

## ARTICLE

# Timing of Antiviral Treatment Initiation is Critical to Reduce SARS-CoV-2 Viral Load

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We modeled the viral dynamics of 13 untreated patients infected with severe acute respiratory syndrome-coronavirus 2 to infer viral growth parameters and predict the effects of antiviral treatments. In order to reduce peak viral load by more than two logs, drug efficacy needs to be > 90% if treatment is administered after symptom onset; an efficacy of 60% could be sufficient if treatment is initiated before symptom onset. Given their pharmacokinetic/pharmacodynamic properties, current investigated drugs may be in a range of 6–87% efficacy. They may help control virus if administered very early, but may not have a major effect in severely ill patients.

## Study Highlights

### WHAT IS THE CURRENT KNOWLEDGE ON THE TOPIC?

☑ Repurposed drugs are being evaluated in clinical trials but little is known about their efficacy on severe acute respiratory syndrome-coronavirus 2 viral kinetics.

### WHAT QUESTION DID THIS STUDY ADDRESS?

☑ Our study aims to combine pharmacokinetic/pharmacodynamic and viral kinetics modeling to anticipate the effects of lopinavir/ritonavir, hydroxychloroquine (HCQ), IFN- $\beta$ -1a, and remdesivir.

### WHAT DOES THIS STUDY ADD TO OUR KNOWLEDGE?

☑ Given the predicted efficacy of lopinavir/ritonavir, HCQ, IFN- $\beta$ -1a, and remdesivir, it is unlikely that these drugs will have a major effect on viral kinetics if they are administered as monotherapy after symptom onset.

### HOW MIGHT THIS CHANGE DRUG DISCOVERY, DEVELOPMENT, AND/OR THERAPEUTICS?

☑ Our results suggest that these drugs should be evaluated in persons exposed to the virus but prior to appearance of the first symptoms.

The outbreak of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), which originated in Wuhan, China, has become a global pandemic. By May 7, 2020, this virus had infected more than 3,700,000 people worldwide and caused more than 260,000 deaths. To readily propose a first line of defense and combat the virus in hospitalized patients, the World Health Organization relies on already existing drugs (“repurposed”) that are immediately available in large quantities and have a good safety profile. In coordination with other European institutions, France is implementing a randomized clinical trial in hospitalized patients (“DisCoVery,” NCT04315948) comparing the efficacy of lopinavir/ritonavir  $\pm$  IFN- $\beta$ -1a, remdesivir, and hydroxychloroquine (HCQ) in hospitalized patients. However, the clinical efficacy of currently available therapies is unknown and could be limited.<sup>1</sup>

Here, we fit mathematical models of viral dynamics to *in vivo* data to estimate parameters driving viral replication. We then use these models to predict the needed efficacy

of treatments.<sup>2</sup> By combining the expected drug concentrations and half-maximal effective concentration ( $EC_{50}$ ) of drug candidates, we also use the model to predict the effects of various dosing regimens (doses and timing of treatment initiation) on viral load dynamics.

## METHODS

### Data used for fitting

We used published data from 13 untreated patients infected with SARS-CoV-2 that were followed in 4 Singapore hospitals.<sup>3</sup> Patients were hospitalized in median at day 3 after onset of symptoms (range 1–10 days) and had a median symptomatic period of 12 days (range 5–24 days). Viral loads in nasopharyngeal swabs were measured by real-time reverse transcriptase polymerase chain reaction (lower limit of quantification: 38 cycles-threshold, CT) at multiple time points with an observed peak of viral load at day 5 post onset of symptoms (range 2–27 days).

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Data presented in CT were transformed to  $\log_{10}$  copies/mL using a published relationship in Zou *et al.*<sup>4</sup> and the model was fit to the  $\log_{10}$  viral load. Of note, the transformation from CT to  $\log_{10}$  copies/mL does not affect the estimates of parameters of interest, in particular  $R_0$  and the death rate of productively infected cells. Time since infection was assumed to be 5 days before the onset of symptoms.<sup>5</sup> In a sensitivity analysis, we also examined values of 2 and 10 days.

### Model

Viral dynamics was fitted using a target cell limited model with an eclipse phase.

$$\begin{aligned} \frac{dT}{dt} &= -\beta VT \\ \frac{dI_1}{dt} &= \beta VT - kI_1 \\ \frac{dI_2}{dt} &= kI_1 - \delta I_2 \\ \frac{dV}{dt} &= pI_2 - cV - \beta VT \end{aligned} \quad (1)$$

The model considers three populations of cells: target cells,  $T$ , infected cells in the eclipse phase,  $I_1$ , and productively infected cells,  $I_2$ . Given the timescale of the infection, we neglect target cell proliferation and natural death, and we focused on the process of cell depletion by virus infection. We assumed target cells become infected with rate constant  $\beta$ . After an average time of  $1/k$ , these cells start producing virus and are cleared with per capita rate  $\delta$ . Virions are released from productively infected cells  $I_2$  at rate  $p$  per cell and are cleared from the circulation at per capita rate  $c$  or lost by infecting a target cell. Based on this model, the basic reproduction number,  $R_0$ , the average number of cells infected by a single infected cell at the beginning of the infection, is<sup>6,7</sup>

$$R_0 = \frac{p\beta T_0}{\delta(c + \beta T_0)} \quad (2)$$

To determine the target cell concentration, the following calculation was done. We assumed that the total number of epithelial cells in the upper respiratory tract was  $4 \times 10^8$  cells, distributed in a volume of 30 mL.<sup>8</sup> Assuming that 1% of these cells express the ACE2 receptor and associated proteases needed for viral entry,<sup>9</sup> the target cell concentration,  $T_0$ , was fixed to  $1.33 \times 10^5$  cells/mL. Following what was found in other viral infections, including acute infection,<sup>6</sup> the clearance rate of virus,  $c$ , was assumed to be fast and equal to  $10 \text{ day}^{-1}$  but values of 5 and  $20 \text{ day}^{-1}$  were also evaluated.

### Model building strategy

Because not all parameters can be identified when only viral load data are available, parameters  $V_0$  and  $k$  were fixed at  $10^{-1}$  copies/mL and  $3 \text{ day}^{-1}$ , respectively, which corresponds to an initiation of the viral production 8 hours after cell infection on average.<sup>10</sup> A sensitivity analysis was also performed with different values of  $k = \{1, 3, 5\} \text{ day}^{-1}$  and  $V_0 = \{10^{-3}, 10^{-2},$

$10^{-1}\}$  copies/mL to assess the robustness of the parameter estimates. The parameter  $R_0$  was estimated instead of the infection rate  $\beta$  by a change of variables in Eq. 1.

Parameters were estimated in a nonlinear mixed-effect modeling framework using the SAEM algorithm implemented in Monolix (www.lixoft.com). The model providing the best description of the data was used for the predictions and the individual data fitting, and model averaging was used to correct for the model uncertainty when calculating confidence intervals of estimated parameters.<sup>11</sup>

### Predicting the effects of treatment according to the antiviral efficacy and the time of treatment initiation

We assumed that antivirals with a constant effectiveness  $\epsilon$  could reduce  $R_0$  by a factor  $(1-\epsilon)$ , with  $\epsilon$  taking values from 50–99% in Eq. 2. We considered different times of treatment initiation, from the time of infection to 3 days after the symptom onset. For each treatment strategy, we calculated the reduction in viral load at the peak of infection in the absence of treatment (i.e., 5 days after symptom onset).

### Model including an innate immune response

We also examined the possibility that cell infection is limited by an innate immune response that renders cells refractory to infection, as was proposed for other acute viral infections.<sup>8,12</sup> In this model, two additional compartments are added, one for a cytokine (e.g., IFN) released in response to antigen, and one representing cells in an antiviral state that cannot be infected (**Supplementary Information**).

### Pharmacokinetic/pharmacodynamic drug properties of lopinavir/ritonavir, HCQ, IFN- $\beta$ -1a, and remdesivir

We relied on the literature to find pharmacokinetic (PK) population models and parameter values of lopinavir/ritonavir,<sup>13</sup> plasma HCQ,<sup>14</sup> IFN- $\beta$ -1a,<sup>15</sup> and remdesivir, as well as reported  $EC_{50}$  values *in vitro* (see **Table 1**). For lopinavir  $EC_{50}$ , specific results were obtained as follows. Vero E6 cells were infected by SARS-CoV-2 (strain BetaCoV/France/IDF0571/2020) at a multiplicity of infection of 0.01 and treated with several concentrations of lopinavir 1 hour after infection. Supernatant samples were collected at 48 and 72 hours postinfection. Relative quantification of viral genome was performed by real-time quantitative polymerase chain reaction RT-qPCR from RNA extracted using QIAamp viral RNA Mini Kit (Qiagen).  $IC_{50}$  values of lopinavir (5.246  $\mu\text{M}$  and 4.941  $\mu\text{M}$  at 48 and 72 hours postinfection, respectively) were calculated from dose-response curve using a four-parameter logistic regression model.

To determine the mean antiviral efficacy of these drugs, we simulated their plasma PK profiles considering clinical regimens used in the Discovery trial, namely 400–100 mg twice daily (b.i.d.) for lopinavir/ritonavir, 400 mg b.i.d. the first day (loading dose) followed by 400 mg once daily (q.d.) for HCQ and 12 MIU for IFN- $\beta$ -1a. For each regimen, we simulated 100 PK profiles according to the reported parameter distributions. Then, we calculated for each simulated individual the mean inhibitory coefficient, sometimes called the mean antiviral effectiveness,  $\epsilon = \frac{1}{V_0} \times \int_0^7 \frac{C(u)}{C(u) + EC_{50}} du$  during the first week of treatment, and the mean value over the  $N$  profiles are given in **Table 1**. For comparison purposes, we based the analysis

Table 1 PK/PD properties of candidate antiviral drugs

Drug	PK parameter	EC <sub>50</sub>	Dosing regimen D0–D7	$\bar{\varepsilon} = \frac{1}{N} \times \frac{1}{7} \times \int_0^7 \frac{C(u)}{C(u)+EC_{50}} du$
Lopinavir/ritonavir	Wang et al. <sup>13</sup>	5.2 μM (unpublished)	400/100 b.i.d.	66%
Hydroxychloroquine	Morita et al. <sup>14</sup>	4.2 μM <sup>27</sup>	400 mg b.i.d. at D0, followed by 400 mg q.d.	6%
IFN-β-1a	Hu et al. <sup>15</sup>	175 IU/mL <sup>29</sup>	12 MIU at D0, D2, D5	18%
Remdesivir	EMA guidelines <sup>16</sup>	1 μM <sup>17</sup>	200 mg q.d. at D0, followed by 100 mg q.d.	87%

We assume that the total concentrations were the driver of efficacy, and we did not consider intracellular metabolites or free drug concentrations. D, day; EC<sub>50</sub>, half-maximal effective concentration; EMA, European Medicines Evaluation Agency; PK/PD, pharmacokinetic/pharmacodynamic.

on total plasma concentrations and did not adjust for plasma protein binding when computing efficacy.

Because the PK parameters of remdesivir have not been reported yet in humans, the same method could not be applied. We used the summary statistics reported in the summary for compassionate use of remdesivir filed by Gilead to the European Medicines Evaluation Agency (EMA)<sup>16</sup> to derive the mean concentration of the active metabolite in peripheral blood mononuclear cell, mean serum concentration (C<sub>mean</sub>), using the area under the curve (AUC) after a loading dose of 200 mg and after repeated doses of 100 mg q.d., and we assumed that the EC<sub>50</sub> for the metabolite was equal to the EC<sub>50</sub> of the parent.<sup>17</sup>

## RESULTS

We used a “target-cell limited” model with an eclipse phase.<sup>8</sup> given by Eq. 1 to characterize the viral load

dynamics of 13 hospitalized patients in Singapore for which data obtained from frequent nasopharyngeal swabs were available.<sup>3</sup> Because this model needs to incorporate a date of infection, an incubation period of 5 days was used as the most plausible date of infection in each patient.<sup>5</sup> The model fitted the data well (**Figure 1**) and using a model-averaging approach to take into account model uncertainty,<sup>11</sup> the within-host basic reproductive number, R<sub>0</sub>, was found equal to 8.6 (95% confidence interval (CI) = 1.9–17.6), and the death rate of productively infected cells was estimated as 0.60 day<sup>-1</sup> (95% CI = 0.22–0.97), corresponding to a median half-life of 1.2 days (**Table 2** and **Figure S1**). In influenza A, another respiratory infectious disease, estimates of the within host R<sub>0</sub> varied greatly, but the half-life of infected cells was shorter than 10 hours (see more details in ref. 18), suggesting a faster clearance of influenza-infected cells than SARS-CoV-2. The viral production rate p was also

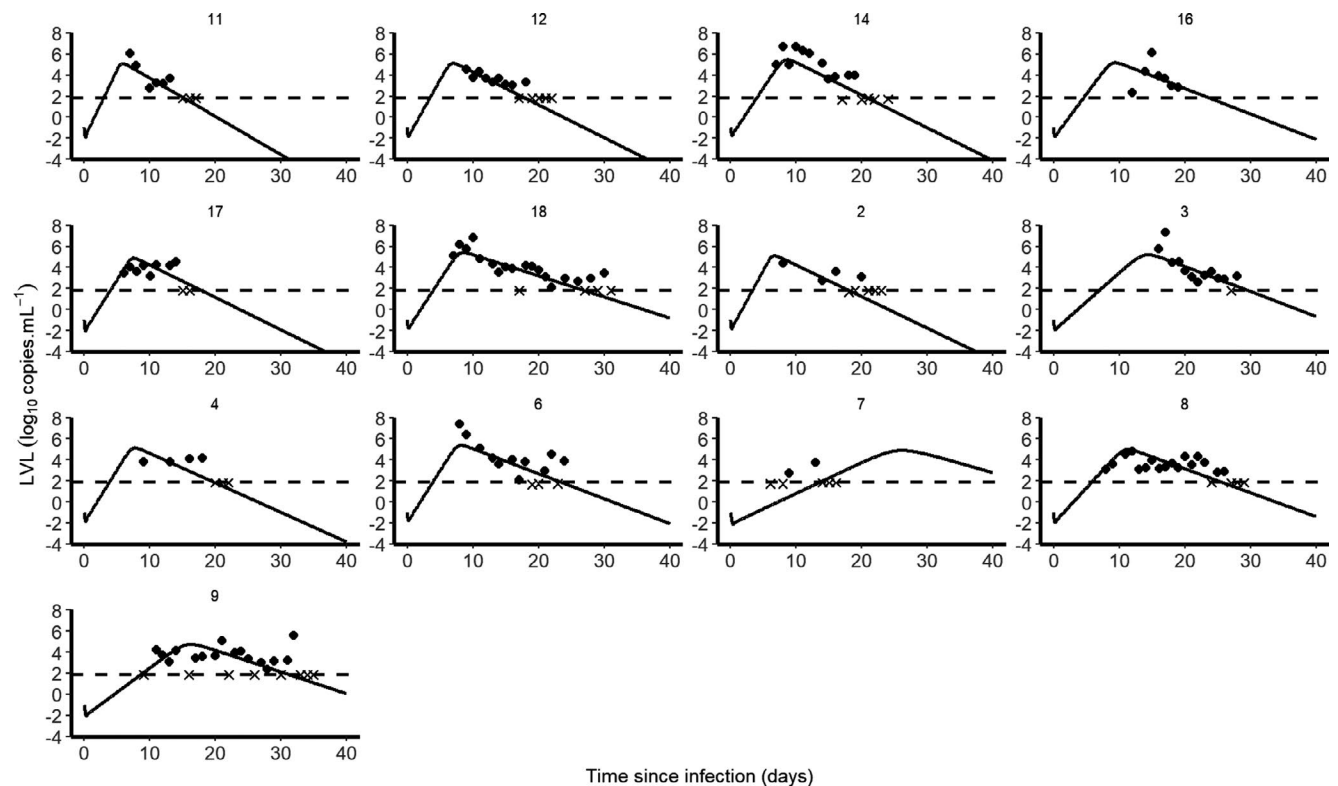


Figure 1 Individual predictions of severe acute respiratory syndrome-coronavirus 2 (SARS-CoV-2) of 13 untreated patients from Young et al.<sup>3</sup>

**Table 2** Median and confidence intervals of  $R_0$ ,  $\delta$ , and  $p$  across models and following model averaging procedure

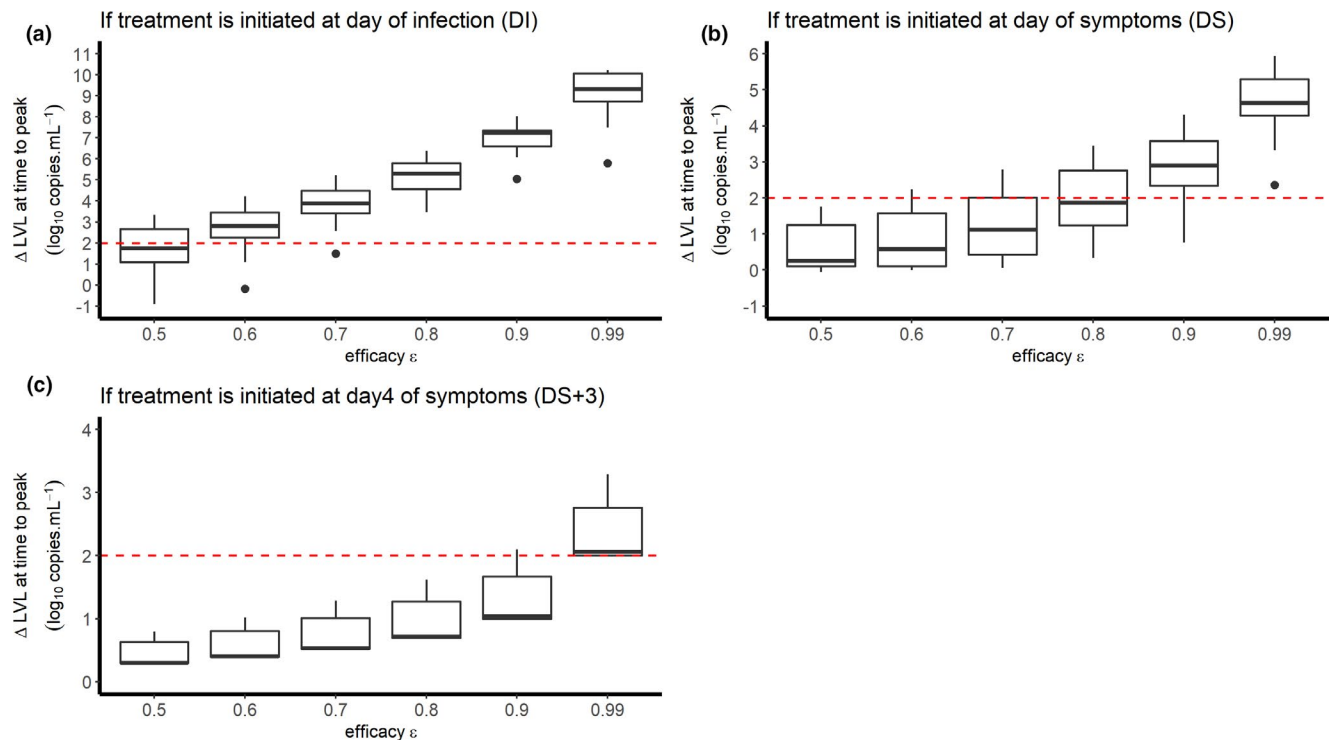
$k$ (day <sup>-1</sup> )	$V_0$ (log <sub>10</sub> cp/mL)	$R_0$ [95% CI]	$\delta$ [95% CI] (day <sup>-1</sup> )	$p$ [95% CI] (day <sup>-1</sup> )
1	10 <sup>-1</sup>	13.1 [5.1–21.8]	0.68 [0.44–0.9]	26.55 [0–57.32]
	10 <sup>-2</sup>	15.4 [7.9–23.2]	0.71 [0.46–0.94]	32 [8.38–56.26]
	10 <sup>-3</sup>	19.1 [10.3–28.8]	0.71 [0.46–0.94]	35.18 [12.46–60.06]
3	10 <sup>-1</sup>	8.2 [3.5–13.1]	0.6 [0.38–0.82]	21.36 [0–60.35]
	10 <sup>-2</sup>	9.8 [4.1–15.4]	0.58 [0.08–1.15]	20.37 [0–85.01]
	10 <sup>-3</sup>	12.5 [4.8–20]	0.58 [0.42–0.74]	23.37 [6.36–39.57]
5	10 <sup>-1</sup>	7.1 [0–13.9]	0.6 [0.19–1.06]	22.07 [0–58.57]
	10 <sup>-2</sup>	8.9 [4.7–13.2]	0.57 [0.43–0.74]	22.42 [1.9–41.84]
	10 <sup>-3</sup>	10.2 [4.8–16.2]	0.58 [0.46–0.71]	22.31 [5.48–41.46]
Model averaging		8.6 [1.9–17.6]	0.6 [0.22–0.97]	22.71 [0–59.64]

CI, confidence interval.

estimated as 22.7 copies/day (95% CI = 0–59.6; **Table 2**). However, as shown previously,  $p$  cannot be uniquely identified unless the initial target cell density  $T_0$  is known.<sup>19,20</sup> Therefore, the only quantity that can be reliably estimated is the product  $p \times T_0$ , equal to  $3.0 \times 10^6$  copies/day (95% CI = 0– $7.9 \times 10^6$ ). Parameter estimates and CIs were also consistent across models assuming a viral clearance  $c$  of 5 or 20 day<sup>-1</sup> (see **Table S1**).

Our model, along with the parameter estimates given above, also inform us both on the time to initiate antiviral treatment, and the level of efficacy that needs to be achieved to reduce viral load.<sup>6</sup> As limited information is available on the mechanisms leading to viral clearance, and how they

may be modulated by treatment, we used our model to predict the effects of treatment at day 5 post-symptoms, which corresponds to the time the viral load tends to peak in the absence of treatment in these data.<sup>3</sup> We considered a simple case where the drug effectiveness is assumed to be constant after therapy initiation (see Methods) and we calculated the minimal efficacy that would be needed to generate more than 2 logs of viral decline at peak viral load in the 13 studied patients (**Figure 2**). As predicted by viral kinetic modeling theory,<sup>2</sup> we found that the impact of treatment on peak viral load is inversely correlated with the time of treatment initiation. For a putative treatment blocking the viral production  $p$  and initiated at the time of infection, symptom



**Figure 2** Reduction in viral load at day 5 post-symptom onset according to the level of antiviral effectiveness of a treatment blocking the viral production and the timing of treatment initiation (a) at time of infection; (b) at time of symptom onset; (c) 3days after symptom onset). We assumed an incubation period of 5 days.

onset, or 3 days post-symptom onset, a median efficacy of at least 60%, 90%, and 99% in reducing viral replication would be needed, respectively, to generate more than 2 logs of decline in the peak viral load (Figure 2). The results obtained assuming 2 or 10 days of incubation are presented in Figures S2 and S3. We also considered the case of a drug like hydroxychloroquine blocking viral infection (parameter  $\beta$  in Eq. 1). Results were similar to those obtained before, as long as the treatment was initiated before or at symptom onset. However, initiating a treatment 3 days post-symptom onset could not reduce peak viral load, regardless of the drug effectiveness (see Figure S4).

The model, including an innate immune response, did not improve the data fitting and, therefore, was not selected for inclusion in the main text. However, the same analysis on viral dynamics and treatment was conducted and the conclusions remained unchanged (see Figures S5, S6 and Table S2).

How do these levels of effectiveness compare with the antiviral drugs that are currently being investigated? To study this question, we assumed that the treatment antiviral effectiveness at time  $t$  after treatment initiation,  $\epsilon(t)$ , was related to the plasma total drug concentration,  $C(t)$ :  $\epsilon(t) = \frac{C(t)}{C(t) + EC_{50}}$  (except for remdesivir, see Methods) and the mean antiviral effectiveness during the first 7 days of treatment is given by  $\bar{\epsilon} = \frac{1}{N} \times \frac{1}{7} \times \int_0^7 \frac{C(u)}{C(u) + EC_{50}} du$ . Given their PK and pharmacodynamic properties (Table 1), we calculated a mean antiviral effectiveness of up to 66% for lopinavir/ritonavir, 6% for hydroxychloroquine, 18% for IFN- $\beta$ -1a, and 87% for remdesivir. Given these estimates, these compounds are unlikely to have a dramatic effect on peak viral load if administered after the onset of symptoms. In fact, the effective concentrations will presumably be lower in patients, as drug availability may be further limited by protein binding (in particular for lopinavir, which has a protein binding rate > 98%) or capability to penetrate respiratory compartments. Importantly, levels of antiviral efficacy of ~ 50% could nonetheless be relevant in a prophylactic setting, before symptom onset, to reduce viral replication in the upper respiratory tract and reduce the risk of large infiltration to the lungs before an effective immune response is mounted to clear the virus.<sup>2</sup> Note, above, we calculated the effectiveness of drugs administered in monotherapy for their usual dosing regimen. We did not consider drugs that could directly target infected cells and lead to their elimination, such as some monoclonal antibodies.

## DISCUSSION

Overall, our results emphasize that the PK/pharmacodynamic properties of lopinavir/ritonavir, HCQ, IFN- $\beta$ -1a, and remdesivir make them unlikely to have a dramatic impact on nasopharyngeal viral load kinetics if they are administered after symptom onset. However, these drugs may be relevant to reduce viral replication if administered early (i.e., as a pre-exposure or post-exposure prophylaxis). This conclusion, however, depends on a number of hypotheses that we discuss below.

First, we focused on the capability of drugs to reduce the peak viral load making the implicit hypothesis that reducing peak viral load would likely reduce symptoms and disease

severity. However, the relationship between viral kinetics and disease severity is still debated, with several studies suggesting that viral load did not differ between patients with severe and nonsevere disease.<sup>21,22</sup> Related to this question, our study had some limitations. Our calculations relied on blood or plasma drug concentrations. Except for HCQ, for which the ratio of lung-to-plasma concentration is known to be high,<sup>23,24</sup> the lung exposure of the other drugs that we considered is unknown, and their effect on viral load in the lower respiratory tract, as measured from broncho-alveolar aspirates, for instance, may differ. In addition, the drug  $EC_{50}$  that we used were determined on Vero E6 cells, an *in vitro* system that may not reflect the *in vivo*  $EC_{50}$ . For instance, we found in another study that HCQ had no antiviral activity in a more physiological model of reconstituted human airway epithelium, and this may explain the absence of antiviral activity of HCQ *in vivo* against SARS-CoV-2.<sup>24,25</sup> Finally, we focused solely on the antiviral effects of these drugs, and did not consider other potential effects of these drugs, such as their immunomodulatory effects. Such effects have been suggested for drugs that are not purely antivirals, such as HCQ and IFN- $\beta$ -1a.<sup>26,27</sup>

Another implicit implication of our work is the benefit of drugs used for prophylaxis (i.e., before exposure to the virus). In that case, the objective of the treatment may be to “flatten the peak viral load” (by analogy with the now popular terminology of epidemiological models) but also to prevent infection. Our deterministic, ordinary differential equation-based model, cannot reproduce virus extinction, but this can be captured using a stochastic version of the same model.<sup>28</sup>

Our modeling provides estimates of viral kinetic parameters, in particular  $R_0$  and the loss rate of infected cells,  $\delta$ , and the limit of extrapolation of these parameters also need to be well understood. The advantage of studying this series of patients was the fact that viral load was sampled extremely frequently and early in the infection. However, the number of patients we studied was quite small (only 13), and the patients were rather young (37 years, range 31–56 years) and may not be representative of patients who typically evolve to have severe forms of the disease.<sup>21</sup> Therefore, more data will be needed from various populations to estimate precisely the parameter distribution in the population of patients that are most in need of antiviral therapy. Finally, our estimate of the loss rate of free virus and infected cells are constant over time, which neglects the effects of the adaptive immune response. For instance, antibodies that emerge in the second week after symptom onset,<sup>29,30</sup> may contribute to accelerate viral clearance or reduce viral infectivity. However, to include these effects would require more complex models and quantitative data on these antibodies and their *in vivo* effects, which are currently lacking. Further, our analysis of the antiviral effects of repurposed drugs focused on their ability to reduce the peak viral load, which typically occurs well before the antibody response emerges and, hence, the analysis we presented here should not be affected in any major way by our neglect of antibody responses.

Future models of drug efficacy may need to account for viral resistance, as it is possible that continued viral replication in the presence of drug will select for drug-resistant mutations,<sup>31</sup> although coronaviruses are unusual in that they appear to have low mutation rates due to RNA proofreading capability.<sup>32</sup> Drug combination therapy and more aggressive dosing, including



consideration of loading doses to rapidly achieve therapeutic exposures, may be beneficial to maximize efficacy of these repurposed antiviral agents.<sup>33</sup> For all these reasons, the outcome of randomized clinical trials remains urgently needed, and the analysis of their impacts not only on viral clearance but also on disease severity will be critical to design more potent drugs.

**Supporting Information.** Supplementary information accompanies this paper on the *CPT: Pharmacometrics & Systems Pharmacology* website ([www.psp-journal.com](http://www.psp-journal.com)).

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**Conflict of Interest.** J.G. has consulted for F. Hoffman-La Roche. All other authors declared no competing interests for this work.

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**Ethical Statement.** Data were originally provided in Young *et al.* (<https://doi.org/10.1001/jama.2020.3204>) where “waiver of informed consent for collection of clinical data from infected individuals was granted by the Ministry of Health, Singapore” and “written informed consent was obtained from study participants.”

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