * $P < 0.05$; 1-way ANOVA with Tukey's test (B). ** $P \leq 0.01$, unpaired, 2-tailed Student's t test (G).

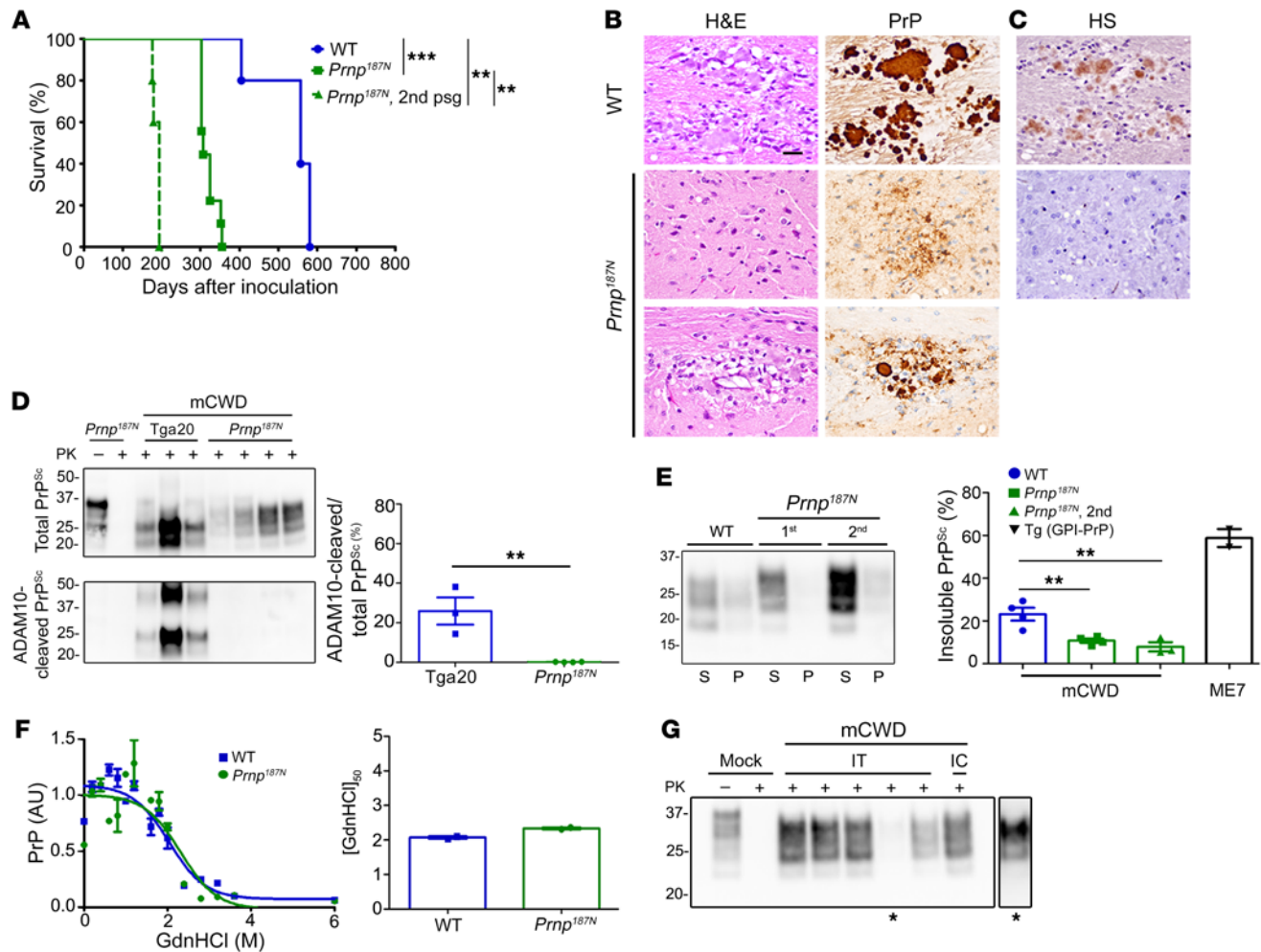


Figure 7. Knockin mice expressing triglycosylated PrP show a reversal in the clinical, histologic, and biochemical phenotype when infected with the plaque-forming prion strain, mCWD. (A) Survival curves of *Prnp*^{187N} mice inoculated with mCWD prions show a profound decrease in the survival time as compared with WT mice; *n* = 5–9/group. (B) The spongiform degeneration is similar in *Prnp*^{187N} mice as compared with WT mice; however, PrP deposits switched from the typical large, dense plaques in WT mice to primarily diffuse aggregates, with occasional rare small plaques in the corpus callosum (lower panel). (C) HS immunolabelling of the diffuse aggregates or plaque-like structures was not evident in the *Prnp*^{187N} brain. (D) Western blot reveals that the PrP^{Sc} in the *Prnp*^{187N} brain is no longer composed of ADAM10-cleaved PrP^{Sc} that predominated in the original mCWD strain in tga20 mice (overexpressed WT mouse PrP); *n* = 3–5/mouse strain. (E) Western blot of soluble versus insoluble PK-resistant PrP^{Sc} reveals a decrease in insoluble PrP^{Sc} in the *Prnp*^{187N} mice; *n* = 4–7 WT or *Prnp*^{187N} mice and *n* = 2 Tg (GPI-PrP) positive control brain samples. (F) Representative PrP^{Sc} stability assay and quantification show that the resistance to unfolding in chaotropes is similar for PrP^{Sc} aggregates in the mCWD-infected *Prnp*^{187N} and WT brain; *n* = 2/group. (G) Western blot of *Prnp*^{187N} mice inoculated intratongue (IT) or intracerebrally (IC) with mCWD-passaged *Prnp*^{187N} brain homogenate shows that all clinically terminal mice (5/5 mice) harbored PrP^{Sc} in the brain. *A longer exposure of one sample is shown on the right. ***P* ≤ 0.01, ****P* ≤ 0.001; 1-way ANOVA with Tukey's test (A and E), unpaired, 2-tailed Student's *t* test (D). Scale bar: 100 μm (B and C).

neuroinvasive prion (Figure 7G). Therefore, by multiple histological and biochemical measures, the mCWD prion had undergone a profound transformation to a GPI-anchored, non-HS binding, highly soluble, neuroinvasive prion causing a rapidly progressive disease. Collectively, these data support the conclusion that PrP glycans are linked to a decreased tendency for plaque formation.

Discussion

In contrast to Alzheimer's disease, amyloid plaques in the brain are rare in prion disease and are often associated with longer disease durations, as observed in Gerstmann-Sträussler-Scheinker disease, for example (3). Prions more commonly form diffuse, synaptic, or punctate aggregates associated with an extraordi-

narily rapid disease progression, and the molecular determinants that underlie these disease differences are unclear. Our work here indicates that the PrP glycans and GPI-anchor, together with HS, orchestrate major components of the disease phenotype in prion disease. The absence of glycans was linked to the formation of parenchymal plaques, particularly for prions lacking a GPI-anchor. Therefore, our results support a model in which the PrP posttranslational state markedly influences the fate of a prion aggregate, with cleaved, unglycosylated prions efficiently binding to extracellular HS to form parenchymal plaques.

The molecular mechanisms that underlie extracellular fibril formation in prion disease have not been previously identified and tested. Our approach to determining these mechanisms was

prompted by studies of knockin mice expressing unglycosylated PrP, known as G3 mice, which revealed that glycosylation is not essential for prion conversion or transmission of infectivity (21, 24). G3 mice also showed a tendency to form thioflavin-positive plaques, suggestive of fibrillar prions (24). Similar to the G3 mice, transgenic mice expressing GPI-anchorless PrP^C also develop fibrillar, thioflavin-positive prion aggregates, but fibrils accumulate perivascularly (19). Our use of prion-infected knockin mice expressing unglycosylated and triglycosylated PrP, together with WT mice, also reveal a correlation between a lack of PTMs and fibrillar prion formation. Using recently developed antibodies that recognize ADAM10-cleaved PrP (38), we now identify ADAM10-cleaved PrP as a major component of the unglycosylated PrP^{Sc} in plaques. Thus, prions with the capacity to replicate ADAM10-cleaved PrP^C, such as ME7, preferentially recruit unglycosylated, cleaved PrP^C and form large plaques. In contrast, largely GPI-anchored prions, such as RML, recruit unglycosylated, GPI-anchored PrP^C and show a minimally altered disease phenotype. Collectively, these findings suggest that the presence or absence of glycans is relevant for prion conformers that can convert ADAM10-cleaved PrP^C, in which case the unglycosylated PrP^C is recruited by cleaved, GPI-anchorless PrP^{Sc}.

Glycans stabilize the prion protein structure (65, 66) and impede prion conversion and fibril formation in vitro (66–68), potentially through modifying or blocking PrP^C/PrP^{Sc} recognition domains (69). The diverse glycans on PrP are bi-, tri-, and tetra-antennary and are highly sialated (14, 17, 70, 71), with high sialation levels negatively impacting the prion replication rate (27). In the GPI-anchored prions RML and 22L, we found that the glycosylation state did not seem to markedly affect prion conversion, as the incubation period was only modestly affected. Additionally, for RML prions, the aggregate morphology, aggregate stability, and solubility were remarkably similar in the unglycosylated state. For 22L, however, the prion aggregate morphology was subtly altered, as PrP^{Sc} recruited ADAM10-cleaved PrP^C and formed some plaques. Consistent with a morphological change, alterations in the h-FTAA binding environment also suggested differences in the structure of unglycosylated PrP^{Sc}, in agreement with reports that suggest the glycans may affect the tertiary and quaternary structure, but do not significantly affect the secondary structure (72). Thus, in contrast with in vitro reports (67, 68), our data suggest that the glycans do not impede conversion in vivo, as switching from glycosylated to unglycosylated PrP^C did not accelerate prion conversion. Moreover, triglycosylated PrP was efficiently converted and led to a rapidly progressive prion disease.

In contrast to RML, ME7 prions were markedly altered by the glycosylation state, as unglycosylated ME7 PrP^{Sc} formed large extracellular plaques, recruited ADAM10-cleaved PrP^C, was more stable in chaotropes, had more insoluble PrP^{Sc}, and showed an altered h-FTAA decay profile, suggestive of an altered structure due to either adaptation or a switch in the dominant PrP^{Sc} structure. Why did unglycosylated ME7 prion aggregates assemble as highly stable, large parenchymal plaques? HS serves as an extracellular scaffold (73–75) and is widely recognized to bind prion aggregates, both in vitro and in vivo (76–79). In accordance with these observations, we found that HS localizes to parenchymal plaques, but not to diffuse aggregates within the same brain. Addi-

tionally, mass spectrometry quantification revealed 4 times more HS bound to unglycosylated versus glycosylated ME7 prions. This inverse correlation of PrP glycosylation state with HS binding and plaque formation may be explained by the effect of PrP glycans on HS affinity, potentially decreased by electrostatic repulsion between the anionic glycans of PrP and HS. Our experiments indicating that unglycosylated PrP has the highest affinity for heparin support this idea. Another possible explanation is that PrP glycans sterically block heparin and HS binding sites. Further research will be required to distinguish among these possibilities.

Is highly glycosylated PrP compatible with parenchymal plaque formation? We tested this possibility by challenging mice that express triglycosylated PrP with mCWD prions, which are ADAM10-cleaved, monoglycosylated, and form large plaques. Notably, the first mCWD passage in *Prnp*^{187N} mice resulted in a near abrogation of plaques. Instead, prions formed mostly diffuse aggregates concurrent with a sharp reduction in the survival period. Although it remains a possibility that the change in the amino acid side chain and not the third glycans caused a switch in the strain phenotype, this would more likely have led to a lengthening of the incubation period. Instead, these findings suggest that highly glycosylated PrP may not form large, fibrillar parenchymal plaques, at least for certain strains, further supporting a link between PTM-deficient PrP and fibrillar parenchymal plaque formation.

We propose that a triad of factors act in concert to determine parenchymal prion plaque formation: (a) lack of a GPI-anchor, (b) one or no glycans, and (c) extracellular HS. Therefore, prions that replicate ADAM10-cleaved, unglycosylated PrP can bind HS and assemble into fibrils, forming extracellular parenchymal plaques. A key role for HS in parenchymal plaque formation is further supported by the finding that shortening HS chains in mice haploinsufficient in HS polymerase exostosin 1 decreases parenchymal plaques and increases vascular deposition of ADAM10-cleaved PrP^{Sc} (51). Similarly, in an AD mouse model, depleting neuronal HS chains leads to a reduction in parenchymal plaques and an increase in vascular plaques (80), also indicating a role for HS in amyloid- β parenchymal plaque formation. Future studies will be necessary to understand the exact role of HS in prion parenchymal plaque formation.

Our findings support and provide a possible explanation for the parenchymal plaques observed histologically in certain familial prion diseases, for example those caused by the F198S mutation in which the plaque core consists of an unglycosylated PrP fragment (81). Additional familial mutations associated with parenchymal plaque formation are also composed of mono- or unglycosylated PrP, including V180I (fCJD) (82) and T183A (1, 83, 84). Whether these prions are composed of GPI-anchored or ADAM10-cleaved PrP remains unclear, and antibodies that can distinguish the posttranslationally modified forms of PrP will be useful for future studies. Since most PrP^C in mammals is posttranslationally modified with a GPI-anchor and N-linked glycans, these results may help explain the rarity of fibrillar plaques in prion disease and the predominance of small aggregates associated with the highest levels of infectivity.

Methods

Additional Methods are included in the Supplemental Material.

Prnp^{180Q/196Q} and *Prnp*^{187N} mouse generation using the CRISPR/CAS9 system. The *Prnp*^{180Q/196Q} and *Prnp*^{187N} knockin mice were generated by the UC Irvine Mouse Genetics core by microinjection of Cas9 ribonucleoprotein (PNA Bio) into B6SJL/F1 × C57BL/6NJ zygotes. Briefly, Cas9 (20 ng/μL), gRNA-1 (20 ng/μL), and ssDNA HDR template (10 ng/μL) were mixed in injection buffer (10 mM Tris, 0.1 mM EDTA) and incubated on ice for 10 minutes, as per the manufacturer's instructions. The Cas9 mixture was microinjected into the pronucleus of single-cell zygotes isolated from super-ovulated females. All founders and select progeny were genotyped by Sanger sequencing genomic DNA from proteinase K digested tail tissue. Six founder mice carried the mutated *Prnp*^{180Q/196Q} allele and 10 founders carried the *Prnp*^{187N}. Select lines were backcrossed and bred to homozygosity. Aged mice investigated for spontaneous aggregation were 100–600 days old.

Statistics. A Student's *t* test (2-tailed, unpaired) was used to determine the statistical significance between the *Prnp*^{180Q/196Q} and *Prnp*^{187N} versus WT mouse brain samples for the PrP^C level of expression, PK digestion of PrP^C, PrP^{Sc} conformational stability, PrP^C solubility, levels of ADAM10-cleaved PrP^C and PrP^{Sc}, and levels of HS bound to PrP^{Sc} in ME7-infected WT and *Prnp*^{180Q/196Q} brains. Student's *t* test (2-tailed, unpaired) was also used to compare numbers of PrP^{Sc} plaques found in ME7-infected WT and *Prnp*^{180Q/196Q} mice, PrP^C cell surface levels from WT and *Prnp*^{180Q/196Q} cortical neurons, and cerebellar PrP^C levels from WT and *Prnp*^{180Q/196Q} mice.

One-way ANOVA with Tukey's post test was performed to assess survival differences between mouse groups, PrP^{Sc} solubility differences in the prion-infected mouse brain over multiple passages (*Prnp*^{180Q/196Q} and *Prnp*^{187N} as compared with WT mice), and to compare the binding affinity of unglycosylated, monoglycosylated, and diglycosylated isoforms of WT PrP^C to heparin.

Two-way ANOVA with Bonferroni's post hoc test was performed to assess differences in the lesion profiles, PrP^{Sc} levels in each brain region, plaque size of the prion-infected WT and *Prnp*^{180Q/196Q} brain, the heparin-binding affinity of RK13-derived PrP^{180Q/196Q}, PrP^{180Q}, PrP^{187N}, and WT PrP^C, the heparin-binding affinity of full-length and ADAM10-cleaved PrP^C from *Prnp*^{180Q/196Q} and WT mouse brains, and the composition of HS bound to PrP^{Sc} in ME7-infected *Prnp*^{180Q/196Q} and WT mouse brain. GraphPad Prism 5 software was used for statistical analyses. No measurement was excluded for statistical analysis. For all analyses, *P* less than or equal to 0.05 was considered significant. Data displayed in graphs represent mean ± SEM.

Study approval. All animal studies were performed following procedures to minimize suffering and were approved by the Institutional Animal Care and Use Committee at UC San Diego. Protocols were performed in strict accordance with good animal practices, as described in the Guide for the Use and Care of Laboratory Animals published by the National Institutes of Health.

Author contributions

AMS, PAC, TDK, JAL, KPRN, CJS designed experiments, HA and MG provided reagents, AMS, PAC, TDK, JAL, KS, THN, BC, TS, DPP, SN performed the experiments, AMS, PAC, TDK, JAL, THN, TS, DPP, HA, JDE, MG, KPRN, and CJS analyzed the experiments, AMS, PAC and CJS wrote the manuscript. The order of the co-first authors was determined by the study design, data, and experimental interpretation contributed by each co-author.

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