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Transfer RNA methyltransferase gene *NSUN2* mRNA expression modifies the effect of T cell activation score on patient survival in head and neck squamous carcinoma

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Abstract

Objectives: To investigate how T-cell activation interacts with *NSUN2* to influence HNSCC patient survival.

Materials and Methods: The relationships between T-cell activation status (Activation, Intermediate, and Exhaustion), *NSUN2* expression, and patient survival were evaluated using Kaplan-Meier survival curves and multivariate Cox regression models in a public dataset with 520 HNSCC patients. HPV status was determined based on a VirusScan analysis of RNA-seq data.

Results: Among the patients with high *NSUN2* expression, the Activation group exhibited longer survival than the Exhaustion group (trend $P=0.056$). Adjusted hazards ratios (HRs) were 0.77 (95% CI: 0.49–1.19) for the Intermediate vs Exhaustion, and 0.61 (0.36 – 1.03) for Activation vs. Exhaustion. In contrast, there is a positive association between T-cell activation score and mortality in the patients with low *NSUN2* expression (trend $P=0.016$). The adjusted HRs were 1.97 (1.12–3.47) for the Intermediate vs Exhaustion, and 2.06 (1.16–3.68) for the Activation vs Exhaustion. In multivariate cox models with or without HPV status, the interaction between T-cell activation status and *NSUN2* expression was statistically significant ($P=0.004$ for with HPV status, and $P=0.002$ for without, respectively). When not controlling for *NSUN2* expression, there was no significant association between T-cell activation score and patient mortality ($P=0.84$).

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Declarations:

Ethics approval and consent to participate: no formal consent is required for this type of study. The ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards were followed in performing all procedures in this study involving human subjects. The study presented here complies with the current laws of the United States of America.

Consent for publication: not applicable.

Conclusions: An interaction between *NSUN2* expression and T-cell activation status affects patient survival in HNSCC regardless of HPV status, suggesting that *NSUN2* is a potential precision marker for immune-checkpoint blockade, and a potential therapeutic target.

Keywords

head and neck squamous carcinoma; NSUN2; prognosis; T-cell activation score

Introduction

Immune escape is a hallmark of human cancer including head and neck squamous-cell carcinoma (HNSCC). T-cell exhaustion is one of a plethora of mechanisms that can underlie the escape of human cancer from immune surveillance. Both cell-intrinsic and cell-extrinsic negative regulatory pathways play important roles in capitalizing on T-cell dysfunction. Cytotoxic T-lymphocyte antigen 4 (CTLA4) and programmed cell death-1 receptor (PD-1) are two extensively investigated negative regulators that dampen the function of effector T cells by engaging them as receptors of ligands [1]. Early proof-of-concept of these regulatory interactions led to approval of anti-CTLA4 (ipilimumab, tremelimumab) and anti-PD-1 (nivolumab, pembrolizumab) immunotherapy by the United States Food and Drug Administration (FDA) for the clinical management of several types of human cancer. The principle of immune checkpoint-based antibody immunotherapies is to reinvigorate effector CD8⁺ T-cells by releasing the ‘brake’ on the immune system’s proclivity to kill tumor cells. Burtness and colleagues recently reported that pembrolizumab alone or with chemotherapy significantly improved overall survival in a phase 3 study of recurrent and metastatic HNSCC with a relatively large sample size [2]. However, not all patients gain benefit from these therapies. A single-arm phase-II trial of nivolumab in 44 patients with recurrent and metastatic nasopharyngeal carcinoma provided an overall objective response rate (ORR) of 20.5%, and a 1-year overall survival rate of 59% (95% CI: 44.3–78.5%) [3]. Another phase-II trial of pembrolizumab included 27 patients with PD-L1-positive nasopharyngeal carcinoma, wherein the ORR was 25.9% (11.1–46.3%) over a median follow-up of 20 months [4]. An open-label phase-3 trial of nivolumab vs. standard chemotherapy (2:1) in 361 patients with recurrent HNSCC provided 2.4 months of overall survival benefit in the nivolumab group compared to standard therapy, and a response rate of 13.3% for nivolumab vs. 5.8% for standard therapy [5]. A phase-1b trial of pembrolizumab with 60 PD-L1-positive HNSCC patients provided 18% (8–32%) overall response [6]. Another phase 1b trial of pembrolizumab in 132 patients with recurrent and/or metastatic HNSCC produced 18% ORR (12–26%) [7]. A recent phase I/II single-arm trial of durvalumab (anti-PD-L1) and tremelimumab (anti-CTLA4) in combination is ongoing, and includes 35 patients with metastatic HNSCC, and no primary end-point results other than adverse events have been reached thus far [8]. These observations have motivated the search for other negative regulators of the immune response, such as LAG3, TIM3, and TIGIT, three regulators under active investigation [9, 10]. Numerous studies are investigating other cellular signals, such as the signals that affect the host response to immune-checkpoint blockade. For instance, CDK4/6 inhibitor treatment enhances antitumor efficacy of PD-1 immunotherapy, dramatically improving overall survival in animal models [11]. ADAR1 is an adenosine deaminase, which can bind and limit the sensing of endogenous double-stranded RNAs

(dsRNAs) that leads to inflammation. Loss of *ADAR1* can suppress PD-1 immunotherapy resistance by increasing tumor inflammation [12].

NOP2/Sun domain family member 2 protein (*NSUN2*), encoded in chromosome 10 by the *NSUN2* gene, is a RNA-modification protein for m⁵C methylation in both cytoplasmic and mitochondrial tRNAs [13–15]. The methylation of tRNAs prevents tRNAs from endonucleolytic cleavage by angiogenin to generate tRNA-derived fragments (tRFs), consequently controlling the efficacy of protein translation [16–20], and leading to cytokine production and cellular metabolic changes in response to stress [21]. *NSUN2*-deficient cells exhibit reduced protein synthesis, dysregulation of cell cycle, abnormal cell differentiation and proliferation [17–20, 22]. *NSUN2*-knockout testes are completely absent of spermatids and sperm [23]. Over-expression of *NSUN2* promotes the migration of neural cells toward the chemoattractant growth factor [24]. Both *NSUN2* and *IGF-II* increase ovarian cancer mortality risk when expressed at high levels [25]. By interacting with ribosomal protein L6 (*RPL6*), *NSUN2* drives the progression of gallbladder carcinoma [26]. Chen and colleagues recently demonstrated that *NSUN2* promoted pathogenesis of bladder cancer via stabilizing mRNAs, and that overexpression of *NSUN2* predicted poor prognosis of bladder cancer [27]. A previous study showed that high *NSUN2* expression associated with poor prognosis in HNSCC [28]. In addition, *NSUN2* has also been shown to modify mRNAs. High *NSUN2* enhances *CDK1* translation mediated by *CDK1* mRNA methylation, consequently stimulating cell growth [29]. The stability of p16ink4 mRNA, a CDK inhibitor blocking the formation of cyclin D-CDK4/6 complex, increases due to *NSUN2*-mediated mRNA methylation [30, 31]. Moreover, RNA methylation-mediated switch in the secondary structure between double-stranded and single-stranded RNAs can affect immune response [12]. Given that immune cells sense tumors by utilizing pathogen and damage receptors, by which tumor-specific immunity is elicited, we hypothesized that *NSUN2* expression level modifies the effect of T-cell activation on patients' survival in HNSCC.

Methods

Data sources

This study includes 520 individuals with primary HNSCC, whose gene expression information and clinical data were retrieved from a TCGA dataset at cBioPortal (<https://www.cbioportal.org>). Among 518 patients with age information available, the average age was 60.9 years old (standard deviation 11.9, range 19–90). Among 519 patients whose sex was reported, 73.8% were men, and 26.2% were women. Patient ethnicity was Caucasian for 87.9% of reports (444 out of 505), with the remaining 12.1% reported as African American, Asian, and American Native. There were 117 non-smokers and 389 smokers. The majority of patients were diagnosed with the disease at advanced stage (282 at stage IV, 105 at stage III, 98 at stage II and 20 at stage I). Poor differentiation was observed in patient tumor tissues, at an incidence of 12.0% (62 out of 515) for grade I, 58.8% for grade II and 29% for grade III. Tumor sites ($n = 519$) included tongue (38.3%), pharynx (26.0%), and other (35.7%). Approximately half (52.6%, 271/515) had lymph invasion, and 35.2% (122/347) of patients had vascular invasion. Follow-up information was available for 517 patients, and the average overall survival was 21.5 months (range 0.07–210.8 months).

mRNA levels in Fragments Per Kilobase of transcript per Million mapped reads (FPKM), which were normalized by the upper quartile expectation maximization (RNA-seq V2 RSEM), were retrieved for *NSUN2* as well as a panel of 14 genes that are associated with M2 macrophage and myeloid cells: *NKG7*, *CCL4*, *CST7*, *PRF1*, *GZMA*, *GZMB*, *IFNG*, *CCL3*, *PDCD1* (aka *PD-1*), *TIGIT*, *LAG3*, *TIM3*, *CTLA4* and *CD274* (aka *PD-L1*) [32, 33]. Experimental data generation and processing were conducted as previously described [34]. No patients received neoadjuvant treatment.

No formal written consent is required for this type of study. The ethical standards of the institutional and/or national research committee and the 1964 Helsinki declaration and its later amendments or comparable ethical standards were followed in performing all procedures in this study involving human subjects. The study presented here complies with the current laws of USA.

RNA-seq-based HPV status determination

RNA sequencing-based HPV status determination was performed using VirusScan as described previously [35]. A cutoff of 100 HPV RNA viral transcript reads per hundred million (TPM) was specified as determinate of the HPV status for a tumor, with an HPV positive determination when there were ≥ 100 TPM, and HPV negative determination when there were <100 TPM. Using this cutoff, the concordance is 100% between RNA-seq-based HPV status and the HPV status determined by in situ hybridization performed on 248 tumors in the TCGA clinical data.

Gene-set enrichment analysis

Co-expression of genome-wide genes with *NSUN2* was analyzed using Spearman correlation with multiple comparison correction. Gene set enrichment analysis was performed for the genes that had an adjusted *P* value < 0.001 [36].

Statistical analyses

A T-cell activation score was calculated for each subject as described previously [37], by taking the weighted average of log-transformed expression (FPKM + 1) across the gene panel. The ‘overall survival’ was defined as the number of months from the initial diagnosis until death or the last follow-up. Spearman correlation was used to evaluate correlations. Survival analyses were performed using Kaplan-Meier survival curves and multivariate Cox proportional hazards, in which three groups—high (activation), intermediate, and low (exhaustion)—were assigned using the tertiles of the T-cell activation score. To investigate whether the *NSUN2* expression modifies the effect of T-cell activation status on patients’ survival, we used the median of the *NSUN2* expression level as the cutoff value to stratify the models. Patient age at diagnosis, disease stage, tumor grade, gender, and smoking history were included in the models to estimate adjusted hazards ratios (HRs) and their 95% confidence intervals (95% CIs). We also assessed the interaction between T-cell activation score and *NSUN2* level across all patients by including their interactions in the Cox regression models. The proportional hazards assumption was examined. In all statistical analyses, a *P* value less than 0.05 was considered significant. Statistical analyses were performed using SAS version 9.4 (SAS Institute, Inc), and *cgdsr* and *ReactomPA* packages

for R version 3.5.0 (<https://www.R-project.org>) were used for gene retrieval and pathway analysis.

Results

T-cell activation score and gene expression levels

Table 1 shows the distribution of T-cell activation score, and the expression of *NSUN2*. The averages were 0.21 with a standard deviation of 0.09 for T-cell activation score, 2221 FPKM for *NSUN2*. A significant negative correlation was found between *NSUN2* expression and T cell activation score ($P = 0.002$). The Spearman correlation coefficient was -0.14 (95% CI: $-0.22, -0.05$).

Table 2 and Figure 1 show the correlation and scatter plots of *NSUN2* expression with CD163, CD33, and PDCD1 and myeloid cell marker CD45, respectively. There were significant negative correlations between *NSUN2* expression and the expressions of M2-like macrophage markers *CD163* ($P = 0.0003$), *CD33* ($P < 0.0001$), *PDCD1* ($P < 0.0001$), and *CD45* ($P < 0.0001$; Table 2; Figure 1). The Spearman correlation coefficients were -0.16 ($-0.24, -0.07$) for *CD163*, -0.27 ($-0.34, -0.18$) for *CD33*, -0.31 ($-0.47, -0.22$) for *PDCD1*, and -0.25 ($-0.32, -0.16$) for *CD45*.

Interaction between T-cell activation status and NSUN2 expression level in patient survival

In the survival analysis, we first tested the proportional-hazards assumption based on 1000 simulations for T-cell activation status, and use of the assumption was validated ($P = 0.676$ for all patients, $P = 0.185$ in the *NSUN2* low expression group, and $P = 0.922$ in the *NSUN2* high expression group).

We investigated the relationship between survival and the T-cell activation status (Activation, Intermediate, and Exhaustion) in the whole sample. No significant association was observed between T-cell activation status and overall survival in HNSCC ($P = 0.97$). The medians of overall survival were 65.8 months (32.2 – 89.3 months) for patients with a low T-cell activation score (Exhaustion), 49.4 months (34.5 – 71.2 months) for those with an intermediate T-cell activation score (Intermediate), and 54.9 months (95% CI: 37.3 – 68.4 months) for those with a high T-cell activation score (Activation; data not shown).

In the *NSUN2* low expression group, patients in the Activation group showed inferior overall survival compared to those in the Exhaustion group (Figure 2A). The median of overall survival was 88.8 months (52.3–210.8 months) in the Exhaustion group, 49.4 months (35.5–156.4 months) in the Intermediate group, and 54.9 months (28.0–67.8 months) in the Activation group. Patients in the Activation group lived on average 34.9 months (almost 3 years) shorter than those in the Exhaustion group. The Activation group showed significantly increased risk of death (trend $P = 0.012$) compared to the Exhaustion group. The HRs of death were 1.67 (1.00–2.78) for Intermediate vs Exhaustion ($p = 0.05$), and 1.96 (1.16–3.29) for Activation vs Exhaustion ($p = 0.012$). In contrast, in the *NSUN2* high expression group, patients with an Activation status showed better overall survival compared to those with an Exhaustion status (Figure 2B). The median of overall survival was 26.4 months (17.9–76.2 months) in the Exhaustion group, 45.8 months (27.5–84.4 months) in the

Intermediate group, and 159.5 months (42.4–159.5 months) in the Activation group, respectively. On average, patients in the Activation group lived 133.1 months (over 11 yrs) longer than those in the Exhaustion group. The HRs of death were 0.75 (0.49–1.13) for Intermediate vs Exhaustion ($p = 0.168$), and 0.61 (0.38–0.99) for Activation vs Exhaustion ($p = 0.044$). The Activation group tended to have a lower risk in comparison to the Exhaustion group (trend $P = 0.037$). In the whole sample, a significant interaction was observed between the expression level of *NSUN2* and the T-cell activation status in the interaction test ($P = 0.004$).

We then performed multivariate Cox proportional-hazards models to adjust for potential confounding variables including the patient's age at surgery, disease stage, tumor grade, gender, smoking status, and tumor site ("model 1"; Table 3). Similarly, there was no significant association between the T-cell activation status and death risk among all patients without accounting for the expression level of *NSUN2* (Table 2). The adjusted HRs were 1.08 (0.77–1.50) for Intermediate vs Exhaustion ($P = 0.664$), and 1.03 (0.73–1.47) for Activation vs. Exhaustion ($P = 0.852$). However, when adjusting for covariates and including expression of *NSUN2* in model 1, a significant interaction term is estimated between the expression level of *NSUN2* and the T-cell activation status ($P = 0.002$). In the group with low *NSUN2* expression, T-cell activation score was positively associated with increased death risk (trend $P = 0.009$). The adjusted HRs were 2.01 (1.15 – 3.52) for Intermediate vs Exhaustion ($P = 0.015$), and 2.16 (1.22–3.82) for Activation vs. Exhaustion ($P = 0.008$). In contrast, in the group with high expression of *NSUN2*, T-cell activation score was negatively associated with the disease risk (trend $P = 0.028$). The adjusted HRs were 0.74 (0.48–1.14) for Intermediate vs Exhaustion ($P = 0.174$), and 0.57 (0.34–0.96) for Activation vs Exhaustion ($P = 0.033$). Moreover, the interaction between the *NSUN2* expression and T-cell activation status remained significant ($P = 0.004$) after addition of HPV status to the set of covariates ("model 2"). As with model 1, in the group with low *NSUN2* expression, T-cell activation score was positively associated with an increased death risk (trend $P = 0.016$), whereas in the group with high *NSUN2* expression, the Activation group trended toward a lower risk of death ($P = 0.056$, not significant). Among patients with low *NSUN2* expression, the adjusted HRs were 1.97 (1.12–3.47) for Intermediate vs Exhaustion ($p = 0.019$), and 2.06 (1.16–3.68) for Activation vs. Exhaustion ($p = 0.014$). Among patients with high *NSUN2* expression, adjusted HRs were 0.77 (0.49–1.19) for Intermediate vs Exhaustion ($p = 0.240$), and 0.61 (0.36–1.03) for Activation vs Exhaustion ($p = 0.063$).

Pathway analysis for gene co-expressed with *NSUN2*

Among the 15 pathways that are enriched for genes coexpressed with *NSUN2*, the majority (11 pathways) are immune function- or inflammation-related pathways (such as interleukins, PD-1 signaling, TCR signaling, neutrophil degranulation, NF- κ B and immunoregulatory interaction) (Figure 3) (adjusted $p < 0.001$). Gene-set enrichment analysis showed that high expression of *NSUN2* was associated with low expression of genes in pathways such as IFN- γ signaling (normalized enrichment score (NES) = - 2.94, false-discovery rate (FDR) = 0.006), TCR signaling (NES = -2.80, FDR = 0.006), innate immune system (NES = -3.13, FDR = 0.006), adaptive immune system (NES = -3.75, FDR = 0.006), PD-1 signaling (NES = -3.75, FDR = 0.006), Immunoregulatory interaction (NES = -4.29, FDR = 0.006), IL-2

signaling (NES = -2.67, FDR = 0.006), IL-3/IL-5 and GM-CSF signaling (NES = -2.37, FDR = 0.006). High expression of *NSUN2* was associated with high expression of genes in pathways such as cell cycle checkpoint (NES = 2.60, FDR = 0.006), epigenetic regulation of gene expression (NES = 2.49, FDR = 0.006), gene silencing by RNA (NES = 2.46, FDR = 0.006), mRNA splicing (NES = 2.81, FDR = 0.006), unfolded protein response (NES = 2.27, FDR = 0.006), and mitochondrial translation (NES = 2.27, FDR = 0.006) and tRNA modification (NES = 2.44, FDR = 0.006) and processing (NES = 2.49, FDR = 0.006) (Supplementary Table S1).

Discussion

In this study, we investigated the association of T-cell activation score with patient survival and the interaction between *NSUN2* expression and the T-cell activation status in HNSCC patient survival. In our analysis of 520 patients with primary HNSCC, the relationship between T-cell activation score and patient survival was in the opposite direction for tumors with high expression of *NSUN2* as for tumors with low expression of *NSUN2*. We found that in the subgroup with a low expression of *NSUN2*, patients with a high T-cell activation score lived 34.9 months shorter on average than those with a low T-cell activation score, whereas in the subgroup with high expression of *NSUN2*, patients with a high T-cell activation score lived 133.1 months longer on average than those with a low T-cell activation score. There was no significant difference in median survival between patients with high and low T-cell activation scores when not stratifying by *NSUN2* expression level. Interaction analysis with a multivariate Cox model suggested a statistically significant interaction between expression of *NSUN2* and T-cell activation score in HNSCC. When we included HPV status in the model—which reduced the sample size to 500 patients—the relationship remained statistically significant. This consistency implies that the *NSUN2* expression level modifies the effect of T-cell activation on patient survival in HNSCC regardless of HPV status.

T-cell exhaustion is a hallmark of human cancer—including HNSCC—and is characterized by high expression of deactivating immune checkpoint proteins such as PD-1/PD-L1, CTLA4, LAG3, TIM3 and TIGIT. Reinvigoration of effector CD8+ T cells by immune-checkpoint blockades such as anti-PD-1/anti-PD-L1 and anti-CTLA4 improves patient survival in several human solid cancers, including HNSCC [3–5]. However, not all patients respond well to the treatment. Unexpectedly, even some patients treated with immunotherapy paradoxically showed hyper-progression of tumor growth [38, 39]. Lo Russo and colleagues reported that hyper-progression occurred in about 25% patients (39 out of 152) with non-small cell lung cancer (NSCLC) who received PD-1/PD-L1 blockade [38]. They found enriched tumor-associated macrophages (TAM) with M2-like CD163+CD33+PD-L1+ phenotype in the hyper-progression patients. Patient-derived xenograft (PDX) mouse models reproduced the phenotype and the clustering of TAMs. Saada-Bouزيد and colleagues retrospectively evaluated the response of patients with recurrent and/or metastatic HNSCC to anti-PD-1/PD-L1 immunotherapy, and found 29% patients with hyper-progression, which was positively associated with a shorter progression-free survival [40]. By performing next-generation sequencing analysis on 155 patients with a stage-IV cancer who received immunotherapy, Kato and colleagues demonstrated that those

with hyper-progression showed *MDM2* family amplification or EGFR aberrations or DNMT3A alterations [41]. A potential mechanism was hypothesized: that anti-PD-1/PD-L1 induced IFN- γ -activated JAK-STAT signaling, consequently increasing interferon regulator factor (IRF)-8 expression that further resulted in expression of MDM2 [41–43]. MDM2 overexpression inhibits the p53 tumor suppressor [41]. Champiat and colleagues retrospectively evaluated 218 consecutive patients enrolled and treated in phase-I clinical trials of anti-PD-1/PD-L1 therapy, and found that 9% (12 of 131) patients had hyperprogression after the immunotherapy treatment, and that hyper-progression was associated with an elevated death risk [44]. Ferrara and colleagues reported that 13.8% (56 of 406) of patients treated with immunotherapy and 5.1% (3 of 59) of patients treated with single-agent chemotherapy had hyper-progression in NSCLC, and that these hyper-progressive patients had poor prognosis [45]. However, the potential molecular mechanisms underlying the hyper-progression are unclear and have yet to be characterized. The reactivation of CD8⁺ T-cells by PD-1 blockade results in elevated IFN- γ , which has been shown to activate tumor immunosuppressive myeloid cells, and increased inhibitory metabolites (such as indoleamine 2,3 –dioxygenase) controlling Treg differentiation, or leading to aberrant inflammation and a microenvironment that favors immune escape and tumor growth [46–49].

NSUN2, as a tRNA methyltransferase, modifies both tRNAs and mRNAs by adding m⁵C methylation to RNAs. NSUN2 is essential factor for maintenance of self-renewal and differentiation in stem cells [24, 50, 51]. CD8⁺ T-cells of sufficient stemness provide the capacity within the immune system to respond the stimuli of tumor neoantigens, and keep CD8⁺ T cells at the enough ‘ammunition’ even with chronic antigen exposure. It has been reported that NSUN2 promotes the translation of intercellular adhesion molecule 1 (ICAM-1) by methylating ICAM-1 mRNA [52], which can further suppress tumor metastasis by inhibiting M2-macrophage polarization. In this study, we found a statistically significant negative association between expression of *NSUN2* and the expression of markers of M2 macrophages, including *CD163*, *CD33*, *PDCD1* (aka *PD-1*), and myeloid cell marker *CD45*. This observation suggests that a low *NSUN2* expression is accompanied with a high level of M2 macrophage and myeloid cells. We found that, conditioning on low expression of *NSUN2*, patients with a high T-cell activation score exhibited inferior overall survival than those with a low T-cell activation score, suggesting that a phenomenon similar to hyper-progression could be occurring at high immune response. We do not know what molecular mechanisms underlie this phenomenon—it may be complicated, but it warrants exploration. Indeed, in the context of tumors, the immune system may have dual functions of both eradicating ‘foreign antigens’, and contributing to tumor growth through direct or indirect mechanisms such as DNA damage and angiogenesis/tumor tissue remodeling induced by inflammation and free radicals [53–55]. Recently, research utilizing animal models demonstrated that RNA-editing induced RNA structure switching from single-strand RNA (ssRNA) to dsRNA could increase tumor inflammation and enhance the efficacy of PD-1 blockade [12]. Gene-set enrichment analysis for *NSUN2* co-expressed genes suggests that low expression of *NSUN2* could lead to excessive inflammation and immune response (negatively associated with IFN- γ signaling), and could synergize with high T-cell activation score to result in increased mortality.

Conclusions

This study is the first one to demonstrate the interaction between T-cell activation score and expression of *NSUN2* affects patient survival in HNSCC. T-cell activation score was negatively associated with *NSUN2* expression, but was not associated with patient survival overall. However, in the subgroup of patients with low *NSUN2* expression, high T-cell activation score significantly increased the risk of mortality, whereas an opposite association was observed in the subgroup of patients with high expression of *NSUN2*, where a (non-significant) reduction in the risk of mortality manifested for those with high T-cell activation score. These findings suggest that immune checkpoint inhibitors could benefit patients with high *NSUN2* expression by reinvigorating effector T cells, but may be detrimental to patients who have a low *NSUN2* expression. *NSUN2* expression is a potential marker for precision immunotherapy in HNSCC, and the outcome of immunotherapy might be able to be improved by targeting *NSUN2*. Studies with larger sample sizes are warranted to further examine how *NSUN2* expression level affects the patients' response to immune checkpoint blockade.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Availability of data and material: The data that support the findings of this study are available from head and neck squamous carcinoma (TCGA, Provisional) at cBioPortal for Cancer Genomics (<https://www.cbioportal.org>).

References:

- [1]. Chen L, Han X. Anti-PD-1/PD-L1 therapy of human cancer: past, present, and future. *J Clin Invest.* 2015;125:3384–91. [PubMed: 26325035]
- [2]. Burtness B, Harrington KJ, Greil R, Soulieres D, Tahara M, Castro G, et al. Pembrolizumab alone or with chemotherapy versus cetuximab with chemotherapy for recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-048): a randomized, open-label, phase 3 study. *Lancet.* 2019.
- [3]. Ma BBY, Lim WT, Goh BC, Hui EP, Lo KW, Pettinger A, et al. Antitumor Activity of Nivolumab in Recurrent and Metastatic Nasopharyngeal Carcinoma: An International, Multicenter Study of the Mayo Clinic Phase 2 Consortium (NCI-9742). *J Clin Oncol.* 2018;36:1412–8. [PubMed: 29584545]
- [4]. Hsu C, Lee SH, Ejadi S, Even C, Cohen RB, Le Tourneau C, et al. Safety and Antitumor Activity of Pembrolizumab in Patients With Programmed Death-Ligand 1-Positive Nasopharyngeal Carcinoma: Results of the KEYNOTE-028 Study. *J Clin Oncol.* 2017;35:4050–6. [PubMed: 28837405]
- [5]. Ferris RL, Blumenschein G Jr., Fayette J, Guigay J, Colevas AD, Licitra L, et al. Nivolumab for Recurrent Squamous-Cell Carcinoma of the Head and Neck. *N Engl J Med.* 2016;375:1856–67. [PubMed: 27718784]

- [6]. Seiwert TY, Burtneß B, Mehra R, Weiss J, Berger R, Eder JP, et al. Safety and clinical activity of pembrolizumab for treatment of recurrent or metastatic squamous cell carcinoma of the head and neck (KEYNOTE-012): an open-label, multicentre, phase 1b trial. *Lancet Oncol.* 2016;17:956–65. [PubMed: 27247226]
- [7]. Chow LQM, Haddad R, Gupta S, Mahipal A, Mehra R, Tahara M, et al. Antitumor Activity of Pembrolizumab in Biomarker-Unselected Patients With Recurrent and/or Metastatic Head and Neck Squamous Cell Carcinoma: Results From the Phase Ib KEYNOTE-012 Expansion Cohort. *J Clin Oncol.* 2016;34:3838–45. [PubMed: 27646946]
- [8]. Bahig H, Aubin F, Stagg J, Gologan O, Ballivy O, Bissada E, et al. Phase I/II trial of Durvalumab plus Tremelimumab and stereotactic body radiotherapy for metastatic head and neck carcinoma. *BMC Cancer.* 2019;19:68. [PubMed: 30642290]
- [9]. Fourcade J, Sun Z, Pagliano O, Guillaume P, Luescher IF, Sander C, et al. CD8(+) T cells specific for tumor antigens can be rendered dysfunctional by the tumor microenvironment through upregulation of the inhibitory receptors BTLA and PD-1. *Cancer Res.* 2012;72:887–96. [PubMed: 22205715]
- [10]. Wherry EJ. T cell exhaustion. *Nat Immunol.* 2011;12:492–9. [PubMed: 21739672]
- [11]. Zhang J, Bu X, Wang H, Zhu Y, Geng Y, Nihira NT, et al. Cyclin D-CDK4 kinase destabilizes PD-L1 via cullin 3-SPOP to control cancer immune surveillance. *Nature.* 2018;553:91–5. [PubMed: 29160310]
- [12]. Ishizuka JJ, Manguso RT, Cheruiyot CK, Bi K, Panda A, Iracheta-Vellve A, et al. Loss of ADAR1 in tumours overcomes resistance to immune checkpoint blockade. *Nature.* 2019;565:43–8. [PubMed: 30559380]
- [13]. Brzezicha B, Schmidt M, Makalowska I, Jarmolowski A, Pienkowska J, Szweykowska-Kulinska Z. Identification of human tRNA:m5C methyltransferase catalysing intron-dependent m5C formation in the first position of the anticodon of the pre-tRNA Leu (CAA). *Nucleic Acids Res.* 2006;34:6034–43. [PubMed: 17071714]
- [14]. Shinoda S, Kitagawa S, Nakagawa S, Wei FY, Tomizawa K, Araki K, et al. Mammalian NSUN2 introduces 5-methylcytidines into mitochondrial tRNAs. *Nucleic Acids Res.* 2019;47:8734–45. [PubMed: 31287866]
- [15]. Van Haute L, Lee SY, McCann BJ, Powell CA, Bansal D, Vasiliauskaite L, et al. NSUN2 introduces 5-methylcytosines in mammalian mitochondrial tRNAs. *Nucleic Acids Res.* 2019;47:8720–33. [PubMed: 31276587]
- [16]. Blanco S, Dietmann S, Flores JV, Hussain S, Kutter C, Humphreys P, et al. Aberrant methylation of tRNAs links cellular stress to neuro-developmental disorders. *EMBO J.* 2014;33:2020–39. [PubMed: 25063673]
- [17]. Ivanov P, Emara MM, Villen J, Gygi SP, Anderson P. Angiogenin-induced tRNA fragments inhibit translation initiation. *Mol Cell.* 2011;43:613–23. [PubMed: 21855800]
- [18]. Sobala A, Hutvagner G. Small RNAs derived from the 5' end of tRNA can inhibit protein translation in human cells. *RNA Biol.* 2013;10:553–63. [PubMed: 23563448]
- [19]. Spriggs KA, Bushell M, Willis AE. Translational regulation of gene expression during conditions of cell stress. *Mol Cell.* 2010;40:228–37. [PubMed: 20965418]
- [20]. Tuorto F, Liebers R, Musch T, Schaefer M, Hofmann S, Kellner S, et al. RNA cytosine methylation by Dnmt2 and NSun2 promotes tRNA stability and protein synthesis. *Nat Struct Mol Biol.* 2012;19:900–5. [PubMed: 22885326]
- [21]. Liu S, Chen Y, Ren Y, Zhou J, Ren J, Lee I, et al. A tRNA-derived RNA Fragment Plays an Important Role in the Mechanism of Arsenite -induced Cellular Responses. *Sci Rep.* 2018;8:16838. [PubMed: 30442959]
- [22]. Popis MC, Blanco S, Frye M. Posttranscriptional methylation of transfer and ribosomal RNA in stress response pathways, cell differentiation, and cancer. *Curr Opin Oncol.* 2016;28:65–71. [PubMed: 26599292]
- [23]. Hussain S, Tuorto F, Menon S, Blanco S, Cox C, Flores JV, et al. The mouse cytosine-5 RNA methyltransferase NSun2 is a component of the chromatoid body and required for testis differentiation. *Mol Cell Biol.* 2013;33:1561–70. [PubMed: 23401851]

- [24]. Flores JV, Cordero-Espinoza L, Oeztuerk-Winder F, Andersson-Rolf A, Selmi T, Blanco S, et al. Cytosine-5 RNA Methylation Regulates Neural Stem Cell Differentiation and Motility. *Stem Cell Reports*. 2017;8:112–24. [PubMed: 28041877]
- [25]. Yang JC, Risch E, Zhang M, Huang C, Huang H, Lu L. Association of tRNA methyltransferase NSUN2/IGF-II molecular signature with ovarian cancer survival. *Future Oncol*. 2017;13:1981–90. [PubMed: 28829218]
- [26]. Gao Y, Wang Z, Zhu Y, Zhu Q, Yang Y, Jin Y, et al. NOP2/Sun RNA methyltransferase 2 promotes tumor progression via its interacting partner RPL6 in gallbladder carcinoma. *Cancer Sci*. 2019.
- [27]. Chen X, Li A, Sun BF, Yang Y, Han YN, Yuan X, et al. 5-methylcytosine promotes pathogenesis of bladder cancer through stabilizing mRNAs. *Nat Cell Biol*. 2019;21:978–90. [PubMed: 31358969]
- [28]. Lu L, Zhu G, Zeng H, Xu Q, Holzmann K. High tRNA Transferase NSUN2 Gene Expression is Associated with Poor Prognosis in Head and Neck Squamous Carcinoma. *Cancer investigation*. 2018;36:246–53. [PubMed: 29775108]
- [29]. Xing J, Yi J, Cai X, Tang H, Liu Z, Zhang X, et al. NSun2 Promotes Cell Growth via Elevating Cyclin-Dependent Kinase 1 Translation. *Mol Cell Biol*. 2015;35:4043–52. [PubMed: 26391950]
- [30]. Zhang X, Liu Z, Yi J, Tang H, Xing J, Yu M, et al. The tRNA methyltransferase NSun2 stabilizes p16INK(4) mRNA by methylating the 3'-untranslated region of p16. *Nat Commun*. 2012;3:712. [PubMed: 22395603]
- [31]. Sharpless NE. INK4a/ARF: a multifunctional tumor suppressor locus. *Mutat Res*. 2005;576:22–38. [PubMed: 15878778]
- [32]. Cancer Genome Atlas N Comprehensive molecular portraits of human breast tumours. *Nature*. 2012;490:61–70. [PubMed: 23000897]
- [33]. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal*. 2013;6:p11. [PubMed: 23550210]
- [34]. Cancer Genome Atlas Research N. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature*. 2014;513:202–9. [PubMed: 25079317]
- [35]. Cannataro VL, Gaffney SG, Sasaki T, Issaeva N, Grewal NKS, Grandis JR, et al. APOBEC-induced mutations and their cancer effect size in head and neck squamous cell carcinoma. *Oncogene*. 2019.
- [36]. Yu G, He QY. ReactomePA: an R/Bioconductor package for reactome pathway analysis and visualization. *Mol Biosyst*. 2016;12:477–9. [PubMed: 26661513]
- [37]. Lu L, Bai Y, Wang Z. Elevated T cell activation score is associated with improved survival of breast cancer. *Breast Cancer Res Treat*. 2017;164:689–96. [PubMed: 28488141]
- [38]. Lo Russo G, Moro M, Sommariva M, Cancila V, Boeri M, Centonze G, et al. Antibody-Fc/FcR Interaction on Macrophages as a Mechanism for Hyperprogressive Disease in Non-small Cell Lung Cancer Subsequent to PD-1/PD-L1 Blockade. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2019;25:989–99. [PubMed: 30206165]
- [39]. Knorr DA, Ravetch JV. Immunotherapy and Hyperprogression: Unwanted Outcomes, Unclear Mechanism. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2019;25:904–6. [PubMed: 30397179]
- [40]. Saada-Bouzid E, Defaucheux C, Karabajakian A, Coloma VP, Servois V, Paoletti X, et al. Hyperprogression during anti-PD-1/PD-L1 therapy in patients with recurrent and/or metastatic head and neck squamous cell carcinoma. *Annals of oncology : official journal of the European Society for Medical Oncology*. 2017;28:1605–11. [PubMed: 28419181]
- [41]. Kato S, Goodman A, Walavalkar V, Barkauskas DA, Sharabi A, Kurzrock R. Hyperprogressors after Immunotherapy: Analysis of Genomic Alterations Associated with Accelerated Growth Rate. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2017;23:4242–50. [PubMed: 28351930]
- [42]. Zhou JX, Lee CH, Qi CF, Wang H, Naghashfar Z, Abbasi S, et al. IFN regulatory factor 8 regulates MDM2 in germinal center B cells. *J Immunol*. 2009;183:3188–94. [PubMed: 19648273]

- [43]. Waight JD, Netherby C, Hensen ML, Miller A, Hu Q, Liu S, et al. Myeloid-derived suppressor cell development is regulated by a STAT/IRF-8 axis. *J Clin Invest*. 2013;123:4464–78. [PubMed: 24091328]
- [44]. Champiat S, Derclé L, Ammari S, Massard C, Hollebecque A, Postel-Vinay S, et al. Hyperprogressive Disease Is a New Pattern of Progression in Cancer Patients Treated by Anti-PD-1/PD-L1. *Clinical cancer research : an official journal of the American Association for Cancer Research*. 2017;23:1920–8. [PubMed: 27827313]
- [45]. Ferrara R, Mezquita L, Texier M, Lahmar J, Audigier-Valette C, Tessonier L, et al. Hyperprogressive Disease in Patients With Advanced Non-Small Cell Lung Cancer Treated With PD-1/PD-L1 Inhibitors or With Single-Agent Chemotherapy. *JAMA Oncol*. 2018;4:1543–52. [PubMed: 30193240]
- [46]. Akbay EA, Koyama S, Liu Y, Dries R, Bufe LE, Silkes M, et al. Interleukin-17A Promotes Lung Tumor Progression through Neutrophil Attraction to Tumor Sites and Mediating Resistance to PD-1 Blockade. *J Thorac Oncol*. 2017;12:1268–79. [PubMed: 28483607]
- [47]. Baban B, Chandler PR, Sharma MD, Pihkala J, Koni PA, Munn DH, et al. IDO activates regulatory T cells and blocks their conversion into Th17-like T cells. *J Immunol*. 2009;183:2475–83. [PubMed: 19635913]
- [48]. Spranger S, Spaapen RM, Zha Y, Williams J, Meng Y, Ha TT, et al. Up-regulation of PD-L1, IDO, and T(regs) in the melanoma tumor microenvironment is driven by CD8(+) T cells. *Sci Transl Med*. 2013;5:200ra116.
- [49]. Huang B, Pan PY, Li Q, Sato AI, Levy DE, Bromberg J, et al. Gr-1+CD115+ immature myeloid suppressor cells mediate the development of tumor-induced T regulatory cells and T-cell anergy in tumor-bearing host. *Cancer Res*. 2006;66:1123–31. [PubMed: 16424049]
- [50]. Frye M, Harada BT, Behm M, He C. RNA modifications modulate gene expression during development. *Science*. 2018;361:1346–9. [PubMed: 30262497]
- [51]. Blanco S, Kurowski A, Nichols J, Watt FM, Benitah SA, Frye M. The RNA-methyltransferase Misu (NSun2) poises epidermal stem cells to differentiate. *PLoS genetics*. 2011;7:e1002403. [PubMed: 22144916]
- [52]. Luo Y, Feng J, Xu Q, Wang W, Wang X. NSun2 Deficiency Protects Endothelium From Inflammation via mRNA Methylation of ICAM-1. *Circ Res*. 2016;118:944–56. [PubMed: 26838785]
- [53]. Guo X, Zhai L, Xue R, Shi J, Zeng Q, Gao C. Mast Cell Tryptase Contributes to Pancreatic Cancer Growth through Promoting Angiogenesis via Activation of Angiotensin-1. *Int J Mol Sci*. 2016;17.
- [54]. Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer*. 2004;4:71–8. [PubMed: 14708027]
- [55]. DeNardo DG, Andreu P, Coussens LM. Interactions between lymphocytes and myeloid cells regulate pro- versus anti-tumor immunity. *Cancer Metastasis Rev*. 2010;29:309–16. [PubMed: 20405169]

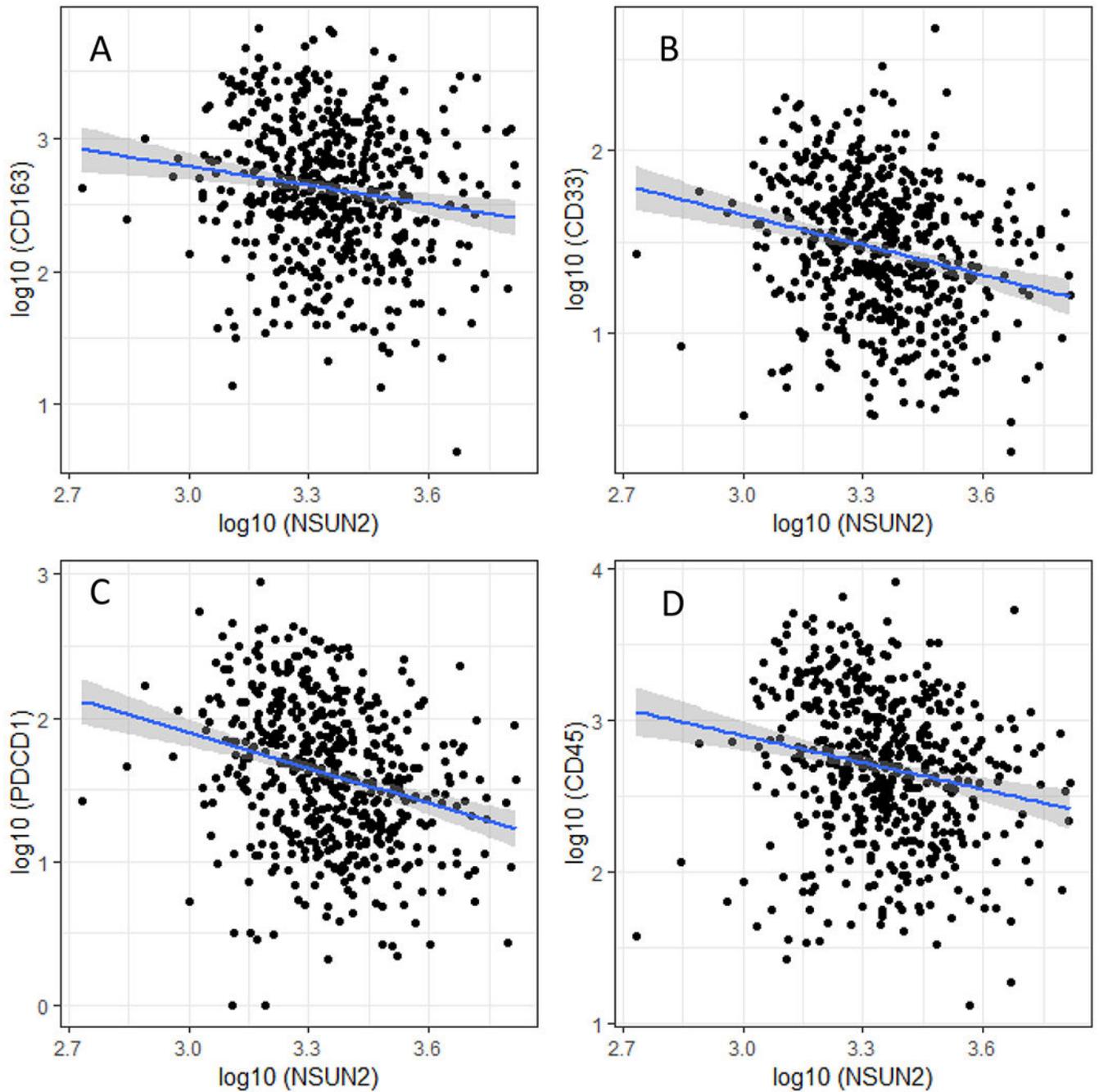
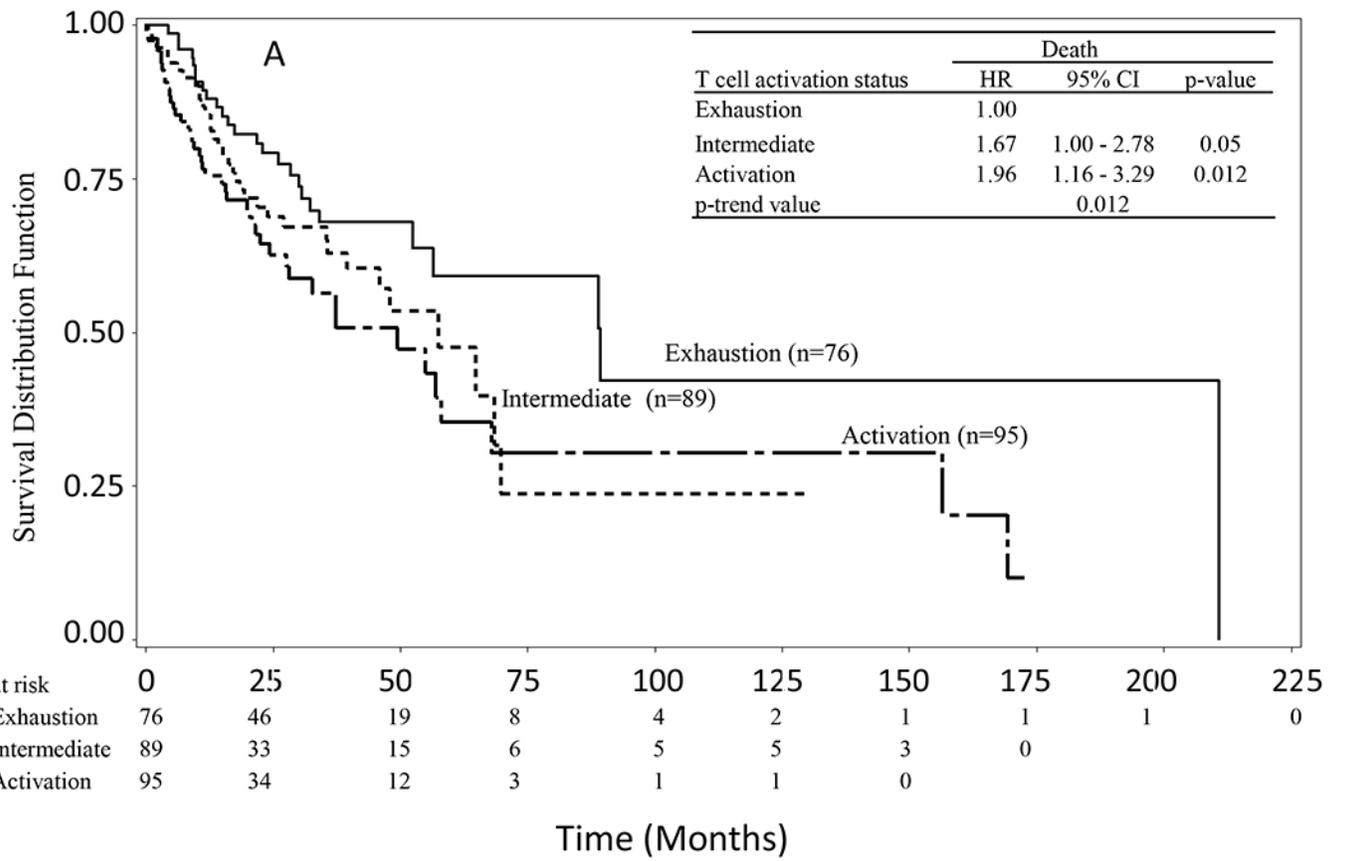


Figure 1. Scatter plot of *NSUN2* expression with the markers for macrophage and myeloid cells. Expression (\log_{10} FPKM) of *NSUN2* is plotted against A) *CD163*, B) *CD33*, C) *PDCD1* and D) *CD45*. The linear trend line is in blue, and the grey bound represents the 95% confidence interval.



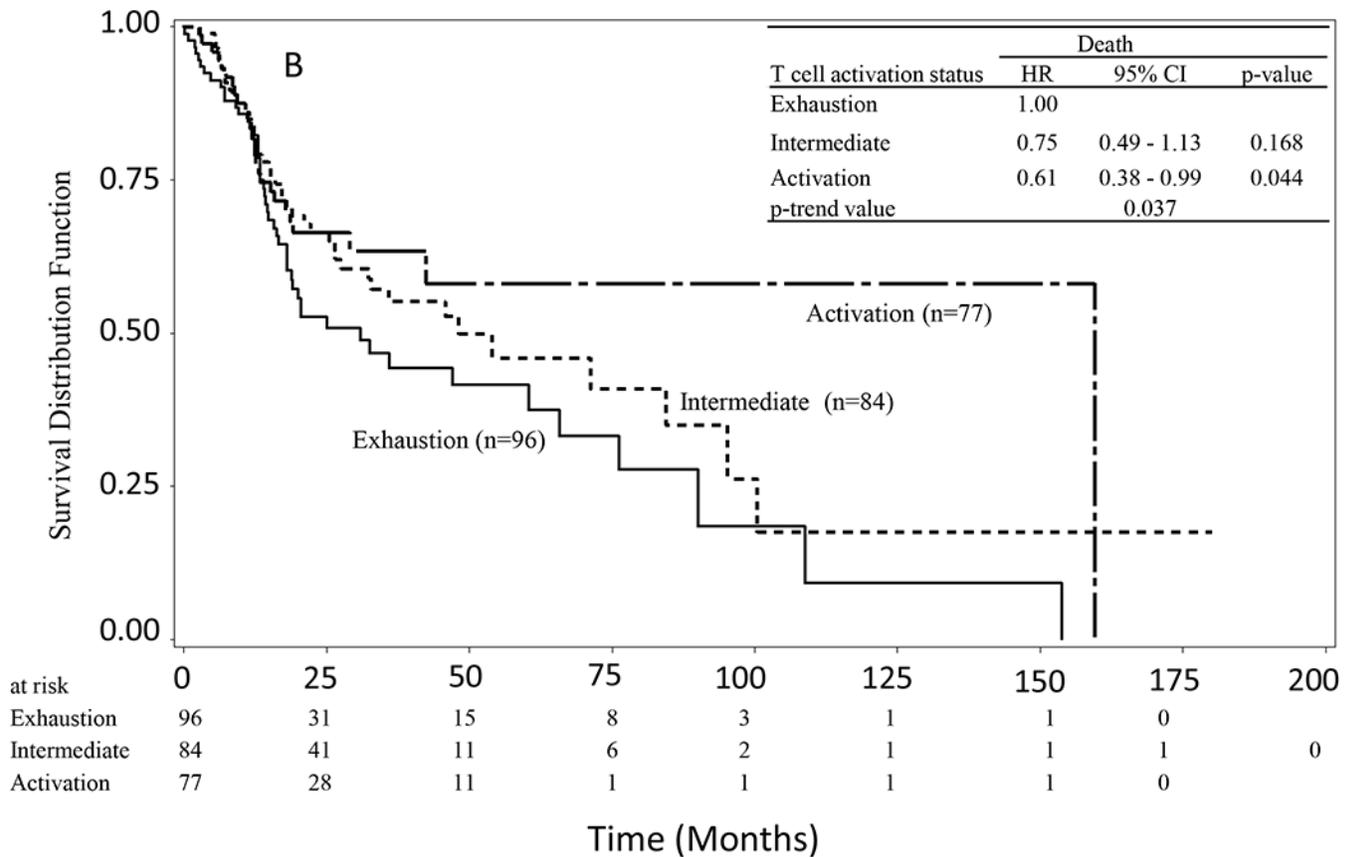


Figure 2.

Kaplan-Meier survival curves of HNSCC stratified by T-cell activation status (score). A) In the subgroup with a low expression of *NSUN2*, patients in the Activation group had reduced overall survival compared to those in the Exhaustion group ($P=0.012$). B) In the subgroup with a high expression of *NSUN2*, patients in the Activation group had longer overall survival than those in the Exhaustion group ($P=0.037$).

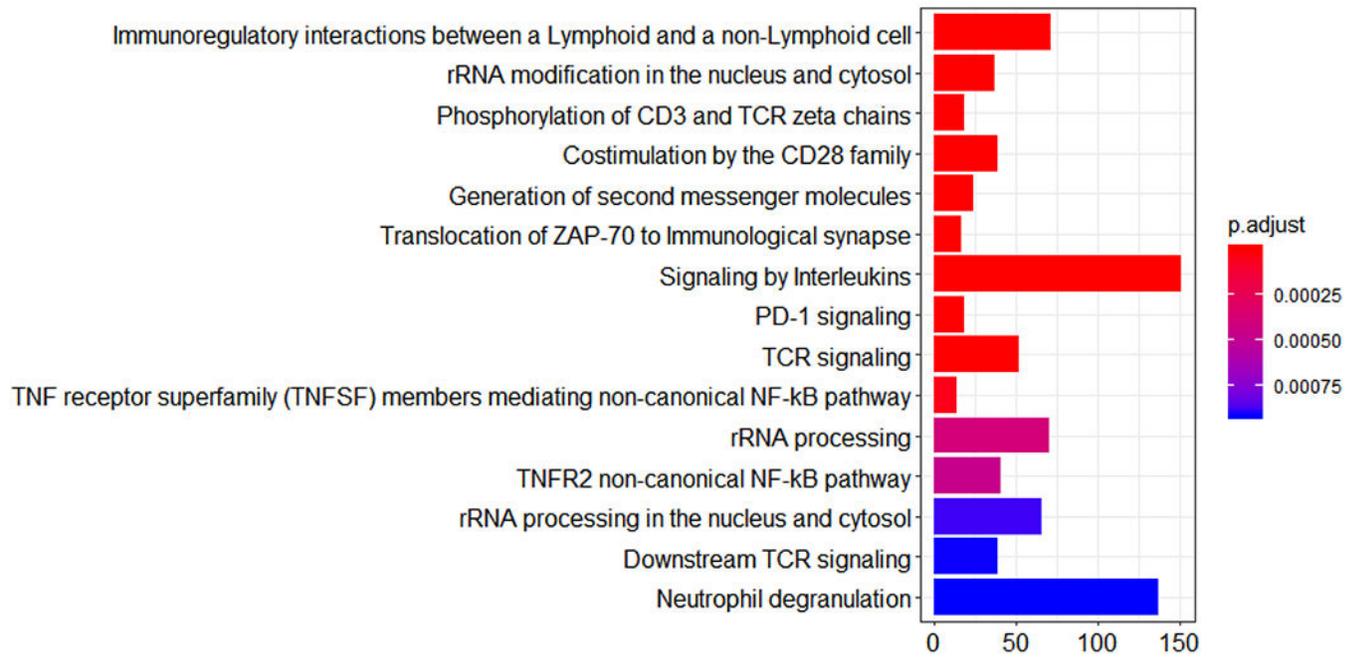


Figure 3.

Bar plot of the pathways enriched for genes that are coexpressed with *NSUN2*. The size of the bar quantifies the number of genes in the pathway that are significantly coexpressed with *NSUN2*.

Table 1.Distribution of the expression level of *NSUN2* and the T-cell activation score, and their spearman correlation

Variable	<i>n</i>	Mean	SD ^a	Median	Range
<i>NSUN2</i>	520	2395	969	2221	(540, 6577)
T cell activation score	520	0.21	0.09	0.21	(-0.01, 0.50)
Correlation coefficient (95% CI ^b)				-0.14	(-0.22, -0.05)
<i>P</i> value				0.002	

^{1.}SD: standard deviation^{2.}CI: confidence interval

Table 2.Spearman correlation between *NSUN2* and markers of macrophage and myeloid cells

Gene	<i>n</i>	Correlation coefficient	95% CI ^a	<i>P</i> value
<i>CD163</i>	520	-0.16	(-0.24, -0.07)	0.0003
<i>CD33</i>	520	-0.27	(-0.34, -0.18)	<0.0001
<i>PDCD1</i>	520	-0.31	(-0.47, -0.22)	<0.0001
<i>CD45</i>	520	-0.25	(-0.32, -0.16)	<0.0001

^aCI: confidence interval

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Table 3.T-cell activation status and mortality from HNSCC, stratified by expression level of *NSUN2*

Stratification		Death		
Variable	Variable	HR ^a	95% CI ^b	P value
Model 1	T-cell activation			
	Exhaustion	1.00		
	Intermediate	1.08	0.77–1.50	0.664
	Activation	1.03	0.73–1.47	0.852
	<i>P</i> (for trend)		0.84	
Low <i>NSUN2</i>	T-cell activation			
	Exhaustion	1.00		
	Intermediate	2.01	1.15–3.52	0.015
	Activation	2.16	1.22–3.82	0.008
	<i>P</i> (for trend)		0.009	
High <i>NSUN2</i>	T-cell activation			
	Exhaustion	1.00		
	Intermediate	0.74	0.48–1.14	0.174
	Activation	0.57	0.34–0.96	0.033
	<i>P</i> (for trend)		0.028	
<i>P</i> value for the interaction between <i>NSUN2</i> level and T-cell activation		0.002		
Model 2	T-cell activation			
	Exhaustion	1.00		
	Intermediate	1.97	1.12–3.47	0.019
	Activation	2.06	1.16–3.68	0.014
	<i>P</i> (for trend)		0.016	
High <i>NSUN2</i>	T-cell activation			
	Exhaustion	1.00		
	Intermediate	0.77	0.49–1.19	0.240
	Activation	0.61	0.36–1.03	0.063
	<i>P</i> (for trend)		0.056	
<i>P</i> value for the interaction between <i>NSUN2</i> level and T-cell activation		0.004		

^{1.}^aHR: adjusted hazard ratio, which was obtained from a multivariate Cox proportional hazards regression model with covariates of patient age at diagnosis (per 5 yrs), disease stage, tumor grade, gender (male vs female), smoking status (yes vs no) and primary tumor site (tongue, pharynx and other) in the model 1 (n =247 for low *NSUN2* group and n =240 for high *NSUN2*). In model 2, HPV status (positive vs negative) was also included as a covariate (n = 237 for low *NSUN2* group, and n =234 for high *NSUN2*).

^{2.}^bCI: confidence interval.