

HHS Public Access

Author manuscript *Muscle Nerve*. Author manuscript; available in PMC 2020 June 11.

Published in final edited form as: *Muscle Nerve*. 2019 March ; 59(3): 357–362. doi:10.1002/mus.26378.

Novel *SPEG* mutations in congenital myopathies: Genotypephenotype correlations

Anita E. Qualls¹, Sandra Donkervoort, MS, CGC², Johanna C. Herkert, MD³, Alissa M. D'Gama, MD, PhD¹, Diana Bharucha-Goebel, MD^{2,4}, James Collins, MD, PhD⁵, Katherine R. Chao, BS⁶, A. Reghan Foley, MD², Mirthe H. Schoots, MD⁷, Jan D.H. Jongbloed, PhD³, Carsten G. Bönnemann, MD², Pankaj B. Agrawal, MD¹

¹Division of Newborn Medicine, Division of Genetics and Genomics, and The Manton Center for Orphan Disease Research, Boston Children's Hospital and Harvard Medical School, Boston, MA ²Neuromuscular and Neurogenetic Disorders of Childhood, National Institutes of Health, Bethesda, MD ³University of Groningen, University Medical Centre Groningen, Department of Genetics, Groningen, the Netherlands ⁴Division of Neurology, Children's National Health System, Washington, DC. ⁵Mercy Clinic Pediatric Neurology, Springfield, MO ⁶Center for Mendelian Genomics at the Broad Institute of MIT and Harvard, Boston, MA ⁷Department of Pathology, University of Groningen, University Medical Centre Groningen, the Netherlands

Abstract

Introduction: Centronuclear myopathies (CNMs) are a subtype of congenital myopathies (CMs) characterized by muscle weakness, predominant type 1 fibers, and increased central nuclei. *SPEG* (striated preferentially expressed protein kinase) mutations have recently been identified in seven CM patients (six with CNMs). We report two additional patients with *SPEG* mutations expanding the phenotype and evaluate genotype-phenotype correlations associated with *SPEG* mutations.

Methods/Results: Using whole exome/genome sequencing in CM families, we identified novel recessive *SPEG* mutations in two patients. Patient 1, with severe muscle weakness requiring respiratory support, dilated cardiomyopathy, ophthalmoplegia, and findings of non-specific CM on muscle biopsy carried a homozygous SPEG mutation (p.Val3062del). Patient 2, with milder muscle weakness, ophthalmoplegia, and CNM carried compound heterozygous mutations (p.Leu728Argfs*82) and (p.Val2997Glyfs*52).

Discussion: The two patients add insight into genotype-phenotype correlations of *SPEG*-associated CMs. Clinicians should consider evaluating a CM patient for *SPEG* mutations even in the absence of CNM features.

Corresponding Author: Pankaj B. Agrawal, MD, Address: 300 Longwood Ave, Boston MA 02115, pagrawal@enders.tch.harvard.edu.

Ethical Publication Statement: We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

Disclosure of Conflicts of Interest: None of the authors has any conflict of interest to disclose.

Keywords

Centronuclear myopathies (CNMs); Striated preferentially expressed protein kinase (SPEG); Congenital myopathies (CMs); Next generation sequencing (NGS); Cardiomyopathy; Myotubularin (MTM1)

Introduction

Congenital myopathies (CMs) are a group of muscle diseases that commonly present at birth or during infancy with muscle weakness and hypotonia. The clinical presentation ranges from mild hypotonia causing delays in achieving motor skills to severe muscle weakness causing death from respiratory involvement¹. Centronuclear myopathies (CNMs) are a subtype characterized by increased central nuclei within myofibers, and often associated with disruption of excitation-contraction coupling^{1,2}. Approximately 60–80% of CNMs are caused by dominant *DNM2* mutations, dominant and recessive *RYR1* and *CACNA1S* mutations, recessive *BIN1* mutations, and X-linked recessive *MTM1* mutations^{3–8}. Recently, recessive *SPEG* mutations have been identified in six CNM patients and one patient with non-CNM CM^{9–12}. Here, we report two additional unrelated patients with CMs caused by recessive *SPEG* mutations, compare the clinical findings of all nine patients, and discuss genotype-phenotype correlations thereby improving the understanding of *SPEG*-related CM.

Methods

Patient Recruitment and Genetic Analysis

For patient 1, a CGH array was initially performed and then whole exome sequencing (WES) was performed in a diagnostic setting with a parent–offspring trio approach as previously described¹³. For Patient 2, the patient and her family were enrolled in an IRB-approved study (NINDS Protocol 12-N-0095). WES was initially performed through the NIH Intramural Sequencing Center using the Nimblegen SeqCap EZ Exome+UTR Library and Illumina HiSeq, and variants were analyzed using Varsifter¹⁴. Whole genome sequencing (WGS) was then performed by the Genomics Platform at the Broad Institute using Illumina HiSeq X Ten v2 chemistry, and variants were analyzed using Variant Effect Predictor.

Histopathology Studies

The muscle biopsy samples were frozen and processed using standard histological techniques¹⁵.

Results

Clinical Description

Patient 1 was the first child of healthy consanguineous parents, with normal intellect and no family history of neuromuscular disease. He has been reported in a large series of cardiomyopathy patients with minimal clinical information¹³. The pregnancy was reportedly uncomplicated, and he was delivered by vacuum extraction at 37 weeks gestation. At birth,

he presented with severe hypotonia and left-sided inguinal hernia. At age 4, he developed progressive proximal muscle weakness and was noted to have marked atrophy of his lower leg muscles, pes planovalgus, and a high-arched palate. His history was significant for recurrent abdominal pain and diarrhea, recurrent otitis media, frequent upper airway infections, recurrent pneumonias, and multiple bone fractures (distal ulna, medial condyle, distal tibia, all after trauma). His serum creatine kinase level ranged from 9–60 U/l (normal <171 U/l)¹⁶. At age 6, an electrocardiogram (EKG) showed biventricular hypertrophy and an echocardiogram demonstrated severe left ventricular dilation with poor muscle contractility. He was started on digoxin, captopril and diuretics, tube feeding, and nocturnal noninvasive ventilation. At age 7, a gastrostomy tube was inserted. At age 12, ophthalmoplegia and mild lumbar torsion-scoliosis was diagnosed. His dilated cardiomyopathy was progressive; his shortening fraction decreased from 20% at age 10 to 9% at age 16 with severe mitral valve insufficiency. Despite maximum support, he died at age 17 due to cardiopulmonary insufficiency.

Patient 2 is a 6.5-year-old female. She was born at term via Cesarean section and presented with a weak cry, respiratory distress, hypotonia, and reduced deep tendon reflexes. She had bilateral vocal cord paralysis, and a gastrostomy tube was placed at age 4 weeks due to swallowing concerns. She attained head control at 4 months, rolled over at 6–9 months, got into a sitting position at 9-12 months, crawled at 18 months, pulled to stand at 18-20 months, and walked at 2 years. EKG at 3 years and 10 months revealed sinus tachycardia; an echocardiogram was normal. Her serum creatine kinase level was within normal limits at 89 IU/l. At age 4, she had mild lower facial weakness, axial hypotonia and proximal muscle weakness (MRC 3-4/5 range) with subgravity neck flexion. She has nearly complete ophthalmoplegia, bilateral ptosis, and intermittent strabismus. She has a high-arched palate and nasal speech. She has a weak cough and has had recurrent respiratory infections. She has difficulty feeding by mouth and receives all nutrition via a gastrostomy tube. Although she has a history of delayed motor milestones, she continues to demonstrate improvements. At age 4.5, she was still unable to run and jump. A nerve conduction study at age 5 showed a reduced compound muscle action potential (CMAP) amplitude of 2.3 mV (normal > 3.0 mV) of the ulnar motor nerve recorded at the abductor digiti minimi muscle.

Muscle Biopsy Findings

Patient 1 had a quadriceps muscle biopsy at age 9, which is consistent with non-CNM CM and shows a mild increase in fiber size variability, several atrophic fibers, and only a few internal/central nuclei (<20% of fibers) (Figure 1a). No clear fiber size hypertrophy is noted. Patient 2 had a quadriceps muscle biopsy at age 3, which is consistent with CNM and shows good fiber type differentiation without clear fiber type predominance, hypotrophic Type 1 fibers and hypertrophic Type 2 fibers, and many central nuclei (~50% of fibers and 60% of Type 1 fibers) (Figure 1b–c). Electron microscopy for Patient 2 shows a few myofibers with unstructured cores.

Genetic Results

Copy number variant analysis for Patient 1 using array-CGH identified a deletion of chromosome 4q35.2 (190,462,807–191,041,681; 579 kb), a deletion of chromosome

7q11.22 (66,692,376–68,103,955; 1,412 Mb), and copy neutral homozygosity of 6 regions >10 Mb, confirming consanguinity. Both deletions did not correlate to a phenotype and were identified in his father; the first includes *BC087857* and the second does not include any genes. Sanger sequencing of *FKRP*, *SEPN1*, and *RYR1* was unrevealing. Trio WES identified a homozygous mutation in exon 38 of *SPEG*, c.9185_9187delTGG (p. (Val3062del)). The amino acid at this position is highly conserved and located in the protein kinase domain, which is critical for SPEG function. This variant was heterozygous in the parents and unaffected sister.

WES analysis for Patient 2 initially identified a maternally inherited c.2183delT (p. (Leu728Argfs*82)) mutation in exon 10 of *SPEG*. Due to regions of low coverage, WGS was then performed, and identified the same maternally inherited mutation in compound heterozygosity with a paternally inherited 25 base pair insertion in exon 38, c.8962_8963insCGGGGGGGAACGTTCGTGGCCAAGAT (p.(Val2997Glyfs*52)). These variants result in frameshifts and thus are classified as loss-of-function. The variants identified in both patients were predicted deleterious by MutationTaster and absent from ExAC, gnomAD, and 1000 Genomes databases.

Discussion

We report two additional patients with *SPEG*-associated CMs, Patient 1 with muscle pathology consistent with non-specific CM and Patient 2 with muscle pathology consistent with CNM (P1 and P2 in Figure 1d). The clinical and molecular findings of all nine patients reported so far including ours are summarized in Table 1 and pathological findings are described in Supplementary Table 1.

SPEG is alternatively spliced into 4 tissue-specific isoforms: APEG (aortic preferentially expressed gene), BPEG (brain preferentially expressed gene), and SPEGa and SPEG β (expressed in skeletal and cardiac muscle)¹⁷. SPEG β is the longer isoform with SPEGa missing amino acids 1–854⁹. Clinical data from Patients 6 and 8 suggests that SPEGa may partially rescue mutations affecting only SPEG β , possibly preserving cardiac function^{10,12}. This appears to be the case for Patient 2, who carries one variant sparing SPEGa, and has not yet developed signs of cardiac dysfunction. In contrast, Patient 1 carried a homozygous mutation affecting SPEGa and SPEG β , and developed dilated cardiomyopathy, also seen in Patients 3, 4, 5 and 7 carrying mutations affecting both isoforms^{9,10}. Interestingly, Patient 9, who also carries a mutation affecting both isoforms, developed non-compaction cardiomyopathy¹¹.

Skeletal muscle dysfunction seems more severe in patients with mutations affecting both isoforms, as seen in Patients 1, 3, and 9 dying early, and Patient 4 needing constant mechanical ventilation^{9,11}. The other two patients with both isoforms affected are Patients 5 and 7^{9,10}. In Patient 5, the disease was relatively mild, likely due to one variant being missense while all *SPEG* variants described so far have been loss-of-function, suggesting haploinsufficiency⁹. In Patient 7, the milder phenotype may be due to the mutation being very close to the C-terminus, thereby escaping nonsense mediated decay and potentially having less effect on protein function¹⁰. Overall, these findings suggest SPEGa has a critical

role in skeletal and cardiac function and the disease is more severe when both isoforms are affected. Future studies should investigate the role of SPEGa in compensating for mutant SPEG β . The clinical features of all patients have phenotypic similarities, most notably the presence of respiratory problems (Patients 1–7, and 9), eye involvement (Patients 1,2, 4, and 6), and scoliosis (Patients 1, 6, and 8)^{9–12}.

In summary, this study expands the genetic heterogeneity of *SPEG*-associated CMs and further elucidates genotype-phenotype correlations to help guide appropriate clinical screening and management. The phenotype of *SPEG*-associated CM is varied and expanding, including ophthalmoplegia, and diagnostic markers that were initially considered, such as the presence of centralized nuclei on muscle biopsy and dilated cardiomyopathy, do not capture all cases. Thus, it is important for clinicians to consider evaluating a patient with congenital myopathy for *SPEG* mutations using WES even in the absence of type 1 fiber predominance, central nuclei, or cardiomyopathy.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

The authors thank the families for their participation in the study, Daniel Ezzo for help with data analysis, Dr. Anne Rutkowski and CureCMD for help with patient recruitment, and Gilberto (Mike) Averion and Christopher Mendoza for clinical support. For Patient 2, initial whole exome sequencing was funded by the Clinical Center Genomics Opportunity, which is sponsored by the National Human Genome Research Institute, the NIH Deputy Director for Intramural Research, and the NIH Clinical Center. Whole genome sequencing for the same patient was performed at the Broad Center for Mendelian Genomics (CMG) (UM1 HG008900), funded by the National Human Genome Research Institute with supplemental funding provided by the National Heart, Lung, and Blood Institute under the Trans-Omics for Precision Medicine program and the National Isstitute. CGB was supported by NIH Intramural Research Program funding from the National Institute of Neurological Disorders and Stroke. AMD was supported by the National Institute of General Medical Sciences (T32GM007753). PBA was supported by NIH/NIAMS 1R01AR068429-01.

Abbreviations

CGH	comparative genomic hybridization
CMs	congenital myopathies
CMG	Center for Mendelian Genomics
CNMs	centronuclear myopathies (CNMs)
EKG	electrocardiography
SPEG	Striated preferentially expressed gene
WES	whole exome sequencing
WGS	whole genome sequencing

References:

- 1. Nance JR, Dowling JJ, Gibbs EM, Bonnemann CG. Congenital myopathies: an update. Curr Neurol Neurosci Rep 2012;12(2):165–174. [PubMed: 22392505]
- 2. Pierson CR, Tomczak K, Agrawal P, Moghadaszadeh B, Beggs AH. X-linked myotubular and centronuclear myopathies. J Neuropathol Exp Neurol 2005;64(7):555–564. [PubMed: 16042307]
- Bevilacqua JA, Monnier N, Bitoun M, Eymard B, Ferreiro A, Monges S, Lubieniecki F, Taratuto AL, Laquerriere A, Claeys KG, Marty I, Fardeau M, Guicheney P, Lunardi J, Romero NB. Recessive RYR1 mutations cause unusual congenital myopathy with prominent nuclear internalization and large areas of myofibrillar disorganization. Neuropathol Appl Neurobiol 2011;37(3):271–284. [PubMed: 21062345]
- 4. Bitoun M, Maugenre S, Jeannet PY, Lacene E, Ferrer X, Laforet P, Martin JJ, Laporte J, Lochmuller H, Beggs AH, Fardeau M, Eymard B, Romero NB, Guicheney P. Mutations in dynamin 2 cause dominant centronuclear myopathy. Nat Genet 2005;37(11):1207–1209. [PubMed: 16227997]
- Ceyhan-Birsoy O, Agrawal PB, Hidalgo C, Schmitz-Abe K, DeChene ET, Swanson LC, Soemedi R, Vasli N, Iannaccone ST, Shieh PB, Shur N, Dennison JM, Lawlor MW, Laporte J, Markianos K, Fairbrother WG, Granzier H, Beggs AH. Recessive truncating titin gene, TTN, mutations presenting as centronuclear myopathy. Neurology 2013;81(14):1205–1214. [PubMed: 23975875]
- Laporte J, Hu LJ, Kretz C, Mandel JL, Kioschis P, Coy JF, Klauck SM, Poustka A, Dahl N. A gene mutated in X-linked myotubular myopathy defines a new putative tyrosine phosphatase family conserved in yeast. Nat Genet 1996;13(2):175–182. [PubMed: 8640223]
- Nicot AS, Toussaint A, Tosch V, Kretz C, Wallgren-Pettersson C, Iwarsson E, Kingston H, Garnier JM, Biancalana V, Oldfors A, Mandel JL, Laporte J. Mutations in amphiphysin 2 (BIN1) disrupt interaction with dynamin 2 and cause autosomal recessive centronuclear myopathy. Nat Genet 2007;39(9):1134–1139. [PubMed: 17676042]
- 8. Schartner V, Romero NB, Donkervoort S, Treves S, Munot P, Pierson TM, Dabaj I, Malfatti E, Zaharieva IT, Zorzato F, Abath Neto O, Brochier G, Lornage X, Eymard B, Taratuto AL, Bohm J, Gonorazky H, Ramos-Platt L, Feng L, Phadke R, Bharucha-Goebel DX, Sumner CJ, Bui MT, Lacene E, Beuvin M, Labasse C, Dondaine N, Schneider R, Thompson J, Boland A, Deleuze JF, Matthews E, Pakleza AN, Sewry CA, Biancalana V, Quijano-Roy S, Muntoni F, Fardeau M, Bonnemann CG, Laporte J. Dihydropyridine receptor (DHPR, CACNA1S) congenital myopathy. Acta Neuropathol 2017;133(4):517–533. [PubMed: 28012042]
- Agrawal PB, Pierson CR, Joshi M, Liu X, Ravenscroft G, Moghadaszadeh B, Talabere T, Viola M, Swanson LC, Haliloglu G, Talim B, Yau KS, Allcock RJ, Laing NG, Perrella MA, Beggs AH. SPEG Interacts with Myotubularin, and Its Deficiency Causes Centronuclear Myopathy with Dilated Cardiomyopathy. Am J Hum Genet 2014;95(2):218–226. [PubMed: 25087613]
- Wang H, Castiglioni C, Kacar Bayram A, Fattori F, Pekuz S, Araneda D, Per H, Erazo R, Gumus H, Zorludemir S, Becker K, Ortega X, Bevilacqua JA, Bertini E, Cirak S. Insights from genotypephenotype correlations by novel SPEG mutations causing centronuclear myopathy. Neuromuscul Disord 2017;27(9):836–842. [PubMed: 28624463]
- Wang H, Schanzer A, Kampschulte B, Daimaguler HS, Logeswaran T, Schlierbach H, Petzinger J, Ehrhardt H, Hahn A, Cirak S. A novel SPEG mutation causes non-compaction cardiomyopathy and neuropathy in a floppy infant with centronuclear myopathy. Acta Neuropathol Commun 2018;6(1):83. [PubMed: 30157964]
- Lornage X, Sabouraud P, Lannes B, Gaillard D, Schneider R, Deleuze JF, Boland A, Thompson J, Bohm J, Biancalana V, Laporte J. Novel SPEG Mutations in Congenital Myopathy without Centralized Nuclei. J Neuromuscul Dis 2018;5(2):257–260. [PubMed: 29614691]
- 13. Herkert JC, Abbott KM, Birnie E, Meems-Veldhuis MT, Boven LG, Benjamins M, du Marchie Sarvaas GJ, Barge-Schaapveld D, van Tintelen JP, van der Zwaag PA, Vos YJ, Sinke RJ, van den Berg MP, van Langen IM, Jongbloed JDH. Toward an effective exome-based genetic testing strategy in pediatric dilated cardiomyopathy. Genet Med 2018.
- Teer JK, Green ED, Mullikin JC, Biesecker LG. VarSifter: visualizing and analyzing exome-scale sequence variation data on a desktop computer. Bioinformatics 2012;28(4):599–600. [PubMed: 22210868]
- 15. Dubowitz V, Sewry CA, Oldfors A. Muscle biopsy: A practical approach: Elsevier; 2013.

- 16. Schumann G, Bonora R, Ceriotti F, Clerc-Renaud P, Ferrero CA, Ferard G, Franck PF, Gella FJ, Hoelzel W, Jorgensen PJ, Kanno T, Kessne A, Klauker R, Kristiansen N, Lessinger JM, Linsinger TP, Misaki H, Panteghini M, Pauwels J, Schimmel HG, Vialle A, Weidemann G, Siekmann L. IFCC primary reference procedures for the measurement of catalytic activity concentrations of enzymes at 37 degrees C. Part 2. Reference procedure for the measurement of catalytic concentration of creatine kinase. Clin Chem Lab Med 2002;40(6):635–642. [PubMed: 12211662]
- Hsieh CM, Fukumoto S, Layne MD, Maemura K, Charles H, Patel A, Perrella MA, Lee ME. Striated muscle preferentially expressed genes alpha and beta are two serine/threonine protein kinases derived from the same gene as the aortic preferentially expressed gene-1. J Biol Chem 2000;275(47):36966–36973. [PubMed: 10973969]

D





Figure 1:

Histological examination of patients' muscle biopsies and *SPEG* schematic. (A) Hematoxylin & eosin (H&E) staining of Patient 1's muscle biopsy specimen, performed at 9 years of age. The muscle biopsy shows a mild increase in fiber size variability, several atrophic fibers, and only a few internal/central nuclei, consistent with non-CNM CM. (B) SDH and (C) H&E staining of Patient 2's muscle biopsy, performed at 3 years of age. The muscle biopsy reveals marked variability in fiber size with hypotrophic type 1 fibers and hypertrophic type II fibers with many central nuclei, consistent with CNM. Scale bar 50μm for all images. (D) Schematic of SPEGβ domain organization with positions of identified mutations generated by IBS (Illustrator for Biological Sequences). Mutations affecting both SPEGα and SPEGβ are in black, while mutations affecting only SPEGβ are in pink.

Author Manuscript

Author Manuscript

SPEG Mutations	
arrying	1
Ű	
ıals	
idu	
div	
l In	
sir	
ding)
Εï	
ecular	
D	
2	
and	
äl	
Clinic	

Patient/Sex	P1/M (this study)	P2/F (this study)	P3/F9	P4/F9	PS/M ⁹	$P_{6/M}10$	P7/M10	P8/M12	P9/M11
Age (when reported)	Died at 17 years	6.5 years	Died at 3 weeks	6 years	1.5 years	3 years	7 years	10 years	Died at 19 weeks
SPEG exons	Exon 38	Exon 10 and 38	Exon 30	Exons 18 and 13	Exons 10 and 35	Exon 4	Exon 40	Exon 4 and 20	Exon 30
Allele 1 (maternal)	c.9185_9187delTGG; p.Val3062del	c.2183defT; p.Leu728fs	c.6697C>T; p.Gln2233*	c.4276C>T; p.Arg1426*	c.2915_2916delCCinsA; p.Ala972fs	c.1627–1628insA; p.Thr544fs	c.9586C>T; p.Arg3196*	c.1071_1074dup; p.Lys359fs	c.7119C>A p.Tyr2373*
Allele 2 (Paternal)	same as above	c.8962_8963ins25; p.Val2997fs	same as above	c.3709_3715+29del36; p.Thr1237fs	c.8270G>T; p.Gly2757Val	same as above	same as above	c.4399C>T; p.Arg1467 ^a	same as above
Family History	Consanguineous parents, one healthy sister	No known consanguinity	Consanguineous parents, two sisters died early	No known consanguinity	No known consanguinity, sibling died early	parents from village in Turkey	Likely consanguineous	Non-consanguineous	Consanguineous parents
Birth History	Full term, severely hypotonic	Full-term, hypotonic	Full-term, breech delivery, severely hypotonic	Severely hypotonic	Born at 36 weeks of gestation, severely hypotonic	Full-term, hypotonic	Full-term, poor fetal movements	Uneventful pregnancy, hypotonic	Uneventful pregnancy, severely hypotonic
Neurological Findings	symmetric atrophy of lower extremities, wheel chair bound at 17 years	normal early motor milestones, walked at 2 years, unable to run or jump	Died of severe muscle weakness	Sit unsupported at 2.5 years, unable to walk unsupported	Head control at 16 months, sit unsupported at 18 months	Head control - 6 months, sit unsupported - 12 months, unable to walk	Head control at 18 months, sitting at 30 months, walking- 4 years	sit-11 months, walk - 30 months, short distances	Contracture of right ankle and lacked deep tendon reflex, antigravity movement at 1 week
Eye Findings	Ophthalmoplegia	Ophthalmoplegia, bilateral ptosis	No known evaluation	Ophthalmoplegia	None	ophthalmoplegia, mild ptosis	None	None	None
Respiratory Issues	non-invasive ventilation during night, recurrent pneumonia	Weak cough	Insufficient respiratory efforts	Trache ostomy, mechanical ventilation dependent	brief NICU stay for respiratory issues, no assisted ventilation	NICU for apnea, no intubations, recurrent lung infections	non-invasive ventilation during first 48 hours of life	None	Intubation required immediately after birth, weaned at 10 weeks for palliative care
Feeding Issues	Gastrostomy tube from age 6	gastrostomy tube	Gastrostomy tube early in life	Gastrostomy tube early in life	NG feeding	None	NG feeding until day 13	Gastrostomy tube from age 9	Gastrostomy tube
Cardiac Issues	Dilated cardiomyopathy at age 7, severe mitral valve insufficiency	No cardiomyopathy at 3 years 10 months, sinus tachycardia	No cardiac evaluation	Dilated cardiomyopathy	Dilated cardionyopathy, mitral valve insufficiency	None	Dilated cardionyopathy mild mitral insufficiency	Reduced myocardial contraction, no ventricular dilation at 5 year	Enlarged atria, abnormal trabeculation of left ventricle
Skeletal Issues	Torsion scoliosis	Ultrar fracture at age 4, condyle fracture at age 5, tibia fracture at age 11 (all after trauma)	Not applicable	None	None	Pectus excavatum and mild scoliosis	None	Scoliosis developed at age 4	Not applicable