SARS-CoV-2-related myocarditis-like syndromes Shakespeare's question: what's in a name?

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The current pandemic coronavirus disease 2019 (COVID-19) is primary a respiratory infection caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Epidemiological data from China show that approximately 20% of COVID-19 patients have concomitant established cardiovascular diseases and are more likely to develop life-threatening complications in the course of infection. In some cases, the development of myocarditis, a cardiac disorder characterized by inflammatory cell infiltration of the heart and greater risk of deterioration of cardiac function, has been clinically suggested. However, endomyocardial biopsy (EMB) analysis as the gold standard diagnostic tool to verify the clinical diagnosis and to understand the underlying pathomechanisms at the cellular level, had rarely been used.

To the best of our knowledge, only 11 single cases, including the case of Tavazzi et al.,4 have been reported so far of probable SARS-CoV-2-associated myocarditis (Table 1).4-14 In most of the cases, a SARS-CoV-2-related myocarditis was clinically suspected based on cardiac biomarkers, combined with echocardiography and electrocardiogram, and in five cases combined with magnetic resonance imaging findings. One group also provided a case in the absence of any clinical symptoms or cardiac functional limitations. 12 In a post-mortem examination, endothelial inflammation was found in a patient with suspected myocarditis.¹⁴ In the same case series, global endotheliitis was associated with viral inclusion structures in endothelial cells in severe COVID-19-diseased patients. This indicates that the findings and definitions are not completely consistent and that several clinical-based 'myocarditis-like syndromes' are described. Thus, in most of these reports, no pathophysiological confirmation of the suspected diagnosis occurred. Only in one report, EMB analysis was performed, describing a mild lymphocytic myocarditis in the absence of myocardial SARS-CoV-2 RNA presence.⁹

In the previous issue of the Journal, Tavazzi et al.4 provide the first case of EMB-proven localization of viral particles in the heart with morphology and size typical of SARS-CoV-2 in a COVID-19 patient presenting with cardiogenic shock. Viral particles are observed in interstitial macrophages and their surroundings, but not in cardiomyocytes or endothelial cells, excluding the direct damage of the heart due to virus replication. In contrast to the clinical presentation suggestive for severe and necrotizing acute myocarditis, only low-grade myocardial inflammation and absence of myocyte necrosis was observed. This is in agreement with Xu et al. 15 who also only found low-grade inflammatory responses in cardiac autopsy species of a COVID-19 patient with acute respiratory distress syndrome. Furthermore, cardiac myocytes showed only non-specific features such as myofibrillar lysis, and no signs of myocyte hypertrophy. There were no indications of vasculitis or thrombosis, and fibrosis was mainly perivascular.

This case report is of important value since it demonstrates for the first time the cardiac presence of SARS-CoV-2 in a COVID-19 patient with cardiogenic shock. Despite the tropism of SARS-CoV-2 for angiotensin-converting enzyme 2 (ACE2), which is expressed by cardiomyocytes, fibroblasts, endothelial cells and particularly by pericytes, ¹⁶ viral particles were interestingly not found in these cells, but in macrophages, which may have reached the heart during transient viremia or by migration, e.g. from the infected lung. The cellular serine proteases TMPRSS2, which is co-expressed on lung cells expressing ACE2, and essential for viral entry, ¹⁷ is not co-expressed in cardiomyocytes, pericytes and fibroblasts, ¹⁸ suggesting also its relevance for viral uptake in cardiac cells, without excluding the potential need of other co-receptors like cathepsin B and L, for spike protein priming. Though, the

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Table 1	Table 1 Case reports of myocarditis-like syndromes	ol IIIyocarun								
Study	COVID-19 nasopharyngeal swab	MC diagnosis after clinical onset	Clinical parameters	Cardiac biomarker	Æ	ω Σ	Ш В			Diagnosis
					-		Histology	Immunohistology	Viral PCR or EM	
Inciardi et al. ⁵		7 days	No respiratory involvement	Troponin: elevated	Yes (T1/T2 pos)	o Z	1	ı	1	COVID-19 myo-pericarditis
Zeng et al. ⁶	Positive (Cycle threshold)	~-	EF 33% ARDS, shock LVEDD 61mm EF 32%	N - probin: elevated Troponin: elevated NT-proBNP: elevated	° Z	°Z	1	1	I	COVID-19-complicated fulminant myocarditis
Hu et al.7	Positive	3 days	Er 32.% Pneumonia, shock LVEDD 58 mm	Troponin: ? BNP elevated	° Z	°Z	I	1	I	Coronavirus infection-caused fulminant myocarditis
Kim et al. ⁸	Positive	~-	Er 27 % Pneumonia, severe LV systolic dvefunction	Troponin: elevated	Yes (T1/T2 pos)	°Z	1	1	1	COVID-19-related myocarditis
Sala et al.º	Positive	3 days	Pneumonia, no LV dilatation EF 43%	Troponin: elevated NT-proBNP: elevated	Yes (T1/T2 pos)	Yes	×	Mild inflammation	Negative for SARS-CoV-2	Acute virus-negative lymphocytic myocarditis associated with SARS-CoV-2
Tavazzi et al. ⁴	Positive	4 days	ARDS, shock LVEDD 56 mm EF 34%	Troponin: elevated NT-proBNP: ?	o Z	Yes	Dallas criteria negative	Low-grade inflammation	SARS-CoV-2 particle detection in macrophages, no myocyte necrosis in	Low grade myocarditis in COVID-19 cardiogenic shock
Irabien-Ortiz et al. ¹⁰	Positive	5 days	Shock, severe biventricular fail ure	Troponin: elevated NT-proBNP: elevated	o Z	°Z	ı	ı		Fulminant COVID-19 myocarditis
Hua et al. ¹¹	Positive	~-	Shock due to LV perforation Normal EF	Troponin: elevated NT-proBNP: ?	o Z	° Z	ı	ı	ı	COVID-19 myo-pericarditis
Paul et al. ¹²	Positive	~-	Absolutely no clinical signs Normal EF	Troponin: elevated NT-proBNP: ?	Yes	o Z	ı	ı	I	COVID-19 myocarditis
Doyen et al. ¹³	positive	~:	ARDS Normal EF	Troponin: elevated NT-proBNP: ?	Yes	°Z	ı	ı	ı	COVID-19 myocarditis
Varga et al. ¹⁴	Positive	N/A	Severe hypoxic failure Preserved EF	Troponin: elevated NT-proBNP: elevated	o Z	Post-mortem tissue	Endothelial inflammation in heart, small bowel lung	ı	SARS-CoV-2 particle detection in renal endothelial cells in EM PCR not provided	Global endotheliitis in COVID-19
	Positive	16 days	ARDS Normal EF	Troponin: elevated NT-proBNP:?	° Z	Post-mortem tissue	Endothelial inflammation in lung, heart, kidney and liver	I		

_, not performed; ARDS, acute respiratory distress syndrome; COVID-19, coronavirus disease 2019; EF, ejection fraction; EM, electron microscopy; EMB, endomyocardial biopsy; UX, left ventricular; UEDD, left ventricular end-diastolic dimension; MC, myocardisis; MRI, magnetic resonance imaging; N/A, not applicable; NT-proBNP, N-terminal pro brain natriuretic peptide; POR, polymerase chain reaction; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

924 Editorial comment

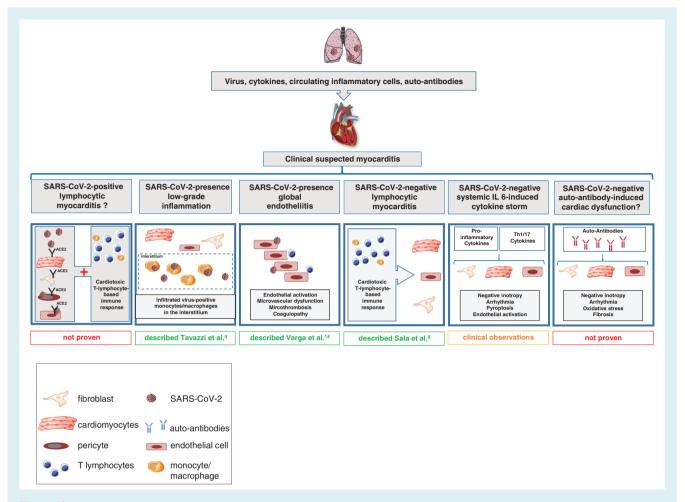


Figure 1 Hypothetical cardiac inflammatory-like pathomechanisms triggered by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). So far, different cardiac inflammatory-like pathomechanisms triggered by SARS-CoV-2 have been suggested: (i) lymphocytic myocarditis induced by SARS-CoV-2 infection of cardiac cells (SARS-CoV-2-positive lymphocytic myocarditis), (ii) cardiac SARS-CoV-2 presence with low-grade inflammation, (iii) cardiac SARS-CoV-2 presence and global endotheliitis, (iv) lymphocytic myocarditis in the absence of cardiac infection of SARS-CoV-2 (SARS-CoV-2-negative lymphocytic myocarditis), (v) cardiac injury elicited by a SARS-CoV-2 triggered interleukin (IL)-6-induced cytokine storm, and (vi) cardiac dysfunction due to auto-antibodies in the absence of SARS-CoV-2. Differentiation of cardiac inflammatory-like pathologies triggered by SARS-CoV-2 is required to allow specific treatment strategies (antiviral vs. immunosuppressive/modulating). Therefore, biomarkers are needed and model systems, including basic virology studies.

absence of viral particles in cardiomyocytes, pericytes and fibroblasts seen in this patient may not be generalized, not knowing the cardiac (cellular)-specific expression levels of ACE2 in this patient, the clinical history of this patient — ACE2 is increased in heart failure patients, ¹⁹ — and the SARS-CoV-2 viral load in the heart or other organs (lung). Investigations in model systems are needed to assess correlations between ACE2 and co-receptor expression and viral uptake in specific cardiac cells and to evaluate whether SARS-CoV-2 can damage cardiac cells dependent or independent of virus replication in the myocardium.

The low-grade cardiac inflammation on the one hand, and the presence of viral particles only in macrophages, as source of inflammation on the other hand, suggest that virus presence in the heart of COVID-19 patients is rare. This advocates that the clinical

presentation of fulminant myocarditis is rather caused by an indirect, e.g. via a cytokine storm, than a direct SARS-CoV-2-mediated viral effect, or even by other concomitant diseases/infections. It further supports the relevance to exclude SARS-CoV-2 in the blood and the need to quantify SARS-CoV-2 copy numbers in EMB, allowing comparison with other cases and primary with virus load in nasopharyngeal swab material, to enable interpretation of the clinical and laboratory observations and to assess the severity of the infection and viremic response.

Despite the clinical manifestation similar to this of fulminant myocarditis, no lymphocytic myocarditis in heart tissue was diagnosed. This resembles the clinical presentation by influenza-associated septic shock, which also mimics fulminant myocarditis, but occurs in the absence of inflammatory cell infiltration or necrosis of cardiomyocytes.²⁰ Acute respiratory

infections as well as sepsis are often associated with a rise in troponin, indicative for cardiac injury²¹ and in frame with the present case.

This case report underpins the importance of performing EMB analysis for quantitative SARS-CoV-2 RNA evaluation and specific immune cell quantification by immunohistological staining to enable correct diagnosis and insights in the potential association, or not, between SARS-CoV-2 and myocarditis. Since myocarditis is a cardiac disorder with a heterogeneous aetiology, associations between SARS-CoV-2 and myocarditis based on sole cardiac imaging should be interpreted with caution. At the same time, this case as an example of a SARS-CoV-2-induced septic cardiomyopathy does not exclude the occurrence of SARS-CoV-2-associated cardiac inflammation. Beyond in vitro and in vivo models enabling further insights related to direct or indirect (cytokine storm, autoantibody production) SARS-CoV-2-mediated injury and/or cardiac immune cell presence, profound characterization and follow-up of COVID-19 patients are needed. In patients with suspected SARS-CoV-2-associated myocarditis, this includes quantitative evaluation of SARS-CoV-2 RNA in nasopharyngeal swab, and in EMB in combination with immunohistological analysis of EMB according to European Society of Cardiology guidelines³ and biomarker analysis. In COVID-19 patients without any signs of cardiovascular involvement, SARS-CoV-2 copy numbers in nasopharyngeal swab and the collection of plasma/serum for potential subsequent (antibody) analyses are needed, to rule out the potential occurrence of myocarditis due to a SARS-CoV-2-triggered autoantibody production, which might take place 3 to 4 weeks post-infection.

In conclusion, this case reports for the first time cardiac presence of SARS-CoV-2 in a COVID-19 patient with cardiogenic shock mimicking fulminant myocarditis. Many questions about COVID-19-related heart disease remain unanswered, calling for profound COVID-19 patient characterization and model systems, stressing also basic virology studies enabling to study the pathogenesis of multifaced SARS-CoV-2-related heart disease and to differentiate cardiac inflammatory-like pathologies triggered by SARS-CoV-2 (*Figure 1*). These insights are needed to allow appropriate therapeutic regimens (antiviral vs. immunosuppressive/immunomodulatory).

Conflict of interest: none declared.

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