

Circulating angiogenic cells in glioblastoma: toward defining crucial functional differences in CAC-induced neoplastic versus reactive neovascularization

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Abstract

Background. In order to identify suitable therapeutic targets for glioma anti-angiogenic therapy, the process of neovascularization mediated by circulating angiogenic cells (CACs) needs to be scrutinized.

Methods. In the present study, we compared the expression of neovascularization-related genes by 3 circulating CAC subsets (hematopoietic progenitor cells [HPCs], CD34⁺, and KDR⁺ cells; internal controls: peripheral blood mononuclear cells and circulating endothelial cells) of treatment-naïve patients with glioblastoma (GBM) to those of patients undergoing reactive neovascularization (myocardial infarction (MI)). CACs from umbilical cord (representing developmental neovascularization) and healthy subjects served as controls. Fluorescent-activated cell sorting was used to isolate CACs, RT-PCR to determine the expression levels of a panel of 48 neovascularization-related genes, and Luminex assays to measure plasma levels of 21 CAC-related circulating molecules.

Results. We found essential differences in gene expression between GBM and MI CACs. GBM CACs had a higher expression of proangiogenic factors (especially, *KITL*, *CXCL12*, and *JAG1*), growth factor and chemotactic receptors (*IGF1R*, *TGFBR2*, *CXCR4*, and *CCR2*), adhesion receptor monomers (*ITGA5* and *ITGA6*), and matricellular factor *POSTN*. In addition, we found major differences in the levels of neovascularization-related plasma factors. A strong positive correlation between plasma MMP9 levels and expression of *CXCR4* in the CAC subset of HPCs was found in GBM patients.

Conclusions. Our findings indicate that CAC-mediated neovascularization in GBM is characterized by more efficient CAC homing to target tissue and a more potent proangiogenic response than in physiologic tissue repair in MI. Our findings can aid in selecting targets for therapeutic strategies acting against GBM-specific CACs.

Key Points

- Glioblastoma CACs have a more potent homing and angiogenic capacity than controls.
- CACs are programmed in the circulation by target tissue-specific requirements.
- Unique CAC characteristics in different diseases translate to therapeutic targets.

High-grade gliomas are among the most vascularized tumors and are characterized by an abundance of leaky vessels. Despite the high degree of vascularization, anti-angiogenic therapies have remained without the expected success.¹ Anti-angiogenic

drugs like bevacizumab interfere with Vascular Endothelial Growth Factor A (VEGFA) and the process of sprouting angiogenesis. However, the contribution of circulating cells engaged in the formation of blood vessels may be overlooked as

Importance of the Study

Prior literature on circulating angiogenic cells (CACs) in glioblastoma (GBM) uncovered their potent proangiogenic effects *in vitro/vivo* and their increased numbers in GBM patients. Our study is the first to show that GBM CACs are qualitatively different from non-neoplastic CACs (ie, in reactive [myocardial infarction], developmental [umbilical cord blood], and steady-state adult [healthy control] neovascularization). GBM CACs exhibit a gene expression profile compatible with increased tumor-homing capacity (higher expression of *CXCR4*, *CCR2*, *ITGA5*, and *ITGA6*) and a more potent proangiogenic

potential (higher expression of *KITL*, *CXCL12*, *JAG1*, *IGF1R*, *TGFBR2*, and *POSTN*). Plasma levels of tumor-derived mobilization factor MMP9 correlate positively with both circulating hematopoietic progenitor cell (HPC) levels and HPC *CXCR4* gene expression in GBM patients, illustrating that GBM tissue is capable of pre-programming CACs. GBM, though non-metastatic, should thus be considered a systemic disease requiring systemic treatment. Our results can be translated toward developing disease-specific therapies targeting CAC-induced neovascularization in GBM.

a significant component of neovascularization in gliomas. This could partially explain the failing of anti-angiogenic therapies in glioma patients. Vasculogenesis is defined as *de novo* formation of blood vessels by endothelial progenitor cells (EPCs) that differentiate into endothelial cells and become part of the newly formed vessel wall.² Although characteristic for embryogenesis, the process of vasculogenesis also contributes to neovascularization in adults.³ Whereas in embryogenesis differentiation into endothelial cells by EPCs is widespread, this process is limited in adulthood.⁴ In adulthood, circulating cells stimulate neovascularization by invading the target tissue and secreting proangiogenic factors that fuel angiogenesis⁴. Since these cells do not differentiate into endothelial cells, they do not fit the definition of EPC and are better termed “circulating angiogenic cells” (CACs). Various stages of CAC-mediated neovascularization exist. CACs are mobilized from the bone marrow by factors secreted by the target tissue and/or bone marrow microenvironment, or in an autocrine fashion by CACs themselves. In the bloodstream CACs migrate towards the target tissue through chemotaxis where they adhere to endothelial cells mediated by integrins and invade the tissue by expressing proteinases such as matrix metalloproteases (MMPs). Once in the target tissue CACs differentiate and start to secrete growth factors thus creating an environment permissive for angiogenesis.

In adulthood, neovascularization is stimulated on demand and is activated during revascularization after trauma or ischemia. In myocardial infarction (MI), a well-described and potent mobilization of CACs is induced early after the ischemic event.⁵ Other ischemic states, such as ischemic stroke, have been less extensively studied. The literature on CACs in ischemic stroke shows less consistent results regarding the mobilization of CACs, with some studies showing no increase^{6,7} or even a decrease of CACs.⁸ Since the CAC response to ischemic brain appears to be far less extensive than to ischemic myocardium,⁹ we chose to use MI patients rather than stroke patients as representing CAC-induced neovascularization in response to ischemia.

While in MI revascularization aids in recovery, new blood vessels in tumors are associated with propagation and

contribute to the decrease of the organism.¹⁰ In patients suffering from MI, CAC-based therapies have been implemented with promising results.¹¹ In cancer, however, CAC-directed therapies have only been applied in animal studies where significant decreases in tumor sizes were reached.¹² Little is known about functional differences in CAC trafficking and function in the contexts of acute ischemia, cancer, and development. A better understanding of CAC biology in these different situations is necessary to design therapies acting on CAC-related neovascularization in cancer.

Here we compared the expression in CAC subsets of genes involved in neovascularization of glioblastomas (GBMs) and MI. Umbilical cord blood (UCB) and blood from adult healthy controls (HC) served as references for embryonic/fetal and steady-state adult neovascularization, respectively. Genes and 21 circulating plasma factors were chosen based on their functional roles (mobilization, chemo-attraction, homing, and growth factors secretion).¹³ The expressional profiles of the respective CACs and the plasma factors of patients with GBM and MI were compared and correlated. The findings show profound differences between CAC-mediated neovascularization in GBM and MI patients.

Material and Methods

This study was approved by the Medical Ethics Committee of the Erasmus Medical Center, Rotterdam, The Netherlands (MEC-2011-313) and carried out in adherence to the Code of Good Conduct of the Federation of Medical Scientific Societies in the Netherlands (<http://www.federa.org/codes-conduct>). Informed consent was obtained from all subjects.

Blood Samples and Preparation: See [Supplementary Materials and Methods](#).

Selection and FACS Sorting of CAC Subsets: See [Supplementary Materials and Methods](#).

RNA Isolation and RT-PCR and Gene Expression Analysis: Quality Control: See [Supplementary Materials and Methods](#).

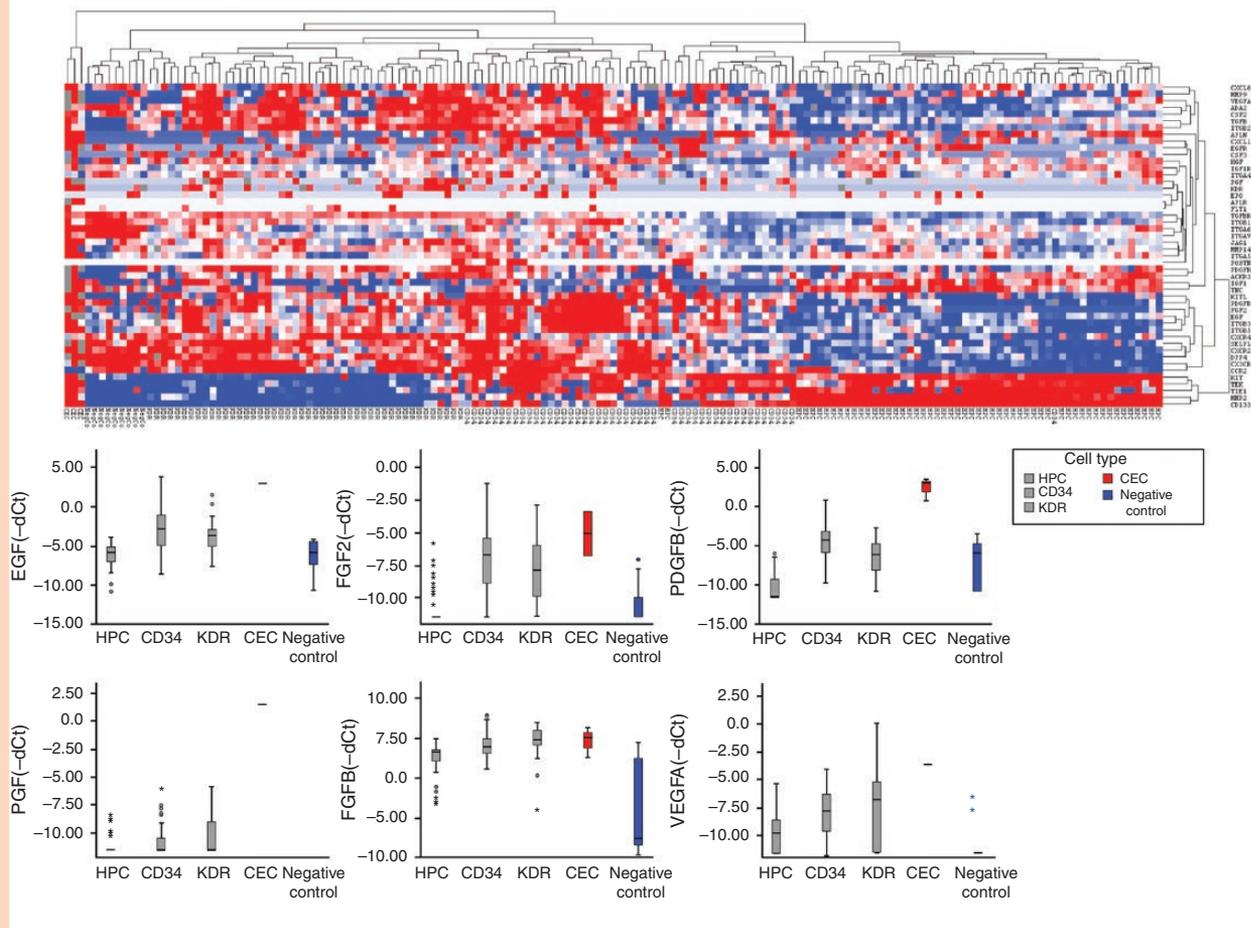


Figure 1. Unsupervised hierarchical cluster analysis of gene expression in all samples and boxplots of expression levels. *Upper panel:* Unsupervised hierarchical cluster analysis of gene expression in all samples (city block distance with complete linkage). Blue = low expression and red = high expression. Clustering is seen based on CAC type: CECs display the most conspicuous phenotype (high expression). CD34⁺ cells partially cluster with HPCs and partially with KDR⁺ cells. Negative control leukocytes cluster with KDR⁺ cells. The HPC cluster in general shows lower gene expression than the other CACs or CECs. *Lower panel:* Boxplots showing gene expression levels (-dCt) of proangiogenic factors in HPCs ($n = 54$), CD34⁺ cells ($n = 47$), KDR⁺ cells ($n = 46$), CECs ($n = 3$), and negative control PBMCs ($n = 9$). Proangiogenic factors overall are expressed highest in CECs and lowest in negative control PBMCs. In general, CD34⁺ and KDR⁺ cells express higher levels of proangiogenic factors than HPCs.

RT-PCR Data Analysis: See [Supplementary Materials and Methods](#).

Data Analysis: See [Supplementary Materials and Methods](#).

Results

Hierarchical Cluster Analysis: Gene Expression Patterns of CAC Subsets From All Subjects

The expression patterns of the CAC subsets, negative control peripheral blood mononuclear cells (PBMCs), and circulating endothelial cells (CECs) in the various patient and control groups clustered according to the respective cell types (Figure 1). CECs expressed genes from almost all functional groups at a much higher level than the other CACs, except for chemotactic receptors, which were only

expressed at a higher level in CECs compared to hematopoietic progenitor cells (HPCs). HPCs showed relatively low overall expression of neovascularization-related genes. Overall expression levels of the investigated genes were lower in HPCs than in KDR⁺ cells, CD34⁺ cells, and CECs. HPCs were most homogenous regarding gene expression, irrespective of the source of the blood samples. CD34⁺ cells clustered with HPCs for growth factor receptors and *CD133* expression while they resembled KDR⁺ cells by their high expression of proangiogenic molecules and molecules operative in (de)adhesion and invasion. KDR⁺ cells clustered with negative control leukocytes for all functional groups, suggesting the closest kinship of all subsets investigated with negative control PMBCs. CACs from GBM patients expressed neovascularization-related genes at a higher level than those from MI patients or HC. Following unsupervised hierarchical cluster analysis on individual CAC subsets, we found that HPCs from UCB and MI clustered together, as opposed to GBM HPCs (Figure 2).

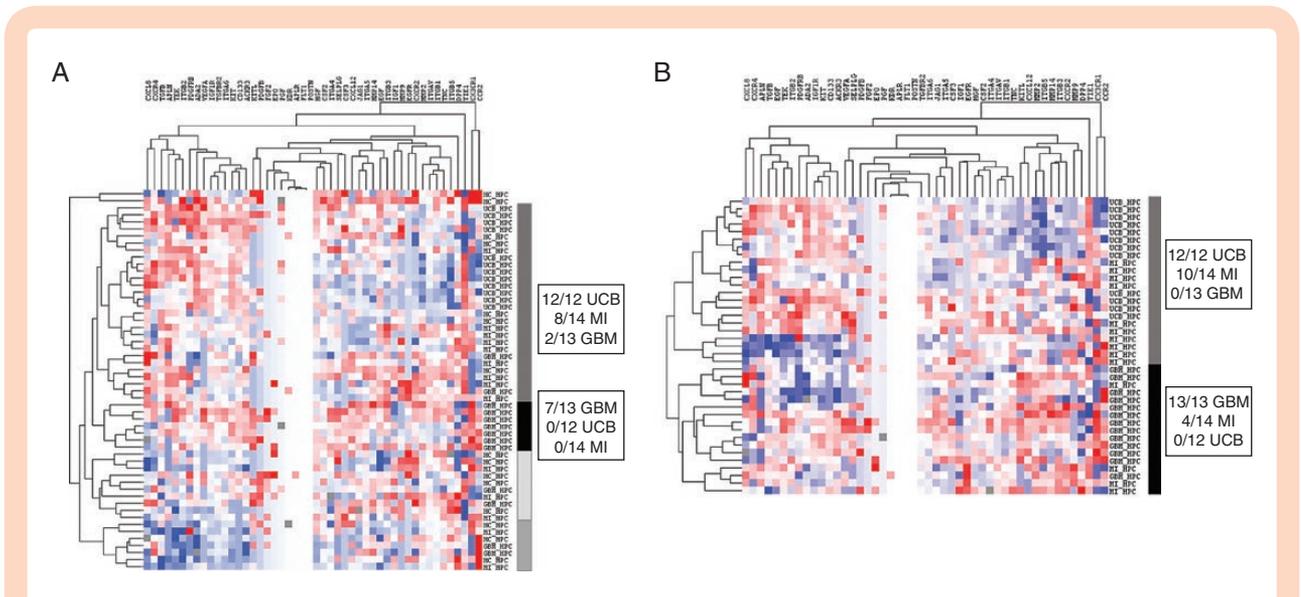


Figure 2. Unsupervised hierarchical cluster analysis of gene expression in HPCs. (A) Unsupervised hierarchical cluster analysis (city block distance, complete linkage) of gene expression in HPCs (all samples). UCB and MI HPCs cluster together, while GBM HPCs are in a separate cluster. There is higher overall gene expression in the GBM HPCs cluster compared to the other clusters. (B) Unsupervised hierarchical cluster analysis of HPCs (city block distance, complete linkage) after removing HC samples from the analysis: clustering of UCB and MI HPCs is more obvious. There is higher gene expression in the GBM HPCs cluster compared to the other clusters.

Differences in Expression of Individual Genes in CACs Between GBM and MI Patients

The genes that showed differential expression in CACs between the GBM and MI group represented all distinct functional groups (Figures 3–5). *CXCR4* and *KITL* were overexpressed in all CAC subsets of GBM patients as compared to patients with MI. Reversely, *IGF1* was underexpressed in GBM compared to MI. Higher RNA levels of *APLN* were detected in MI CACs as compared to GBM, while *CXCL12* and *ITGA5* transcript levels were lower in MI. The activity of some genes was consistently different for all CAC subtypes (eg, *CXCR4* was overexpressed in GBM HPCs, CD34⁺, and KDR⁺ cells as compared to these cells in MI), while the differential activity of other genes appeared to be confined to specific CAC subtypes (eg, overexpression of *JAG1* in GBM vs MI HPCs only, not in CD34⁺ or KDR⁺ CACs (Figures 3–5). Deviations from the reference HC expression levels (whether upregulated or downregulated) consistently followed the direction of UCB gene expression levels with the exception of *KITL* expression in GBM CACs (upregulated in GBM, downregulated in UCB compared to HC (Figures 4 and 5).

Plasma Factors

In GBM patients the overall levels of all plasma factors were higher than those in MI patients and HC subjects. Unsupervised hierarchical cluster analysis of the concentrations of all plasma factors measured in all samples yielded 3 main clusters: one containing only UCB samples, one with the large majority of GBM and HC samples, and one with the large majority of MI samples (lower overall

levels of plasma factors) (Figure 6). Spearman correlation analysis between plasma factor concentrations and gene expression in CACs revealed a strong positive correlation between plasma MMP9 levels and the expression of *CXCR4* in HPCs in GBM patients (Spearman's rho = 0.77; $P < .01$). In MI patients no correlation between HPC *CXCR4* gene expression and plasma MMP9 levels was found (Supplementary Figure 3). When lowering the correlation threshold to at least 0.5, multiple significant correlations were detected between CAC gene expression and plasma factor levels (eg, a positive correlation between HPC *CSF2* gene expression and plasma *CXCL12* levels; positive correlation significant for both GBM and MI patients, not for HC).

Discussion

In the present study, we investigated alterations in the expression of neovascularization-related genes in circulating CAC subsets between GBM and MI patients and sought correlations with circulating chemo-attractants and mobilization factors. Where in previous studies we observed that levels of circulating CACs differ in GBM patients as compared to HC and patients suffering from recent MIs,^{7,14} in the present study we explored the expression of 48 neovascularization-related genes in 3 CAC subsets in these groups. We found major differences in expressional profiles. There was close similarity between the gene expression patterns of HPCs in MI and UCB, indicative of reactivation of embryonal/fetal mechanisms for CAC-mediated neovascularization following acute myocardial ischemia. In circulating CACs from GBM (where neovascularization is disordered and haphazard) this coordinated CAC

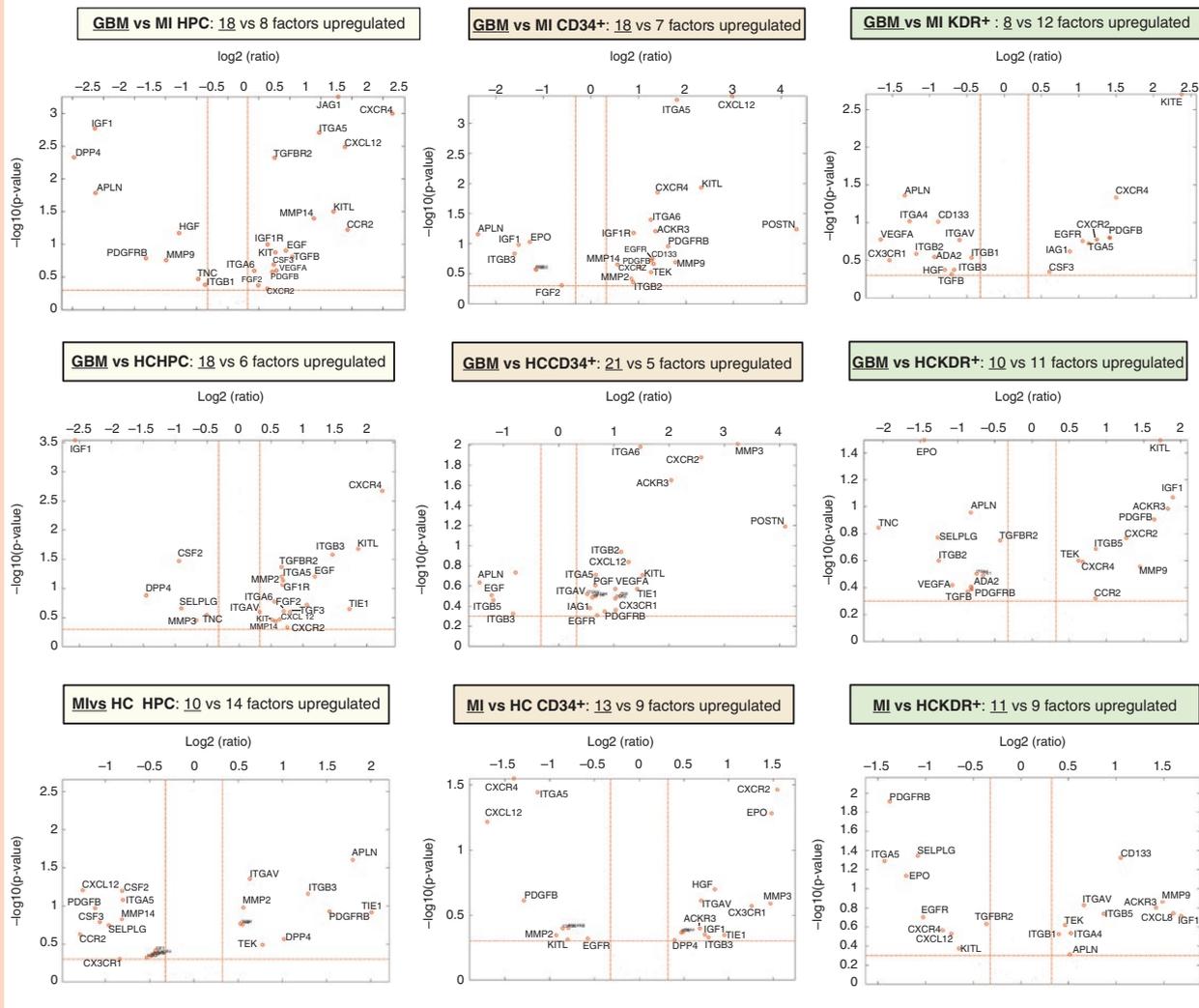


Figure 3. Volcano plots of gene expression differences between patients and controls by CAC subset. *Upper row (A–C):* Volcano plots ($-\log_{10} P$ -value vs \log_2 fold change (FC) with the following cutoff values: FC > 1.1251, $P < .5$) of GBM versus MI CACs. More genes are overexpressed in GBM versus MI CACs. Overexpressed genes belong to all functional groups. Specifically, there is higher expression in GBM versus MI CACs (especially, HPCs and CD34⁺ cells) of growth factor receptors (GFRs), chemotactic receptors (CRs), and mobilization factors (MFs). There is higher expression in GBM versus MI HPCs of proangiogenic factors (PAFs). Z-scores and P-values of gene expression in GBM versus MI CACs are given in Figure 5. *Middle row (D–F):* Volcano plots ($-\log_{10} P$ -value vs \log_2 FC with the following cutoff values: FC > 1.1251, $P < .5$) of GBM versus HC CACs. A similar overall pattern of higher gene expression is seen as in the comparison of GBM versus MI CACs. Overexpressed genes belong to all functional groups. Higher expression in GBM versus HC CACs (especially, HPCs and CD34⁺ cells) of GFRs, CRs, MFs, adhesion factors (ITGs), PAFs. *Lower row (G–I):* Volcano plots ($-\log_{10} P$ -value vs \log_2 FC with the following cutoff values: FC > 1.1251, $P < .5$) of MI versus HC CACs. Overall gene expression is similar/lower in MI CACs versus HC CACs. Lower expression is seen in MI versus HC HPCs for PAFs, CRs, and MFs.

gene expression program was absent. We also discovered significant variations in the concentrations of 21 neovascularization-related plasma factors between GBM and MI patients, reflecting considerable differences in the “microenvironment” of the peripheral circulation, in which circulating CACs reside. Furthermore, we found strong correlations between the levels of specific plasma factors and gene expression levels in CACs. Altogether, these findings suggest that the difference in “blood microenvironment” as a result of MI or neoplastic growth drives alterations in gene expression in circulating CACs.

HPCs are capable of trafficking back and forth between the bone marrow, peripheral blood, (extra)-medullary tissues, and the lymphatic system.¹⁵ We know from the literature that HPCs mobilized to peripheral blood have different gene expression profiles than bone marrow (BM)-resident HPCs.¹⁶ Hypothetically, residing in target tissues will alter HPC (and other CAC) gene expression profiles dependent on target tissue/lesion-specific microenvironments. Hence, another explanation for our findings of altered gene expression patterns in CACs between GBM and MI patients is the reentrance of CACs

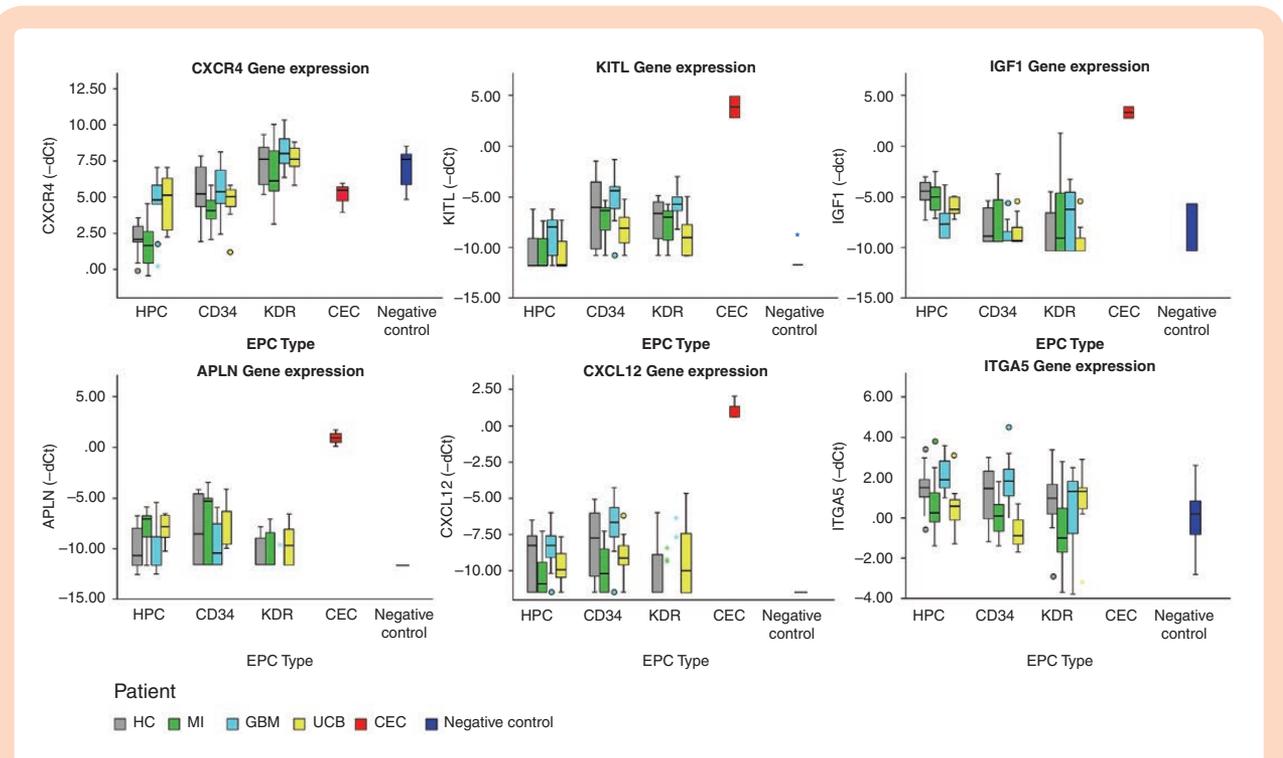


Figure 4. Boxplots of gene expression levels ($-dCt$) of significantly differentially expressed genes between GBM and MI CACs. Boxplots showing gene expression levels ($-dCt$) of significantly differentially expressed genes between GBM and MI CACs (data shown for gene expression differences present in ≥ 2 CAC subsets). *CXCR4* and *KITL* are overexpressed in GBM CACs compared to both MI and HC CACs. *IGF1* is underexpressed in GBM CACs (HPCs and CD34⁺) compared to MI and HC CACs. *ITGA5* is underexpressed in MI CACs compared to GBM and HC CACs. *APLN* is overexpressed in MI CACs compared to GBM and HC CACs. *CXCL12* is underexpressed in MI CACs compared to GBM and HC CACs. Deviations from the reference HC expressions levels (whether upregulated or downregulated) follow the pattern of UCB CAC gene expression levels (except for the overexpression of *KITL* in GBM CACs). For exact *P*-values and *Z*-scores for each CAC subtype, see Figure 5.

that were reprogrammed in such target tissues into the bloodstream. Differences in the trafficking speed of CACs between the bone marrow, peripheral blood, and target tissues can also contribute to changes in particular gene expression patterns¹⁶ and could be another factor contributing to our findings. The trafficking speed is dependent on various circumstances, such as levels of mobilization factors in the circulation and sympathetic innervation of BM.¹⁷ The latter could be altered in the presence of malignant glioma. Various combinations of cues like adhesion/chemotactic receptors, not single molecules themselves, drive the attraction and retention of HPCs to specific niches in the bone marrow.^{18,19} It is likely that similar cue patterns govern the attraction and retention of CACs to specific target tissues. We found that these cues differ in the context of GBM and MI, pointing to disease-driven alterations in gene expression in circulating CACs. The CAC gene expression profile in GBM patients suggests that they have a more potent capacity to home to GBM tissue and are capable of a stronger proangiogenic response than CACs in MI. Overall, the influence of GBM tumor tissue on circulating CAC biology justifies the notion that GBM should be considered as a systemic disease, rather than a disease which is limited to the brain.

The expression level of *CXCR4* in GBM CACs was similar to that in UCB CACs, but significantly higher than in MI CACs. *CXCR4* is a chemokine receptor expressed on the surface of leukocytes and HPCs.²⁰ *CXCR4* binds to its ligand

CXCL12, which acts as a mobilization factor and chemoattractant of *CXCR4*⁺ cells, including HPCs. Because *CXCL12* is highly expressed in GBM tumor cells, endothelial cells, neurons, and white matter we included this protein in our panel of plasma factors.^{21–24} We found decreased plasma *CXCL12* levels in both GBM and MI patients. The lower *CXCL12* levels in MI patients are in line with the existing literature,²⁵ while in glioma patients elevated, not reduced, plasma levels of *CXCL12* have been reported.²⁶ A technical explanation for the reported elevated levels could be the release of α -granule factors including *CXCL12* into plasma following blood sample cooling.^{26,27} In our study plasma values represent the free *CXCL12* fraction, not the platelet α -granule stored fraction. While high free plasma *CXCL12* mobilizes CACs from the bone marrow, homing of *CXCR4*⁺ cells to target tissues is less efficient due to the lower target tissue-to-plasma *CXCL12* ratio.^{28,29} Reversely, low plasma *CXCL12* levels allow for more efficient homing of *CXCR4*⁺ cells to *CXCL12*-expressing target tissues due to a high target tissue-to-plasma *CXCL12* ratio.³⁰ The low plasma level of *CXCL12* in GBM patients therefore facilitates homing of *CXCR4*⁺ cells to *CXCL12*-expressing GBM tissue.^{30,31} The present finding of increased expression of *CXCR4* in UCB HPCs was previously reported in the literature,³² but increased *CXCR4* expression in GBM HPCs (and other CACs) was not described earlier. Higher expression of *CXCR4* in cultured CACs increases migration triggered by *CXCL12* and enhances their capacity to exit blood vessels

	GBM>MI						GBM vs MI						GBM>HC						GBM vs HC						MI>HC						MI vs HC											
	MI>GBM		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR											
	PAF	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P													
PAF	APLN	-2.4	0.02	-1.9	0.05	-1.7	0.09	APLN	-0.6	0.54	-1.3	0.20	-1.4	0.15	APLN	2.4	0.02	0.4	0.66	0.8	0.45	CXCL12	2.8	0.01	3.3	0.00	0.1	0.93	CXCL12	0.6	0.53	1.3	0.19	-0.6	0.56	CXCL12	-1.7	0.08	-1.8	0.07	-0.8	0.41
	CXCL8	-0.2	0.82	-1.3	0.20	-0.8	0.40	CXCL8	1.0	0.34	0.1	0.91	0.2	0.83	CXCL8	1.4	0.15	1.7	0.09	1.3	0.20	EGF	0.8	0.45	-1.0	0.30	0.2	0.83	EGF	1.4	0.17	-0.9	0.37	1.7	0.09	EGF	0.3	0.74	0.4	0.71	0.5	0.64
	EGF	0.8	0.45	-1.0	0.30	0.2	0.83	EPO	0.0	1.00	-1.8	0.07	-1.3	0.20	EPO	0.1	0.94	2.2	0.03	-1.7	0.09	FGF2	0.0	1.00	-1.8	0.07	-1.3	0.20	FGF2	1.4	0.15	-0.2	0.88	0.5	0.64	FGF2	0.6	0.57	-0.1	0.95	-0.1	0.93
	EPO	0.0	1.00	-1.8	0.07	-1.3	0.20	FGF2	0.0	1.00	-1.8	0.07	-1.3	0.20	HGF	-1.8	0.08	0.2	0.82	-1.3	0.19	HGF	-1.8	0.08	0.2	0.82	-1.3	0.19	HGF	1.3	0.21	1.1	0.25	0.7	0.51	HGF	1.3	0.21	1.1	0.25	0.7	0.51
	FGF2	0.0	1.00	-1.8	0.07	-1.3	0.20	HGF	-1.8	0.08	0.2	0.82	-1.3	0.19	IGF1	-2.9	0.00	-1.3	0.18	0.2	0.82	IGF1	-2.9	0.00	-1.3	0.18	0.2	0.82	IGF1	-1.0	0.30	0.5	0.64	1.1	0.27	IGF1	-1.0	0.30	0.5	0.64	1.1	0.27
	HGF	-1.8	0.08	0.2	0.82	-1.3	0.19	JAG1	3.5	0.00	1.0	0.31	1.2	0.23	JAG1	3.5	0.00	1.0	0.31	1.2	0.23	JAG1	3.5	0.00	1.0	0.31	1.2	0.23	JAG1	-1.6	0.12	-0.4	0.67	-1.0	0.34	JAG1	-1.6	0.12	-0.4	0.67	-1.0	0.34
	IGF1	-2.9	0.00	-1.3	0.18	0.2	0.82	KITL	2.2	0.03	2.7	0.01	3.0	0.00	KITL	2.2	0.03	2.7	0.01	3.0	0.00	KITL	2.2	0.03	2.7	0.01	3.0	0.00	KITL	0.4	0.73	-0.7	0.48	-0.9	0.35	KITL	0.4	0.73	-0.7	0.48	-0.9	0.35
	JAG1	3.5	0.00	-1.3	0.18	0.2	0.82	PDGFB	2.0	0.31	1.7	0.08	1.6	0.11	PDGFB	2.0	0.31	1.7	0.08	1.6	0.11	PDGFB	2.0	0.31	1.7	0.08	1.6	0.11	PDGFB	-1.3	0.19	-1.2	0.23	0.2	0.85	PDGFB	-1.3	0.19	-1.2	0.23	0.2	0.85
	KITL	2.2	0.03	2.7	0.01	3.0	0.00	PGF	0.1	0.96	1.2	0.22	-0.3	0.76	PGF	0.1	0.96	1.2	0.22	-0.3	0.76	PGF	0.1	0.96	1.2	0.22	-0.3	0.76	PGF	1.0	0.32	0.1	0.95	-1.1	0.26	PGF	1.0	0.32	0.1	0.95	-1.1	0.26
	PDGFB	2.0	0.31	1.7	0.08	1.6	0.11	TGF	0.1	0.96	1.2	0.22	-0.3	0.76	TGF	0.1	0.96	1.2	0.22	-0.3	0.76	TGF	0.1	0.96	1.2	0.22	-0.3	0.76	TGF	-0.6	0.54	-0.1	0.95	-0.8	0.40	TGF	-0.6	0.54	-0.1	0.95	-0.8	0.40
PGF	0.1	0.96	1.2	0.22	-0.3	0.76	VEGFA	1.0	0.30	0.4	0.70	-1.4	0.17	VEGFA	1.0	0.30	0.4	0.70	-1.4	0.17	VEGFA	1.0	0.30	0.4	0.70	-1.4	0.17	VEGFA	-0.6	0.58	0.8	0.42	0.4	0.70	VEGFA	-0.6	0.58	0.8	0.42	0.4	0.70	
TGF	0.1	0.96	1.2	0.22	-0.3	0.76																																				
VEGFA	1.0	0.30	0.4	0.70	-1.4	0.17																																				
GFR	GBM vs MI						GBM vs HC						MI vs HC						MI vs HC																							
	HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR													
	GFR	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P													
	ADA2	0.3	0.74	0.3	0.74	-1.1	0.28	ADA2	-1.0	0.29	-0.3	0.80	-1.4	0.18	ADA2	-1.2	0.25	-0.9	0.39	0.2	0.87	APLR	0.0	1.00	0.0	1.00	0.0	1.00	APLR	-1.0	0.33	0.0	1.00	-1.0	0.30							
	APLR	0.0	1.00	0.0	1.00	0.0	1.00	APLR	-0.9	0.35	0.0	1.00	-0.9	0.36	APLR	-1.0	0.33	0.0	1.00	-1.0	0.30	CXCR4	3.1	0.00	2.4	0.02	1.9	0.06	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29							
	CXCR4	3.1	0.00	2.4	0.02	1.9	0.06	CXCR4	2.7	0.01	-0.1	0.90	0.9	0.39	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29	EGFR	0.2	0.81	1.1	0.29	1.4	0.15	EGFR	-0.2	0.82	-0.7	0.48	-1.2	0.25							
	EGFR	0.2	0.81	1.1	0.29	1.4	0.15	EGFR	0.0	0.98	0.5	0.64	0.2	0.88	EGFR	-0.2	0.82	-0.7	0.48	-1.2	0.25	FLT1	0.0	1.00	-1.1	0.26	-0.4	0.71	FLT1	0.0	1.00	1.0	0.30	1.4	0.17							
	FLT1	0.0	1.00	-1.1	0.26	-0.4	0.71	FLT1	0.0	1.00	0.0	1.00	1.1	0.27	FLT1	0.0	1.00	1.0	0.30	1.4	0.17	IGF1R	1.7	0.09	1.9	0.05	0.5	0.60	IGF1R	0.1	0.91	-0.4	0.69	-0.1	0.91							
	IGF1R	1.7	0.09	1.9	0.05	0.5	0.60	IGF1R	1.8	0.07	1.2	0.25	0.6	0.57	IGF1R	0.1	0.91	-0.4	0.69	-0.1	0.91	KDR	1.0	0.30	-0.8	0.43	-0.4	0.66	KDR	-1.0	0.33	0.7	0.47	0.6	0.57							
	KDR	1.0	0.30	-0.8	0.43	-0.4	0.66	KDR	0.2	0.88	-0.2	0.86	-0.1	0.94	KDR	-1.0	0.33	0.7	0.47	0.6	0.57	KIT	1.6	0.11	0.1	0.91	0.3	0.80	KIT	-0.5	0.63	0.0	1.00	0.1	0.93							
KIT	1.6	0.11	0.1	0.91	0.3	0.80	KIT	0.7	0.50	0.4	0.72	0.5	0.58	KIT	-0.5	0.63	0.0	1.00	0.1	0.93	PDGFRB	-1.1	0.27	1.6	0.11	0.8	0.44	PDGFRB	1.6	0.10	-1.1	0.28	-2.6	0.01								
PDGFRB	-1.1	0.27	1.6	0.11	0.8	0.44	PDGFRB	-0.5	0.65	0.6	0.55	-0.1	0.95	PDGFRB	1.6	0.10	-1.1	0.28	-2.6	0.01	TEK	0.4	0.70	1.0	0.30	0.2	0.81	TEK	1.0	0.34	-0.2	0.85	1.1	0.27								
TEK	0.4	0.70	1.0	0.30	0.2	0.81	TEK	1.2	0.22	1.2	0.25	1.2	0.22	TEK	1.0	0.34	-0.2	0.85	1.1	0.27	TGFBR2	2.7	0.01	1.0	0.30	-0.2	0.88	TGFBR2	0.0	0.98	-0.6	0.55	-1.0	0.34								
TGFBR2	2.7	0.01	1.0	0.30	-0.2	0.88	TGFBR2	2.1	0.04	0.3	0.78	-0.7	0.47	TGFBR2	0.0	0.98	-0.6	0.55	-1.0	0.34	TIE1	0.4	0.72	0.5	0.58	-0.7	0.51	TIE1	1.2	0.21	0.5	0.60	-0.5	0.58								
TIE1	0.4	0.72	0.5	0.58	-0.7	0.51	TIE1	1.3	0.18	1.1	0.28	-1.3	0.21	TIE1	1.2	0.21	0.5	0.60	-0.5	0.58																						
CR	GBM vs MI						GBM vs HC						MI vs HC						MI vs HC																							
	HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR													
	CR	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P													
	ACKR3	0.6	0.54	2.0	0.04	1.5	0.14	ACKR3	0.3	0.80	2.2	0.03	1.6	0.11	ACKR3	-0.6	0.53	0.9	0.38	0.9	0.37	CCR2	2.0	0.04	0.7	0.48	0.2	0.83	CCR2	-1.2	0.25	0.4	0.67	0.8	0.43							
	CCR2	2.0	0.04	0.7	0.48	0.2	0.83	CCR2	1.0	0.32	0.8	0.40	0.6	0.55	CCR2	-1.2	0.25	0.4	0.67	0.8	0.43	CX3CR1	0.7	0.46	-0.1	0.96	-0.8	0.44	CX3CR1	-0.2	0.86	1.5	0.13	0.7	0.46							
	CX3CR1	0.7	0.46	-0.1	0.96	-0.8	0.44	CX3CR1	0.4	0.71	0.8	0.44	-0.5	0.60	CX3CR1	-0.2	0.86	1.5	0.13	0.7	0.46	CXCR2	0.4	0.70	1.4	0.16	1.3	0.18	CXCR2	0.2	0.86	1.9	0.06	0.0	1.00							
CXCR2	0.4	0.70	1.4	0.16	1.3	0.18	CXCR2	0.6	0.68	2.6	0.01	1.5	0.13	CXCR2	0.2	0.86	1.9	0.06	0.0	1.00	CXCR4	3.1	0.00	2.4	0.02	1.9	0.06	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29								
CXCR4	3.1	0.00	2.4	0.02	1.9	0.06	CXCR4	2.7	0.01	-0.1	0.90	0.9	0.39	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29	SELPLG	0.1	0.94	0.7	0.48	0.4	0.71	SELPLG	-1.3	0.20	-0.2	0.85	-2.0	0.05								
SELPLG	0.1	0.94	0.7	0.48	0.4	0.71	SELPLG	1.2	0.24	0.3	0.80	-1.1	0.29	SELPLG	-1.3	0.20	-0.2	0.85	-2.0	0.05																						
DIF	GBM vs MI						GBM vs HC						MI vs HC						MI vs HC																							

	GBM>MI						GBM vs MI						GBM>HC						GBM vs HC						MI>HC						MI vs HC											
	MI>GBM		HPCs		CD34		KDR		HC>GBM		HPCs		CD34		KDR		HC>MI		HPCs		CD34		KDR		HC>MI		HPCs		CD34		KDR											
	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P												
PAF	APLN	-2.4	0.02	-1.9	0.05	-1.7	0.09	APLN	-0.6	0.54	-1.3	0.20	-1.4	0.15	APLN	2.4	0.02	0.4	0.66	0.8	0.45	APLN	2.4	0.02	0.4	0.66	0.8	0.45	APLN	2.4	0.02	0.4	0.66	0.8	0.45	APLN	2.4	0.02	0.4	0.66	0.8	0.45
	CXCL12	2.8	0.01	3.3	0.00	0.1	0.93	CXCL12	0.6	0.53	1.3	0.19	-0.6	0.56	CXCL12	-1.7	0.08	-1.8	0.07	-0.8	0.41	CXCL12	-1.7	0.08	-1.8	0.07	-0.8	0.41	CXCL12	-1.7	0.08	-1.8	0.07	-0.8	0.41							
	CXCL8	-0.2	0.82	-1.3	0.20	-0.8	0.40	CXCL8	1.0	0.34	0.1	0.91	0.2	0.83	CXCL8	1.4	0.15	1.7	0.09	1.3	0.20	CXCL8	1.4	0.15	1.7	0.09	1.3	0.20	CXCL8	1.4	0.15	1.7	0.09	1.3	0.20							
	EGF	0.8	0.45	-1.0	0.30	0.2	0.83	EGF	1.4	0.17	-0.9	0.37	1.7	0.09	EGF	0.3	0.74	0.4	0.71	0.5	0.64	EGF	0.3	0.74	0.4	0.71	0.5	0.64	EGF	0.3	0.74	0.4	0.71	0.5	0.64							
	EPO	0.0	1.00	-1.8	0.07	-1.3	0.20	EPO	0.0	1.00	0.9	0.35	-2.2	0.02	EPO	0.1	0.94	2.2	0.03	-1.7	0.09	EPO	0.1	0.94	2.2	0.03	-1.7	0.09	EPO	0.1	0.94	2.2	0.03	-1.7	0.09							
	FGF2	0.8	0.40	-0.2	0.87	0.6	0.55	FGF2	1.4	0.15	-0.2	0.88	0.5	0.64	FGF2	0.6	0.57	-0.1	0.95	-0.1	0.93	FGF2	0.6	0.57	-0.1	0.95	-0.1	0.93	FGF2	0.6	0.57	-0.1	0.95	-0.1	0.93							
	HGF	-1.8	0.08	0.2	0.82	-1.3	0.19	HGF	-0.6	0.56	1.4	0.17	-0.3	0.74	HGF	1.3	0.21	1.1	0.25	0.7	0.51	HGF	1.3	0.21	1.1	0.25	0.7	0.51	HGF	1.3	0.21	1.1	0.25	0.7	0.51							
	IGF1	-2.9	0.00	-1.3	0.18	0.2	0.82	IGF1	-3.3	0.00	-1.3	0.21	1.7	0.08	IGF1	-1.0	0.30	0.5	0.64	1.1	0.27	IGF1	-1.0	0.30	0.5	0.64	1.1	0.27	IGF1	-1.0	0.30	0.5	0.64	1.1	0.27							
	JAG1	3.5	0.00	1.0	0.31	1.2	0.23	JAG1	1.4	0.15	0.6	0.52	0.5	0.64	JAG1	-1.6	0.12	-0.4	0.67	-1.0	0.34	JAG1	-1.6	0.12	-0.4	0.67	-1.0	0.34	JAG1	-1.6	0.12	-0.4	0.67	-1.0	0.34							
	KITL	2.2	0.03	2.7	0.01	3.0	0.00	KITL	2.3	0.02	1.1	0.29	2.0	0.05	KITL	0.4	0.73	-0.7	0.48	-0.9	0.35	KITL	0.4	0.73	-0.7	0.48	-0.9	0.35	KITL	0.4	0.73	-0.7	0.48	-0.9	0.35							
	PDGFB	1.0	0.31	1.7	0.08	1.6	0.11	PDGFB	-0.4	0.68	0.3	0.74	1.6	0.11	PDGFB	-1.3	0.19	-1.2	0.23	0.2	0.85	PDGFB	-1.3	0.19	-1.2	0.23	0.2	0.85	PDGFB	-1.3	0.19	-1.2	0.23	0.2	0.85							
	PGF	0.1	0.96	1.2	0.22	-0.3	0.76	PGF	1.1	0.28	1.2	0.23	-1.3	0.18	PGF	1.0	0.32	0.1	0.95	-1.1	0.26	PGF	1.0	0.32	0.1	0.95	-1.1	0.26	PGF	1.0	0.32	0.1	0.95	-1.1	0.26							
TGFB	1.0	0.33	-0.2	0.81	-0.2	0.88	TGFB	0.2	0.82	-0.1	0.92	-1.1	0.28	TGFB	-0.6	0.54	-0.1	0.95	-0.8	0.40	TGFB	-0.6	0.54	-0.1	0.95	-0.8	0.40	TGFB	-0.6	0.54	-0.1	0.95	-0.8	0.40								
VEGFA	1.0	0.30	0.4	0.70	-1.4	0.17	VEGFA	0.6	0.56	1.2	0.22	-1.1	0.28	VEGFA	-0.6	0.58	0.8	0.42	0.4	0.70	VEGFA	-0.6	0.58	0.8	0.42	0.4	0.70	VEGFA	-0.6	0.58	0.8	0.42	0.4	0.70								
GFR	GBM vs MI						GBM vs HC						MI vs HC																													
	HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR													
	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P														
	ADA2	0.3	0.74	0.3	0.74	-1.1	0.28	ADA2	-1.0	0.29	-0.3	0.80	-1.4	0.18	ADA2	-1.2	0.25	-0.9	0.39	0.2	0.87	ADA2	-1.2	0.25	-0.9	0.39	0.2	0.87	ADA2	-1.2	0.25	-0.9	0.39	0.2	0.87							
	APLR	0.0	1.00	0.0	1.00	0.0	1.00	APLR	-0.9	0.35	0.0	1.00	-0.9	0.36	APLR	-1.0	0.33	0.0	1.00	-1.0	0.30	APLR	-1.0	0.33	0.0	1.00	-1.0	0.30	APLR	-1.0	0.33	0.0	1.00	-1.0	0.30							
	CXCR4	3.1	0.00	2.4	0.02	1.9	0.06	CXCR4	2.7	0.01	-0.1	0.90	0.9	0.39	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29							
	EGFR	0.2	0.81	1.1	0.29	1.4	0.15	EGFR	0.0	0.98	0.5	0.64	0.2	0.88	EGFR	-0.2	0.82	-0.7	0.48	-1.2	0.25	EGFR	-0.2	0.82	-0.7	0.48	-1.2	0.25	EGFR	-0.2	0.82	-0.7	0.48	-1.2	0.25							
	FLT1	0.0	1.00	-1.1	0.26	-0.4	0.71	FLT1	0.0	1.00	0.0	1.00	1.1	0.27	FLT1	0.0	1.00	1.0	0.30	1.4	0.17	FLT1	0.0	1.00	1.0	0.30	1.4	0.17	FLT1	0.0	1.00	1.0	0.30	1.4	0.17							
	IGF1R	1.7	0.09	1.9	0.05	0.5	0.60	IGF1R	1.8	0.07	1.2	0.25	0.6	0.57	IGF1R	0.1	0.91	-0.4	0.69	-0.1	0.91	IGF1R	0.1	0.91	-0.4	0.69	-0.1	0.91	IGF1R	0.1	0.91	-0.4	0.69	-0.1	0.91							
	KDR	1.0	0.30	-0.8	0.43	-0.4	0.66	KDR	0.2	0.88	-0.2	0.86	-0.1	0.94	KDR	-1.0	0.33	0.7	0.47	0.6	0.57	KDR	-1.0	0.33	0.7	0.47	0.6	0.57	KDR	-1.0	0.33	0.7	0.47	0.6	0.57							
	KIT	1.6	0.11	0.1	0.91	0.3	0.80	KIT	0.7	0.50	0.4	0.72	0.5	0.58	KIT	-0.5	0.63	0.0	1.00	0.1	0.93	KIT	-0.5	0.63	0.0	1.00	0.1	0.93	KIT	-0.5	0.63	0.0	1.00	0.1	0.93							
	PDGFRB	-1.1	0.27	1.6	0.11	0.8	0.44	PDGFRB	-0.5	0.65	0.6	0.55	-0.1	0.95	PDGFRB	1.6	0.10	-1.1	0.28	-2.6	0.01	PDGFRB	1.6	0.10	-1.1	0.28	-2.6	0.01	PDGFRB	1.6	0.10	-1.1	0.28	-2.6	0.01							
TEK	0.4	0.70	1.0	0.30	0.2	0.81	TEK	1.2	0.22	1.2	0.25	1.2	0.22	TEK	1.0	0.34	-0.2	0.85	1.1	0.27	TEK	1.0	0.34	-0.2	0.85	1.1	0.27	TEK	1.0	0.34	-0.2	0.85	1.1	0.27								
TGFB2	2.7	0.01	1.0	0.30	-0.2	0.88	TGFB2	2.1	0.04	0.3	0.78	-0.7	0.47	TGFB2	0.0	0.98	-0.6	0.55	-1.0	0.34	TGFB2	0.0	0.98	-0.6	0.55	-1.0	0.34	TGFB2	0.0	0.98	-0.6	0.55	-1.0	0.34								
TIE1	0.4	0.72	0.5	0.58	-0.7	0.51	TIE1	1.3	0.18	1.1	0.28	-1.3	0.21	TIE1	1.2	0.21	0.5	0.60	-0.5	0.58	TIE1	1.2	0.21	0.5	0.60	-0.5	0.58	TIE1	1.2	0.21	0.5	0.60	-0.5	0.58								
CR	GBM vs MI						GBM vs HC						MI vs HC																													
	HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR													
	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P														
	ACKR3	0.6	0.54	2.0	0.04	1.5	0.14	ACKR3	0.3	0.80	2.2	0.03	1.6	0.11	ACKR3	-0.6	0.53	0.9	0.38	0.9	0.37	ACKR3	-0.6	0.53	0.9	0.38	0.9	0.37	ACKR3	-0.6	0.53	0.9	0.38	0.9	0.37							
	CCR2	2.0	0.04	0.7	0.48	0.2	0.83	CCR2	1.0	0.32	0.8	0.40	0.6	0.55	CCR2	-1.2	0.25	0.4	0.67	0.8	0.43	CCR2	-1.2	0.25	0.4	0.67	0.8	0.43	CCR2	-1.2	0.25	0.4	0.67	0.8	0.43							
	CX3CR1	0.7	0.46	-0.1	0.96	-0.8	0.44	CX3CR1	0.4	0.71	0.8	0.44	-0.5	0.60	CX3CR1	-0.2	0.86	1.5	0.13	0.7	0.46	CX3CR1	-0.2	0.86	1.5	0.13	0.7	0.46	CX3CR1	-0.2	0.86	1.5	0.13	0.7	0.46							
	CXCR2	0.4	0.70	1.4	0.16	1.3	0.18	CXCR2	0.4	0.68	2.6	0.01	1.5	0.13	CXCR2	0.2	0.86	1.9	0.06	0.0	1.00	CXCR2	0.2	0.86	1.9	0.06	0.0	1.00	CXCR2	0.2	0.86	1.9	0.06	0.0	1.00							
	CXCR4	3.1	0.00	2.4	0.02	1.9	0.06	CXCR4	2.7	0.01	-0.1	0.90	0.9	0.39	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29	CXCR4	-0.7	0.50	-2.1	0.03	-1.1	0.29							
	SELPLG	0.1	0.94	0.7	0.48	0.4	0.71	SELPLG	1.2	0.24	0.3	0.80	-1.1	0.29	SELPLG	-1.3	0.20	-0.2	0.85	-2.0	0.05	SELPLG	-1.3	0.20	-0.2	0.85	-2.0	0.05	SELPLG	-1.3	0.20	-0.2	0.85	-2.0	0.05							
	DIF	GBM vs MI						GBM vs HC						MI vs HC																												
		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR		HPCs		CD34		KDR												
		Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P	Z	P													
DPP4		-2.8	0.01	-0.1	0.91	-0.6	0.58	DPP4	-1.2	0.24	0.2	0.85	-0.5	0.62	DPP4	1.7	0.09	1.1	0.28	-0.1	0.91	DPP4	1.7	0.09	1.1	0.28	-0.1	0.91	DPP4	1.7	0.09	1.1	0.28	-0.1	0.91							
MMP14		2.1	0.04	1.2	0.24	1.0	0.33	MMP14	0.9	0.39	-0.1	0.96	-0.8	0.42	MMP14	-1.4	0.17	-1.0	0.34	-1.2	0.25	MMP14	-1.4	0.17	-1.0	0.34	-1.2	0.25	MMP14	-1.4	0.17	-1.0	0.34	-1.2	0.25							
MMP2		0.4	0.70	0.8	0.44	-0.2	0.81	MMP2	1.8	0.07	0.0	1.00	0.0	1.00	MMP2	1.2	0.25	-0.4	0.71	0.0	0.97	MMP2	1.2	0.25	-0.4	0.71	0.0	0.97	MMP2	1.2	0.25	-0.4	0.71									

and improve endothelial recovery.³³ In MI strategies to increase the expression of *CXCR4* by circulating progenitor cells lead to improved homing to ischemic myocardium resulting in restoration of the blood flow and a reduction of cardiac damage following the infarction.²⁴ MMP9 not only induces mobilization of HPCs by cleaving the CXCL12–CXCR4 interaction,³⁴ but also increases the expression of CXCR4 by bone marrow progenitor cells.^{35,36} The increased plasma levels of MMP9 in GBM patients found in the present study corroborate the literature.³⁷ The elevated levels of tumor-derived MMP9 could cause upregulation of *CXCR4* in CACs of GBM patients. Furthermore, the reduced expression of *DPP4* in GBM HPCs is also associated with a more efficient homing of HPCs to CXCL12-expressing target tissue.³⁸ It is therefore likely that the elevated expression of *CXCR4* and the reduced expression of *DPP4* by GBM CACs, combined with the high GBM tissue-to-plasma CXCL12 gradient, translate into a highly efficient homing process of CXCR4⁺ CACs to GBM tumor. Interference with the MMP9/DPP4/CXCR4/CXCL12 axis in CACs in GBM patients seems a very promising therapeutic option for targeting CAC-mediated neovascularization.

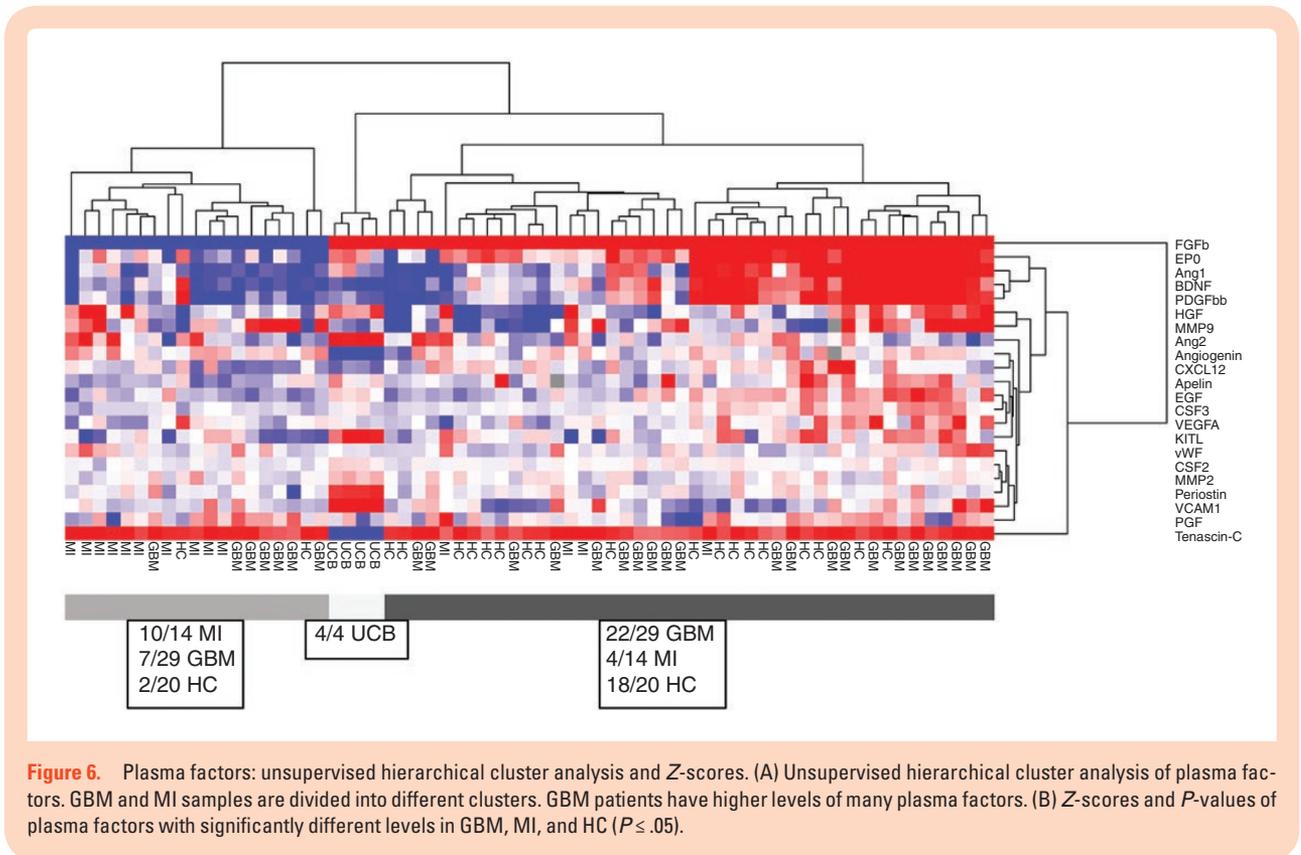
In GBM CACs, gene expression of *KITL* was significantly higher than in MI and HC. *KIT* was expressed higher in GBM than in MI HPCs. *KITL* is a cytokine that binds to the *KIT* receptor; the *KIT/KITL* receptor/ligand pair is important for hematopoiesis and for the mobilization, chemotaxis/homing, and maintenance of HPCs,^{39,40} as well as for angiogenesis.^{41–43} The *KIT/KITL* axis is also essential for neovascularization in glial tumors.⁴¹ In GBM tissue, *KITL* is not only produced by glial tumor cells, but also by neurons.⁴¹ Silencing of *KITL* in glioma cells leads to a decrease in angiogenesis and tumor growth and improved survival.⁴¹ The *KIT* receptor is widely expressed in GBM endothelial cells and in tumor cells present around foci of necrosis.⁴⁴ *KITL* exists in a soluble (s*KITL*) and membrane bound (m*KITL*) form.⁴⁵ s*KITL* results from proteolytic cleavage of m*KITL*.⁴² Transmembrane *KITL* is formed by alternative mRNA splicing. The proteolytic cleavage of m*KITL* to s*KITL* by MMPs (in particular MMP9) is crucial for the mobilization of HPCs from the bone marrow in a similar fashion as for CXCR4/CXCL12.^{46,47} Indeed, we previously found a strong correlation between plasma MMP9 levels and circulating levels of HPCs in GBM patients.¹⁴ In the present study, the primer set used to determine *KITL* mRNA levels did not distinguish between the soluble and transmembrane forms. Hence, we do not yet know if the increased *KITL* gene expression translates to higher levels of s*KITL*, m*KITL*, or both in GBM CACs. Importantly, m*KITL* can act as a chemotactic membrane bound ligand to *KIT*⁺ cells in the target tissue,⁴⁸ mediating the homing of m*KITL*⁺ cells to *KIT*⁺ target tissue. Reversely, *KIT*⁺ circulating progenitor cells home to *KITL*⁺ target tissue.⁴⁹

Hence, the high *KITL* expression by GBM CACs, and the high *KIT* expression by GBM HPCs, is expected to facilitate homing to *KIT*⁺/*KITL*⁺ GBM tissue and stimulate tumor angiogenesis. The role of *KIT/KITL* in GBM CACs therefore deserves further investigations in the search for targets for CACs-induced neovascularization in GBM.

The functional meaning of our findings should be explored further using *in vitro* and *ex vivo* experimental systems, in animal models and finally in clinical trials on humans. FACS or immunomagnetic bead-isolated CACs could be used in chemotaxis/invasion assays (transwell) to determine the potential of GBM versus MI/HC CACs to migrate along gradients of chemoattractants (eg, CXCL12, CCL2, s*KITL*, s*KIT*, and s*VCAM1*) and/or to GBM cells. Silencing of *CXCR4*, *KIT/KITL*, and *ITGA5/ITGA4* in CACs or the addition of CXCR4 blockers (such as AMD3100) or *KITL/KIT/Intα5β1/Intα4β1* inhibitors could be used to validate the importance of these factors in the chemoattraction/homing response. Additionally, CACs could be treated with MMP9 to determine its effect on CAC CXCR4 expression and chemotaxis. The angiogenic function of GBM CACs in GBM could be confirmed using 3D angiogenesis assays.⁵⁰ Labeled CACs (GBM vs MI/HC) could be injected into the circulation and tumor tissue of a GBM xenograft orthotopic mouse model to determine their tumor-homing capacity and their effect on tumor neovascularization and growth. CACs could be isolated from GBM tissue after having homed to tumor, and their expression profile compared to the original CACs to determine the effect of the GBM microenvironment on CAC gene expression. Inhibition of homing molecules like CXCR4, *KITL/KIT*, and *Intα5β1/Intα4β1* prior to peripheral administration of CACs would validate the function of these molecules *in vivo*. Finally, clinical trials can be developed investigating the effect of blocking the mobilization and/or tumor homing of CACs on GBM neovascularization and growth (eg, by blocking circulating MMP9 or VCAM1, both elevated in GBM patient plasma and correlating positively with levels of HPCs and *KDR*⁺ cells, respectively¹⁴). Lowering the levels of plasma MMP9 would reduce CAC CXCR4 expression^{35,36} and diminish their homing capacity to tumor CXCL12. Similarly, blockage of CXCR4 using, eg, AMD3100 could abrogate the homing potential of CACs.⁵¹ Since AMD3100 also mobilizes CACs from the bone marrow, alternative homing mechanisms than the CXCR4/CXCL12 axis may need to be targeted simultaneously to prevent CACs from reaching GBM tissue using alternative routes (eg, *KIT/KITL*, *Intα4β1/VCAM1*).

Our results can eventually be translated toward developing disease-specific therapies targeting CAC-induced neovascularization. Crucial to the development of these targeted therapies is maintaining the balance between effective anti-angiogenic therapy and preservation of the necessary regenerative capacities of the organism.

Figure 5. Differential gene expression between GBM, MI, and control groups. Z-scores and P-values of CAC subset gene expression (–dCt values) differences in patients and controls (Mann–Whitney U-test; SPSS version 25). Comparisons are made for each CAC subset included (HPCs, CD34⁺ cells, *KDR*⁺ cells) between patients (GBM, MI) and controls (HC, UCB). Genes are organized based on their function: PAFs, proangiogenic factors; GFRs, growth factor receptors; CRs, chemotactic receptors; DIFs, de-adhesion and invasion factors; MFs, mobilization factors; ITG, integrins (adhesion factors); Other, matricellular modulators of angiogenesis (*POSTN/TNC*) and the progenitor cell marker *CD133*.



Supplementary Data

Supplementary data are available at *Neuro-Oncology Advances* online.

Keywords

angiogenesis | circulating angiogenic cell | endothelial progenitor cell | glioma | hematopoietic progenitor cell | myocardial infarction | neovascularization.

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