

ORIGINAL ARTICLE

Initial detections and spread of invasive *Spodoptera frugiperda* in China and comparisons with other noctuid larvae in cornfields using molecular techniques

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Abstract The fall armyworm, *Spodoptera frugiperda*, is a species native to the Americas and has spread to many countries in Africa and Asia in recent years. Proactive actions for potential invasion of *S. frugiperda* to China coordinated by government agencies and agricultural extension systems resulted in timely detection in January 2019 in Yunnan province neighboring onto Myanmar. The extensive monitoring in southern provinces of China since February 2019 resulted in dynamic tracking of *S. frugiperda* spreading to 13 provincial regions in China within 4 months by May 10, 2019, which is crucial for timely management actions in the fields. The first detections of *S. frugiperda* (corn strain) in China were confirmed using cytochrome oxidase subunit 1 (CO1) and triosephosphate isomerase (*Tpi*) genes molecular marker method. In addition to *S. frugiperda*, larvae of three other noctuid species with similar morphological appearance (*S. litura, S. exigua* and *Mythimna separata*) can occur simultaneously and cause similar damage in cornfields in southern China. Thus, we can use both morphological and molecular marker methods to compare larval stages of four noctuid species. Further, we discuss the risk of potential spread of invasive *S. frugiperda* to other regions and impact on corn production in China.

Key words corn; molecular identification; *Mythimna separata*; *Spodoptera exigua*; *Spodoptera frugiperda*; *Spodoptera litura*

Introduction

Fall armyworm, *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae), is one of the most devastating pests of corn in the Americas. *S. frugiperda* is a polyphagous pest with a broad range of hosts, over 80 plant species, and it is highly migratory (Pogue, 2002; CABI, 2016). It is known that *S. frugiperda* feeds on corn leaves, tassel

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and ears during the larval stage, which causes serious damage to corn (Sena *et al.*, 2003). In Brazil, *S. frugiperda* attacks on corn reduced the yield up to 34%, causing annual losses of 400 million US dollars (Sena *et al.*, 2003; Figueiredo *et al.*, 2005; Lima *et al.*, 2010). It is native to tropical and sub-tropical areas in America but has spread rapidly across almost all sub-Saharan countries after it was first detected in western and central Africa in early 2016, and 2018 in Uganda (Goergen *et al.*, 2016; Otim *et al.*, 2018). Then it was confirmed in several Asian countries such as India (Mallapur *et al.*, 2018), Sri Lanka, Thailand, Yemen and Myanmar (CABI, 2019) as well as Bangladesh (Farmer, 2019) in 2018.

When *S. frugiperda* invasion was confirmed in India in July 2018, we submitted a report "Rapid spread of crop-devastating fall armyworm in Africa and

invasion in India" to the government agencies of Ministry of Agriculture and Rural Affairs (MARA), China in August 2018, and then published a review paper "Potential invasion of the crop-devastating insect pest fall armyworm Spodoptera frugiperda in China" in Plant Protection in November 2018 (Guo et al., 2018). When S. frugiperda was confirmed in Myanmar in December 2018, the notification was issued by MARA for field monitoring in Yunnan and Guangxi provinces neighboring Myanmar for timely detection of S. frugiperda. The first suspected detection of S. frugiperda in China was confirmed based on morphological methods in January 2019 [(China National Agro-Tech Extension and Service Center (NATESC), 2019a; International Plant Protection Convention (IPPC, 2019)] in Yunnan province adjacent to Myanmar. It was also confirmed by methods using molecular makers (Zhang et al., 2019).

The females lay eggs on abaxial leaf surfaces of host leaves and an individual adult can lay 100 to 200 eggs per egg mass (Kumela *et al.*, 2018). Immediately after hatching, neonates start feeding on the leaf which leads to complete skeletonization (Brévault *et al.*, 2018). Duration of larval stage (usually six instars) is about 14 days during the summer and may prolong to 30 days during cold weather and the last instar moves onto the ground and pupates (Sparks, 1979). The adult of *S. frugiperda* has strong flight ability and has spread rapidly in Africa, India and other Asian countries (Early *et al.*, 2018).

To control S. frugiperda, an effective identification method is needed because similar species like S. litura (Fabricius), S. exigua (Hübner) and Mythimna separata (Walker) also occur simultaneously in cornfields. In our previous report, the morphological differences among the four noctuid species were described (Guo et al., 2019). However, the morphological characteristics of these pests and the damage symptoms they cause on corn leaves are very similar, and sometimes this may lead to misidentification with other noctuid species, especially for the 1st and 2nd instars. Also, egg masses of the three Spodoptera species are very difficult to distinguish based on morphological characteristics. In addition, S. frugiperda consists of "rice" and "corn" subpopulations and differs in host plant distribution without difference in morphological features (Pashley, 1986; Pashley et al., 1987; Nagoshi et al., 2017). Molecular identification methods could be more accurate than the morphological identification. With the rapid advancement in molecular biology techniques, various biological macromolecules containing evolutionary information have been used as molecular markers, which are widely used in species identification and phylogenetic analysis (Tautz et al., 2002). Mitochondrial DNA (mtDNA) has many advantages for intraspecific identifications due to maternal inheritance, high copy number, and both conservative and high-mutation rate DNA segments (Mauro *et al.*, 2006). Currently, the cytochrome oxidase subunit 1 (*CO1*) gene has been used as a standard universal sequence used to elucidate the species uncertainty in morphological taxonomy (Paul *et al.*, 2003; Murray & Prowell, 2005; Solovyeva *et al.*, 2014) and this technology has been adapted to distinguish *S. frugiperda* (Nagoshi, 2007a; Nagoshi, 2007b; Kergoat *et al.*, 2012; Cock *et al.*, 2017). In addition, polymorphisms in triosephosphate isomerase (*Tpi*) gene (Knowles, 1991) helps to identify specific host strains and the efficiency of this gene during species identification has been explained in many reports (Nagoshi, 2010, 2012; Nagoshi *et al.*, 2017).

In this study, we reported the initial detections and sequential spread of *S. frugiperda* in 13 provincial regions (Yunnan, Guangxi, Guangdong, Guizhou, Hunan, Hainan, Fujian, Zhejiang, Hubei, Sichuan, Jiangxi, Chongqing, and Henan) of China within 4 months by May 10, 2019 and confirmation of the species using molecular markers based on *CO1* and *Tpi* genes. We also compared molecular differences among four similar noctuid larvae of *S. frugiperda*, *S. exigua*, *S. litura* and *M. separata* from cornfields.

Materials and methods

Initial detections of S. frugiperda in China

On January 3, 2019, NATESC (2019b) of MARA published a notification to provincial Plant Protection Stations nationwide in China for proactive actions for potential invasion of S. frugiperda into China, with special requirement to set up key monitoring sites for all yearround monitoring in the four southern provinces (i.e., Yunnan, Guangxi, Guangdong, Hainan) and other regions with high invasion risks. The major monitoring methods were: (1) on-farm monitoring by national agricultural extension agencies, National Crop Pest Monitoring and Forecasting Regional Stations (>1000 in total), with special attention in winter cornfields in Yunnan and Guangxi starting January 2019, in the southern provinces starting February 2019, and nationwide starting April 2019; (2) adult trapping using black light traps and vertical-pointing searchlight traps in the southern provinces starting February 2019 and 220 counties of 26 provinces since April 2019; (3) sex pheromone trapping using equipment confirmed by NATESC on key host crops in the four southern provinces since February 2019. NATESC published a standardized method "Methods of Investigation and Forecast for Fall Armyworm in 2019 (Preliminary)" in March 2019 for data collection in China (NATESC, 2019c).

Sample collection and molecular identification and analysis of CO1 and Tpi genes

In January, 2019 the first suspected *S. frugiperda* infestation was detected in a cornfield in Jiangcheng county (of Pu'er City), Yunnan province; we surveyed *S. frugiperda* on winter corn in Pu'er and Dehong (Dehong Dai and Jingpo Autonomous prefecture) of Yunnan province during January 12–24 and confirmed the *S. frugiperda* invasion by morphological identification. The *S. litura*, *M. separata* and *S. exigua* larvae, as well as suspected *S. frugiperda* larvae were collected from a winter cornfield in Dehong, Yunnan province (24.42°N, 98.60°E) on January 20, 2019. Totally, 250–350 larvae were collected from Yunnan province. The collected larvae were transported to the laboratory and reared in a controlled environment chamber at 27 ± 1 °C, with 60%–70% relative humidity (RH), a 14 : 10 L : D photoperiod.

The CO1 primer of *S. frugiperda*, *M. separata* (Mehrdad *et al.*, 2006) and *S. exigua* (Hebert *et al.*, 2003) were designed based on previous reports. Total DNA was extracted from whole body of 4th instars using a DNA kit (Tianmoboi, China) and stored at -20 °C for further analysis. Twenty individuals from each species were selected and genomic DNA was extracted from every individual larva (n = 20). Polymerase chain reaction (PCR) amplifications were carried out in a volume of 20 μ L reaction containing: 10 μ L of Super HF PCR Master Mix (HeroGen Biotechnology, USA), 2 μ L primers (Forward and Reverse), 1 μ L DNA template and 7 μ L sterile H₂O.

S. litura primers were designed according to the sequence from the National Center for Biotechnology Information (NCBI) (GenBank No. KF022223.1). On other hand, based on single-nucleotide polymorphisms at e₄183 site the Tpi gene was selected for analysis and this site explains strain differentiation. In females, the Tpi e₄183 site possesses either a C or T nucleotide, which can differentiate the two S. frugiperda strains. The corn strain possesses C nucleotide (Tpi e₄183-C) and rice strain with T (Tpi e₄183-R). These *Tpi* genes are likely to be Z-linked in Lepidoptera (Yasukochi et al., 2006). However, there are more possibilities of overlap between Tpi genes in male genomic DNA during PCR product sequencing, which possess both C and T nucleotides at e₄183 site (Nagoshi, 2010; Nagoshi et al., 2017). To overcome this problem, primer design and methods were followed according to Nagoshi et al. (Fig. 1) (Nagoshi, 2010; Nagoshi et al., 2017). Twenty larvae were selected randomly, and DNA was isolated. All specific primer details are mentioned in Table 1. Amplifications were performed in Techne-5000 (UK) and programmed according to the following conditions: initial denaturation at 94 °C for 5 min; 30 amplification cycles consisting of 1 min at 94 °C, 30 s at 50 °C, 45 s of extension at 72 °C, and final extension at 72 °C for 10 min.

Sequencing and species identification

The PCR products from four noctuid species were outsourced to Shenggong Biotec (Shanghai, China) for sequencing. The assembled sequences from four samples were subjected to molecular identification using the online NCBI Basic Local Alignment Search tool (BLAST) (https://www.ncbi.nlm.nih.gov/). The

Exon-3	
<u>GTGAAATCTCCCCTGCTATG</u> ATTAAGGACATCGGAGCCAACTGGGTCATCCTTGGTCACT	60
Tpi-282F //Intron-3	
CTGAACGTAGGACTATCTTCGGTGAGAAAGATGATCTTGTTGCAGAGAAA GTAAGTACTA 1	.20
AACATAAATGAGATTTATAATATATATATTTTTTTTTTT	80
TTTCTAG GTTGCCCATGCTCTTGAGTCCGGACTGAAGGTTATCGCTTGCATTGGAGAGAC 2	240
<i>Tpi-</i> 412F	
TCTTGAAGAACGTGAGGCTGGAAAGACTGAAGAAGTTGTGTTCAGGCACACTAAGGCCCT 3	300
TTTACCAGCTATTGGCAGCAACTGGGACAAGGTCGTACTAGCTTATGAACCCGTTTGGGC 3 e4183	360
	394
	124
C: Tpi-C: T: Tpi-R: C/T: Tpi-h	

Fig. 1 Triosephosphate isomerase (*Tpi*) gene mapping at $e4_{183}$ polymorphic site for a C or T nucleotide. Polymerase chain reaction amplification was done using primer *Tpi*-282F and *Tpi*-850R. DNA sequencing was performed using primer *Tpi*-412F, which initiates in the same exon as $e4_{183}$. [Correction added on 12 September 2019, after first online publication: Figure 1 has been corrected to include a missing 'C' in the sequence.]

Table 1 Selected species-specific cytochrome oxidase subunit 1 and triosephosphate isomerase (*Tpi*) primers.

Species	Primer	Sequence $(5'-3')$
Spodoptera frugiperda/Mythimna separata	Forward	ATTCAACCAATCATAAAGATATTGG
	Reverse	TAAACTTCTGGATGTCCAAAAAATCA
S. exigua	Forward	GGTCAACAAATCATAAAGATATTGG
-	Reverse	TAAACTTCAGGGTGACCAAAAAATCA
S. litura	Forward	ACAAATCATAAAGATATTGGAACATTA
	Reverse	TAAACTTCGGGATGTCCAAAAAATCA
<i>Tpi</i> -282F	_	GGTGAAATCTCCCCTGCTATG
<i>Tpi</i> -850R	_	AATTTTATTACCTGCTGTGG
<i>Tpi</i> -412F	_	CCGGACTGAAGGTTATCGCTTG

assembled nucleotide using the CO1 method was submitted to NCBI (GenBank No. MK790611). A phylogenetic tree was constructed using the maximum likelihood method (MEGA 7.0 software, USA) with 1000 replications.

Results

The initial detection of S. frugiperda in China

Larvae detections The first suspected S. frugiperda infestation was detected in a cornfield in Jiangcheng county, Yunnan province on January 11, 2019. Additional field surveys on wide range areas in Yunnan province confirmed the presence of S. frugiperda larvae in cornfields in three cities or autonomous prefectures (Pu'er, Dehong and Baoshan) of Yunnan province by the end of January 2019. There was an average of 0.05-0.20 larvae per corn plant and maximum two larvae per plant. There were six cities or autonomous prefectures in Yunnan with S. frugiperda larvae in cornfields by the end of February 2019 (Fig. 2). By April 2019, there were 63 counties from 12 cities or autonomous prefectures in Yunnan with the infestation of larvae detected on 5200 ha of corn fields and 3700 ha in sugarcane fields, with highest larvae infestation of 100% corn plants and up to 1.5-2.0 larvae per plant in selected fields in Longchuan county.

By the end of April 2019, there were six provincial regions in China with *S. frugiperda* larvae detected (Fig. 2); Guangxi Zhuang Autonomous Region was the second provincial area with *S. frugiperda* larvae detected in two cities (Chongzuo and Baise) by April 2019. The highest larvae infestation was 50% of corn plants in selected cornfields. Guangdong was the third province with *S. frugiperda* larvae detected in Zengcheng and Conghua districts of Guangzhou City. In late April 2019 the *S. frugiperda* larvae were detected in the fourth and fifth provincial regions of Guizhou (Qianxinan Prefecture) and Hunan province (Chenzhou City). On April 30, 2019 the *S. frugiperda* larvae were detected in Haikou of Hainan as the sixth province, which is the major region of winter research nurseries for corn breeding in China. By May 10, 2019, it was also found in seven additional provincial regions (Fujian, Zhejiang, Hubei, Sichuan, Jiangxi, Chongqing and Henan) resulting in 13 provincial regions in total within four months (Table 2).

Adult detections In Yunnan from February 19 to April 1, 2019 in Ruili of Dehong, the total number of moths per sex pheromone traps and sticky-card traps with food bait were 32 and 159, respectively. There were fewer moths, only 20, detected using vertical-pointing searchlight traps from January to March from Jiangcheng of Pu'er. In Guangxi, the first *S. frugiperda* moth was detected in Hechi of Yizhou on March 11, 2019 (Table 2). Subsequently, more moths were trapped in Hechi (Du'an and Donglan counties), Baise (Youjiang and Tianlin counties) and Qinzhou (Pubei county) before larvae were detected in these regions.

Molecular identification and phylogenetic analysis

The sequences obtained from our samples by CO1 marker were subjected to NCBI BLAST, the results showed positive hits and the species were identified as *S. frugiperda*, *S. litura*, *S. exigua* and *M. separata*. Especially, *S. frugiperda* from Yunnan of China belonged to a haplotype that showed 100% match with that of India (GenBank No. MH753324-27 and MH190445). In addition, other lepidopterans showed sequence similarity above 90% when comparing with *S. frugiperda* (Fig. 3). Based on polymorphisms at *Tpi* e4₁₈₃ site, 15 larval samples out of 20 belonged to the corn strain (*Tpi* e4₁₈₃- C)

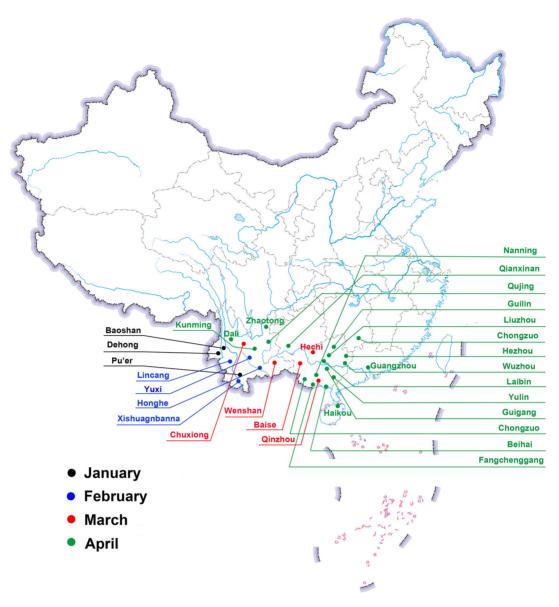


Fig. 2 Map showing the location of *Spodoptera frugiperda* in China by April 30, 2019. The dots in black, blue, red and green represent areas where *S. frugiperda* were found in respective months of 2019. GS(2019)3426.

and five samples showed heterozygous polymorphisms (*Tpi*-h $e4_{183}$ C and T).

Discussion

The exotic *S. frugiperda* has been detected in some countries in Africa and Asia in recent years. The spread or invasion of *S. frugiperda* is not only related to global trade and transport, but also strong long-distance migration ability of the *S. frugiperda* moths. Proactive actions for potential invasion of *S. frugiperda* to China coordinated by govern-

ment agencies and agricultural extension systems resulted in rapid detection of the first larvae on January 11, 2019 in a cornfield in Yunnan province near Myanmar. Scientists, especially those from the Chinese Academy of Agricultural Sciences (CAAS), provided timely and important support on the invasive pest alert, migration forecasting, monitoring and identification methods, and pest management techniques (Guo *et al.*, 2018; Guo *et al.*, 2019; Wu *et al.*, 2019; Zhang *et al.*, 2019). The extensive monitoring in the southern provinces resulted in dynamic tracking of the *S. frugiperda* spreading in China, which is crucial for

Provincial region	Month of first detection	Insect stage	No. of cities	No. of counties	Crop type	Infestation and crop acreage
Yunnan	January	Larvae	15	112	Corn	20 000 ha, up to 3 larvae per plant and normally 1%–10% plants infested.
	April	Larvae	3	3	Sugarcane	3680 ha, up to 40% plants infested in Longchuan county.
Guangxi	March	Adults	3	6	_	_
	April	Larvae	14	65	Corn/sugarcane	Corn: 46 600 ha, up to 6 larvae per plant and normally 5%–9% plants infested.
Guangdong	April	Larvae	7	16	Corn	187 ha, up to 0.2 larvae per plant and 10%–20% plants infested.
Guizhou	April	Larvae	6	21	Corn	1200 ha, up to 0.4 larvae per plant and normally 9%–11% plants infested.
Hunan	April	Larvae	6	12	Corn	1300 ha, up to 1.2 larvae per plant and normally 5%–41% plants infested.
Hainan	April	Larvae	_	18	Corn	30 ha, up to 0.7 larvae per plant and 80% plants infested in Wanning county.
Fujian	May	Larvae	6	9	Corn	6.6 ha, up to 1.2 larvae per plant and 20%–40% plants infested.
Zhejiang	May	Larvae	2	2	Corn	5.3 ha, up to 0.02–0.1 larvae per plant and 1%–10% plants infested.
Hubei	May	Larvae	2	1	Corn	333 ha, up to 0.2–0.6 larvae per plant and 9.5%–18% plants infested in Tongshan county.
Sichuan	May	Larvae	1	1	Corn	<1 ha, up to 0.1 larvae per plant and 1.3% plants infested in Xichang City.
Jiangxi	May	Larvae	1	1	Corn	33 ha, up to 0.3 larvae per plant and3%–30% plants infested inGanzhou City.
Chongqing	May	Larvae	1	1	Corn	2 ha, up to 0.6 larvae per plant and up to 50% plants infested in Beibei District.
Henan	May	Larvae	1	1	Corn	<1 ha in Xinyang City.

Table 2 The severity of Spodoptera frugiperda infestations in 13 provinces in China by May 10, 2019 (NATESC, 2019e, f).

coordinating management of this pest in new locations. It was detected in >260 counties of 13 provinces within 4 months by May 10, 2019, suggesting it is spreading rapidly.

Wu *et al.* (2019) simulated the migration patterns from Myanmar to spread to Yunnan and Guangxi. Their flight toward other provinces in southern China (Guizhou, Guangdong, Hainan and Hunan Provinces) was also predicted. If prevention and control is not timely, the pest will be able to spread toward the main corn producing areas of China and even reach the Korean peninsula and Japan in June or July of 2019. Currently, *S. frugiperda* management mainly relies on the use of chemical insecticides and *Bacillus thuringiensis* (Bt) corn (Cook *et al.*, 2004). However, with the repeated use of pesticides and highly

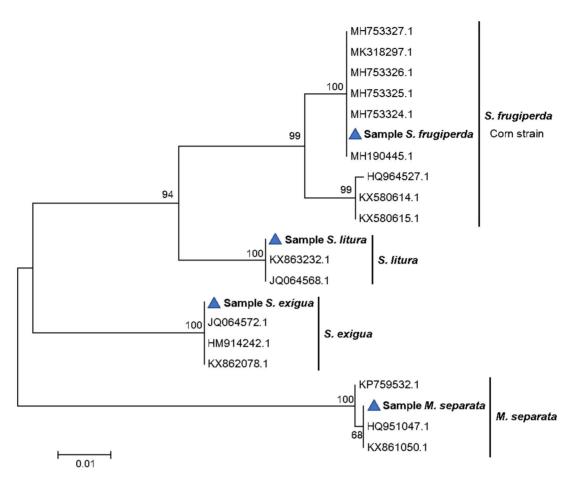


Fig. 3 The phylogenetic tree was constructed using cytochrome oxidase subunit 1 (CO1) sequences from four Lepidoptera noctuid species. Maximum likelihood method was adopted based on the Tamura-Nei model (Tamura & Nei, 1993). The nucleotide sequences were aligned using the Clustal W program and the evolutionary analyses were conducted in MEGA 7.0 (Kumar *et al.*, 2016). The triangle indicates the samples collected from China used for this study.

adaptable capabilities of this species, S. frugiperda can develop resistance to those chemicals (Yu, 1991, Gutiérrez-Moreno et al., 2019) and to the plants expressing a Cry1A or Cry1F Bt protein (Farias et al., 2014; Huang et al., 2014; Omoto et al., 2016). It is therefore essential to use an integrated pest management (IPM) approach to manage S. frugiperda and other Lepidoptera pests in corn. In March 2019, NATESC (2019d) published the IPM Guide on Fall Armyworm in China (2019 Preliminary Version), which will be updated based on current studies. Based on the experiences in North and South America (Huang et al., 2014; Burtet et al., 2017) and South Africa (Botha et al., 2019), Bt corn plants can be highly effective on S. frugiperda when utilized as part of an IPM program after registration for cultivation in China in the future. The use of pyramided traits expressing effective Bt proteins on S. frugiperda with different sites of action (Burtet et al., 2017; Margues et al., 2019) should be prioritized and then deployed with appropriate refuge, resistance monitoring, and grower education. In southern China, three similar noctuid larvae of native lepidopteran species (S. litura, S. exigua and M. separata) are often found in the same field and each species can cause considerable damage on important agricultural crops, such as corn, rice, wheat, cotton and vegetables (Jiang et al., 2011). However, on-farm identification of larvae of these pests using morphological characteristics is challenging, and often results in misidentifications, especially for early instars. Moreover, the traditional morphological classification requires a high level of knowledge and experience by researchers or extension specialists. Therefore, a more accurate and timely identification method is needed to differentiate these species and that can be achieved through molecular characterization. In this research, four noctuid larvae of lepidopteran pests from cornfields with three of them belonging to the same genus were collected and identified by both morphology and molecular methods. From the morphological identification, we can separate *S. frugiperda* from the other three species (*S. litura, S. exigua* and *M. separata*). Molecular methods were used to further confirm the morphological identification results (Fig. 3). Fast and accurate molecular methods are very important for timely identifying and distinguishing the larvae with similar morphology in cornfields and designing the control measures on newly invaded species. Meanwhile, the molecular methods may help greatly to understand its distant migratory and invasion pattern around the globe.

Three native noctuid species (S. litura, S. exigua and *M. separata*) are important pests infesting corn in China. The invasive S. frugiperda has the potential to become a hugely important pest of corn, if not the most important pest of corn at least in certain regions of China. It is particularly problematic for corn plants because female moths oviposit on corn leaves and larvae have strong serrated mandibles that allow them to feed on most corn tissues (Brown & Dewhurst, 1975; Pogue, 2002). The cannibalistic nature of S. frugiperda larvae also is an important factor because it allows them to dominate interspecific competitors and reduce intraspecific rivals (Chapman et al., 2000). In addition, larvae usually bore into corn whorls or ears and develop under protected conditions, which allows them to avoid direct exposure to insecticides. This greatly increases the chance of survival for S. frugiperda during insecticidal sprays and low-dose exposure to insecticides probably has contributed to their resistance to several classes of insecticides (Adamczyk et al., 1999; Gutiérrez-Moreno et al., 2019).

Two S. frugiperda strains were identified, one preferentially infests corn and the other preferentially infests rice (Pashley et al., 1985). The rice strain also prefers to infest millet and grass and the corn strain prefers corn and sorghum (Nagoshi & Meagher, 2004a,b). Both strains can infest corn and are morphologically indistinguishable (Pashley, 1986). According to strain identification results, only corn strain moths were discovered in Yunnan province, China. The invasion path of S. frugiperda is likely India, Myanmar then Yunnan province, China, after which they migrated to the eastern and northern regions of China. This possibility is due to its long-distance flight ability and adaptability to environments. Supporting our hypothesis, migration of S. frugiperda inside the American states and its adaptability to different climatic conditions are well documented (Raulston & Sparks, 1986). Sequencing results of CO1 are consistent with the sequence of S. frugiperda found in India, and this result strongly supports our hypothesis on the invasion path to

China. China as the second largest corn growing region in the world has not yet approved commercial planting of Bt corns. As a new invasive species, it will take time to sort out the spread potential, migratory dynamics, and ultimately the impact this moth will have on crops in China. As a new invasion species, S. frugiperda will enhance its reproductive capacity and spreading performance as part of its life history strategy (Johnson, 1987). Hence, if S. frugiperda is not well controlled, it particularly may have a serious impact on corn production in China. In addition, there are other major pests of corn in China, including Ostrinia furnacalis (He et al., 2003), Conogethes punctiferalis (Lu et al., 2010) and M. separata. A sustainable and effective IPM strategy to mitigate the impact of these insect pests of corn, especially S. frugiperda, is urgently needed.

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Disclosure

The authors declare they have no conflict interests.

References

- Adamczyk, J.J., Leonard, B.R. and Graves J.B. (1999) Toxicity of selected insecticides to fall armyworms (Lepidoptera: Noctuidae) in laboratory bioassay studies. *Florida Entomologist*, 82, 230–236.
- Botha, A.S., Erasmus, A., du Plessis, H. and Van den Berg, J. (2019) Efficacy of Bt maize for control of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in South Africa. *Journal* of *Economic Entomology*, https://doi.org/10.1093/jee/toz048.
- Brévault, T., Ndiaye, A., Badiane, D., Bal, A.M., Sembène, M., Silvie, P. *et al.* (2018) First records of the fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae), in Senegal. *Entomologia Generalis*, 37, 129–142.
- Brown, E.S. and Dewhurst C.F. (1975) The genus *Spodoptera* (Lepidoptera, Noctuidae) in Africa and the Near East. *Bulletin* of *Entomological Research*, 65, 221–262.
- Burtet, L.M., Bernardi, O., Melo, A.A., Pes, M.P, Strahl, T.T. and Guedes, J.V. (2017) Managing fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae), with Bt maize and insecticides in southern Brazil. *Pest Management Science*, 73, 2569–2577.

CABI (2016) Datasheet *Spodoptera frugiperda* (fall armyworm). *Invasive Species Compendium*, http://www.cabi.org/ isc/datasheet/29810 (accessed April 26, 2019).

CABI (2019) Datasheet *Spodoptera frugiperda* (fall armyworm). *Invasive Species Compendium*, https://www.cabi. org/isc/datasheet/29810#94987198-9f50-4173-8bbd-30bd93840e73?tdsourcetag=s_pcqq_aiomsg (accessed

April 26, 2019). Chapman, J.W., Williams, T., Martínez, A.M., Cisneros, J., Ca-

- ballero, P., Cave, R.D. *et al.* (2000) Does cannibalism in *Spodoptera frugiperda* (Lepidoptera: Noctuidae) reduce the risk of predation? *Behavioral Ecology and Sociobiology*, 48, 321–327.
- Cock, M.J.W., Beseh, P.K., Buddie, A.G., Cafáet, G. and Crozier, J. (2017) Molecular methods to detect *Spodoptera frugiperda* in Ghana, and implications for monitoring the spread of invasive species in developing countries. *Scientific Reports*, https://doi.org/10.1038/s41598-017-04238-y.
- Cook, D.R., Leonard, B.R. and Gore, J. (2004) Field and laboratory performance of novel insecticides against armyworms (Lepidoptera: Noctuidae). *Florida Entomologist*, 87, 433– 439.
- Early, R., Pablo, G.M., Murphy, S.T. and Day, R. (2018) Forecasting the global extent of invasion of the cereal pest *Spodoptera frugiperda*, the fall armyworm. *NeoBiota*, 40, 25– 50.
- Farias, J.R., Andow, D.A., Horikoshi, R.J., Sorgatto, R.J., Fresia, P., Santos, C.D. *et al.* (2014) Field-evolved resistance to Cry1F maize by *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in Brazil. *Crop Protection*, 64, 150–158.
- Farmer, B. (2019) Fall armyworm marches on as pest that devastated African crops spreads in Asia. *The Telegraph*, https://www.telegraph.co.uk/news/2019/01/09/fallarmyworm-marches-pest-devastated-african-crops-spreads/ (accessed April 26, 2019).
- Figueiredo, M.L.C., Penteado-Dias, A.M. and Cruz, I. (2005) Danos provocados por *Spodoptera frugiperda* na produção de matéria seca e nos rendimentos de grãos, na cultura do milho. *Sete Lagoas, MG: Embrapa, CNPMS*, 130. https:// ainfo.cnptia.embrapa.br/digital/bitstream/CNPMS/18881/1/ Com_130.pdf (accessed May 30, 2019).
- Goergen, G., Kumar, P.L., Sankung, S.B., Togola, A. and Tamò, M. (2016) First report of outbreaks of the fall armyworm *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera, Noctuidae), a new alien invasive pest in west and central Africa. *PLoS ONE*, 11, e0165632.
- Guo, J.F., Jing, D.P., Tai, H.K., Zhang, A.H., He, K.L. and Wang, Z.Y. (2019) Morphological characteristics of *Spodoptera frugiperda* in comparison with three other lepidopteran species with similar injury characteristics and morphology in cornfields. *Plant Protection*, 45, 7–12.

- Guo, J.F., Zhao, J.Z., He, K.L., Zhang, F. and Wang, Z.Y. (2018) Potential invasion of the crop-devastating insect pest fall armyworm *Spodoptera frugiperda* to China. *Plant Protection*, 44, 1–10.
- Gutiérrez-Moreno, R.G., Mota-Sanchez, D., Blanco, C.A., Whalon, M.E., Terán-Santofimio, H., Rodriguez-Maciel, J.C. *et al.* (2019) Field-evolved resistance of the fall armyworm (Lepidoptera: Noctuidae) to synthetic insecticides in Puerto Rico and Mexico. *Journal of Economic Entomology*, 112, 792–802.
- He, K.L., Wang, Z.Y., Zhou, D.R., Wen, L.P., Song, Y.Y. and Yao, Z.Y. (2003) Evaluation of transgenic Bt corn for resistance to the Asian corn borer (Lepidoptera: Pyralidae). *Journal of Economic Entomology*, 96, 935–940.
- Hebert, P.D.N, Cywinska, A., Ball S.L. and deWaard, J.R. (2003) Biological identifications through DNA barcodes. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 270, 313–321.
- Huang, F.N., Qureshi, J.A., Meagher, R.L., Reisig, D.D., Head, G.P., Andow, D.A. *et al.* (2014) Cry1F resistance in fall armyworm *Spodoptera frugiperda*: single gene versus pyramided Bt maize. *PLoS ONE*, 9, e112958.
- IPPC (2019) First detection of fall armyworm in China. https:// www.ippc.int/en/news/first-detection-of-fall-armyworm-inchina/ (accessed March 28, 2019).
- Jiang, X., Luo, L. and Zhang, L. (2011) Regulation of migration in *Mythimna separata* (Walker) in China: a review integrating environmental, physiological, hormonal, genetic, and molecular factors. *Environmental Entomology*, 40, 516–533.
- Johnson, S.J. (1987) Migration and the life history strategy of the fall armyworm, *Spodoptera frugiperda* in the western hemisphere. *International Journal of Tropical Insect Science*, 8, 543–549.
- Kergoat, G.J., Prowell, D.P., Le Ru, B.P., Mitchell, A., Dumas, P., Clamens, A.L. *et al.* (2012) Disentangling dispersal, vicariance and adaptive radiation patterns: a case study using armyworms in the pest genus *Spodoptera* (Lepidoptera: Noctuidae). *Molecular Phylogenetics and Evolution*, 65, 855–870.
- Knowles, J.R. (1991) Enzyme catalysis: not different, just better. *Nature*, 350, 121–124.
- Kumar, S., Stecher, G. and Tamura, K. (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33, 1870–1874.
- Kumela, T., Simiyu, J., Sisay, B., Likhayo, P., Mendesil, E., Gohole, L. *et al.* (2018) Farmers' knowledge, perceptions, and management practices of the new invasive pest, fall armyworm (*Spodoptera frugiperda*) in Ethiopia and Kenya. *International Journal of Pest Management*, 65, 1–9.
- Lima, M.S., Silva, P.S.L., Oliveira, O.F., Silva, K.M.B. and Freitas, F.C.L. (2010) Corn yield response to weed and fall armyworm controls. *Planta Daninha*, 28, 103–111.

- Lu, J.Q, Wang, Z.Y., He, K.L. and Liu, Y. (2010) Research history, progresses and prospects in the yellow peach moth, *Conogethes punctiferalis. Plant Protection*, 36, 31–38.
- Mallapur, C.P., Naik, A.K., Hagari, S., Prabhu, S.T. and Patil, P.K. (2018) Status of alien pest fall armyworm, *Spodoptera frugiperda* (J.E. Smith) on maize in Northern Karnataka. *Journal of Entomology and Zoology Studies*, 6, 432–436.
- Marques, L. M., Santos, A.C., Castro, B.A., Moscardini, V.F., Rosseto, J., Silva, O.A.B.N. *et al.* (2019) Assessing the efficacy of *Bacillus thuringiensis* (Bt) pyramided proteins Cry1F, Cry1A.105, Cry2Ab2, and Vip3Aa20 expressed in Bt maize against Lepidopteran pests in Brazil. *Journal of Economic Entomology*, 112, 803–811.
- Mauro, D.S., Gower, D.J., Zardoya, R. and Wilkinson, M. (2006) A hotspot of gene order rearrangement by tandem duplication and random loss in the vertebrate mitochondrial genome. *Molecular Biology and Evolution*, 23, 227.
- Mehrdad, H., Janzen, D.H., Burns, J.M., Hallwachs, W. and Hebert, P.D.N. (2006) DNA barcodes distinguish species of tropical Lepidoptera. *Proceedings of the National Academy* of Sciences USA, 103, 968–971.
- Murray, D. and Prowell, D.P. (2005) Molecular phylogenetics and evolutionary history of the neotropical satyrine subtribe *Euptychiina* (Nymphalidae: Satyrinae). *Molecular Phylogenetics and Evolution*, 34, 67–80.
- Nagoshi, R.N. (2010) The fall armyworm triose phosphate isomerase (*Tpi*) gene as a marker of strain identity and interstrain mating. *Annals of the Entomological Society of America*, 103, 283–292.
- Nagoshi, R.N. (2012) Improvements in the identification of strains facilitate population studies of fall armyworm subgroups. *Annals of the Entomological Society of America*, 105, 351–358.
- Nagoshi, R.N. and Meagher, R.L. (2004a) Seasonal distribution of fall armyworm (Lepidoptera: Noctuidae) host strains in agricultural and turf grass habitats. *Environmental Entomol*ogv, 33, 881–889.
- Nagoshi, R.N. and Meagher, R.L. (2004b) Behavior and distribution of the two fall armyworm host strains in Florida. *Florida Entomologist*, 87, 440–449.
- Nagoshi, R.N., Koffi, D., Agboka, K., Tounou, K.A., Banerjee, R., Jurat-Fuentes, J.L. *et al.* (2017) Comparative molecular analyses of invasive fall armyworm in Togo reveal strong similarities to populations from the eastern United States and the Greater Antilles. *PLoS ONE*, 12, e0181982.
- Nagoshi, R.N., Silvie, P., Meagher, R.L., Lopez, J. and Machado, V. (2007a) Identification and comparison of fall armyworm (Lepidoptera: Noctuidae) host strains in Brazil, Texas, and Florida. *Annals of the Entomological Society of America*, 100, 394–402.
- Nagoshi, R.N., Silvie, P. and Meagher, R.L. (2007b) Comparison of haplotype frequencies differentiate fall armyworm (Lep-

idoptera: Noctuidae) corn-strain populations from Florida and Brazil. *Journal of Economic Entomology*, 100, 954–961.

- NATESC (2019a) Fall armyworn, the major pest has invaded in Yunnan, the field scouting and monitoring for this pest to start immediately within China. https://www.natesc. org.cn/Html/2019_01_29/28092_151760_2019_01_29_457 209.html (accessed April 26, 2019).
- NATESC (2019b) Notification for prevention and precaution of invasion and damage of fall armyworm. https://www.natesc.org.cn/Html/2019_01_03/28092_52304_2019_01_03_456907.html (accessed April 26, 2019).
- NATESC (2019c) Methods of investigation and forecast for fall armyworm in 2019 (Preliminary). https://www. natesc.org.cn/Html/2019_03_01/28092_52304_2019_03_01 _457510.html (accessed April 26, 2019)
- NATESC (2019d) Technical guideline for fall armyworm control in 2019 (Preliminary). https://www.natesc.org.cn/Html/ 2019_03_13/2_1878_2019_03_13_457664.html (accessed April 26, 2019).
- NATESC (2019e) Recent occurrence and infection of *Spodopt era frugiperda*. https://www.natesc.org.cn/Html/2019_05_06/28092_151760_2019_05_06_458341.html (accessed May 25, 2019).
- NATESC (2019f) *Spodoptera frugiperda* invaded 13 provinces and infested spring corn. https://www.natesc.org. cn/Html/2019_05_14/28092_151760_2019_05_14_458487. html?from=singlemessage&isappinstalled=0 (accessed May 25, 2019).
- Omoto, C., Bernardi, O., Salmeron, E., Sorgatto, R.J., Dourado, P.M., Crivellari, A. *et al.* (2016) Field-evolved resistance to Cry1Ab maize by *Spodoptera frugiperda* in Brazil. *Pest Management Science*, 72, 1727–1736.
- Otim, M.H., Tay, W.T., Walsh, T.K., Kanyesigye, D., Adumo, S., Abongosi, J. *et al.* (2018) Detection of sister-species in invasive populations of the fall armyworm *Spodoptera frugiperda* (Lepidoptera: Noctuidae) from Uganda. *PLoS ONE*, 13, e0194571.
- Pashley, D.P. (1986) Host-associated genetic differentiation in fall armyworm (Lepidoptera: Noctuidae): a sibling species complex? *Annals of Entomological Society of America*, 79, 898–904.
- Pashley, D.P., Johnson, S.J. and Sparks, A.N. (1985) Genetic population structure of migratory moths: the fall armyworm (Lepidoptera: Noctuidae). *Annals of the Entomological Society of America*, 78, 756–762.
- Pashley, D.P., Sparks, T.C. and Quisenberry, S.S. (1987) Two fall armyworm strains feed on corn, rice and bermudagrass. *Louisiana Agriculture Louisiana Agricultural Experiment Station*, 30, 8–9.
- Paul, D.N., Ratnasingham, S. and Dewaard, J.R. (2003) Barcoding animal life: cytochrome c oxidase subunit 1 divergences

among closely related species. *Proceedings Biological sciences*, 270, S96–S99.

- Pogue, G.M. (2002) A world revision of the genus Spodoptera Guenée (Lepidoptera: Noctuidae). Memoirs of the American Entomological Society, 43, 1–202.
- Raulston, J.R. and Sparks, A.N. (1986) Fall armyworm distribution and population dynamics in the southeastern states. *Florida Entomologist*, 69, 468–487.
- Sena Jr, D.G., Pinto, F.A.C., Queriroz, D.M. and Viana, P.A. (2003) Fall armyworm damaged maize plant identification using digital images. *Biosystems Engineering*, 85, 449– 454.
- Solovyeva, E.N., Poyarkov, N.A., Dunayev, E.A., Nazarow, R.A., Lebedev, V.S. and Bannikova, A.A. (2014) Phylogenetic relationships and subgeneric taxonomy of toadheaded agamas *Phrynocephalus* (Reptilia, Squamata, Agamidae) as determined by mitochondrial DNA sequencing. *Doklady Biological Sciences Proceedings of the Academy* of Sciences of the USSR Biological Sciences, 455, 119– 124.
- Sparks, A.N. (1979) A review of the biology of the fall armyworm. *Florida Entomologist*, 62, 82–87.
- Tamura, K. and Nei, M. (1993) Estimation of the number of nucleotide substitutions in the control region of mitochondrial

DNA in humans and chimpanzees. *Molecular Biology and Evolution*, 10, 512–526.

- Tautz, D., Arctander, P., Minwlli, A., Thomas, R.H. and Vogler, A.P. (2002) DNA points the way ahead in taxonomy. *Nature*, 418, 479–479.
- Wu, Q.L., Jiang, Y.Y. and Wu, K.M. (2019) Analysis of migration routes of the fall armyworm *Spodoptera frugiperda* (J.E. Smith) from Myanmar to China. *Plant Protection*, 45, 1–6.
- Yasukochi, Y., Ashakumary, L.A., Baba, K., Yoshido, A. and Sahara, K. (2006) A second-generation integrated map of the silkworm reveals synteny and conserved gene order between lepidopteran insects. *Genetics*, 173, 1319–1328.
- Yu, S.J. (1991) Insecticide resistance in the fall armyworm, Spodoptera frugiperda (J.E. Smith). Pesticide Biochemistry and Physiology, 39, 84–91.
- Zhang, L., Jin, M.H., Zhang, D.D., Jiang, Y.Y., Liu, J., Wu, K.M. *et al.* (2019) Molecular identification of invasive fall armyworm *Spodoptera frugiperda* in Yunnan province. *Plant Protection*, 45, 19–24.

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