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# Letters to the Editor

mortality and in an analogous fashion to patterns seen in sepsis.<sup>10</sup> We still encourage clinicians to remain vigilant for drug-induced liver injury, and for liver damage in high-risk groups (*i.e.* drug/alcohol abusers, family history *etc.*), but not to get overly distracted by raised liver aminotransferases in this context.

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## **Conflict of interest**

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## **Authors' contributions**

All authors contributed equally to the content and production of the letter, all authors checked and approved the final draft of the letter.

## Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.jhep.2020.05.035.

## References

Author names in bold designate shared co-first authorship

- Wang Y, Liu S, Liu H, Li W, Lin F, Jiang L, et al. SARS-CoV-2 infection of the liver directly contributes to hepatic impairment in patients with COVID-19. J Hepatol 2020;73(4):807–816.
- [2] Bangash MN, Patel J, Parekh D. COVID-19 and the liver: little cause for concern. Lancet Gastroenterol Hepatol 2020;5(6):529–530.
- [3] Li W, Deng M, Loughran PA, Yang M, Lin M, Yang C, et al. LPS induces active HMGB1 release from hepatocytes into exosomes through the coordinated activities of TLR4 and caspase-11/GSDMD signaling. Front Immunol 2020;11:229.
- [4] Grasl-Kraupp B, Ruttkay-Nedecky B, Koudelka H, Bukowska K, Bursch W, Schulte-Hermann R. In situ detection of fragmented DNA (TUNEL assay) fails to discriminate among apoptosis, necrosis, and autolytic cell death: a cautionary note. Hepatology 1995;21(5):1465–1468.
- [5] Miller SE, Brealey JK. Visualization of putative coronavirus in kidney. Kidney Int 2020;98(1):231–232.

- [6] Wang A, Medzhitov R. Not the usual suspect: type I interferon-responsive T cells drive infection-induced cachexia. Nat Immunol 2019;20(6): 666–667.
- [7] Siddall E, Khatri M, Radhakrishnan J. Capillary leak syndrome: etiologies, pathophysiology, and management. Kidney Int 2017;92(1):37–46.
- [8] Zuo Y, Yalavarthi S, Shi H, Gockman K, Zuo M, Madison JA, et al. Neutrophil extracellular traps in COVID-19. JCI Insight 2020;5:e138999.
- [9] Liu Y, Fang Y, Chen X, Wang Z, Liang X, Zhang T, et al. Gasdermin Emediated target cell pyroptosis by CAR T cells triggers cytokine release syndrome. Sci Immunol 2020;5(43):eaax7969.
- [10] Kobashi H, Toshimori J, Yamamoto K. Sepsis-associated liver injury: incidence, classification and the clinical significance. Hepatol Res 2013;43(3):255–266.

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# Reply to: Correspondence relating to "SARS-CoV-2 infection of the liver directly contributes to hepatic impairment in patients with COVID-19"

To the Editor:

We appreciate the thoughtful comments on our paper. Our study focused on coronavirus disease 2019 (COVID-19)-related liver

enzyme abnormalities, rather than acute liver injury. Although severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) could infect the liver, it is not a hepatotropic virus that induces pronounced hepatitis. In keeping with this, abnormal liver function tests during the course of COVID-19 are common and primarily limited to aminotransferases, while clinically severe

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acute liver injury is rare.<sup>1,2</sup> This also explains the mild transaminase elevation and other well-preserved parameters, such as lactate, glucose and international normalized ratio, which most likely result from direct virus infection.

Regarding biochemical parameters, the median gamma glutamyltransferase (GGT) levels in both groups were in the normal range and there were no significant differences in levels of total or direct bilirubin and alkaline phosphatase, suggesting that cholangiocytes are unlikely the target sites. Aspartate aminotransferase was recently reported to highly correlate with alanine aminotransferase and reflect disease severity throughout the COVID-19 course, whereas the correlation with markers of muscle injury was weak.<sup>3</sup> Meanwhile, we noted that the median creatinine kinase (CK) was still in the normal range in the abnormal aminotransferases group, and the frequency of elevated CK was comparable between the 2 groups (17.86 % vs. 13.51%, p = 0.89), indicating that elevated CK and abnormal transaminases might not have a causal relationship. Leakage of albumin from blood vessels due to severe vasculitis was usually accompanied by hydrothorax or ascites. However, the albumin reduction in patients with COVID-19 was usually mild. Taken together, these aberrant blood variables appear to be predominantly the result of hepatic impairment.

We stated that SARS-CoV-2 directly contributed to cytopathy based on the ultrastructural findings of conspicuous mitochondria swelling, endoplasmic reticulum dilatation, glycogen granule decrease, and impaired cell membranes. Apoptotic and bi- or multi-nucleated syncytial hepatocytes were also signs of viral effects.<sup>4,5</sup> Previous studies demonstrated direct coronaviruses exerted extensive cytopathic effects through apoptosis, via a caspase-dependent pathway.<sup>4,6–8</sup> The 'coronalike' particles are characteristic of coronavirus particles, but not cholesterol crystals which are normally highly electron dense. structureless and inhomogeneous. Hepatocytes manifested no megamitochondria with linear crystalline inclusions, cristae loss, or increased matrix electrondensity, which would be expected in non-alcoholic fatty liver disease (NAFLD).9-11 None of the reported autolytic microscopic modifications in the liver<sup>12</sup> were observed in our biopsies. Clathrins are generally located at the plasma membrane of synapses and play a role in synaptic vesicle reconstitution.<sup>13</sup> Clathrin possesses variable morphology, such as multiple coated buds which are formed on endosomes, distinctive from typical coronavirus.

In light of the low percentage of hepatocytes expressing ACE2, but not the absolute absence, it is not surprising that SARS-CoV-2 is capable of causing liver injury directly. On the other hand, there exist wide discrepancies between organ symptomatology and ACE2 expression levels.<sup>14,15</sup> For instance, ACE2 expression in the respiratory tract is only moderate compared with that in intestinal epithelia, but respiratory symptomatology is substantially more severe than intestinal.<sup>14,15</sup> It is therefore assumed that direct viral effects on the liver might not operate absolutely through ACE2 expression, and other receptors cannot be excluded. Alternatively, the finding of upregulated ACE2 expression in hepatocytes in cirrhotic liver inspired us to speculate that SARS-CoV-2 infection might induce compensatory hyperplasia of hepatocytes, which are possibly derived from cholangiocytes (with high ACE2 expression).<sup>16</sup> This procedure might result in a higher ACE2 expression in regenerative hepatocytes and facilitate viral infection in turn.

We acknowledge that fatty liver disease, drug-induced liver injury (DILI), sepsis and multiple organ dysfunction syndromes could be involved in liver damage in the setting of COVID-19, but our clinical and pathological findings do not favor these possibilities, as thoroughly described in our paper. Firstly, the prevalence of transaminase abnormalities was up to 23.5% even in the context of mild cases, precluding the role of COVID-19-related comorbidities. Secondly, despite steatosis, both biopsied cases had neither NAFLD nor other metabolic disorders. The pathological features of no obvious eosinophil infiltration, cholestasis, fibrin deposition, vasculitis, granuloma, massive central necrosis, or interface hepatitis did not comply with DILI. Thirdly, only 7 of 64 (10.9%) cases had sepsis and other severe comorbidities, which could not explain the apparently high rate (41.0%) of liver enzyme abnormality in our cohort. Additionally, the liver pathology in our septic case showed no pathological hallmark of sepsis, suggesting sepsis was not the main insult. All in all, our data fully support a direct role of SARS-CoV-2 in COVID-19related hepatic impairment.

In conclusion, we believe SARS-CoV-2 is capable of infecting the liver and is a crucial factor in liver dysfunction, though other factors cannot be fully excluded.

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### **Authors' contributions**

JZ and FL contributed to study concept, critical revision of the manuscript; YW contributed to literature search and writing of the manuscript.

## Supplementary data

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#### References

Author names in bold designate shared co-first authorship

- [1] Garrido I, Liberal R, Macedo G. Review article: COVID-19 and liver disease what we know on 1st May 2020. Aliment Pharmacol Ther 2020;52(2): 267–275.
- [2] Kukla M, Skonieczna-Zydecka K, Kotfis K, Maciejewska D, Loniewski I, Lara LF, et al. COVID-19, MERS and SARS with concomitant liver injurysystematic review of the existing literature. J Clin Med 2020;9(5):1420.
- [3] Bloom PP, Meyerowitz EA, Reinus Z, Daidone M, Gustafson J, Kim AY, et al. Liver biochemistries in hospitalized patients with COVID-19. Hepatology 2020.
- [4] Chau TN, Lee KC, Yao H, Tsang TY, Chow TC, Yeung YC, et al. SARS-associated viral hepatitis caused by a novel coronavirus: report of three cases. Hepatology 2004;39:302–310.
- [5] He L, Ding Y, Zhang Q, Che X, He Y, Shen H, et al. Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected ACE2+ cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS. J Pathol 2006;210:288–297.
- [6] Tan YJ, Fielding BC, Goh PY, Shen S, Tan TH, Lim SG, et al. Overexpression of 7a, a protein specifically encoded by the severe acute respiratory syndrome coronavirus, induces apoptosis via a caspase-dependent pathway. J Virol 2004;78:14043–14047.

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- [7] Eleouet JF, Slee EA, Saurini F, Castagne N, Poncet D, Garrido C, et al. The viral nucleocapsid protein of transmissible gastroenteritis coronavirus (TGEV) is cleaved by caspase-6 and -7 during TGEV-induced apoptosis. J Virol 2000;74:3975–3983.
- [8] Liu C, Xu HY, Liu DX. Induction of caspase-dependent apoptosis in cultured cells by the avian coronavirus infectious bronchitis virus. J Virol 2001;75:6402–6409.
- [9] Lotowska JM, Sobaniec-Lotowska ME, Bockowska SB, Lebensztejn DM. Pediatric non-alcoholic steatohepatitis: the first report on the ultrastructure of hepatocyte mitochondria. World J Gastroenterol 2014;20:4335–4340.
- [10] Le TH, Caldwell SH, Redick JA, Sheppard BL, Davis CA, Arseneau KO, et al. The zonal distribution of megamitochondria with crystalline inclusions in nonalcoholic steatohepatitis. Hepatology 2004;39:1423–1429.
- [11] Caldwell SH, Swerdlow RH, Khan EM, Iezzoni JC, Hespenheide EE, Parks JK, et al. Mitochondrial abnormalities in non-alcoholic steatohepatitis. J Hepatol 1999;31:430–434.
- [12] Cocariu EA, Mageriu V, Staniceanu F, Bastian A, Socoliuc C, Zurac S. Correlations between the autolytic changes and postmortem interval in refrigerated cadavers. Rom J Intern Med 2016;54:105–112.
- [13] Heuser JE, Reese TS. Evidence for recycling of synaptic vesicle membrane during transmitter release at the frog neuromuscular junction. J Cell Biol 1973;57:315–344.
- [14] Hamming I, Timens W, Bulthuis ML, Lely AT, Navis G, van Goor H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus.

A first step in understanding SARS pathogenesis. J Pathol 2004;203:631–637.

- [15] Harmer D, Gilbert M, Borman R, Clark KL. Quantitative mRNA expression profiling of ACE 2, a novel homologue of angiotensin converting enzyme. FEBS Lett 2002;532:107–110.
- [16] Guan GW, Gao L, Wang JW, Wen XJ, Mao TH, Peng SW, et al. Exploring the mechanism of liver enzyme abnormalities in patients with novel coronavirus-infected pneumonia. Zhonghua Gan Zang Bing Za Zhi 2020;28:100–106.

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