

ORIGINAL RESEARCH

Mitochondrial DNA sequence analysis reveals multiple Pleistocene glacial refugia for the Yellow-spotted mountain newt, *Neurergus derjugini* (Caudata: Salamandridae) in the mid-Zagros range in Iran and Iraq

Maryam Malekoutian | Mozafar Sharifi  | Somaye Vaissi 

Department of Biology, Baghabrisham, Razi University, Kermanshah, Iran

Correspondence

Somaye Vaissi, Department of Biology, Baghabrisham, Razi University, Kermanshah, Iran.

Email: veisi.somaye@gmail.com

Funding information

Razi University

Abstract

Phylogeography is often used to investigate the effects of glacial cycles on current genetic structure of various plant and animal species. This approach can also identify the number and location of glacial refugia as well as the recolonization routes from those refugia to the current locations. To identify the location of glacial refugia of the Yellow-spotted mountain newt, *Neurergus derjugini*, we employed phylogeography patterns and genetic variability of this species by analyzing partial ND4 sequences (867 bp) of 67 specimens from 15 sampling localities from the whole species range in Iran and Iraq. Phylogenetic trees concordant with haplotype networks showed a clear genetic structure among populations as three groups corresponding to the populations in the north, center, and south. Evolutionary ages of clades north and south ranging from 0.15 to 0.17 Myr, while the oldest clade is the central clade, corresponding to 0.32 Myr. Bayesian skyline plots of population size change through time show a relatively slight increase until about 25 kyr (around the last glacial maximum) and a decline of population size about 2.5 kyr. The presence of geographically structured clades in north, center, and south sections of the species range signifies the disjunct populations that have emerged in three different refugium. This study illustrates the importance of the effect of previous glacial cycles in shaping the genetic structure of mountain species in the Zagros range. These areas are important in terms of long-term species persistence and therefore valuable areas for conservation of biodiversity.

KEYWORDS

demography, multiple glacial refugia, phylogeography patterns, pleistocene climatic fluctuations, population genetic structure

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1 | INTRODUCTION

Quaternary glacial-interglacial periods had a significant impact on the current pattern of species distribution and interpopulation gene flow (Fedorov & Stenseth, 2002; Ikeda et al., 2017). Generally, glacial plates at higher latitude (and elevation) made habitats inappropriate to inhabit, while climate at the same location at lower latitude (and elevation) was habitable (Ohlemüller, Huntley, Normand, & Svenning, 2012). Populations at higher latitude may cope through adaptation or phenotypic plasticity, or alternatively they may track suitable habitat (Hoffmann & Sgro, 2011). However, a more likely outcome is that such populations go extinct (Stewart, Lister, Barnes, & Dalén, 2009). At lower latitudes, populations can endure glacial periods relatively unaffected in so called glacial refugia (Bennett & Provan, 2008). Glacial refugium refers to a location where the mountain species can survive in the glacial periods, regardless of the geographical position or spatial extent of the location (Holderegger & Thiel-Egenter, 2009). Biogeographic patterns are one approaches for identifying and describing refugia and suggesting that refugia existed in an area at some stage in the past (Keppel et al., 2012). Numerous studies are now revealing that within each of the main refugial areas, intraspecific lineages exhibited differential responses to climatic changes, often surviving in distinct, allopatric local refugia and exhibiting population fragmentation, allopatric differentiation, demographic expansion, and population admixture (Branco, Monnerot, Ferrand, & Templeton, 2002; García-París, Alcobendas, Buckley, & Wake, 2003; Mattocchia, Marta, Romano, & Sbordoni, 2011). On a broader scale, in the case of widespread species a detailed knowledge of phylogeographic patterns within refugial areas is crucial for deriving robust inferences about the evolutionary history of species at a continental scale (Gómez & Lunt, 2007). On a finer scale, this approach depicts the existence of multiple refugial areas and the succession of events of population fragmentation, allopatric differentiation, demographic expansion, and population admixture (Sequeira, Alexandrino, Rocha, Arntzen, & Ferrand, 2005).

In higher latitudes in North American and Northern Europe, the influence of Quaternary climatic fluctuations on plant and animal species is well documented (Hewitt, 2011); however, such influence of climate change on range dynamics of species in other parts of the world is much less reported (Jandzik et al., 2018). Various geomorphic studies in Iran, including lake sediments, deserts, glacial moraines and periglacial features, salt domes, alluvial sediments, pediments and alluvial fans, fluvial and marine terraces, and loess-soil sequences have shown that Iran was affected by Pleistocene and Holocene climate fluctuations (Kehl, 2009). The available paleoecological and palynological records in Iran indicate that the last glacial maximum (LGM) was characterized by a cooler and more arid climate compared to the Holocene (Djamali et al., 2012; Kehl, 2009). At the present time, deep valleys of several high altitude mountain areas in the Iranian Plateau are covered by glaciers. In the LGM, the expansion of such climatic conditions encouraged the spread of steppe habitats in lower altitudes, while high altitudes in the mountain areas were covered by glaciers (Ghahroudi Tali, Naeimi, & Gharnaie, 2016). In Zagros range, there are reports of isolated populations of

various temperate evergreen plant species such as *Olea europaea* and *Myrtus communis* (Sharifi, Najafi, Yossefshahi, & Hemmati, 2001) and pockets of plant communities such as mires and patterned mires (Wasylikowa & Walanus, 2004), floating meadow and highland peatlands (Sharifi, Rezaii, Hosseini, & Raji, 2004). These relict individuals and plant communities are assumed to persist in enclaves of environmental conditions within an inhospitable regional climate and left behind during range shifts caused by Pleistocene climatic fluctuations (Hampe & Jump, 2011; Wright, McAndrews, & Van Zeist, 1967). According to Djamali et al. (2008), the vegetation composition during the LGM in the Lake Urmia, in northwestern Iran, demonstrates lower winter temperatures than today, and higher July temperatures about 11–12°C (Djamali et al., 2008).

The impact of climate fluctuations on several Iranian animal species has been investigated. These studies have shown that impact of climate fluctuation is more complicated and may not follow a prevailing scenario of glacial retraction and postglacial expansion as has been reported from the more northerly located regions of the western Palearctic (Chiocchio, Bisconti, Zampiglia, Nascetti, & Canestrelli, 2017). Indeed, Southern Europe, Turkey, the southern parts of the Caucasus and Alborz act as multiple glacial refugia due to topographical heterogeneity and low latitude (Ahmadzadeh, Flecks, et al., 2013a; Asadi et al., 2019; Rossiter, Benda, Dietz, Zhang, & Jones, 2007). In Iran, some species experienced range contractions to glacial refugia located, for example, in the Zagros, Alborz, and Kope Dagh Mountains, which were followed by postglacial expansion for Brandt's Persian lizard, *Iranolacerta brandtii* (Ahmadzadeh, Carretero, et al., 2013b), oriental green lizards of the *Lacerta trilineata* (Ahmadzadeh, Flecks, et al., 2013a), two sympatric moth species, *Gnopharmia colchidari* and *G. kasrunensis* (Rajaei Sh et al., 2013), Persian jird, *Meriones persicus* (Dianat, Darvish, Cornette, Aliabadian, & Nicolas, 2017), greater horseshoe bat, *Rhinolophus ferrumequinum* (Shahabi, Akmal, & Sharifi, 2017), Asian pit viper, *Gloydius halys caucasicus* (Asadi et al., 2019), and Eastern rock nuthatch, *Sitta tephronota*, (Yousefi, Shabani, & Azarnivand, 2019). Some species did not change range such as Greek tortoise, *Testudo graeca* (Javanbakht et al., 2017), and Caspian turtles, *Mauremys caspica*, *M. rivulata* (Vamberger et al., 2017). There are even species that showed more extensive distribution during the glacial and retraction of ranges in interglacial as was evidenced for cold-adapted species such as Yellow-spotted mountain newt, *Neurergus derjugini* (Afroosheh et al., 2019), and Blanford's Semaphore gecko, *Pristurus rupestris* (Saberi-Pirooz et al., 2019).

Neurergus derjugini is a critically endangered species occurring in 42 highland streams of the Zagros Mountain range in western Iran and eastern Iraq (Afroosheh, Akmal, Esmaeili-Rineh, & Sharifi, 2016). This area is characterized by high physiographic complexity, with several large mountain ranges that in some area are large enough to facilitate precipitation and local open woodlands. In areas where mountains are enough large to prevent moist air parcels, similar oak forests develop in western-eastern corridors from northern Mesopotamian Plain to the western edge of the Iranian plateau. Within highland open woodland, 81% of the reported habitats for *N. derjugini* are located in Iran, 19% in

Iraq (Afroosheh et al., 2016). The extent of occurrence of *N. derjugini* as indicated by a minimum convex polygon is 6,366 km² for the 42 known localities (Afroosheh et al., 2016). This polygon is positioned along the western edge of the Zagros range with elevations ranging from 630 to 2057 masl. This area is covered by an oak open woodland. The breeding habitat of *N. derjugini* in the Zagros range has been degraded recently by water pollution, water extraction, and severe droughts, which have led to the extirpation of some populations (Afroosheh et al., 2016).

The present study analyzed spatial variation of mtDNA sequences, aiming to investigate the genetic structure within species, as well as attempting to trace the recent demographic and phylogeographical history of the *N. derjugini* in the Zagros range. We tested the multiple glacial refugia hypothesis for *N. derjugini* using molecular data in the whole specie range in mid-Zagros range at the border of Iran and Iraq. We therefore aim to: (a) examine the extent of interspecific genetic variation and produce a phylogeography pattern of genetic diversity in various population of *N. derjugini*, (b) estimate time divergences among the different lineages, (c) determine if all populations derived from one glacial refugium or multiple refugia, (d) examine the similarity of the emerged pattern with the current subspecific taxonomy. Finally, we (e) match phylogeographic results with available Quaternary range dynamic outputs.

2 | METHODS

2.1 | Population sampling and sequencing

We were sampled from 67 individuals of *Neurergus derjugini* from 15 localities in Iran and Iraq during 2012–2014 (Table 1, Figure 1).

Tissue samples were collected from each individual by removing a small section of tail or toe using sterile equipment. Tissue samples were stored in 95% ethanol at –20°C until DNA was extracted. Genomic DNA was extracted using tissue kit GenNetBio™. We sequenced an 867 bp fragment of a mtDNA generation consisting of 726 bp fragment of NADH subunit 4, the whole tRNA-His and tRNA-Ser and 21 bp from the “5” end of tRNA-Leu using the primers ND4-Leu: ND4, CAC CTA TGA CTA CCA AAA GCT CAT GTA GAA GC and Leu, CAT TAC TTT TAC TTG GAT TTG CAC CA (Arevalo, Davis, & Sites, 1994). Amplification of mtDNA was conducted using denaturation at 94°C for 2 min, 58°C for 45 s, 72°C for 2 min followed by 35 cycles at 94°C for 30 s, 58°C for 45 s, 72°C for 60 s for annealing and extension at 72°C for 3 min. Sequencing was performed by Microsynth Switzerland Laboratories.

DNA sequences were aligned using the programs Clustal W in the BioEdit v.7.0.5.3 (Hall, 1999) and Muscle in MEGA 6 (Tamura, Stecher, Peterson, Filipinski, & Kumar, 2013). As outgroups, we used *Neurergus kaiseri*, *N. strauchii*, *Triturus karelinii*, and *Ommatotriton vittatus* (GenBank accession numbers for outgroups: EU880320, EU880321, HQ697277, EU880338, respectively). Values for the numbers of haplotypes, polymorphic sites, parsimony informative site, haplotype diversity (Hd), nucleotide diversity (Pi) and number of transitions and transversions were calculated using the software Dnasp v 5.10.01 (Rozas, Librado, Sánchez-Del Barrio, Messeguer, & Rozas, 2010) and Arlequin v 3.0 (Excoffier, Laval, & Schneider, 2005). To investigate pairwise sequences, divergence between haplotypes was computed using uncorrected Kimura 2-parameter (K2P) model (Kimura, 1980) with 1,000 bootstrap replicates using the MEGA ver. 6 software package (Tamura et al.,

TABLE 1 List of sampling locations used in this study and haplotypes with genetic diversities and frequencies

| No. | Name of localities | Latitude (N) | Longitude (E) | Elevation (m) | Haplotypes and their frequencies | SS | H | Pi | Hd |
|-------|--------------------|--------------|---------------|---------------|----------------------------------|----|----|---------|--------|
| 1 | Kavat | 34° 52' N | 46° 30' E | 1601 | Hap4(4) | 4 | 1 | 0.00000 | 0.000 |
| 2 | Ghorighale | 34° 52' N | 46° 29' E | 1,600 | Hap4(5) | 5 | 1 | 0.00000 | 0.000 |
| 3 | Gholani | 34° 54' N | 46° 27' E | 1575 | Hap4(3) | 3 | 1 | 0.00000 | 0.000 |
| 4 | Dourisan | 35° 01' N | 46° 23' E | 1,600 | Hap4(4) | 4 | 1 | 0.00000 | 0.000 |
| 5 | Darrenajjar | 35° 05' N | 46° 18' E | 1,472 | Hap4(2), Hap5(1) | 3 | 1 | 0.00076 | 0.666 |
| 6 | Lashkargah | 35° 00' N | 46° 08' E | 1,415 | Hap4(5), Hap6(1) | 6 | 2 | 0.00038 | 0.333 |
| 7 | Nowdeshe | 35° 11' N | 46° 14' E | 1,760 | Hap7(2), Hap1(1) | 3 | 2 | 0.00231 | 0.667 |
| 8 | Hanigarmale | 35° 14' N | 46° 08' E | 1,383 | Hap2(4) | 4 | 1 | 0.00000 | 0.000 |
| 9 | Tawale | 35° 11' N | 46° 11' E | 1,400 | Hap1(2), Hap2(2) | 4 | 2 | 0.00077 | 0.667 |
| 10 | Balkha | 35° 12' N | 46° 09' E | 1,482 | Hap1(3), Hap2(1) | 4 | 2 | 0.00058 | 0.500 |
| 11 | Penjwin | 35° 36' N | 45° 58' E | 1,421 | Hap8(5) | 5 | 1 | 0.00000 | 0.000 |
| 12 | Siyagwez | 35° 47' N | 45° 47' E | 1689 | Hap8(6), Hap11(1) | 7 | 2 | 0.00033 | 0.286 |
| 13 | Shalmash | 36° 05' N | 45° 29' E | 1622 | Hap10(5) | 5 | 1 | 0.00000 | 0.000 |
| 14 | Saqez | 36° 03' N | 46° 02' E | 2,168 | Hap9(5) | 5 | 1 | 0.00000 | 0.000 |
| 15 | Benjun | 36° 32' N | 45° 31' E | 2,152 | Hap3(5) | 5 | 1 | 0.00000 | 0.000 |
| Total | | | | | | 67 | 11 | 0.00324 | 0.8304 |

Note: Abbreviation: SS, sample sizes; H, haplotypes and Pi, nucleotide diversity and Hd, haplotype diversity.

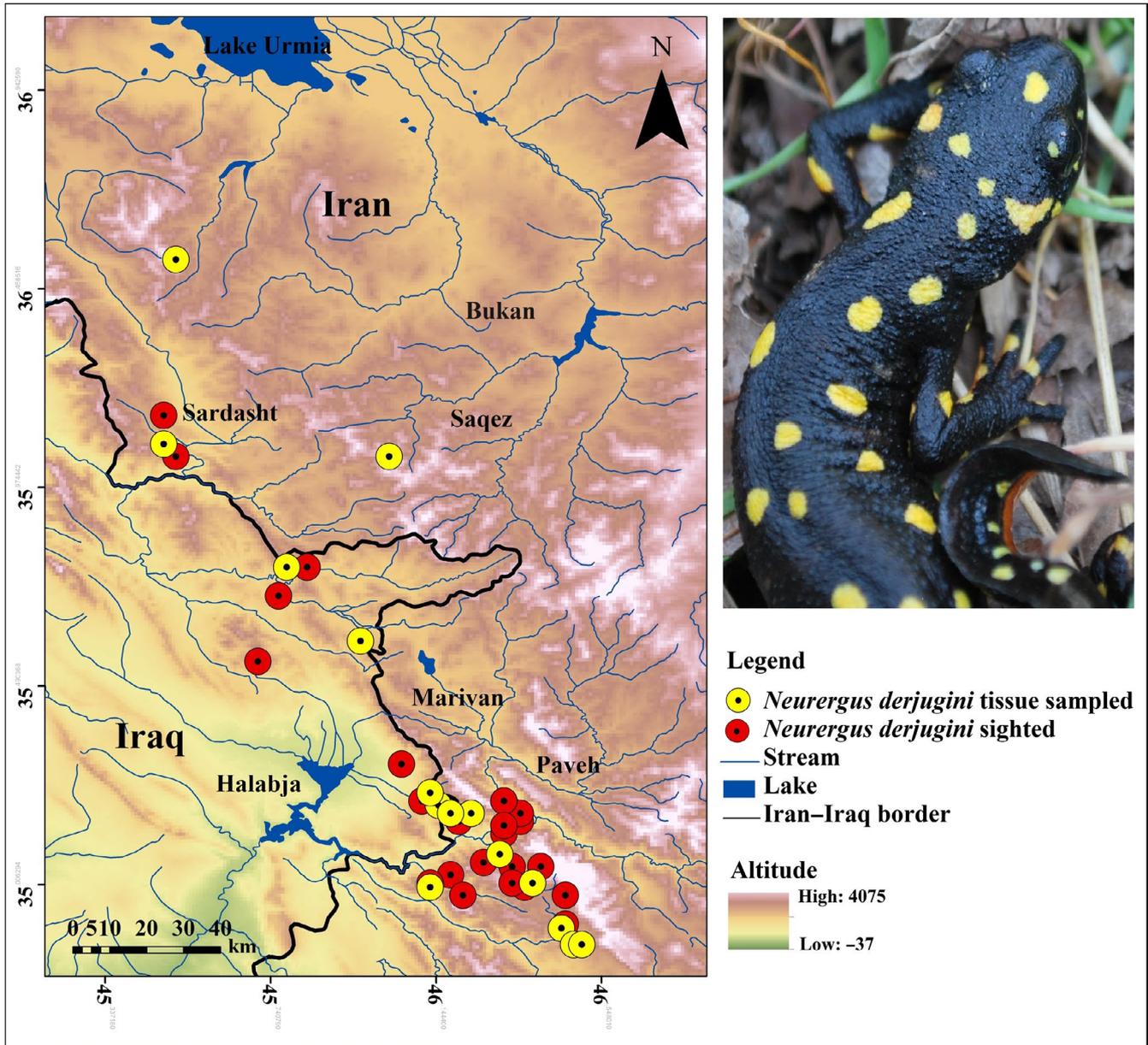


FIGURE 1 Geographical distribution of 42 populations of the Yellow-spotted mountain newt, *Neuregus derjugini*, in Iran and Iraq

2013). MtDNA haplotypes used in this study were deposited in the NCBI Nucleotide Database under accession numbers MN995069 to MN995079.

2.2 | Phylogenetic analyses

Bayesian inference (BI) phylogeny was produced from the ND4 gene haplotypes by means of the program MrBayes, v 3.2.2 program (Ronquist et al., 2012) with 10,000,000 generations and Maximum likelihood (ML) by PhyML, v 3.0 program (Guindon et al., 2010) with 1,500 bootstrap replicates. Also we combined the ND4, ND2, and D-loop sequences to reconstruct a maximum posterior probability tree using the program MrBayes 3.2.2 (Ronquist et al., 2012). ND2 and D-loop sequences have been downloaded from GenBank as follows:

accession numbers MK035716 – MK035726 for ND2 and MK098471 – MK098476 for D-loop. The appropriate model for BI and ML analysis was selected with jModelTest v 0.1.1 program (Posada, 2008) with Akaike information criterion (AIC). The best fit model identified by AIC was TrN. The program FigTree v1.4.0 program (Rambaut, 2016) was used to visualize the phylogenetic tree. A nested clade phylogeographic analysis (NCPA) was constructed for ND4 using the TCS v 1.21 program (Clement, Posada, & Crandall, 2000).

2.3 | Population analysis

Molecular variance was assessed using separate analyses of molecular variance (AMOVA) with 10,000 permutations in Arlequin version 3.0 (Excoffier et al., 2005). The analysis was performed considering

populations from three geographical regions (northern, southern, and central haplogroups) to assess the level of genetic differentiation within populations. Then, all populations are considered as one group to assess the degree of differentiation among regions. Pairwise F_{ST} between populations was generated using Arlequin 3.0 (Excoffier et al., 2005).

2.4 | Environmental data

We chose eight bioclimatic variables environmental characters of so evaluating by Sharifi, Karami, Akmal, Afroosheh, and Vaissi (2017). Bioclimatic variables included isothermality (BIO2/BIO7) ($\times 100$), temperature seasonality (BIO4, standard deviation $\times 100$), temperature annual range (BIO5–BIO6), mean temperature of driest quarter (BIO9), mean temperature of wettest quarter (BIO8), precipitation of warmest quarter (BIO18), precipitation of coldest quarter (BIO19), and elevation. We used ArcMap 10.3 to process variables. The matrix of environmental distances was computed by SPSS version 16.0. We computed geographical distances using DIVA-GIS v 7.5.0 (Hijmans, Guarino, & Mathur, 2004). We performed a Mantel test to evaluate correlations between genetic, geographical, and environmental distances using Arlequin 3.0 (Excoffier et al., 2005). This analysis runs with 10,000 random permutations. In addition, a three-way Mantel test was applied between the matrix of pairwise genetic differentiation and the matrix of environmental distances and geographical distances among populations. We also performed principal components analysis (PCA) to investigate ecological differentiation within *N. derjugini* distribution range.

2.5 | Demographic analysis

Mismatch distribution analysis was performed using DnaSP v 5.10.01 software (Rozas et al., 2010) to estimate population expansion as the distributions of the pairwise nucleotide differences. Haplotype and nucleotide diversity indices, Raggedness index, sum of squared deviations (SSD), and their variances as well as neutrality tests (Tajima's D, Fu's FS) were calculated using DnaSP v 5.10.01 software. The Bayesian skyline plots (BSP) produced from the ND4 gene haplotypes were obtained with a linear model using BEAST v 2.4.5 program (Bouckaert et al., 2014). Markov chain Monte Carlo technique (MCMC) run for 100 million generations sampled every 1,000 steps. Analysis performed with the uncorrelated lognormal relaxed clock and the Bayesian skyline as a coalescent model with the evolutionary rate for salamanders 0.64% per million years (Myr) per lineage (Malyarchuk, Derenko, & Denisova, 2013; Weisrock, Macey, Ugurtas, Larson, & Papenfuss, 2001). To estimate effective population size through time, Tracer v 1.6 (Rambaut, Suchard, Xie, & Drummond, 2016) was used.

2.6 | Divergence time estimate

We estimate times of divergence between lineages of *N. derjugini* using BEAST v 2.4.5 (Bouckaert et al., 2014). We ran for 30,000,000

generations sampled every 1,000 steps, with the first 3,000,000 generations regarded as burn-in 95% highest posterior density (95% HPD) were considered significant support. The maximum clade credibility tree summarized by Tree Annotator v1.8.4 program (Drummond & Rambaut, 2007). The calibration point was derived from molecular phylogenetic studies on the Salamandridae family estimate the divergence times, 0.64% per million years (Myr) per lineage, as proposed by Weisrock et al. (2001) and Malyarchuk et al. (2013). Sequences for outgroups have been downloaded as follows: *Triturus cristatus*: NC_015790; *T. dobrogicus*: HQ697274; *T. carnifex*: HQ697272; *T. marmoratus*: HQ697279; *T. pygmaeus*: HQ697280; *T. macedonicus*: HQ697278, in addition to the previous outgroups.

3 | RESULTS

3.1 | Genetic variation

We identified 11 haplotypes in a sample of 67 individuals of *N. derjugini* from 15 localities based on 863 base pairs of the mitochondrial gene, ND4. Haplotype diversity values range from 0 to 0.667. An indel occurred at the haplotype 5. Haplotype 4 was most widespread and abundant, shared among 6 of the 15 populations (Table 2). Eleven polymorphic sites were recorded including 11 transitions and 9 parsimony informative sites and one Indel site. Mean nucleotide composition were A: 31.79%, T: 30.24%, C: 23.44%, and G: 14.52%. Nucleotide diversity values varied from 0 to 0.00231. Pairwise uncorrected Kimura 2-parameter of genetic distances among 15 populations of *N. derjugini* ranged from 0% to 0.04% (Table 3).

3.2 | Phylogenetic analyses

Phylogenetic relationships are represented by Bayesian and ML trees for 11 haplotypes of *N. derjugini* (Figure 2). Two tree topologies were similar. Monophyly of the *N. derjugini* haplogroups is well supported relative to the outgroup taxa (likelihood bootstrap 99%, posterior probability 1.00). In the phylogenetic trees recorded three groups corresponding to the populations north, center, and south (the northern: Shalmash, Saqez, Benjun; the central: Hani garmale, Tawella, Balkha, Penjwin, Siya gwez; and the southern: Kavat, Ghorighale, Gholani, Dourisan, Darrenajjar, Lashkargah, Nowdeshe), (Figure 2). *Neurergus derjugini* is well supported as sister taxon to *N. kaiseri* (likelihood bootstrap 99%, posterior probability 1.0). Within *N. derjugini*, there is little support for relationships northern populations sampled, with bootstrap 38%. The southern populations of *N. derjugini* from a well-supported clade, with bootstrap 74%. To evaluate the phylogenetic analyses, 867 base pairs of ND4 sequences gene were combined with 718 base pairs of D-loop and 1036 base pairs of ND2 sequences gene. Based on the combined sequences, Bayesian inference trees showed that *N. derjugini* haplotypes form a monophyletic group with a three populations in north, center, and south (Figure 3).

TABLE 2 Variable nucleotide positions and molecular diversity within the sequences (867 bp) of the ND4 for 11 haplotypes of 67 *Neuregus derjugini* sequences in different regions

| Hap | Polymorphic site | | | | | | | | | | | Locality | | | | | | | | | | | Total | | | | | | | |
|---|------------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|----------|-----|----|------|------|----|----|------|----|----|-----|-------|----|------|----|----|------|----|---|
| | 328 | 337 | 376 | 398 | 493 | 576 | 580 | 583 | 634 | 646 | 695 | 779 | 779 | Ka | Ghor | Ghol | Do | Da | La | No | Ha | Ba | | Ta | Pe | Si | Sh | Sa | Be | |
| 1 | G | G | C | G | G | A | C | A | G | - | C | - | C | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 3 | 2 | 0 | 0 | 0 | 0 | 0 | 6 |
| 2 | . | . | . | . | . | . | . | G | . | - | . | . | . | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 4 | 1 | 2 | 0 | 0 | 0 | 0 | 0 | 7 | |
| 3 | . | . | . | . | . | G | G | . | . | - | . | . | . | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 5 | |
| 4 | . | . | . | A | A | . | T | . | . | - | . | . | . | 4 | 5 | 3 | 4 | 2 | 5 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 23 | |
| 5 | . | . | . | A | A | . | T | . | . | A | . | . | . | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | |
| 6 | . | A | . | A | A | . | T | . | . | - | . | . | . | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | |
| 7 | A | . | . | . | . | A | . | . | . | - | . | . | . | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | |
| 8 | . | . | T | . | . | . | . | . | . | - | . | . | . | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 5 | 6 | 0 | 0 | 0 | 11 | |
| 9 | . | . | . | . | . | G | . | . | A | - | T | . | . | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 5 | 0 | 5 | |
| 10 | . | . | . | . | . | G | . | . | A | - | . | . | . | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 5 | 0 | 0 | 5 | |
| 11 | . | . | T | A | . | . | . | . | . | - | . | . | . | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 1 | |
| Sample size | | | | | | | | | | | | | | 4 | 5 | 3 | 4 | 3 | 6 | 3 | 4 | 4 | 4 | 5 | 7 | 5 | 5 | 67 | | |
| Number of polymorphic sites | | | | | | | | | | | | | | 0 | 0 | 0 | 0 | 0 | 1 | 3 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 11 | | |
| Number of transitions | | | | | | | | | | | | | | 0 | 0 | 0 | 0 | 0 | 1 | 3 | 0 | 1 | 1 | 0 | 1 | 0 | 0 | 11 | | |
| Number of transversions | | | | | | | | | | | | | | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | | |
| Number of parsimony informative sites | | | | | | | | | | | | | | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0 | 0 | 9 | | |
| Average number of nucleotide difference | | | | | | | | | | | | | | 0 | 0 | 0 | 0 | 0 | 0.33 | 2 | 0 | 0.5 | 0.66 | 0 | 0.28 | 0 | 0 | 2.81 | | |

Abbreviations: Ka, Kavati; Ghor, Ghorighale; Ghol, Gholani; Do, Dourisan; Da, Darrenaijar; La, Lashkargah; No, Nowdeshe; Ha, Hani garmale; Ta, Tawella; Ba, Balkha; Pe, Penjwin; Si, Siya gwez; Sh, Shalmash; Sa, Saqez; Be, Benjun.

TABLE 3 K2P genetic distances values among 15 populations of *Neuregus derjugini*. Numbers are representative of localities as indicated in Table 1

| No | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 |
|----|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| 1 | | | | | | | | | | | | | | | |
| 2 | 0.000 | | | | | | | | | | | | | | |
| 3 | 0.000 | 0.000 | | | | | | | | | | | | | |
| 4 | 0.000 | 0.000 | 0.000 | | | | | | | | | | | | |
| 5 | 0.000 | 0.000 | 0.000 | 0.000 | | | | | | | | | | | |
| 6 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | | | | | | | | |
| 7 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | | | | | | | |
| 8 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | | | | | | |
| 9 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | | | | | |
| 10 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | | | | |
| 11 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | | | |
| 12 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | | |
| 13 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | | | |
| 14 | 0.003 | 0.003 | 0.003 | 0.003 | 0.003 | 0.003 | 0.003 | 0.003 | 0.003 | 0.003 | 0.003 | 0.004 | 0.003 | | |
| 15 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.003 | 0.000 |

The statistical parsimony haplotype network (Figure 4) suggested three haplotype subnetworks, which were in agreement with the topology described in the phylogenetic trees. The haplotype network pattern suggestive of little or no gene flow between regions because northern, central, and southern haplogroups were not intermingled.

3.3 | Population analysis

The AMOVA analysis showed most genetic variation was significantly explained by differences among regions (74.87% genetic variation, $F_{CT} = 0.74, p < .01$), and genetic difference was detected among populations within regions (19.07 genetic variation, $F_{SC} = 0.75, p < .001$; Table 4). The genetic differences were significant among populations without the grouping. Most of the diversity was observed among populations (92.16%) and among regions (74.78%), while low percentage of variance was detected within populations (6.06%–7.84%; Table 4). The comparisons of pairs of population samples confirmed the high level of heterogeneity. Thirteen pairwise comparisons out of 105 (12.38%) were not significant ($p < .05$) indicating populations sharing the same haplotypes or closely related haplotypes. F_{ST} values between populations range from -0.166 to 1 (Table 5).

3.4 | Demographic analysis

The mismatch distribution for the *N. derjugini* appeared smooth and unimodal, consistent with a model of population expansion (Figure 5). The Harpending's Raggedness Index ($r = .08, p(r) = .1$) and sum of squared deviations (SSD) ($r = .03, p(r) = .1$) were not significant, indicating population expansion. Results of Tajima's D test was positive and not significant (0.60, $P(r) = .10$). However, negative value (-0.63, $P_{Fs} = 0.04$) for Fu's F_s , which is more sensitive to recent population expansions, suggested the recent demographic expansion. Bayesian skyline plots suggest that population size was increased very slightly 25 kyr (around LGM), (Figure 6). BSP_s showed a decline of population size about 2.5 kyr. Thus, the BSP_s suggest that populations went through a bottleneck in the cold period around 2.5 kyr.

3.5 | Environmental data

The analyzed populations revealed significant levels of isolation by distance ($r = .66, p < .0001$; Figure 7a) and correlation between genetic divergence and environmental distance ($r = .68, p < .0001$; Figure 7b). Mantel test detected a signal of isolation by environment had significant influence on genetic distance. Each of two variables in isolation had a significant effect on genetic differentiation. In the three-way Mantel test, the correlation between genetic and geographical ($r = .36, p = .0068$) and environmental distances remained significant ($r = .30, p = .0069$).

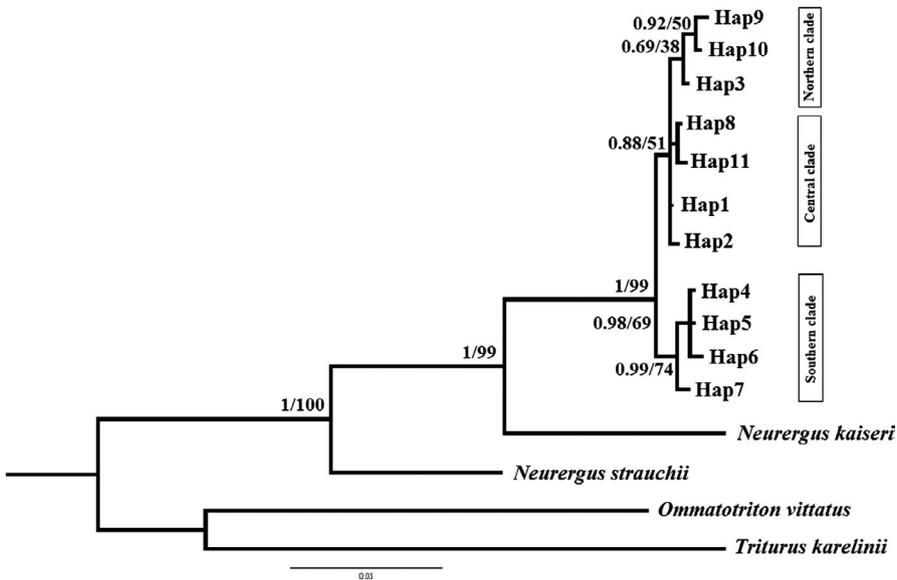


FIGURE 2 Phylogenetic trees of haplotypes implemented in PhyML and MrBayes based on partial ND4 gene sequence of 67 individuals for *Neuregerus derjugini*. Bayesian posterior probability values are the left of the slash, and maximum likelihood bootstrap values are the right

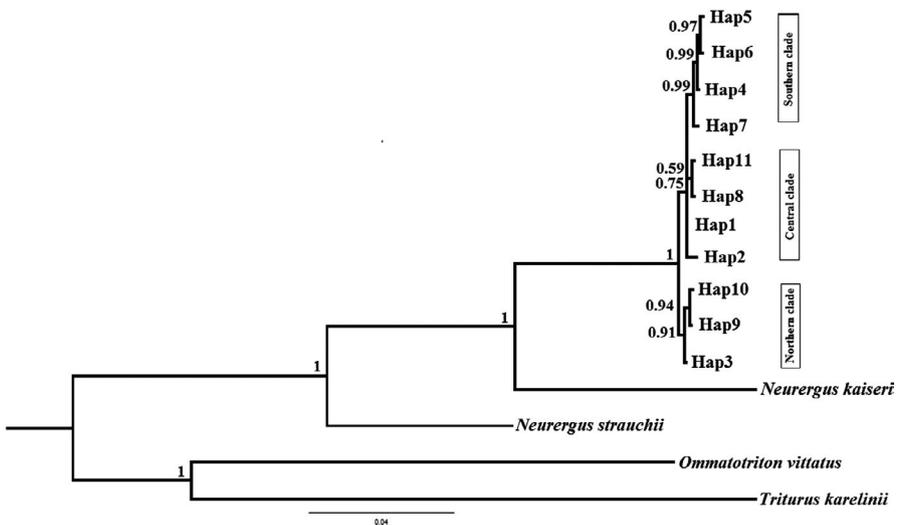


FIGURE 3 Phylogenetic tree of haplotypes implemented in MrBayes based on ND4, ND2, and D-loop sequences for *Neuregerus derjugini*. Numbers above branches indicate Bayesian posterior probability values

The first two principal components (PC1 and PC2) explained 72.51% and 22.17% of the total variation and did not separate the northern, central, and southern localities along temperature and precipitation gradients (Table 6, Figure 8). The occurrence sites were not divided into separated environmental spaces in the Cartesian coordinates formed by the first two principal components (Figure 8).

3.6 | Divergence time estimate

Dating of major mtDNA cladogenetic events within *N. derjugini* haplogroups is presented in Figure 9. The Bayesian estimate of the age of mtDNA diversity in *N. derjugini* between the north-central haplogroups and southern haplogroups was 0.46 Myr and divergence between northern and central haplogroups took place in 0.39 Myr (middle Pleistocene). Evolutionary ages of clades north and south range from 0.15 to 0.17 Myr, while the age of clade center is higher, corresponding to 0.32 Myr.

4 | DISCUSSION

The Yellow-spotted mountain newt exhibited a complex phylogeographic pattern with multiple divergent mtDNA clades across its relatively small range (6,366 km²). A total of 3 allopatric and 11 parapatric clades were identified for ND4, most of them with a very restricted distribution in northern, central, and southern parts of the distribution. Of 11 haplotypes identified by ND4 in present the study, 3, 4 and 4 haplotypes are reciprocally monophyletic for northern, central, and southern sections of the species range. Previous studies on monophyly in *N. derjugini* as evidence for refugia matching include six haplotypes identified in the control region (D-loop) which form a well-supported monophyletic clade within northern and southern populations (Afroosheh et al., 2019) and 11 haplotypes identified in ND2 with three monophyletic clade each 3, 3 and 5 haplotypes in north, center, and south (Salehi, Akmal, & Sharifi, 2019).

The present phylogeographic structure of *N. derjugini* does not match to the current subspecific division of this species as reported

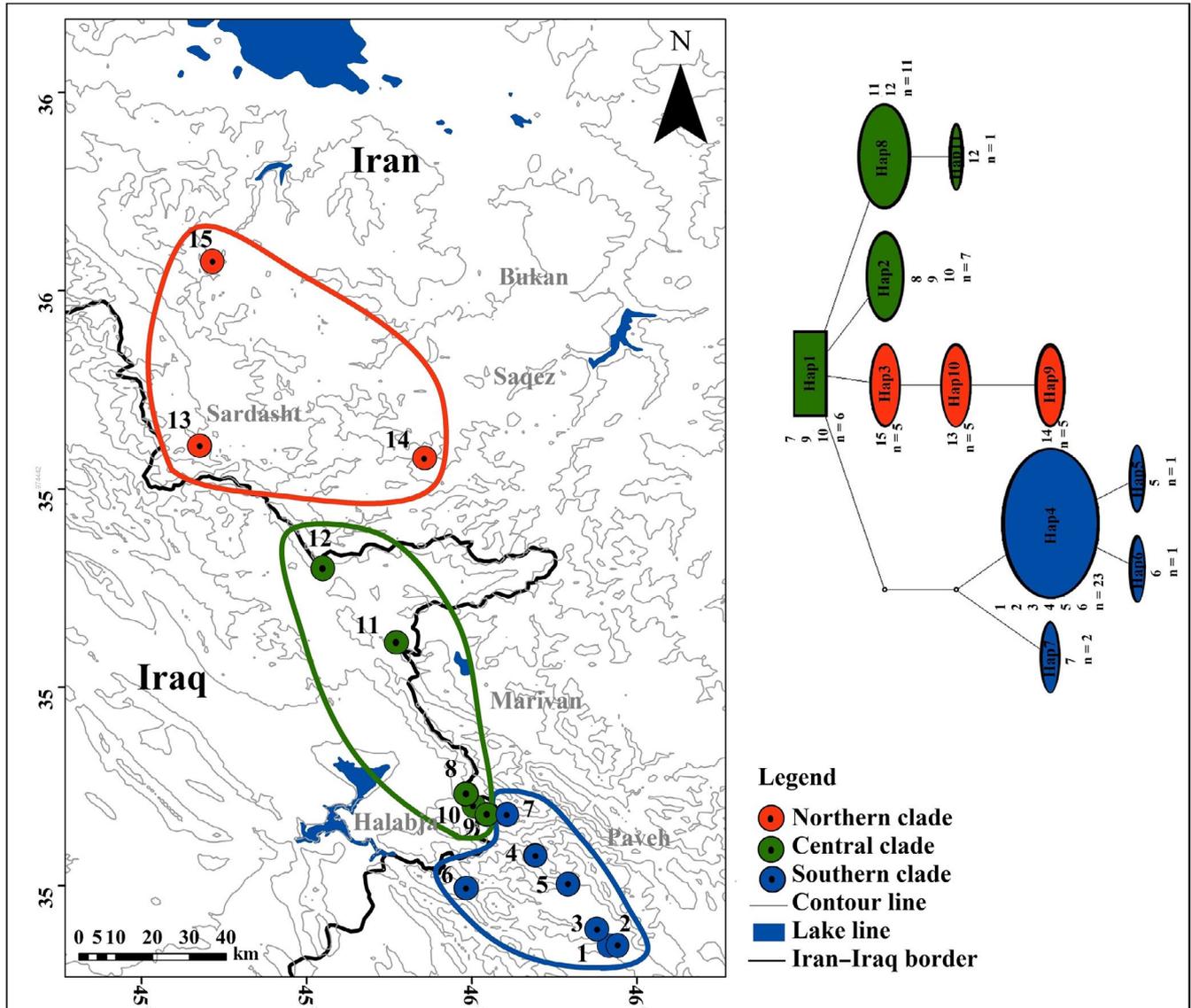


FIGURE 4 The map illustrates the sampling localities of *Neurergus derjugini* in Iran and Iraq, Haplotype network showing the phylogenetic relationships among the 11 haplotypes, Different haplotypes in the haplotype network have different colors, sizes of cycles are representative of the haplotype frequencies, and open dots represent missing intermediate haplotypes

TABLE 4 Analysis of molecular variance (AMOVA) using partial ND4 gene

| Structure | Source of variation | df | Variation (%) | F _{SC} | F _{ST} | F _{CT} |
|---------------------|----------------------------------|----|---------------|-----------------|-----------------|-----------------|
| Three regions | Among regions | 2 | 74.87 | 0.75** | 0.93** | 0.74** |
| | Among populations within regions | 12 | 19.07 | | | |
| | Within populations | 52 | 6.06 | | | |
| The studied samples | Among populations | | 92.16 | | | |
| | Within populations | | 7.84 | | 0.92** | |

Note: Significant values are shown for $p < .05$ “**” and $p < .01$ “***”.

by Nesterov, (1917) and Hendrix et al. (2014): *Neuregus derjugini derjugini* and *N. d. microspilotus*. Frequently, number and size of yellow spots on the dorsal section of body have been used to distinguish different forms of the Yellow-spotted mountain newts (Schneider & Schneider, 2011). However, use of this feature in taxonomy of the

Yellow-spotted newts in Middle East has caused uncertainty because more than one species of newt in Iran and neighboring Iraq and Turkey have yellow spots (Schneider & Schneider, 2011; Sharifi, Naderi, & Hashemi, 2013). Although a taxonomic revision is beyond the scope of this study, our results indicate that the current intraspecific taxonomy

TABLE 5 F_{ST} values between populations for ND4. Numbers are representative of localities as indicated in Table 1

| No | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 |
|----|--------|--------|--------|--------|-------|-------|-------|-------|--------|-------|--------|-------|----|----|----|
| 1 | 0 | | | | | | | | | | | | | | |
| 2 | 0 | 0 | | | | | | | | | | | | | |
| 3 | 0 | 0 | 0 | | | | | | | | | | | | |
| 4 | 0 | 0 | 0 | 0 | | | | | | | | | | | |
| 5 | 0.111 | 0.189 | 0 | 0.111 | 0 | | | | | | | | | | |
| 6 | -0.081 | -0.034 | -0.153 | -0.081 | 0.068 | 0 | | | | | | | | | |
| 7 | 0.641 | 0.690 | 0.571 | 0.641 | 0.5 | 0.648 | 0 | | | | | | | | |
| 8 | 1 | 1 | 1 | 1 | 0.937 | 0.950 | 0.724 | 0 | | | | | | | |
| 9 | 0.904 | 0.917 | 0.886 | 0.904 | 0.826 | 0.874 | 0.506 | 0.333 | 0 | | | | | | |
| 10 | 0.923 | 0.933 | 0.908 | 0.923 | 0.841 | 0.883 | 0.494 | 0.666 | -0.166 | 0 | | | | | |
| 11 | 1 | 1 | 1 | 1 | 0.948 | 0.955 | 0.763 | 1 | 0.805 | 0.825 | 0 | | | | |
| 12 | 0.954 | 0.958 | 0.948 | 0.954 | 0.913 | 0.928 | 0.753 | 0.912 | 0.742 | 0.739 | -0.055 | 0 | | | |
| 13 | 1 | 1 | 1 | 1 | 0.957 | 0.964 | 0.825 | 1 | 0.884 | 0.903 | 1 | 0.945 | 0 | | |
| 14 | 1 | 1 | 1 | 1 | 0.964 | 0.970 | 0.861 | 1 | 0.917 | 0.933 | 1 | 0.958 | 1 | 0 | |
| 15 | 1 | 1 | 1 | 1 | 0.948 | 0.955 | 0.763 | 1 | 0.805 | 0.825 | 1 | 0.920 | 1 | 1 | 0 |

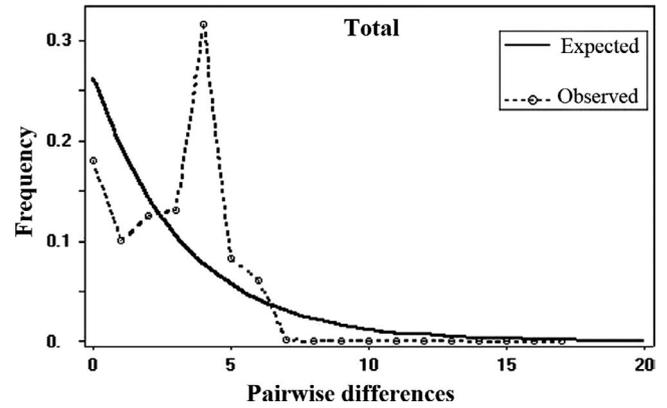


FIGURE 5 Mismatch distributions of the pairwise total population of *Neurergus derjugini*. The dashed lines represent the observed frequency of pairwise differences among sequences, and the lines show the expected distribution

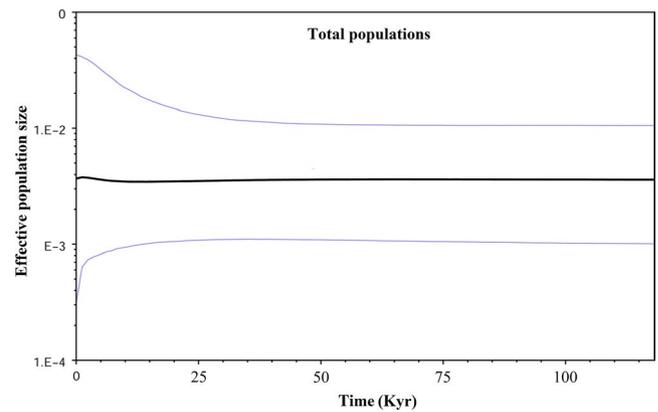
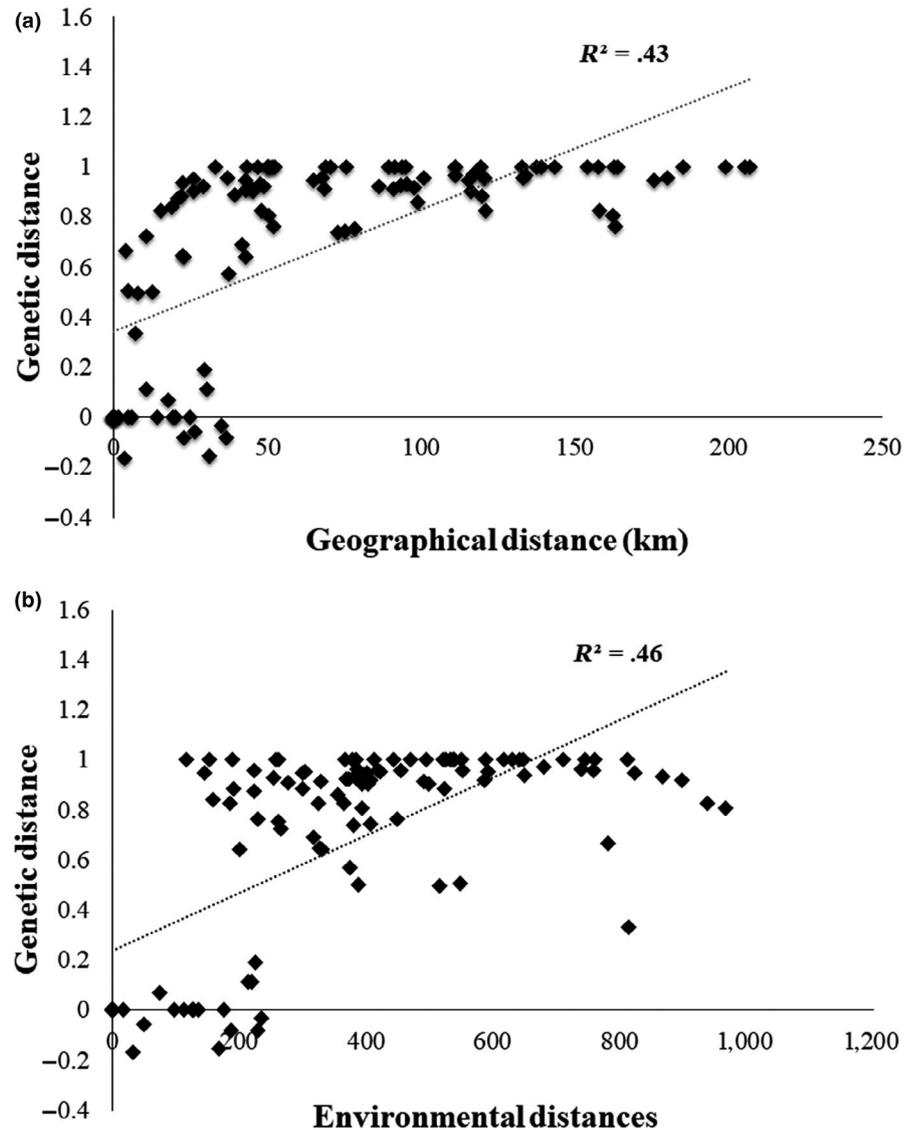


FIGURE 6 Bayesian skyline plots (BSPs) for *Neurergus derjugini*. BSPs derived using a mitochondrial ND4 sequences mutation rate of 0.64 per million years (Myr). The median estimates are shown as blue lines, and the 95% HPD limits are shown by the width areas between blue lines

reflects local phenotypic varieties rather than distinct evolutionary units. According to a study conducted by Vaissi, Parto, and Sharifi (2018), on captive *N. derjugini* spot configuration (numbers, circularity, size, and asymmetry) change drastically during first three years before sexual maturation and remain fairly constant thereafter. In another study on configuration of yellow spots in a free living *N. derjugini* based on photo-identification method a complete spatial randomness (CSR) test using nearest neighbors distances for the spots over dorsal part in adult newt showed a reasonable match with the Poisson distribution indicating that spots are distributed randomly over body of the newt (Sharifi et al., 2013). If such differences prevail, spot features are not adequate external characteristics to firmly determine taxonomic status of various populations of the species groups unless differences in the number, size, and also their stability are determined throughout the distribution range of the species. Therefore, any new arrangement within species of the genus *Neurergus* should be based on detailed intrapopulation examination of morphological features, ontogenetic pattern development, and genetic characteristics of different populations (Vaissi et al., 2018).

FIGURE 7 The plot of simple Mantel test showing the relationships between geographical (a) and environmental distance with genetic distance (b) among 15 populations of *Neurergus derjugini*



The finding of present genetic divergence among geographical clusters of populations of *N. derjugini* has not provided an indication for the existence of cryptic species because the degree of genetic divergence found between various haplogroups are within the range of the distances found in other accepted newt species. Although there appears to be a lack of evidence for sympatric deviations between mitochondrial lineages, it might be suggested that some form of barrier to gene exchange has existed between the distinct lineages. Steinfartz, Hwang, Tautz, Öz, & Veith, 2002, based on two sections of the 12S and 16S mitochondrial ribosomal genes (810 bp), 19 allozyme and three plasma protein loci, showed that intrageneric distances (HKY- Γ 0.5 distances) for *N. strauchii strauchii* and *N. s. barani* was 1.4%. Also, Özdemir, Üzümlü, Avci, & Olgun, 2009 used two mitochondrial genes (478 bp of 12S rRNA and 355 bp of 16S rRNA) from *N. crocatus*, *N. strauchii strauchii*, and *N. s. barani* and found two subspecies of *N. strauchii* differed by 4.5%–5.1% (*N. strauchii strauchii*) and 5.4%–5.5% (*N. s. barani*) of the pairwise uncorrected p-distances of *N. crocatus*. The p-distances between these two subspecies of *N. strauchii* ranged from 0.48% to 1.2%. Pairwise

uncorrected p-distances between clades in present study range from 0 to 0.002 in ND4 and from 0 to 0.004 in ND2 and 0 to 0.001 in D-Loop. Average values of p-distance for these genetic markers is not a level of divergence typically found between recognized species of the genus *Neurergus*; therefore, there may not be adequate evidences to support that *N. derjugini* constitutes a species complex.

The phylogeographic pattern observed in present and two more studies (Afroosheh et al., 2019; Salehi et al., 2019) on population genetic structure of *N. derjugini* with three mitochondrial markers have yielded two (D-Loop) and three (ND2 and ND4) reciprocally monophyletic lineages. Investigation on mitochondrial DNA variation and reconstruction of the potential current and past distribution of *N. derjugini* together with landscape connectivity analysis showed higher gene flow between the breeding streams in the southern part of the range, while the northern populations are more isolated (Afroosheh et al., 2019). This study also showed that during the mid-Holocene and LGM conditions, the range of the species may have been more extended and shifted to lower elevations. These findings show retraction of the *N. derjugini* range during the Quaternary and indicate

TABLE 6 Pearson correlation coefficients between environmental variables and principal component axes describing 15 *Neurergus derjugini* localities

| Environmental variables | PCA1 | PCA2 | PCA3 |
|-------------------------------------|--------|--------|--------|
| BIO3 | | | |
| Isothermality | 0.902 | 0.404 | 0.027 |
| BIO4 | | | |
| Temperature seasonality | 0.879 | 0.473 | 0.048 |
| BIO7 | | | |
| Temperature annual range | 0.895 | 0.442 | 0.03 |
| BIO8 | | | |
| Mean temperature of wettest quarter | 0.781 | -0.473 | 0.402 |
| BIO9 | | | |
| Mean temperature of driest quarter | 0.595 | -0.784 | -0.086 |
| BIO19 | | | |
| Precipitation of coldest quarter | 0.959 | -0.015 | -0.182 |
| Alt | | | |
| Elevation | -0.897 | 0.363 | 0.202 |
| Eigenvalue | 5.076 | 1.552 | 0.246 |
| % of variance | 72.514 | 22.17 | 3.521 |

that range dynamics of the species occupying lower latitudes where they would have a better chance to find glacial refugia (Afroosheh et al., 2019). A reasonable explanation for this phylogeographic pattern in an area with complex topography in mid-Zagros range is one of the larger numbers of refugia occurring in the south to benefit from warmer climate close to the Mesopotamian Plain at lower elevation in the LGM. Phylogeographic analyses on many American and North European species have demonstrated the classic pattern of “southern genetic richness and northern purity” (Fritz et al., 2012, 2008, 2009). Contrarily to this configuration, the pattern of distribution of genetic diversity in *N. derjugini* is an exception.

To our knowledge *N. derjugini* is the first reported case of differentiation into multiple refugia within Zagros range in western Iran. The occurrence of allopatric lineages in northern, central, and southern species range (Figures 2, 3 and 9) which are reciprocally monophyletic provides evidence for a long-term persistence of *N. derjugini* in these areas during Pleistocene glaciations, within separate refugia. Although, the phylogeographic pattern of *N. derjugini* fits the classical southern refugia model this study cannot show if *N. derjugini* used these areas as both glacial refugia and the source areas for northward postglacial colonization. Considering that the highest genetic diversity of these lineages is found in the southern portion of the species range it can be hypothesized that the refugia where these lineages differentiated were located in proximity of this area. In present study, variation in ND4 has revealed notable levels of genetic structuring and a low nucleotide diversity in *N. derjugini*. However, haplotype diversity for the total populations was relatively high. Low nucleotide diversity associated with a relatively high haplotype diversity in species provides evidence for a recent and rapid population expansion (Spear, Peterson, Matocq, & Storfer,

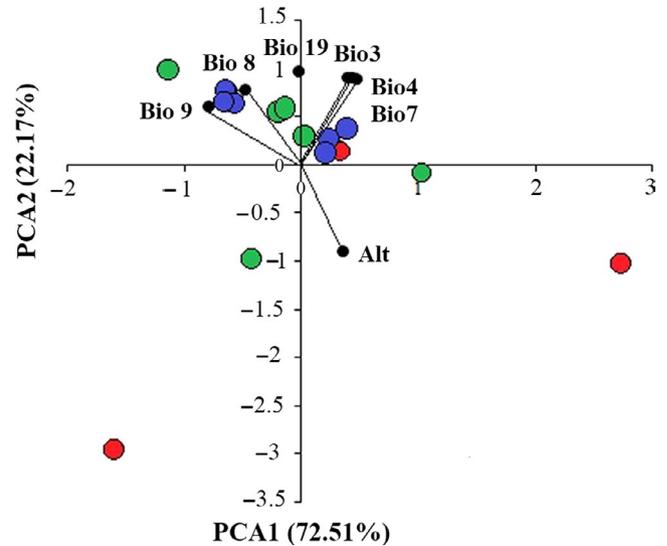


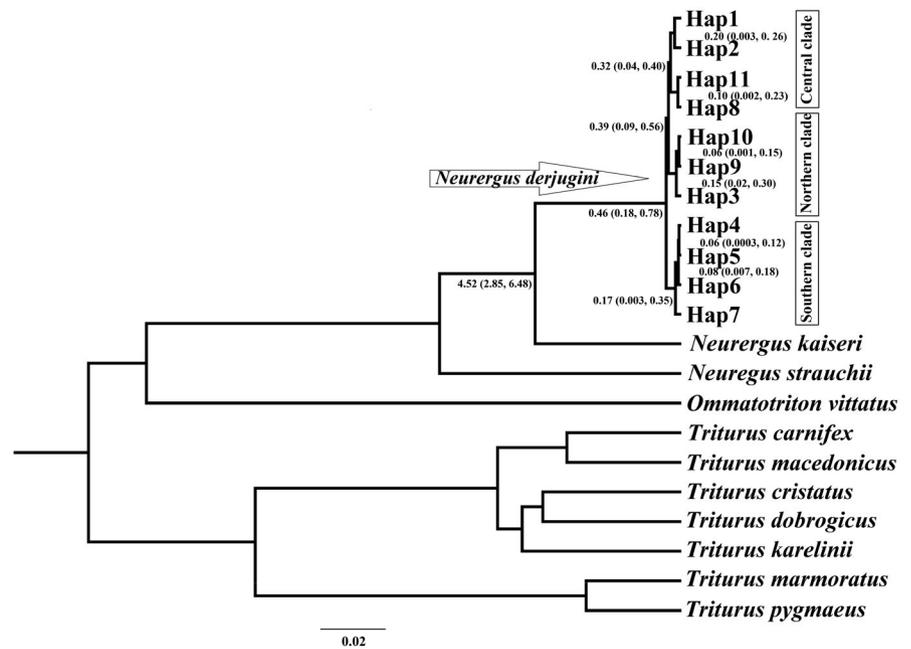
FIGURE 8 Plot of PCA based on environmental variables describing *Neurergus derjugini* localities. The populations in the northern (red circles), central (green circles), and southern (blue circles) regions were not separated along the PC1 and PC2 describing 15 *Neurergus derjugini* localities

2005). However, it may also be due to the limitation caused by using a single-genetic marker and limited sampling (Wang, Jiang, Xie, & Li, 2013).

There are several studies on salamanders and newts that follow multiple refugia scenario. For example, Church, Kraus, Mitchell, Church, and Taylor (2003) analyzed two mitochondrial markers of individuals of the eastern tiger salamander, *Ambystoma tigrinum tigrinum*. The results appear to be multiple Pleistocene refugia with little migration among the remaining populations. Sotiropoulos et al. (2007) performed phylogenetic analyses of *Mesotriton alpestris* populations from the entire range of species distribution, using fragments of two mtDNA genes. Extensive sequence divergence, implying greater isolation in multiple refugia, is found within eastern clades, while the western clades seem to have been involved in the colonization of central, western, and northeastern Europe from a hypothetical refugium in central Europe. Mattocchia et al. (2011) analyzed spatial variation of mtDNA sequences, aiming to investigate the genetic structure within each species, as well as attempting to trace the recent demographic and phylogeographical history of the genus *Salamandrina* in Italy. In this study, three main southern European refugia areas have been identified, corresponding to the Iberian, Italian, and Balkan Peninsulas.

The presence of several monophyletic groups indicate the existence of multiple refugia and has been used for explanation of genetic structure in many amphibian species (Giovannotti, Nisi-Cerioni, & Caputo, 2010). The pattern of haplogroups in present study is, also, confirmed by AMOVA analysis, showing a pronounced geographical partition of *N. derjugini* genetic variation. One reason for a highly structured genetic diversity in populations occurring in close distances may be high fidelity to small home ranges in breeding streams reported for many species of

FIGURE 9 Chronogram of diversification implemented in BEAST based on ND4 gene for *N. derjugini*. The divergence times with 95% highest posterior density (95% HPD) are presented below the branches



amphibians including *N. derjugini*. Using photographic identification method Sharifi and Afroosheh (2014) showed that the average minimum distance covered by recaptured individuals in this species was only 49.19 ± 71.75 m. This value shows that the home range of *N. derjugini* in the breeding streams was estimated to be only 230 m^2 (Sharifi & Afroosheh, 2014). Although these data show that *N. derjugini* has high fidelity to its aquatic environment, there is no evidence showing fidelity to either foraging grounds or to overwintering habitats.

A study on species distribution modeling of *N. derjugini* during current, the mid-Holocene (6 ka BP) and the Last Glacial Maximum (LGM, 21 ka BP) shows that the potential distribution area of *N. derjugini* during the last LGM is characterized by low elevations and dry habitats (Afroosheh et al., 2019). This model anticipates that the species may have expanded north-westwards during mid-Holocene and it may have been moved to lower elevations during LGM (Afroosheh et al., 2019). This study, also, showed that according to Circuitscape analysis, northern populations have become more isolated while the southern populations have more connectivity. The model showed main potential refugia in the Zagros mountain system. These potential refugia are more concentrated in the central part of the species distribution range where central clade is present as the oldest clade with divergence time 0.32 Myr. This study, also, indicates that during the LGM and mid-Holocene, the range of the species may have been more extended and shifted to lower elevations where they would have changed around to find glacial refugees. In present study, the monophyly of the *N. derjugini* populations has been occurred in main refugial areas. The presence of geographically structured clades by AMOVA analysis indicated the disjunct populations of *N. derjugini* may have been survived in three different glacial refugia in the Zagros mountains. Furthermore, the phylogenetic analyses based on another mitochondrial gene (ND2 and D-loop) in *N.*

derjugini (Figure 3) confirmed the presence of three reciprocally subclades and multiple glacial refugia.

5 | CONCLUSIONS

Phylogeographic analysis based on mitochondrial markers in various populations of *N. derjugini* provides evidence for a history of isolation and divergence in allopatry resulting in the diversification of three monophyletic and geographically separate clades. These diversifications are largely concordant with the last glacial maxima. Positive and significant correlation between geographical, environmental, and genetic distances suggests a possible impact of geographical and environmental divergence in shaping the genetic variation of *N. derjugini* that have emerged in three different refugia. Principal component analysis did not separate climatic profiles of the distribution range of *N. derjugini* and demonstrated the homogeneity in different bioclimatic variables in the distribution range of *N. derjugini*. The populations occurring in northern, central, and southern sections of the species range have been separated from one another since 0.46 Myr between the north-central haplogroups and southern haplogroups and 0.39 Myr between northern and central haplogroups took place in middle Pleistocene. Results obtain from current study, also, signifies the role of refugia in conserving populations as well as genetic diversity of *N. derjugini*.

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CONFLICT OF INTEREST

The authors declare that they have no competing interests.

AUTHORS' CONTRIBUTIONS

SV and MSH conceived the main idea with input from MM and SV collected the data; and MM and SV analyzed the data with support from MSH. Sampling was done by SV and MSH. The manuscript was written by SV, MSH, and MM. All authors made substantial contributions to the interpretation of results and the editing of the manuscript.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

This study complied with the appropriate institutional, national, and international guidelines.

DATA AVAILABILITY STATEMENT

MtDNA haplotypes used in this study were deposited in the NCBI Nucleotide Database under accession numbers MN995069 to MN995079.

ORCID

Mozafar Sharifi  <https://orcid.org/0000-0003-0826-3000>

Somaye Vaissi  <https://orcid.org/0000-0003-3389-1877>

REFERENCES

- Afroosheh, M., Akmal, V., Esmaili-Rineh, S., & Sharifi, M. (2016). On the distribution, abundance and conservation status of the endangered yellow spotted mountain newt *Neureergus microspilotus* (Caudata: Salamandridae) in western Iran. *Herpetological Conservation and Biology*, 11(1), 52–60.
- Afroosheh, M., Rödder, D., Mikulicek, P., Akmal, V., Vaissi, S., Fleck, J., ... Sharifi, M. (2019). Mitochondrial DNA variation and Quaternary range dynamics in the endangered Yellow Spotted Mountain Newt, *Neureergus derjugini* (Caudata, Salamandridae). *Journal of Zoological Systematics and Evolutionary Research*, 57(3), 580–590. <https://doi.org/10.1111/jzs.12275>
- Ahmadzadeh, F., Carretero, M. A., Rödder, D., Harris, D. J., Freitas, S. N., Perera, A., & Böhme, W. (2013b). Inferring the effects of past climate fluctuations on the distribution pattern of *Iranolacerta* (Reptilia, Lacertidae): Evidence from mitochondrial DNA and species distribution models. *Zoologischer Anzeiger-A Journal of Comparative Zoology*, 252(2), 141–148. <https://doi.org/10.1016/j.jcz.2012.05.002>
- Ahmadzadeh, F., Flecks, M., Rödder, D., Böhme, W., Ilgaz, Ç., Harris, D. J., ... Carretero, M. A. (2013a). Multiple dispersal out of Anatolia: Biogeography and evolution of oriental green lizards. *Biological Journal of the Linnean Society*, 110(2), 398–408. <https://doi.org/10.1111/bj.12129>
- Arevalo, E., Davis, S. K., & Sites, J. W. Jr (1994). Mitochondrial DNA sequence divergence and phylogenetic relationships among eight chromosome races of the *Sceloporus grammicus* complex (Phrynosomatidae) in central Mexico. *Systematic Biology*, 43(3), 387–418. <https://doi.org/10.1093/sysbio/43.3.387>
- Asadi, A., Montgelard, C., Nazarizadeh, M., Moghaddasi, A., Fatemizadeh, F., Simonov, E., ... Kaboli, M. (2019). Evolutionary history and post-glacial colonization of an Asian pit viper (*Gloydius halys caucasicus*) into Transcaucasia revealed by phylogenetic and phylogeographic analyses. *Scientific Reports*, 9(1), 1224. <https://doi.org/10.1038/s41598-018-37558-8>
- Bennett, K., & Provan, J. (2008). What do we mean by 'refugia'? *Quaternary Science Reviews*, 27(27–28), 2449–2455. <https://doi.org/10.1016/j.quascirev.2008.08.019>
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., ... Drummond, A. J. (2014). BEAST 2: A software platform for Bayesian evolutionary analysis. *PLOS Computational Biology*, 10(4), e1003537. <https://doi.org/10.1371/journal.pcbi.1003537>
- Branco, M., Monnerot, M., Ferrand, N., & Templeton, A. R. (2002). Postglacial dispersal of the European rabbit (*Oryctolagus cuniculus*) on the Iberian Peninsula reconstructed from nested clade and mismatch analyses of mitochondrial DNA genetic variation. *Evolution*, 56(4), 792–803. [https://doi.org/10.1554/0014-3820\(2002\)056\[0792:P-DOTER\]2.0.CO;2](https://doi.org/10.1554/0014-3820(2002)056[0792:P-DOTER]2.0.CO;2)
- Chiocchio, A., Bisconti, R., Zampiglia, M., Nascetti, G., & Canestrelli, D. (2017). Quaternary history, population genetic structure and diversity of the cold-adapted Alpine newt *Ichthyosaura alpestris* in peninsular Italy. *Scientific Reports*, 7(1), 2955. <https://doi.org/10.1038/s41598-017-03116-x>
- Church, S. A., Kraus, J. M., Mitchell, J. C., Church, D. R., & Taylor, D. R. (2003). Evidence for multiple Pleistocene refugia in the postglacial expansion of the eastern tiger salamander, *Ambystoma tigrinum tigrinum*. *Evolution*, 57(2), 372–383. [https://doi.org/10.1554/0014-3820\(2003\)057\[0372:EFMPRI\]2.0.CO;2](https://doi.org/10.1554/0014-3820(2003)057[0372:EFMPRI]2.0.CO;2)
- Clement, M., Posada, D., & Crandall, K. A. (2000). TCS: A computer program to estimate gene genealogies. *Molecular Ecology*, 9(10), 1657–1659. <https://doi.org/10.1046/j.1365-294x.2000.01020.x>
- Dianat, M., Darvish, J., Cornette, R., Aliabadian, M., & Nicolas, V. (2017). Evolutionary history of the Persian jird, *Meriones persicus*, based on genetics, species distribution modelling and morphometric data. *Journal of Zoological Systematics and Evolutionary Research*, 55(1), 29–45. <https://doi.org/10.1111/jzs.12145>
- Djamali, M., Baumel, A., Brewer, S., Jackson, S., Jw, K., López-Vinyallonga, S., Simakova, A. N. (2012). Ecological implications of Cousinia Cass. (Asteraceae) persistence through the last two glacial–interglacial cycles in the continental Middle East for the Irano-Turanian flora. *Review of Palaeobotany and Palynology*, 172, 10–20. <https://doi.org/10.1016/j.revpalbo.2012.01.005>
- Djamali, M., de Beaulieu, J.-L., Shah-hosseini, M., Andrieu-Ponel, V., Ponel, P., Amini, A., ... Brewer, S. (2008). A late Pleistocene long pollen record from Lake Urmia, NW Iran. *Quaternary Research*, 69(3), 413–420. <https://doi.org/10.1016/j.yqres.2008.03.004>
- Drummond, A. J., & Rambaut, A. (2007). BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology*, 7(1), 214. <https://doi.org/10.1186/1471-2148-7-214>
- Excoffier, L., Laval, G., & Schneider, S. (2005). Arlequin (version 3.0): An integrated software package for population genetics data analysis. *Evolutionary Bioinformatics*, 1, 117693430500100003. <https://doi.org/10.1177/117693430500100003>
- Fedorov, V. B., & Stenseth, N. C. (2002). Multiple glacial refugia in the North American Arctic: Inference from phylogeography of the colored lemming (*Dicrostonyx groenlandicus*). *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 269(1505), 2071–2077. <https://doi.org/10.1098/rspb.2002.2126>
- Fritz, U., Alcalde, L., Vargas-Ramirez, M., Goode, E. V., Fabius-Turoblin, D. U., & Prasczag, P. (2012). Northern genetic richness and southern purity, but just one species in the *Chelonoidis chilensis* complex. *Zoologica Scripta*, 41(3), 220–232. <https://doi.org/10.1111/j.1463-6409.2012.00533.x>
- Fritz, U., Ayaz, D., Buschbom, J., Kami, H. G., Mazanaeva, L. F., Aloufi, A. A., ... Hundsdoerfer, A. K. (2008). Go east: Phylogeographies of *Mauremys caspica* and *M. rivulata*—discordance of morphology, mitochondrial and nuclear genomic markers and rare hybridization. *Journal of Evolutionary Biology*, 21(2), 527–540. <https://doi.org/10.1111/j.1420-9101.2007.01485.x>
- Fritz, U., Ayaz, D., Hundsdoerfer, A. K., Kottenko, T., Guicking, D., Wink, M., ... Buschbom, J. (2009). Mitochondrial diversity of European pond turtles (*Emys orbicularis*) in Anatolia and the Ponto-Caspian Region: Multiple old refuges, hotspot of extant diversification and critically endangered endemics. *Organisms Diversity and Evolution*, 9(2), 100–114. <https://doi.org/10.1016/j.ode.2009.02.002>

- García-París, M., Alcobendas, M., Buckley, D., & Wake, D. B. (2003). Dispersal of viviparity across contact zones in Iberian populations of fire salamanders (*Salamandra*) inferred from discordance of genetic and morphological traits. *Evolution*, 57(1), 129–143. <https://doi.org/10.1111/j.0014-3820.2003.tb00221.x>
- Ghahroudi Tali, M., Naeimi, A., & Gharnaie, R. (2016). Glacial landforms in border mountainous areas between Iran and Iraq. *Asian Journal of Earth Sciences*, 9, 16–26. <https://doi.org/10.3923/ajes.2016.16.26>
- Giovannotti, M., Nisi-Cerioni, P., & Caputo, V. (2010). Mitochondrial DNA sequence analysis reveals multiple Pleistocene glacial refugia for *Podarcis muralis* (Laurenti, 1768) in the Italian Peninsula. *Italian Journal of Zoology*, 77(3), 277–288. <https://doi.org/10.1080/1125000903143885>
- Gómez, A., & Lunt, D. H. (2007). Refugia within refugia: Patterns of phylogeographic concordance in the Iberian Peninsula. In *Phylogeography of southern European refugia* (pp. 155–188). Berlin, Germany: Springer. https://doi.org/10.1007/1-4020-4904-8_5
- Guindon, S., Dufayard, J.-F., Lefort, V., Anisimova, M., Hordijk, W., & Gascuel, O. O. (2010). New algorithms and methods to estimate maximum-likelihood phylogenies: Assessing the performance of PhyML 3.0. *Systematic Biology*, 59(3), 307–321. <https://doi.org/10.1093/sysbio/syq010>
- Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, 41, 95–98. https://doi.org/10.14601/Phytopathol_Mediterr-14998u1.29
- Hampe, A., & Jump, A. S. (2011). Climate relicts: Past, present, future. *Annual Review of Ecology, Evolution, and Systematics*, 42, 313–333. <https://doi.org/10.1146/annurev-ecolsys-102710-145015>
- Hewitt, G. M. (2011). Quaternary phylogeography: The roots of hybrid zones. *Genetica*, 139(5), 617–638. <https://doi.org/10.1007/s10709-011-9547-3>
- Hijmans, R. J., Guarino, L., & Mathur, P. (2004). DIVA-GIS. Version 7.5. A geographic information system for the analysis of species distribution data. Retrieved from <http://www.diva-gis.org/download>
- Hoffmann, A. A., & Sgro, C. M. (2011). Climate change and evolutionary adaptation. *Nature*, 470(7335), 479. <https://doi.org/10.1038/nature09670>
- Holderregger, R., & Thiel-Egenter, C. (2009). A discussion of different types of glacial refugia used in mountain biogeography and phylogeography. *Journal of Biogeography*, 36(3), 476–480. <https://doi.org/10.1111/j.1365-2699.2008.02027.x>
- Ikeda, D. H., Max, T. L., Allan, G. J., Lau, M. K., Shuster, S. M., & Whitham, T. G. (2017). Genetically informed ecological niche models improve climate change predictions. *Global Change Biology*, 23(1), 164–176. <https://doi.org/10.1111/gcb.13470>
- Jandzik, D., Jablonski, D., Zinenko, O., Kukushkin, O. V., Moravec, J., & Gvozdík, V. (2018). Pleistocene extinctions and recent expansions in an anigid lizard of the genus *Pseudopus*. *Zoologica Scripta*, 47(1), 21–32. <https://doi.org/10.1111/zsc.12256>
- Javanbakht, H., Ihlow, F., Jablonski, D., Široký, P., Fritz, U., Rödder, D., ... Mikuliček, P. (2017). Genetic diversity and quaternary range dynamics in Iranian and Transcaucasian tortoises. *Biological Journal of the Linnean Society*, 121(3), 627–640. <https://doi.org/10.1093/biolinnean/blx001>
- Kehl, M. (2009). Quaternary climate change in Iran—the state of knowledge. *Erdkunde*, 63(1), 1–17. <https://doi.org/10.3112/erdkunde>
- Keppel, G., Van Niel, K. P., Wardell-Johnson, G. W., Yates, C. J., Byrne, M., Mucina, L., ... Franklin, S. E. (2012). Refugia: Identifying and understanding safe havens for biodiversity under climate change. *Global Ecology and Biogeography*, 21(4), 393–404. <https://doi.org/10.1111/j.1466-8238.2011.00686.x>
- Kimura, M. (1980). A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16(2), 111–120. <https://doi.org/10.1007/BF01731581>
- Malyarchuk, B., Derenko, M., & Denisova, G. (2013). Phylogeny and genetic history of the Siberian salamander (*Salmandrella keyserlingii*, Dybowski, 1870) inferred from complete mitochondrial genomes. *Molecular Phylogenetics and Evolution*, 67(2), 348–357. <https://doi.org/10.1016/j.ympev.2013.02.004>
- Mattocchia, M., Marta, S., Romano, A., & Sbordoni, V. (2011). Phylogeography of an Italian endemic salamander (genus *Salandrina*): Glacial refugia, postglacial expansions, and secondary contact. *Biological Journal of the Linnean Society*, 104(4), 903–992. <https://doi.org/10.1111/j.1095-8312.2011.01747.x>
- Ohlemüller, R., Huntley, B., Normand, S., & Svenning, J. C. (2012). Potential source and sink locations for climate-driven species range shifts in Europe since the Last Glacial Maximum. *Global Ecology and Biogeography*, 21(2), 152–163. <https://doi.org/10.1111/j.1466-8238.2011.00674.x>
- Özdemir, N., Üzümlü, N., Avci, A., & Olgun, K. (2009). Phylogeny of *Neurergus crocatus* and *Neurergus strauchii* in Turkey based on morphological and molecular data. *Herpetologica*, 65, 280–291. <https://doi.org/10.1655/07-047R2.1>
- Posada, D. (2008). jModelTest: Phylogenetic model averaging. *Molecular Biology and Evolution*, 25(7), 1253–1256. <https://doi.org/10.1093/molbev/msn083>
- Rajaei Sh, H., Rödder, D., Weigand, A. M., Dambach, J., Raupach, M. J., & Wägele, J. W. (2013). Quaternary refugia in southwestern Iran: Insights from two sympatric moth species (Insecta, Lepidoptera). *Organisms Diversity and Evolution*, 13, 409–423. <https://doi.org/10.1007/s13127-013-0126-6>
- Rambaut, A. (2016). FigTree version 1.4.0. Retrieved from <http://tree.bio.ed.ac.uk/software/figtree>
- Rambaut, A., Suchard, M., Xie, W., & Drummond, A. (2016). Tracer v1.6. 2013. Retrieved from <http://beast.bio.ed.ac.uk/Tracer>
- Ronquist, F., Teslenko, M., Van Der Mark, P., Ayres, D. L., Darling, A., Höhna, S., ... Huelsenbeck, J. P. (2012). MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology*, 61(3), 539–542. [10.1093/sysbio/sys1029](https://doi.org/10.1093/sysbio/sys1029)
- Rossiter, S. J., Benda, P., Dietz, C., Zhang, S., & Jones, G. (2007). Rangewide phylogeography in the greater horseshoe bat inferred from microsatellites: Implications for population history, taxonomy and conservation. *Molecular Ecology*, 16(22), 4699–4714. <https://doi.org/10.1111/j.1365-294X.2007.03546.x>
- Rozas, J., Librado, P., Sánchez-Del Barrio, J., Messeguer, X., & Rozas, R. (2010). DnaSP version 5 help contents [Help File]. Retrieved from <http://www.ub.edu/dnasp>
- Saberi-Pirooz, R., Shahriari, A., Ahmadzadeh, F., Javidkar, M., Oladi, M., & Asadi-Aghbolaghi, M. (2019). Dispersal beyond geographic barriers: A contribution to the phylogeny and demographic history of *Pristurus rupestris* Blanford, 1874 (Squamata: Sphaerodactylidae) from southern Iran. *Zoology*, 134, 8–15. <https://doi.org/10.1016/j.zool.2019.04.001>
- Salehi, T., Akmalı, V., & Sharifi, M. (2019). Population genetic structure of the endangered yellow spotted mountain newt (*Neurergus derjugini*: Amphibia, Caudata) inferred from mitochondrial DNA sequences. *Herpetological Journal*, 29, 37–47. <https://doi.org/10.33256/hj29.1.3747>
- Schneider, C., & Schneider, W. (2011). The Kurdistan newts of the Genus *Neurergus* in Iraq (Caudata: Salamandridae). *Herpetozoa*, 23(3–4), 3–20.
- Sequeira, F., Alexandrino, J., Rocha, S., Arntzen, J., & Ferrand, N. (2005). Genetic exchange across a hybrid zone within the Iberian endemic golden-striped salamander, *Chioglossa lusitanica*. *Molecular Ecology*, 14(1), 245–254. <https://doi.org/10.1111/j.1365-294X.2004.02390.x>
- Shahabi, S., Akmalı, V., & Sharifi, M. (2017). Taxonomic evaluation of the Greater Horseshoe Bat *Rhinolophus ferrumequinum* (Chiroptera:

- Rhinolophidae) in Iran inferred from the mitochondrial D-Loop gene. *Zoological Science*, 34(4), 361–368. <https://doi.org/10.2108/zs170001>
- Sharifi, M., & Afroosheh, M. (2014). Studying migratory activity and home range of adult *Neurergus microspilotus* (Nesterov, 1916) in the Kavat Stream, western Iran, using photographic identification (Caudata: Salamandridae). *Herpetozoa*, 27(1–2), 77–82.
- Sharifi, M., Karami, P., Akmal, V., Afroosheh, M., & Vaissi, S. (2017). Modeling Geographic Distribution for the Endangered Yellow Spotted Mountain Newt, *Neurergus microspilotus* (Amphibia: Salamandridae) in Iran and Iraq. *Herpetological Conservation and Biology*, 12(2), 488–497.
- Sharifi, M., Naderi, B., & Hashemi, R. (2013). Suitability of the photographic identification method as a tool to identify the endangered yellow spotted newt, *Neurergus microspilotus* (Caudata: Salamandridae). *Russian Journal of Herpetology*, 20(4), 264–270.
- Sharifi, M., Najafi, F., Yossefshahi, H., & Hemmati, Z. (2001). Sacred grove in western Iran. *Journal of Environmental Studies*, 26(5), 49–58.
- Sharifi, M., Rezaei, F., Hosseini, M. M., & Raji, G. (2004). Stratigraphical studies in Hashillan Wetland: Introducing a patterned mire in Western Iran. *Journal of Environmental Studies*, 30(35), 99–107.
- Sotiropoulos, K., Eleftherakos, K., Džukić, G., Kalezić, M. L., Legakis, A., & Polymeni, R. M. (2007). Phylogeny and biogeography of the alpine newt *Mesotriton alpestris* (Salamandridae, Caudata), inferred from mtDNA sequences. *Molecular Phylogenetics and Evolution*, 45(1), 211–226. <https://doi.org/10.1016/j.ympev.2007.03.012>
- Spear, S. F., Peterson, C. R., Matocq, M. D., & Storfer, A. (2005). Landscape genetics of the blotched tiger salamander (*Ambystoma tigrinum melanostictum*). *Molecular Ecology*, 14(8), 2553–2564. <https://doi.org/10.1111/j.1365-294X.2005.02573.x>
- Steinfartz, S., Hwang, U. W., Tautz, D., Öz, M., & Veith, M. (2002). Molecular phylogeny of the salamandrid genus *Neurergus*: Evidence for an intrageneric switch of reproductive biology. *Amphibia-Reptilia*, 23(4), 419–431. <https://doi.org/10.1163/15685380260462338>
- Stewart, J. R., Lister, A. M., Barnes, I., & Dalén, L. (2009). Refugia revisited: Individualistic responses of species in space and time. *Proceedings of the Royal Society B: Biological Sciences*, 277(1682), 661–671. <https://doi.org/10.1098/rspb.2009.1272>
- Tamura, K., Stecher, G., Peterson, D., Filipowski, A., & Kumar, S. (2013). MEGA6: Molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, 30(12), 2725–2729. <https://doi.org/10.1093/molbev/mst197>
- Vaissi, S., Parto, P., & Sharifi, M. (2018). Ontogenetic changes in spot configuration (numbers, circularity, size and asymmetry) and lateral line in *Neurergus microspilotus* (Caudata: Salamandridae). *Acta Zoologica*, 99(1), 9–19. <https://doi.org/10.1111/azo.12187>
- Vamberger, M., Stuckas, H., Vargas-Ramírez, M., Kehlmaier, C., Ayaz, D., Aloufi, A. A., ... Fritz, U. (2017). Unexpected hybridization patterns in Near Eastern terrapins (*Mauremys caspica*, *M. rivulata*) indicate ancient gene flow across the Fertile Crescent. *Zoologica Scripta*, 46(4), 401–413. <https://doi.org/10.1111/zsc.12227>
- Wang, B., Jiang, J., Xie, F., & Li, C. (2013). Phylogeographic patterns of mtDNA variation revealed multiple glacial refugia for the frog species *Feirana taihangnica* endemic to the Qinling Mountains. *Journal of Molecular Evolution*, 76(3), 112–128. <https://doi.org/10.1007/s00239-013-9544-5>
- Wasylkova, K., & Walanus, A. (2004). Timing of aquatic and marshplant successions in different parts of Lake Zeribar, Iran, during the Late Glacial and Holocene. *Acta Palaeobotanica*, 44(2), 129–140.
- Weisrock, D. W., Macey, J. R., Ugurtas, I. H., Larson, A., & Papenfuss, T. J. (2001). Molecular phylogenetics and historical biogeography among salamandrids of the “true” salamander clade: Rapid branching of numerous highly divergent lineages in *Mertensiella luschani* associated with the rise of Anatolia. *Molecular Phylogenetics and Evolution*, 18(3), 434–448. <https://doi.org/10.1006/mpev.2000.0905>
- Wright, H., McAndrews, J. H., & van Zeist, W. (1967). Modern pollen rain in western Iran, and its relation to plant geography and Quaternary vegetational history. *The Journal of Ecology*, 55(2), 415–443. <https://doi.org/10.2307/2257886>
- Yousefi, M., Shabani, A. A., & Azarnivand, H. (2019). Reconstructing distribution of the Eastern Rock Nuthatch during the last glacial maximum and last interglacial. *Avian Biology Research*, 1–7. <https://doi.org/10.1177/1758155919874537>

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