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Balanced Polarizable Drude Force Field Parameters for Molecular Anions: Phosphates, Sulfates, Sulfamates and Oxides

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Abstract

Polarizable force fields are emerging as a more accurate alternative to additive force fields in terms of modeling and simulations of a variety of chemicals including biomolecules. Explicit treatment of induced polarization in charged species such as phosphates and sulfates offers the potential for achieving an improved atomistic understanding of the physical forces driving their interactions with their environments. To help achieve this, in this study we present balanced Drude polarizable force-field parameters for molecular ions including phosphates, sulfates, sulfamates and oxides. Better balance was primarily achieved in the relative values of minimum interaction energies and distances of the anionic model compounds with water at the Drude and quantum mechanical (QM) model chemistries. Parametrization involved reoptimizing available parameters as well as extending the force field to new molecules with the goal of achieving self consistency with respect to the Lennard-Jones and electrostatic parameters targeting QM and experimental hydration free energies. The resulting force fields parameters achieve consistent treatment across the studied anions, facilitating more balanced simulations of biomolecules and small organic molecules in the context of the classical Drude polarizable force field.

Graphical Abstract

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Author Contributions

ADM, AAK and AHA conceived the work. AAK and AHA performed the calculations. AAK, AHA and ADM performed the analysis. AAK, AHA, and ADM wrote and revised the manuscript. AAK and AHA contributed equally.

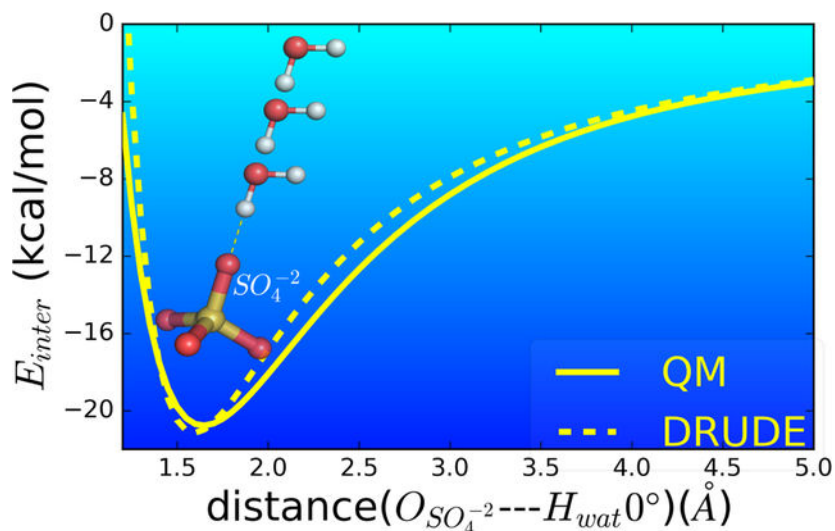
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Supporting Information

Figures and tables showing details of water interaction energy calculations, dipole moments, molecular polarization and final force field parameters for molecular anions.

Competing Financial Interests Statement

ADM Jr. is cofounder and CSO of SilcsBio LLC.



Keywords

Molecular ions; polarizable force field; CHARMM; molecular dynamics; molecular modeling

Introduction

Over last three decades there has been significant increase in computational modeling and simulations of biomolecules to investigate various problems that are experimentally difficult to explore.[1–4] The field has been significantly dependent on empirical potential energy functions that are based on non-polarizable, additive force fields.[5–14] However, the fixed charges on the atoms in these models restrict the investigation of inductive effects on molecules that arise in diverse condensed-phase environments. A notable example of a limitation in additive forces is the inability to correctly treat the distribution of atomic ions at air–water interfaces,[15–17] an issue that was overcome by the explicit inclusion of polarizability in the force field.[18–21] This and other examples,[22–28] point towards the need for polarizable force fields of high accuracy that encompass a wide range of chemical species. While there are a handful of polarizable force fields including induced point dipole or fluctuating charge models [29–31] our lab is focused on development of the CHARMM Drude Force Field based on the classical Drude oscillator.

Phosphate and sulfate moieties occur in a wide range of biomolecules including carbohydrates, glycoproteins, nucleic acids, and lipids. Specifically, phosphates and sulfates play important roles in cellular function including protein-protein interactions facilitated by phosphorylation.[32, 33] Anions such as sulfamates and oxides are present in drug-like molecules and various oxides are intermediates of enzymatic reactions, such as proteolysis. [34, 35] Accordingly, accurate, consistent treatment of molecular anions is essential for a comprehensive force field that will be of utility for modeling and simulation studies of heterogeneous chemical and biomolecular systems.

Our laboratory has been involved in the development of a comprehensive polarizable force field based on the classical Drude Oscillator model; alternative approaches and efforts to treat electronic polarization have been extensively reviewed elsewhere.[20, 36–40] Details of the Drude energy function, parameterization strategies and methodological information for various classes of molecular have been reported in previous studies.[41–47] Notably, the availability of the Drude model in different simulation packages including CHARMM[48], NAMD[49], GROMACS[50], OpenMM[51, 52] and ChemShell QM/MM[53–55] have made it convenient to run polarizable simulations into the microsecond time scale.

The current Drude polarizable force field has a limited number of molecular anions and their optimization in the context of different applications has led to some inconsistencies in the parameters for those molecules. For example, the phosphate parameters were initially developed in the context of the nucleic acids[56] while alternate Lennard-Jones (LJ) and electrostatic parameters were developed in the context of phospholipids.[57, 58] Further, the atom types from nucleic acids were recently used to parameterize the methyl phosphate ions and their interactions with magnesium ions.[59] The imbalance in the force field could cause a divergent effect as future molecules are parameterized with machine learning approaches being developed to predict the parameters for new small molecules. In this work, to achieve more balanced parameters for the Drude molecular anions as well as extend the coverage of the force field for this class of molecules, optimization of sulfate, phosphate, sulfamate and selected oxides are presented and discussed. The molecular anions studied are listed on Table 1 along with their abbreviations.

Computational Methods

Quantum Mechanical (QM) calculations were performed with the PSI4 [60] and Gaussian 03.[61] programs. Solute-water interaction geometries were constructed based on the donor-acceptor properties of the model compound by placing a single water at distances from 1.5 to 5.0 Å in steps of 0.05 Å in selected orientations (Figure 1 and S1–10). Monomer geometries were obtained from gas phase optimizations done at the MP2/6–31G(d) model chemistry to default tolerances. Then single point interaction energies of the model compound-water dimers as well as the individual monomers were calculated at the MP2/cc-pVQZ model chemistry with counterpoise correction for basis set superposition error (BSSE) using the gas phase monomer geometries of the model compounds with the SWM4-NDP geometry for water.[62–64] Interaction energies were obtained based on the difference between the total energy of the dimer and that of the individual monomers in the gas phase. Dipole moments were determined for MP2/6–31+G(d) gas phase optimized structures followed by MP2/cc-pVQZ single point calculations using Gaussian 03.[61] The gas phase molecular polarizabilities were estimated using the finite difference method[65] at the RIMP2/cc-pVQZ//MP2/6–31G(d) model chemistry using Psi4 inputs generated by the program FFParm.[60, 66]

Molecular Mechanics (MM) calculation with Drude polarizable force field involved energy minimization of the model compounds in the gas phase to a tolerance of 10^{-5} kcal·mol⁻¹·Å⁻¹. Model compound-water interaction energies were obtained by relaxing the Drude particles via minimization by steepest-descent (SD) algorithm for 200 steps followed by an

adopted-basis Newton–Raphson (ABNR) algorithm for 500 steps to a final gradient of 10^{-5} kcal·mol⁻¹·Å⁻¹ with the atomic positions restrained with a force constant of 10^7 kcal·mol⁻¹·Å⁻² to the QM gas phase optimized geometry. Interaction energies were again obtained based on the difference between the total energy of the dimer and that of the individual monomers in the gas phase. Dipole moments and molecular polarizabilities were calculated using the QM optimized geometries with only relaxation of the Drude particles as described above. FFParm was used to setup all the calculations above and analyze the results from QM and MM.[66]

Adiabatic dihedral potential energy scans (PES) were performed by geometry optimization at the MP2/6–31G(d) level with the target dihedral constrained followed by RIMP2/cc-pVQZ single point energies (i.e., RIMP2/cc-pVQZ//MP2/6–31G(d) QM model chemistry) using the Psi4 program.[60] PES were performed in 15° increments resulting in 24 conformations for 1D and 576 conformations for 2D surfaces. Analogous adiabatic MM PES were then computed with the target dihedral constrained to the QM values using the Drude force field.[47] Selected dihedral parameters were optimized, including the C-O-P-O dihedral parameter in dimethylphosphate (DMP). Once the QM target PES was calculated the analogous adiabatic MM PES was computed with targeted dihedral force constant set to zero and the MM PES calculated. The energy difference between the MM and QM (MM-QM) PES was then targeted for the fitting of the dihedral parameters using the least-square fitting program developed in this laboratory specifically for use in parameter optimization. [67] During fitting multiplicities n of 1, 2, 3, 4, 5 and 6 were included and the phase angle, δ , was limited to 0 and 180° as required to insure applicability of the parameters to stereoisomers about a chiral center.

The hydration free energy of molecular anions was calculated through alchemical free energy perturbation (FEP) simulations.[68] Final hydration free energies were based on three individual calculations (condensed phase, gas phase and long-range correction (*LRC*)), described and established in previous work by Lin et al.[41, 42] In short, for the condensed phase first the molecule was solvated in a box of SWM4-NDP [62] water and equilibrated for 100 ps in the NPT ensemble at 298.15 K and 1 atm using CHARMM[69]. Then the alchemical FEP was implemented applying coupling parameters to calculate the contributions to free energy by repulsive, dispersive and electrostatic terms based on the approach of Deng and Roux.[70] Each window was equilibrated for 100 ps and data was collected over 200 ps for the production runs. For the charged molecules a contribution arising from the Galvani potential Φ generated from the vacuum/water interface was added as a correction[71, 72], along with an entropic correction term (dS_{corr})[73] as shown in equations 1, 2 and 3.

$$\Delta G_{hyd} = \Delta G_{aq} - \Delta G_{gas} + z.f\Phi + dS_{corr} + LRC \quad (1)$$

$$\Delta G_{aq} = (\Delta G_{aq}^{disp} + \Delta G_{aq}^{rep}) + \Delta G_{aq}^{elec} \quad (2)$$

$$\Delta G_{gas} = (\Delta G_{gas}^{disp} + \Delta G_{gas}^{rep}) + \Delta G_{gas}^{elec} \quad (3)$$

In eq.1 G_{hyd} is the final calculated value of the hydration free energy. G_{aq} and G_{gas} are calculated as sums of the non-polar and electrostatic (ΔG_{gas}^{elec}) contributions (Eq. 2 and 3), where the non-polar contribution is divided into dispersive (ΔG_{gas}^{disp}) and repulsive (ΔG_{gas}^{rep}) components associated with the vdW term. The $z/f\Phi$ term corresponds to interfacial correction where z is the charge on the molecule, f is Faraday's constant and Φ is Galvani potential. For SWM4-NDP water model $\Phi = -0.545$ V.[72] LRC is the long-range correction calculated according to Baker et. al.[74] MD simulations with the Drude model were performed as described in previous studies.[41–43] All molecular mechanics, MD simulations and FEP calculations were performed with the program CHARMM[69].

Results and Discussion

Parameter optimization efforts were designed to extend the treatment of molecular anions in the Drude force field as well as overcome divergent parameters that were developed during optimization of different aspects of the force field. This included divergence of the phosphate parameters based on DMP in nucleic acids versus in phospholipids. For example, the charges on the anionic oxygens in the nucleic acids was -0.776 , while that in the lipids was -0.876 ; the final optimized charge being -0.856 from the present study. The LJ radii, $R_{min}/2$ (i.e. $2^{-5/6} \sigma$), on the anionic oxygens were 1.865 , 1.970 and 1.865 Å in the nucleic acid, lipid and optimized models, respectively, while the LJ ϵ_{min} parameter on the anionic oxygen for the nucleic acids was changed from -0.07 to the lipid value to -0.19 kcal/mol. Similarly, for the ester oxygen the nucleic acid ϵ_{min} was adjusted from -0.02 to the lipid value of -0.17 kcal/mol. Accordingly, in the present work a unified model was developed that will be applied to various systems moving forward. In addition, when considering different molecular anions, optimization in the present study was performed to assure that the differences between the QM and Drude minimum interaction energies and distances were well balanced. For example, there was a tendency for the Drude minimum energy interactions to be more favorable and shorter than QM data involving interactions of water with the nucleic acid DMP anionic phosphates while the opposite occurred with the lipid DMP parameters as discussed in the next section. Accordingly, parameter optimization was focused on minimizing these differences, balancing the agreement between all the interaction orientations and reproducing the hydration free energies of the studied model compounds. Adjustment of the electrostatic parameters also considered the gas phase molecular dipole moments including their tensors.

Water-model compound interactions

Model compound-water interactions were analyzed for various orientations with respect to the different donor/accepter atoms in the compounds. In Figure 1 the example of water-DMP interactions are shown including results from the initial nucleic acid and lipid models, the final optimized model and from the QM calculation. The original nucleic acid parameters yielded minimum energies and distances that were slightly shorter and more favorable than the QM while the opposite was true for the lipid parameters. These differences are

associated with both the partial atomic charges and the LJ parameters. The longer distances with the lipid model are associated with the larger $R_{\min}/2$ on the anionic oxygens, a problem that was fixed with the change from the value of 1.970 to the nucleic acid value 1.865 Å. Simultaneously, the change in ϵ_{\min} and the decrease in the magnitude of the charge from -0.876 to -0.856 corrects for the overestimation seen with the nucleic acid model. This yields the final optimized parameters that are in good agreement with the QM data for both the minimum interaction energies and distances (Table 1). Furthermore, during adjustment of the charge distribution the reproduction of target data on the dipole moment and hydration free energy were considered, as discussed below.

To preserve consistency across different species of molecular anions the diverse model compounds listed in Table 1 were selected and optimized applying the present well-balanced approach. The water-model compound interactions energy orientations along with the corresponding energy surfaces are presented in Figures 1 and S1–11. As shown, Drude energies were computed for the original, previously published parameters, when available, along with those optimized in this work. Of the model compounds in Table 1, HP_1, SO₄, MSO₄, MSNA, NMSM, NESM, MEO and ETO were not previously available in the Drude force field.

In the case of the phosphates, it was important to obtain balanced parameters that were applicable to DMP, methylphosphates (MP_0, MP_1 and MP_2) and phosphates (HP_1 and HP_2). We were able to bring consistency in the charges on the anionic oxygens of DMP, MP_1 and HP_1, which all have a total charge of -1 . Also, in the previous parameters the charges on the anionic oxygens in MP_2, MP_1 and MP_0 decreased irregularly as the magnitude of the total charge decreased.[59] With re-optimization, there is now a consistent decrease in the anionic oxygen charges going from MP_2 to MP_1 to MP_0. Presented in Table 2 are results from the original and optimized models for DMP, all phosphate containing compounds excluding DMP and all non-phosphate containing compounds. The individual minimum interaction energies and distances are reported in Tables S1 to S3 of the supplemental information. As expected, the differences defined by the different metrics in Table 2 show improvements versus the original parameters were available.

Dipole moments

Molecular dipole moments were also included as target data for optimization of the electrostatic parameters. A summary of the results is presented in Table 3 with the differences with respect to QM in the dipole moments before and after optimization. Total dipoles as well as the X, Y and Z components for all the molecules are provided in the supporting Tables S4 and S5. Intramolecular geometries for the Drude model were the QM optimized geometry with only Drude particle positions minimized in the MM calculations. For model compounds in which the electrostatic parameters were optimized as part of the present study, dipole moments were computed for both the original and optimized parameters. Specifically, DMP shows significant improvement in total dipole moment with respect to the nucleic acid version and a slight compromise relative to the lipid version as that model was optimized more recently targeting similar QM data (Table S4). Similarly, MP_1, MP_2 and HP_2 show notable improvement compared to original parameters (Table

S5). An excellent agreement with the QM data was also obtained with all the newly parameterized molecules that were not previously in the Drude force field showing an overall < 0.1 Debye absolute average difference in the total dipole moments (Table S5, legend).

Molecular Polarizabilities

Comparisons were also made with respect to the molecular polarizabilities, though these were not considered directly as target data for the optimization. Table 4 includes the ratios of the Drude to QM molecular polarizabilities, with the Drude values based on the QM geometries. The absolute values of the molecular polarizabilities are in Table S7. Overall, the Drude polarizabilities are smaller than the QM values, consistent with the polarizability scaling used in the Drude FF.[76] With polar neutral species the ratios, or scaling factors, are typically close to 0.85 though smaller values have been obtained. With the anions the average ratio is 0.75. This smaller value may be associated with the more diffuse nature of the electron distribution associated with the negative ions in the gas phase versus neutral species. In the condensed phase the presence of molecules (eg. water) around the anions may limit the extent that the electron cloud can distort leading to smaller effective molecular polarizabilities. However, the approximation that the electric field of the atoms at their nuclei is used to determine the polarization response may also contribute to this effect.[77] Further studies are required to address this issue in detail.

Conformational properties

While the majority of bonded parameters were already available in the Drude FF and not subjected to optimization, it was necessary to improve selected dihedral parameters in molecules that included O-methyl or O-ethyl moieties or hydroxyl groups. Presented in Figure 2 and 3 are the QM and Drude PES. In the case of DMP, a two-dimensional PES was performed for the dihedrals C1-O11-P-O12 and O11-P-O12-C2 to examine the relative stability of *gauche-gauche* (*gg*), *gauche-trans* (*gt*), and *trans-trans* (*tt*) conformations and the barriers between those minima. Figure 3 shows the heatmap of PES with respect to the lowest energy at *gg* conformation at $\pm 75^\circ$. The *gt* and *tt* conformations have slightly unfavorable energies compared to *gg*, though both are comparable to each other as reported previously.[78, 79] As is evident the Drude PES are typically in very good agreement with the QM data, indicating that the overall conformations of the molecules and the barriers between low energy conformations are satisfactorily treated.

Hydration free energies

The final target data were the hydration free energies, which were calculated using the free energy perturbation protocol established by Deng and Roux.[70] For all the anions, target hydration free energies were corrected to account for the vacuum/water interface, entropic terms and long-range dispersion corrections (Equation 1). Two methods for calculation of the correction for target data could be applied according to the type of experimental data where the hydration free energy is measured relative to an ion with the same charge or in the presence of a counterion.[41, 42]

$$\text{When X is coion, } \Delta G_{Y^-}^{\text{target}} = \Delta G_{Y^-}^{\text{expt}} + (\Delta G_{X^-}^{\text{drude}} - \Delta G_{X^-}^{\text{expt}}) \quad \text{Method-1:}$$

$$\text{When X is counterion, } \Delta G_{Y^-}^{\text{target}} = \Delta G_{Y^-}^{\text{expt}} + (\Delta G_{X^+}^{\text{expt}} - \Delta G_{X^+}^{\text{drude}}) \quad \text{Method-2:}$$

Based on which approach was used in the experiments, the appropriate correction/offset is required using the corresponding coion/counterion. Accordingly, we have used Cl^- ($G^{\text{expt}} = -74.60$ kcal/mol and $G^{\text{drude}} = -78.40$ kcal/mol), Na^+ ($G^{\text{expt}} = -100.6$ kcal/mol and $G^{\text{drude}} = -96.3$ kcal/mol), and H^+ ($G^{\text{expt}} = -265.9$ kcal/mol and $G^{\text{drude}} = -258.8$ kcal/mol) to define the target data. For DMP a true experimental value is unavailable and an estimation of $G^{\text{expt}} = -76.0 \pm 4.0$ kcal/mol provided by Klauda et. al.[80] has been used as reference value in this study. Since the estimation uses H^+ as counterion we calculated the target data for DMP using the same; however, the uncertainty in the G^{expt} estimation is high, limiting its utility. For HP_1 three different experimental data are available ranging from -68 to -111 kcal/mol. To be consistent, we have used G^{expt} values -76.0 and -299.0 kcal/mol for HP_1 and HP_2, respectively, from the same study, which uses Na^+ as counterion in the experimental evaluation.[81]

Presented in Table 5 are the experimental and calculated hydration free energies. The overall level of agreement with the Drude model to reproduce the hydration free energies is agreeable (considering the unavailability/ambiguity in experimental data) with an overall RMS difference of 1.33 kcal/mol, excluding the compound HP_2. The hydration free energy of HP_2 was sacrificed to obtain more accurate reproduction of that of HP_1, which is within 1.0 kcal/mol of the target value. Only in the case of HP_2 is the hydration free energy in poorer agreement with the target data as compared to the previous parameters.

Conclusions

The study presents an important refinement of the Drude polarizable FF parameters for modeling of various molecular anions. The effort focused on achieving a better balance in the Drude parameters for these species primarily with respect to their interactions with water while yielding good agreement with molecular dipoles and polarizabilities and the condensed phase hydration free energies. The most important of these were the modifications in the non-bonded parameters of the anionic oxygen of the phosphates, especially in DMP, which is critical for the treatment of nucleic acids and phospholipids. Importantly, the same LJ parameters are used for the methylphosphates, thereby simplifying the force field with respect to the number of atom types.

Final topologies and parameters are provided in the supporting information and can also be accessed from the MacKerell laboratory web site (http://mackerell.umaryland.edu/CHARMM_ff_params.html) with future releases of the program CHARMM and from the Drude Prepper module in the CHARMM-GUI.[87, 88] This includes inclusion of the developed Drude polarizable force field parameters for molecular anions with the remainder of the classical Drude polarizable force, including carbohydrates[46, 47, 89, 90], lipids[58], nucleic acids[91, 92] and proteins[93].

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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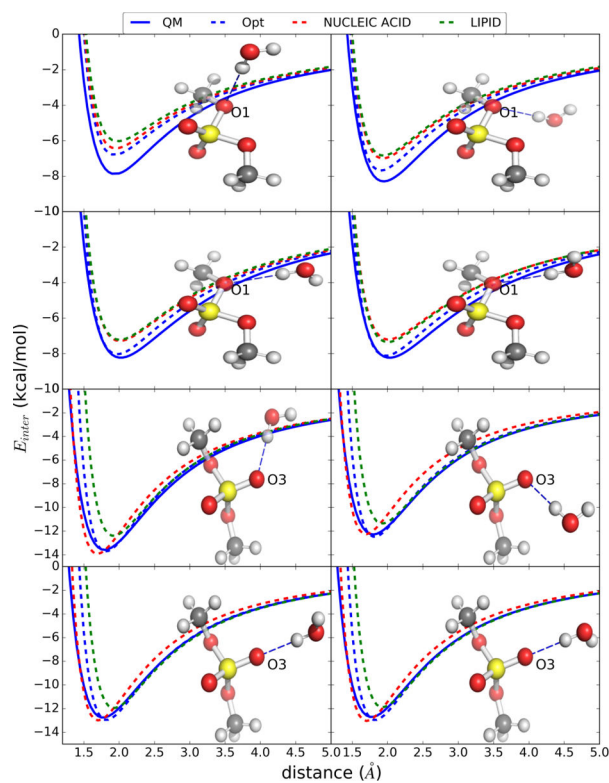


Figure 1.

Water interaction energy surfaces as a function of distance from the QM and Drude models with dimethylphosphate (DMP). Distances are between the hydrogen (H) on the water model and the respective model compound oxygens as shown. Comprehensive interaction energy figures are included in the supplement Figure S1–11. Images rendered using PyMol. [75]

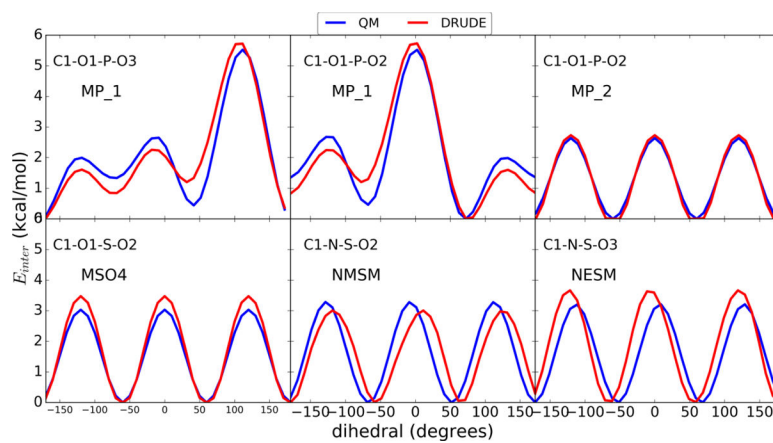


Figure 2.
QM and optimized Drude torsional PES along with target dihedral labels.

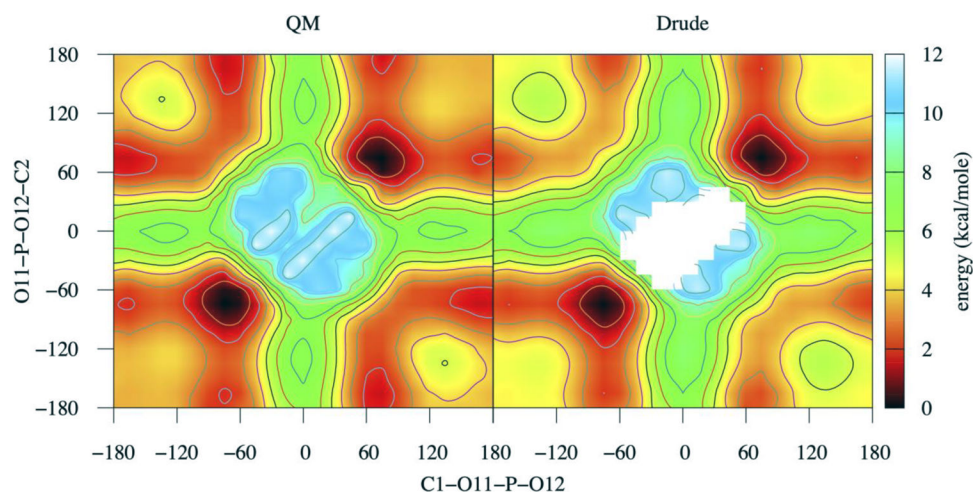


Figure 3. 2D QM and MM conformational energies for DMP along C1-O11-P-O12 and O11-P-O12-C2 dihedrals. On the conformational surface, *gg* (global minima) lies on ± 75.0 degree.

Table 1.

Molecular anions and their abbreviations studied in this work. Molecules not previously in the Drude force field are indicated by *. Molecules ACET, PHET, MES and ETS were optimized in recent study by Lin et. al. [42]

Molecule names	Formula	Abbreviations (Residue ID)
Dimethylphosphate	$(\text{CH}_3)_2\text{PO}_4$	DMP
Methylphosphate (neutral)	$\text{CH}_3\text{PO}_4\text{H}_2$	MP_0
Methylphosphate (anionic)	$\text{CH}_3\text{PO}_4\text{H}^-$	MP_1
Methylphosphate (dianionic)	$\text{CH}_3\text{PO}_4^{2-}$	MP_2
Phosphate (anionic)	H_2PO_4^-	HP_1*
Phosphate (dianionic)	HPO_4^{2-}	HP_2
Sulfate	SO_4^{2-}	SO4*
Methylsulfate	CH_3SO_4^-	MSO4*
Methylsulfonate	CH_3SO_3^-	MSNA*
N-methylsulfamate	$\text{CH}_3\text{NHSO}_3^-$	NMSM*
N-ethylsulfamate	$\text{C}_2\text{H}_5\text{NHSO}_3^-$	NESM*
Methoxide	CH_3O^-	MEO*
Ethoxide	$\text{C}_2\text{H}_5\text{O}^-$	ETO*
Acetate	CH_3COO^-	ACET
Phenolate	$\text{C}_6\text{H}_5\text{O}^-$	PHET
Methanethiolate	CH_3S^-	MES
Ethanethiolate	$\text{C}_2\text{H}_5\text{S}^-$	ETS

Table 2.

Statistical analysis of the differences in the water-model compound minimum interaction energies (E_{\min} , kcal/mol) and distances (R_{\min} , Å) for molecular ions between QM and Drude model (Drude). Complete lists of the water minimum interaction energies and distances can be found in Tables S1 to S3 of the supplemental information.

	DMP						All phosphate containing compounds excluding DMP				Non-phosphate containing compounds	
	Nucleic Acid		Lipid		Opt		Original		Opt		Opt	
	E_{\min}	R_{\min}	E_{\min}	R_{\min}	E_{\min}	R_{\min}	E_{\min}	R_{\min}	E_{\min}	R_{\min}	E_{\min}	R_{\min}
AVG Difference	0.55	0.01	-0.09	-0.01	-0.09	0.01	0.24	0.07	-0.44	0.01	-0.18	0.01
ABS_AVG Difference	0.74	0.06	1.12	0.11	0.34	0.05	0.79	0.11	0.57	0.07	0.58	0.04
STDEV Difference	0.91	0.07	0.35	0.07	0.35	0.07	0.96	0.16	0.63	0.13	0.76	0.07
RMS Difference	1.07	0.11	0.36	0.15	0.36	0.08	0.99	0.17	0.76	0.13	0.78	0.07

Table 3.

Dipole moment differences between the Drude and QM models for molecules in which the electrostatic parameters were updated as part of the present study. QM data at the RIMP2/cc-pVQZ//MP2/6–31G(d) model chemistry. All values are in Debye.

Differences = Drude - QM	DMP			Molecules previously in the Drude FF excluding DMP	
	Nucleic Acid	Lipid	Opt	Original	Opt
AVG Difference	0.21	-0.04	0.12	0.17	0.05
ABS_AVG Difference	0.30	0.04	0.03	0.43	0.16
STDEV Difference	0.79	0.12	0.24	0.30	0.23
RMS Difference	0.83	0.12	0.28	0.54	0.24

Table 4.

Ratio of Drude and QM molecular polarizabilities and their tensors. QM data at the RIMP2/cc-pVQZ//MP2/6-31G(d) model chemistry.

Molecule	Ratio: Drude/QM			
	XX	YY	ZZ	total
DMP	0.72	0.75	0.74	0.74
MP_0	0.82	0.91	0.87	0.87
MP_1	0.85	0.77	0.74	0.79
MP_2	0.75	0.55	0.72	0.66
HP_1	0.91	0.89	0.87	0.89
HP_2	0.74	0.74	0.78	0.75
SO4	0.71	0.71	0.71	0.70
MSO4	0.69	0.77	0.82	0.76
MSNA	0.66	0.73	0.73	0.70
NMSM	0.87	0.78	0.81	0.82
NESM	0.75	0.82	0.84	0.80
MEO	0.56	0.56	0.57	0.56
ETO	1.00	0.61	0.59	0.74
Average	0.77	0.74	0.75	0.75

Table 5.

Calculated, experimental and target free energies of hydration for the molecular anions. Detailed distribution of calculated free energies are provided in Table S6 of the supplemental information. All values are in kcal/mol. Values in parenthesis for DMP corresponds to the calculated hydration free energy based on the current Drude nucleic acid parameters. $G_{\text{drude}_{\text{opt}}}$ is average hydration free energy from 3 independent runs and with errors being standard errors calculated from the 3 independent runs.

Molecule	G_{expt}	G_{target}	$G_{\text{drude}_{\text{orig}}}$	$G_{\text{drude}_{\text{opt}}}$	$G_{\text{Diff}_{\text{orig}}}$	$G_{\text{Diff}_{\text{opt}}}$
DMP	-76.00[80]	-83.10 ^a	-78.48 (-70.99)	-81.04 ± 0.08	-3.32 (-10.81)	2.06
MP_0	N A		-13.83	-10.47 ± 0.22		
MP_1	N A		-87.95	-77.99 ± 0.29		
MP_2	N A		-284.28	-265.27 ± 0.25		
HP_1	-76.00[81-84]	-79.70 ^b		-80.25 ± 0.18		-0.55
HP_2	-299.00[81, 82]	-302.70 ^b	-294.15	-275.54 ± 0.16	-8.65	27.16
SO4	-258.25[83]	-262.05 ^a		-263.19 ± 0.31		-1.14
MSO4	N A			-67.83 ± 0.03		
MSNA	N A			-70.51 ± 0.10		
NMSM	N A			-72.83 ± 0.09		
NESM	N A			-72.77 ± 0.26		
MEO	-95.20[85]	-102.30 ^c		-100.64 ± 0.11		1.66
ETO	-91.20[85]	-98.30 ^c		-95.62 ± 0.32		2.68
ACET*	-77.3[85, 86]	-84.4 ^c		-84.7 ± 0.1		-0.3
PHET*	-71.3[85, 86]	-78.4 ^c		-78.2 ± 0.0		0.2
MES*	-73.7[85, 86]	-80.8 ^b		-80.6 ± 0.1		0.2
ETS*	-71.8[85, 86]	-78.7 ^c		-78.9 ± 0.1		-0.2

^aMethod2 with Proton (H⁺)[85],

^bMethod2 with Na⁺ [81],

^cMethod1 with Cl⁻,

* data from Lin et. al.[42], N A – Not Available