

Mechanistic Causes of Reduced Cardiorespiratory Fitness in Type 2 Diabetes

Layla A. Abushamat,¹ P. Mason McClatchey,² Rebecca L. Scalzo,^{1,3,4} Irene Schauer,^{1,3,4} Amy G. Huebschmann,^{1,4} Kristen J. Nadeau,^{4,5} Zhenqi Liu,⁶ Judith G. Regensteiner,^{1,4,*} and Jane E. B. Reusch^{1,3,4,*}

¹Department of Medicine, University of Colorado, Anschutz Medical Campus, Aurora, Colorado 80045; ²Sigilon Therapeutics, Cambridge, Massachusetts 02142; ³Rocky Mountain Regional VA, Aurora, Colorado 80045; ⁴Center for Women's Health Research, University of Colorado, Anschutz Medical Campus, Aurora, Colorado 80045; ⁵Department of Pediatrics, University of Colorado, Anschutz Medical Campus, Aurora, Colorado 80045; and ⁶Department of Medicine, University of Virginia, Charlottesville, Virginia 22908

*Senior authors.

ORCID numbers: 0000-0001-9291-7390 (R. L. Scalzo); 0000-0001-5077-0941 (Z. Liu); 0000-0001-8620-1003 (J. E. B. Reusch).

Type 2 diabetes (T2D) has been rising in prevalence in the United States and worldwide over the past few decades and contributes to significant morbidity and premature mortality, primarily due to cardiovascular disease (CVD). Cardiorespiratory fitness (CRF) is a modifiable cardiovascular (CV) risk factor in the general population and in people with T2D. Young people and adults with T2D have reduced CRF when compared with their peers without T2D who are similarly active and of similar body mass index. Furthermore, the impairment in CRF conferred by T2D is greater in women than in men. Various factors may contribute to this abnormality in people with T2D, including insulin resistance and mitochondrial, vascular, and cardiac dysfunction. As proof of concept that understanding the mediators of impaired CRF in T2D can inform intervention, we previously demonstrated that an insulin sensitizer improved CRF in adults with T2D. This review focuses on how contributing factors influence CRF and why they may be compromised in T2D. Functional exercise capacity is a measure of interrelated systems biology; as such, the contribution of derangement in each of these factors to T2D-mediated impairment in CRF is complex and varied. Therefore, successful approaches to improve CRF in T2D should be multifaceted and individually designed. The current status of this research and future directions are outlined.

© Endocrine Society 2020.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Key Words: cardiorespiratory fitness, type 2 diabetes, cardiovascular disease, microvascular, mitochondria, endothelium

The prevalence of diabetes mellitus (DM) in the United States has been rising since 1990. In 2018, 34.2 million people, around 10.5% of the US population, had DM (11% of men, 9.5% of women) [1]. In 2016, it was estimated that 91.2% of adults with DM have type 2 diabetes (T2D) [2] and the prevalence of youth-onset T2D is also rising [1]. DM and its related health complications, including cardiovascular disease (CVD), chronic kidney disease (CKD), and congestive heart failure (CHF), are associated with significant morbidity and mortality. People with DM accounted for 7.8 million hospital discharges in 2016 with 1.7

Abbreviations: ACEi, angiotensin-converting enzyme inhibitor; AGE, advanced glycation end product; ARB, angiotensin II receptor blocker; BH4, tetrahydrobiopterin; BMI, body mass index; CHF, congestive heart failure; CKD, chronic kidney disease; CV, cardiovascular; CVD, cardiovascular disease; CRF, cardiorespiratory fitness; DM, diabetes mellitus; eNOS, endothelial nitric oxide synthase; GLP-1, glucagon-like peptide 1 receptor agonists; NO, nitric oxide; NOS, nitric oxide synthase; RAAS, renin-angiotensin activating system; T2D, type 2 diabetes; VO2max, maximal oxygen consumption

Received 23 December 2019

Accepted 4 June 2020

First Published Online 7 June 2020

Corrected and Typeset 3 July 2020

July 2020 | Vol. 4, Iss. 7

doi: 10.1210/endo/bvaa063 | Journal of the Endocrine Society | 1–15

Table 1. Mean cardiorespiratory fitness (CRF) before graded exercise training in women with and without type 2 diabetes (T2D).

	Lean participants (control)	Overweight participants (control)	T2D participants
VO _{2max} (mL/kg/min)	25.1 ±4.7	21.8 ±2.9	17.7 ±4.0 ^a

Values are means ±SD.

^a*P* < .05 for difference between the T2D group and the other 2 groups.

Adapted from Brandenburg SL, Reusch JE, Bauer TA, Jeffers BW, Hiatt WR, Regensteiner JG. Effects of exercise training on oxygen uptake kinetic responses in women with type 2 diabetes. *Diabetes Care*. 1999;22(10):1640–1646 [17].

million of these hospitalizations due to major CVD events [1]. Additionally, premature mortality from CVD is 2- to 6-fold greater in people with DM than for people without diabetes [3]. Lower levels of cardiorespiratory fitness (CRF), measured as maximal oxygen consumption (VO_{2max}), are predictive of greater short-term mortality risk in adults and children with DM [4–6], as in the general population [7–9]. Lower CRF in people with DM has also been associated with future CVD events and as such is an important modifiable CV risk factor [10–12]. In a study by Seyoum et al., men and women with diabetes who developed CVD events within 5 years had lower baseline peak VO₂ than those who did not have CVD events during this follow-up period [10].

Compared with healthy individuals without diabetes, CRF is lower in adults and children with T2D and in children with type 1 diabetes, even in the absence of clinically apparent CVD in either group. These findings were observed in individuals with similar habitual physical activity levels, age, pubertal stage (for youth), and body mass index (BMI) (Table 1) [13–19]. Additionally, the presence of T2D confers a greater CRF deficit in women than in men [15, 20]. Not only does lower CRF predict premature mortality, it may lead to barriers to exercise recommendations by raising the relative intensity of a given work rate [21]. Potential physiological mechanisms that may contribute to lower CRF in people with T2D include insulin resistance and mitochondrial, vascular, and cardiac dysfunction. This review will focus on how each of these factors may contribute to CRF impairment in T2D and conclude with an assessment of the current state of knowledge about sex differences in CRF in people with T2D.

1. Search Methods

Articles included in this narrative review were compiled from original research articles, societal guidelines, and reviews from peer-reviewed journals included in the PubMed database as of 9 April 2020. Search terms included “exercise capacity” OR “cardiorespiratory fitness” PLUS “diabetes AND either “endothelial,” “mitochondria,” “blood flow,” “fibrinolysis,” OR “NOS.” We additionally searched references within publications relevant to the topic.

2. Insulin Action and CRF

Insulin resistance, a defining feature of T2D, correlates with decreased CRF. For example, we demonstrated that insulin resistance measured by the hyperinsulinemic euglycemic clamp strongly and independently correlated with peak VO₂ in adolescents with T2D [18, 19]. In a proof of concept study, we therefore tested the impact of the insulin sensitizer rosiglitazone on CRF in people with T2D. We observed a significant 7% increase in peak VO₂ with rosiglitazone alone compared with a placebo (Table 2) [22]. This finding was corroborated by Kadoglou et al., suggesting that targeting insulin action can improve functional status in people with T2D [23, 24]. Insulin action is associated with cardiac and skeletal muscle function and metabolic flexibility with exercise [25]. In addition, our current

Table 2. Mean cardiorespiratory fitness (CRF) before and after exercise when treated with thiazolidinedione (Rosiglitazone) vs placebo.

	Placebo	Rosiglitazone
VO _{2max} (mL/kg/min)		
Before	19.4 ± 5.2	19.8 ± 5.3
After	18.1 ± 5.3	21.2 ± 5.1 ^a

Values are means ±SD.

^a*P* < 0.05 difference within groups before and after treatment.

Adapted from Regensteiner JG, Bauer TA, Reusch JE. Rosiglitazone improves exercise capacity in individuals with type 2 diabetes. *Diabetes care*. 2005;28(12):2877–2883 [22]. Copyright 2005 by the American Diabetes Association.

work supports a relationship between insulin action and factors related to CRF: cardiac function and skeletal muscle microvascular perfusion [20, 26]. As a vasoactive hormone, insulin has been shown in both rodents and healthy human participants to increase skeletal and cardiac muscle perfusion [27–29]. This perfusion response is blunted in insulin-resistant states [30, 31]. Decreased microvascular blood flow has been shown during insulin infusions in both insulin-resistant animal models and human participants [32]. Moreover, recent studies indicate that decreased microvascular blood flow may limit muscle glucose uptake by limiting the delivery of glucose to the myocyte [33]. Glucose and oxygen are both perfusion limited in their delivery to muscle, suggesting that any underlying microvascular defects are likely to have similar effects on both oxygen and glucose [34]. Even in people with obesity and no family history of DM, insulin-mediated microvascular perfusion in heart and skeletal muscle is decreased [35]. This is important as the microvasculature provides endothelial surface area needed for tissue uptake of oxygen and substrates that are vital to tissue health and function [36]. Indeed, multiple studies have shown that improved skeletal muscle microvascular perfusion via insulin sensitizers/enhancers and agents that cause vasodilation, such as glucagon-like peptide 1 receptor agonists (GLP-1), is associated with improved muscle oxygenation regardless of insulin sensitivity, suggesting that targeting microvascular insulin resistance and overall skeletal muscle and heart perfusion may result in both improved muscle oxygenation and glucose delivery during exercise [22, 37–39]. Taken together, these data support a role for insulin action in the heart, skeletal muscle, and the microvasculature as contributors to CRF.

3. Insulin Resistance and Oxidative Capacity

Insulin resistance is correlated with mitochondrial dysfunction, but the cause and effect relationship is unclear—whether this is a cause or consequence of insulin resistance is a matter of active investigation. For example, skeletal muscle oxidative enzymes are lower in adults with T2D, while glycolytic enzymes are elevated, compared with adults without T2D, suggesting a relationship between mitochondrial dysregulation and insulin resistance [40]. Insulin-resistant adult offspring of people with T2D also have lower mitochondrial activity in vivo and lower expression of mitochondrial and mitochondrial biogenesis genes and proteins than insulin-sensitive adults matched for physical activity, height, weight, and age [41–44]. In a study on healthy participants, a lipid infusion resulted in lower glucose oxidation and muscle glycogen synthesis, as well as a lower glucose-6-phosphate and intracellular glucose concentration, suggesting less glucose transport [45]. Evidence also demonstrates that lipid infusion induces insulin resistance by altering skeletal muscle microvascular perfusion [28]. Lower oxidative capacity has been previously shown in skeletal muscle with decreased NADH:O₂ oxidoreductase activity (measure of respiratory chain activity, when normalized to citrate synthase and creatine kinase activity) in adults with T2D compared with both obese and lean adults [46]. Lower oxidative capacity has also been demonstrated in adolescents with T2D in vivo using in-MRI exercise. In particular, skeletal muscle adenosine diphosphate time constant, a blood flow-dependent mitochondrial

function measure, was slowed and oxidative phosphorylation rates lower in the adolescents with T2D, linking skeletal muscle blood flow and mitochondrial dysfunction in these young people. Moreover, lack of suppression free fatty acids and longer skeletal muscle adenosine diphosphate time constant were independently associated with insulin resistance [47]. Therefore, the interplay between insulin resistance in skeletal muscle and mitochondrial dysfunction may be an adaptation to a chronically hyperglycemic and/or hyperlipemic state and is likely mediated by both microvascular and skeletal muscle changes [25, 48].

Mitochondrial dysfunction is also observed in the myocardium of people with T2D [49]. In human adults with well-controlled, uncomplicated T2D, there is a lower ratio of cardiac phosphocreatine to adenosine triphosphate than in healthy controls. Furthermore, young people and adults with T2D have lower indices of diastolic function, possibly suggestive of early diastolic dysfunction [18, 50]. These findings are similar to those seen in people with clinical heart failure [49, 51, 52]. Peterson et al. found that participants with T2D had lower fractional glucose uptake and oxidation, glycolysis, and glycogen deposition in myocardial tissue [53]. Impaired myocardial mitochondrial function in DM may correlate with future mortality. For instance, in people with dilated cardiomyopathy, the myocardial phosphocreatine-to-adenosine triphosphate ratio has prognostic value predicting total and CV mortality [54]. Our understanding of the complex interaction between insulin resistance, cardiac mitochondrial dysfunction, and CRF is still evolving [55].

4. Glucose, Endothelium, and Vascular Regulation

Hyperglycemia also leads to vascular dysfunction (Fig. 1). The endothelium, or lining of the vasculature, is the first defense of the tissue against toxic metabolites and inflammatory cytokines and is also responsible for moderating oxygen and nutrient access to tissue; both roles are imperative for tissue health and function [28, 56]. The endothelium fine tunes vascular tone via its production of nitric oxide (NO) [57], which leads to calcium-mediated vasodilation and increased perfusion of vital structures [57]. Insulin's vasodilatory effect is NO mediated [58], and insulin increases NO production via Akt phosphorylation of endothelial nitric oxide synthase (eNOS) [59]. In T2D, insulin regulation of nitric oxide synthase (NOS) is impaired [59, 60].

In addition to insulin regulation of perfusion, DM impacts the macrovasculature and microvasculature in many ways. For example, elevation in glucose and fatty acids lead to inflammation and glucose-mediated production of advanced glycation end products (AGEs), activation of the renin–angiotensin activating system (RAAS) pathway, and aldosterone regulation [61]. Hyperglycemia leads to production of AGEs [62]. These accrue in the wall

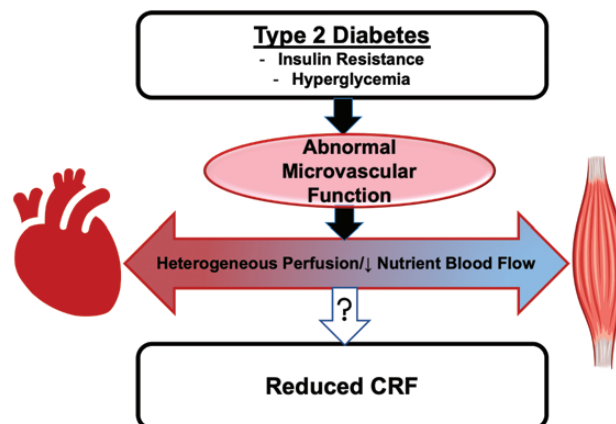


Figure 1. Insulin resistance and hyperglycemia of T2D lead to abnormal microvascular function and heterogeneous microvascular perfusion with lower nutrient blood flow, which may contribute to reduced CRF in people with T2D.

of vessels, leading to integral loss of the structure of the vessel wall and underlying basement membrane, as well as proinflammatory signaling contributing to systemic vascular dysfunction [63–65]. Additionally, in the Multiethnic Study of Artherosclerosis (MESA) study population, higher aldosterone levels were associated with higher fasting glucose, insulin resistance, and risk of incident DM, suggesting a role for RAAS activation in DM [66]. Previous studies have shown that RAAS modulation with angiotensin II receptor blockers (ARBs) and angiotensin-converting enzyme inhibitors (ACEi) lower production of AGEs [67, 68]. Olmesartan, an ARB, has been shown to inhibit AGE-related inflammation in endothelial cells [69]. In the Heart Outcomes Prevention Evaluation (HOPE) study, treatment of participants with DM with ACEi led to lower risk of CVD events, mortality, and nephropathy [70]. Furthermore, Parving et al. showed the renoprotective effect of irbesartan, an ARB, on participants with DM and hypertension [71]. In the IRMA 2 study, inflammatory markers and endothelial dysfunction predicted progression to diabetic nephropathy in T2D [72]; participants on irbesartan, an ARB, had significantly lower markers of inflammation (high sensitivity c-reactive protein, interleukin-6, and fibrinogen) [73]. These findings suggest a role of AGEs and RAAS activation in vascular dysfunction and progression of microvascular disease in diabetes, which are likely contributors to impaired CRF as well.

Vascular dysfunction is present prior to overt CVD and is even detectable in youth with T2D. Decreased limb blood flow correlated with reduced peak VO_2 in young people with T2D [18]. Additionally, arterial stiffness was seen in around half of the Treatment Options for Type 2 Diabetes in Adolescents and Youth (TODAY) cohort [74]. These data support a conceptual model wherein vascular insulin resistance, impaired eNOS regulation, and glucotoxicity- and lipotoxicity-mediated dysregulation of AGEs and vasoactive hormones could each contribute to decreased CRF in T2D.

5. The Role of NOS and NO in DM

Previous studies in healthy participants have shown that aerobic exercise attenuates mitochondrial damage and improves both vascular and skeletal muscle mitochondrial function, both of which are impaired in DM [75–77]. However, when rat models of DM and hypertension (Spontaneously Hypertensive Heart Failure [SHHF] obese) underwent an exercise intervention, there was no induction of vascular mitochondrial content as expected and as seen with control rats [78]. NOS/NO are known upstream regulators of mitochondrial biogenesis [79–81]. Animal models have demonstrated that NOS/NO are pivotal to an expression of vascular mitochondrial content and adaptive response to exercise (Table 3) [38, 39, 78, 82–86]. Lack of NOS expression in these animal studies led to decreased vascular mitochondrial content and impaired vascular mitochondrial response to exercise [84]. Therefore, eNOS has a role in mitochondrial adaptation to exercise, and dysregulation of eNOS in DM may explain the decreased response to exercise.

GLP-1 has been shown to stimulate NOS and cyclic AMP via G-protein-coupled receptor signaling, leading to increased glucose uptake in tissues and insulin-independent vasodilation [88]. In insulin-resistant rat models, treatment with liraglutide, a GLP-1 receptor agonist, rescued insulin-mediated increases in muscle perfusion and oxygenation [39]. Continuous GLP-1 infusion led to increased muscle microvascular blood perfusion, plasma NO, muscle insulin uptake, and muscle glucose extraction [38]. Furthermore, treatment with saxagliptin, a dipeptidyl peptidase 4 inhibitor that inhibits GLP-1 degradation, plus exercise training in an insulin-resistant rat model restored exercise-mediated vascular mitochondrial response and led to increased exercise capacity as measured by endurance [83]. Finally, rats treated with a GLP-1 receptor antagonist had decreased CRF, with and without exercise training. They also had attenuated vascular adaptation to exercise training [85]. However, in people with T2D, sitagliptin improved diastolic cardiac function, but not CRF, in the absence of formal exercise training [89]. Similarly, in people with T2D, treatment with exenatide, a GLP-1 agonist, improved diastolic heart function and arterial stiffness, but did not improve CRF in the absence of formal exercise training [90]. These data suggest

Table 3. Summary of animal studies investigating role of nitric oxide synthase (NOS) and nitric oxide (NO) in vascular mitochondrial adaptation to exercise, muscle perfusion and oxygenation, and muscle glucose extraction.

Animal study	Intervention	Vascular mitochondria				Systemic response	
		Content	Respiration	Adaptation	Endurance	Metabolism	
Goto-Kakizaki (GK) rat ^a [78]	High glucose	↓	↓	none	±↑	—	
eNOS +/- and +/- null mice [84]	None	↓	—	—	—	—	
Sprague Dawley rat ^b [84]	NOS-inhibitor/exercise	↓	—	↓	—	—	
GK rat ^a [83]	Saxagliptin/exercise	↑	—	↑	↑	↑	
Wistar rat ^c [85]	GLP-1 receptor antagonist/exercise	—	↓	↓	↓	↓	
Wistar rat ^c [87]	BH ₄ precursor	↑	—	↑	—	↑	
				Skeletal muscle			
				Mitochondria			
Sprague Dawley rat ^b [39]	High-fat diet/Liraglutide		↑	↑			
Sprague Dawley rat ^b [38]	Continuous GLP-1 infusion		↑	↑			
Streptozotocin (STZ)-induced mouse ^a [82]	BH ₄		↑	—			
eNOS and nNOS null mouse [86]	Acute and chronic exercise	↑	↑training response	—			

^aNonobese DM model.

^bMetabolic syndrome model.

^cControl model.

a role for GLP-1 in improving insulin-mediated defects in mitochondrial function and endothelial function, as well as in mediating adaptive effects of exercise training; to date, these benefits have only been clearly demonstrated in animal models (summarized in [Table 3](#)).

It has been reported that eNOS, as 1 of 3 NOS isoforms, generates NO only if a key cofactor, tetrahydrobiopterin (BH₄) is present [65]. NO originates from L-arginine catabolism by NOS. Reports have demonstrated that intravenous infusion of low-dose L-arginine improved insulin-mediated vasodilation in obese and T2D human participants and improved insulin sensitivity in all participants (healthy, obese, and T2D) [91]. Furthermore, our group treated participants with uncomplicated T2D with oral L-arginine for 7 days and demonstrated an increase in brachial artery diameter response and increased hyperemic forearm blood flow. There was no significant change in response noted in controls after 7 days of L-arginine [92]. In diabetes, oxidation of BH₄ to 7,8-dihydrobiopterin leads to eNOS uncoupling and dysfunction [93]. In a metabolic syndrome mouse model, BH₄ administration lowered glucose by acting as an insulin sensitizer and attenuating eNOS dysfunction [82]. Therefore, both defective insulin signaling to NOS and NOS uncoupling in diabetes can lead to abnormal vasomotion and vascular insulin resistance.

While NOS certainly has a role in vascular adaptation to exercise, it is not fully responsible for the lack of adaptive response to exercise training in animal models. eNOS and nNOS null mice subjected to acute (60 minutes) and chronic (9 days of 60 minutes) exercise training had increases in mitochondrial biogenesis markers during both short-term and long-term exercise training, demonstrating that while eNOS and nNOS are involved in mitochondrial adaptation, there are additional pathways necessary for mitochondrial response in the skeletal muscle to exercise training in nondiabetic models [86]. Sjøberg et al. have shown that these muscular mitochondrial adaptations to exercise are dependent on increased microvascular perfusion and molecular signaling, both induced by insulin, suggesting a role for blood flow-dependent insulin delivery and intact molecular signaling in the endothelium by insulin in exercise adaptive response and consequently decreased CRF in T2D [94].

6. Blood Flow and Muscle Oxygenation in T2D

Adequate perfusion of skeletal and cardiac muscle is necessary for appropriate function during a bout of exercise. Impaired blood flow distribution with exercise in T2D appears to contribute to decreased oxidative capacity by limiting skeletal muscle oxygen for oxidative flux. In a study by Bauer et al. [95], there was a rise in muscle deoxygenation (deoxygenated hemoglobin/myoglobin) that exceeded the rate of oxygen extraction after a bout of exercise in participants with T2D, suggesting a mismatch of supply to demand. There was also a delayed increase in blood flow after onset of exercise in these participants, consistent with perfusion limiting the exercise response [95]. Muscle deoxygenation during exercise in people with T2D was further studied by assessing maximal reactive hyperemic blood flow, peak oxygen utilization during exercise, and assessment of skeletal muscle oxygenation and its relation to blood volume and hematocrit. In control participants, muscle deoxyhemoglobin accumulation correlated inversely with peak VO₂. In contrast, in people with uncomplicated T2D, this correlation was not present; we postulated that this difference in T2D was due to heterogeneous muscle blood flow distribution. In this study, participants with T2D had similar total limb blood flow and tissue hemoglobin content after a bout of exercise as those without T2D, suggesting that the oxygen extraction impairment is independent of limb total blood flow [26].

The role of heterogeneous microvascular perfusion in decreased skeletal muscle oxygen extraction in T2D was first tested in a metabolic syndrome animal model. We demonstrated that heterogeneous blood flow could predict oxygen extraction and that oxygen extraction could be restored with acute antioxidant treatment in obese Zucker rats [34, 96]. The observation that this defect could be corrected led to our next series of clinical studies. In sedentary obese participants with and without T2D, we examined *in vivo* oxidative flux using

³¹P-magnetic resonance spectroscopy. As with previous reports, we observed lower oxidative flux in people with T2D. We tested whether the difference in oxidative flux was due to lower oxygen availability using supplemental oxygen and observed that the supplemental oxygen improved oxidative flux in people with T2D, but not in control participants [97]. These data support the overall working model that perfusion heterogeneity and local muscle hypoxia contribute to in vivo impaired mitochondrial function in people with T2D.

7. Potential Mediators of Heterogeneous and Lower Nutrient Blood Flow in T2D

Heterogeneous blood flow in T2D may be in part due to degradation of the endothelial glycocalyx (Table 4), a semipermeable layer of glycoproteins and proteoglycans at the blood vessel luminal surface [104]. People with DM have been shown to have a decrease in glycocalyx in both acute and chronic exposure of their vessels to hyperglycemia. When there is loss of the glycocalyx, protective enzymes on the surface of the endothelium are lost, leaving endothelial cells vulnerable to oxidative stress and inflammation [98].

DM is also associated with premature atherosclerosis [105] and a prothrombotic state. Increased thrombosis is in part due to augmented platelet activation and creation of compact fibrin networks that are lysis resistant [99]. In our earlier work, we reported that premenopausal women with T2D lose their fibrinolytic potential [100]. Inadequate dynamic resolution of microthrombi is a plausible contributor to heterogeneous blood distribution. Additionally, people with DM have more neutrophil extracellular traps formation. Neutrophil extracellular traps are composed of histones and DNA, which interact with fibrinogen to form thicker fibrin fibers leading to prolongation of clot lysis. This association leads to augmentation of thrombin generation, reduction in permeability of the clot, and consequent decreased fibrinolysis. These complexes are correlated with glycemic control and increased inflammatory state (elevated interleukin-6) [101]. Taken together, these abnormalities are plausible contributors to heterogeneous blood flow in DM.

Lower capillary density, sometimes termed capillary rarefaction, is also present in DM. Reports by Prior et al. have demonstrated increased skeletal muscle capillarization with exercise training in elderly participants and people with impaired glucose tolerance [102, 103]. In a rodent model, we examined the relative contribution of capillary density versus heterogeneous tissue perfusion to muscle oxygen extraction. Only 20% of the decreased muscle oxygen extraction observed in the Zucker rat model could be explained by the lower skeletal muscle capillary density [96]. There is currently not a robust human literature on capillary density changes in uncomplicated T2D so more research is required.

8. Cardiac Dysfunction

We have reported abnormal cardiac function in the setting of exercise in people with uncomplicated T2D. Specifically, women with uncomplicated, recently diagnosed T2D have a significantly and disproportionately greater rise in pulmonary capillary wedge pressure during exercise than in healthy control female participants [20, 106]. This finding suggests a stiff heart, as is also seen with diastolic dysfunction. The augmentation of pulmonary capillary wedge pressure in response to exercise correlated with poorer myocardial perfusion

Table 4. Potential mediators of heterogeneous and lower nutrient blood flow.

T2D effects on blood flow

Endothelial glycocalyx degradation [98]
 Hypofibrinolysis [99, 100]
 Neutrophil extracellular trap formation [101]
 Capillary Rarefaction [96, 102, 103]

across all regions of the heart [20]. Similarly, adolescents with T2D have greater left ventricular mass and abnormal cardiac circumferential strain, along with the finding of lower CRF than in lean and obese healthy control participants. Circumferential strain and CRF correlated with low adiponectin and fat mass, suggestive of obesity factoring into cardiac dysfunction and lower exercise capacity [19]. These findings of subclinical cardiac dysfunction with preserved cardiac output suggest that the noted decline in CRF in people with T2D may be independent of cardiac output and yet still have a cardiac contributor to its multifactorial etiology. Specifically, cardiac microvascular perfusion abnormalities contributing to cardiac stiffness may be implicated, but this question requires further evaluation [20].

9. Sex Differences in CVD Risk and CRF

There are similar absolute risks for CVD among men and women with DM. However, there is a much higher relative risk for CVD conferred by DM in women than in men with DM, despite a similar DM incidence [1, 107–110]. This difference in CV morbidity and mortality risk may be due to T2D dampening the cardioprotection thought to occur in premenopausal women [111–113]. It may also be due to a greater CVD risk factor burden in women with DM than in men with DM—conversely, it may also relate to a lower CVD risk factor burden in women without DM than in men without DM. The multifaceted underlying mechanisms that may account for the excess CVD risk seen in women with T2D are complex and are reviewed more comprehensively elsewhere [111, 114]. Furthermore, as described in the introduction, the presence of T2D confers a greater reduction in CRF in women than in men when compared with their nondiabetic counterparts [15, 20, 115]. Women of all ages have lower physical activity levels than their male counterparts [116, 117]. Women report more barriers to exercise, which may contribute to this difference in physical activity between the sexes [118, 119]. Women with T2D also perceive greater effort during exercise at the same work rate than women without T2D [21, 120]. To date, the physiological explanation for this sex difference is not well understood. Previous work demonstrated that female sex is associated with increased stiffness of the left ventricle during exercise among adults with T2D [121]. Additionally, in the HERITAGE study, women with T2D who underwent aerobic exercise training had a 3-fold smaller improvement in insulin sensitivity than men with T2D who completed the same intensity and frequency of exercise training [122]. It is unknown why these sex differences exist; further investigation is needed to assess underlying mechanisms that may contribute to lower CRF and higher relative risk of CVD in women with T2D.

10. Summary

DM prevalence continues to rise in the United States with associated increased CVD morbidity and mortality. People with T2D have reduced CRF compared with healthy participants, a factor that is associated with increased CV mortality. The mechanisms of lower CRF in T2D are multifaceted and involve interrelated defects in insulin action,

Table 5. Factors contributing to reduced cardiorespiratory fitness (CRF) in type 2 diabetes (T2D) and therapies that may improve these factors.

Factors	Potential interventions
Abnormal insulin action	Exercise [17, 123–125]
Lower oxidative capacity	Thiazolidinedione (rosiglitazone) [22, 23]
Impaired vascular regulation by nitric oxide synthase and nitric oxide	Angiotensin II receptor blockers [67–69, 71, 73]
Lower nutrient blood flow	Glucagon-like peptide 1 receptor agonists [36, 38, 39, 85, 88]
Heterogeneous microvascular perfusion	Dipeptidyl peptidase 4 inhibitor [83]
Cardiac dysfunction	L-arginine (intravenous [91] or oral [92])
Sex	

mitochondrial dysfunction, skeletal muscle microvasculature, and cardiac dysfunction. Various interventions have been studied to counteract these abnormalities; however, further human research is needed to evaluate the effect of these therapies on CRF in T2D (Table 5). Additionally, there is variable response to intervention [123], perhaps due to the multifaceted complexity of exercise training and its effect on the heart, vasculature, and skeletal muscle. Regular exercise typically improves CRF and insulin resistance in healthy controls. However, exercise can have inconsistent therapeutic effect in people with T2D compared with otherwise similar healthy participants. Additionally, there appears to be a greater deficit in CRF and associated physiological dysfunction in women with DM. These sex differences suggest that more research should be done on the role of sex as a moderator of the response to exercise therapy. Therefore, interventions must focus on tackling these mechanisms contributing to reduced CRF, as well as augmenting the beneficial effects of exercise in people with T2D.

Acknowledgments

The authors wish to acknowledge all our study participants and all of the students, fellows, and technicians within our laboratories.

Financial Support: J.E.B.R. reports funding from the U.S. Department of Veterans Affairs (BX002046, CX001532) and AstraZeneca. J.G.R. reports funding from Merck, the Office of Research on Women's Health (HD057022), and the American Diabetes Association.

Additional Information

Correspondence: Layla A. Abushamat, Department of Medicine, University of Colorado, Anschutz Medical Campus, 12801 East 17th Avenue, Mail Stop: 8106, Aurora, CO 80045. E-mail: layla.abushamat@cuanschutz.edu; or Jane E.B. Reusch, University of Colorado School of Medicine, Division of Endocrinology, Metabolism and Diabetes, 12801 East 17th Avenue, Mail Stop: 8106, Aurora, CO 80045. E-mail: jane.reusch@cuanschutz.edu.

Disclosure Summary: J.E.B.R. and J.G.R. disclose AstraZeneca IIT and Merck IIT.

Data Availability: Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

References

1. CDC. *National Diabetes Statistics Report*. Atlanta, GA: Centers for Disease Control and Prevention; 2020.
2. Xu G, Liu B, Sun Y, et al. Prevalence of diagnosed type 1 and type 2 diabetes among US adults in 2016 and 2017: population based study. *BMJ*. 2018;**362**:k1497.
3. Morrish NJ, Wang SL, Stevens LK, Fuller JH, Keen H. Mortality and causes of death in the WHO multinational study of vascular disease in diabetes. *Diabetologia*. 2001;**44**(Suppl (2)):S14-S21.
4. Church TS, Cheng YJ, Earnest CP, et al. Exercise capacity and body composition as predictors of mortality among men with diabetes. *Diabetes Care*. 2004;**27**(1):83-88.
5. Wei M, Gibbons LW, Kampert JB, Nichaman MZ, Blair SN. Low cardiorespiratory fitness and physical inactivity as predictors of mortality in men with type 2 diabetes. *Ann Intern Med*. 2000;**132**(8):605-611.
6. Lyerly GW, Sui X, Lavie CJ, Church TS, Hand GA, Blair SN. The association between cardiorespiratory fitness and risk of all-cause mortality among women with impaired fasting glucose or undiagnosed diabetes mellitus. *Mayo Clin Proc*. 2009;**84**(9):780-786.
7. Myers J, Prakash M, Froelicher V, Do D, Partington S, Atwood JE. Exercise capacity and mortality among men referred for exercise testing. *N Engl J Med*. 2002;**346**(11):793-801.
8. Shah RV, Murthy VL, Colangelo LA, et al. Association of fitness in young adulthood with survival and cardiovascular risk: the coronary artery risk development in young adults (CARDIA) study. *JAMA Intern Med*. 2016;**176**(1):87-95.

9. Farrell SW, Finley CE, Jackson AW, Vega GL, Morrow JR Jr. Association of multiple adiposity exposures and cardiorespiratory fitness with all-cause mortality in men: the Cooper Center Longitudinal Study. *Mayo Clin Proc.* 2014;**89**(6):772-780.
10. Seyoum B, Estacio RO, Berhanu P, Schrier RW. Exercise capacity is a predictor of cardiovascular events in patients with type 2 diabetes mellitus. *Diab Vasc Dis Res.* 2006;**3**(3):197-201.
11. Booth FW, Gordon SE, Carlson CJ, Hamilton MT. Waging war on modern chronic diseases: primary prevention through exercise biology. *J Appl Physiol (1985).* 2000;**88**(2):774-787.
12. Booth FW, Laye MJ, Roberts MD. Lifetime sedentary living accelerates some aspects of secondary aging. *J Appl Physiol (1985).* 2011;**111**(5):1497-1504.
13. Komatsu WR, Gabbay MA, Castro ML, et al. Aerobic exercise capacity in normal adolescents and those with type 1 diabetes mellitus. *Pediatr Diabetes.* 2005;**6**(3):145-149.
14. Nadeau KJ, Regensteiner JG, Bauer TA, et al. Insulin resistance in adolescents with type 1 diabetes and its relationship to cardiovascular function. *J Clin Endocrinol Metab.* 2010;**95**(2):513-521.
15. Regensteiner JG, Sippel J, McFarling ET, Wolfel EE, Hiatt WR. Effects of non-insulin-dependent diabetes on oxygen consumption during treadmill exercise. *Med Sci Sports Exerc.* 1995;**27**(5):661-667.
16. Regensteiner JG, Bauer TA, Reusch JE, et al. Abnormal oxygen uptake kinetic responses in women with type II diabetes mellitus. *J Appl Physiol (1985).* 1998;**85**(1):310-317.
17. Brandenburg SL, Reusch JE, Bauer TA, Jeffers BW, Hiatt WR, Regensteiner JG. Effects of exercise training on oxygen uptake kinetic responses in women with type 2 diabetes. *Diabetes Care.* 1999;**22**(10):1640-1646.
18. Nadeau KJ, Zeitler PS, Bauer TA, et al. Insulin resistance in adolescents with type 2 diabetes is associated with impaired exercise capacity. *J Clin Endocrinol Metab.* 2009;**94**(10):3687-3695.
19. Bjornstad P, Truong U, Dorosz JL, et al. Cardiopulmonary dysfunction and adiponectin in adolescents with type 2 diabetes. *J Am Heart Assoc.* 2016;**5**(3):e002804.
20. Regensteiner JG, Bauer TA, Reusch JE, et al. Cardiac dysfunction during exercise in uncomplicated type 2 diabetes. *Med Sci Sports Exerc.* 2009;**41**(5):977-984.
21. Huebschmann AG, Reis EN, Emsermann C, et al. Women with type 2 diabetes perceive harder effort during exercise than nondiabetic women. *Appl Physiol Nutr Metab.* 2009;**34**(5):851-857.
22. Regensteiner JG, Bauer TA, Reusch JE. Rosiglitazone improves exercise capacity in individuals with type 2 diabetes. *Diabetes Care.* 2005;**28**(12):2877-2883.
23. Kadoglou NP, Iliadis F, Angelopoulou N, Perrea D, Liapis CD, Alevizos M. Beneficial effects of rosiglitazone on novel cardiovascular risk factors in patients with type 2 diabetes mellitus. *Diabet Med.* 2008;**25**(3):333-340.
24. Kadoglou NP, Iliadis F, Sailer N, et al. Exercise training ameliorates the effects of rosiglitazone on traditional and novel cardiovascular risk factors in patients with type 2 diabetes mellitus. *Metabolism.* 2010;**59**(4):599-607.
25. Goodpaster BH, Sparks LM. Metabolic flexibility in health and disease. *Cell Metab.* 2017;**25**(5):1027-1036.
26. Mason McClatchey P, Bauer TA, Regensteiner JG, Schauer IE, Huebschmann AG, Reusch JEB. Dissociation of local and global skeletal muscle oxygen transport metrics in type 2 diabetes. *J Diabetes Complications.* 2017;**31**(8):1311-1317.
27. Barrett EJ, Wang H, Upchurch CT, Liu Z. Insulin regulates its own delivery to skeletal muscle by feed-forward actions on the vasculature. *Am J Physiol Endocrinol Metab.* 2011;**301**(2):E252-E263.
28. Liu Z, Liu J, Jahn LA, Fowler DE, Barrett EJ. Infusing lipid raises plasma free fatty acids and induces insulin resistance in muscle microvasculature. *J Clin Endocrinol Metab.* 2009;**94**(9):3543-3549.
29. Kusters YH, Barrett EJ. Muscle microvasculature's structural and functional specializations facilitate muscle metabolism. *Am J Physiol Endocrinol Metab.* 2016;**310**(6):E379-E387.
30. Baron AD, Tarshoby M, Hook G, et al. Interaction between insulin sensitivity and muscle perfusion on glucose uptake in human skeletal muscle: evidence for capillary recruitment. *Diabetes.* 2000;**49**(5):768-774.
31. Clerk LH, Vincent MA, Barrett EJ, Lankford MF, Lindner JR. Skeletal muscle capillary responses to insulin are abnormal in late-stage diabetes and are restored by angiotensin-converting enzyme inhibition. *Am J Physiol Endocrinol Metab.* 2007;**293**(6):E1804-E1809.
32. Keske MA, Premilovac D, Bradley EA, Dwyer RM, Richards SM, Rattigan S. Muscle microvascular blood flow responses in insulin resistance and ageing. *J Physiol.* 2016;**594**(8):2223-2231.
33. McClatchey PM, Williams IM, Xu Z, et al. Perfusion controls muscle glucose uptake by altering the rate of glucose dispersion in vivo. *Am J Physiol Endocrinol Metab.* 2019;**317**(6):E1022-E1036.

34. McClatchey PM, Frisbee JC, Reusch JEB. A conceptual framework for predicting and addressing the consequences of disease-related microvascular dysfunction. *Microcirculation*. 2017;**24**(6):e12359. doi:[10.1111/micc.12359](https://doi.org/10.1111/micc.12359)
35. Wang N, Tan AWK, Jahn LA, et al. Vasodilatory actions of glucagon-like peptide 1 are preserved in skeletal and cardiac muscle microvasculature but not in conduit artery in obese humans with vascular insulin resistance. *Diabetes Care*. 2020;**43**(3):634-642.
36. Wang N, Alvin T, Jahn LA, et al. Glucagon-like peptide 1's vasodilatory actions are preserved in skeletal and cardiac muscle microvasculature but not in conduit artery in obese humans with vascular insulin resistance [Published online ahead of December 30, 2019]. *Diabetes Care*. 2019. doi:[10.2337/dc19-1465](https://doi.org/10.2337/dc19-1465)
37. Chai W, Wang W, Liu J, et al. Angiotensin II type 1 and type 2 receptors regulate basal skeletal muscle microvascular volume and glucose use. *Hypertension*. 2010;**55**(2):523-530.
38. Chai W, Dong Z, Wang N, et al. Glucagon-like peptide 1 recruits microvasculature and increases glucose use in muscle via a nitric oxide-dependent mechanism. *Diabetes*. 2012;**61**(4):888-896.
39. Chai W, Fu Z, Aylor KW, Barrett EJ, Liu Z. Liraglutide prevents microvascular insulin resistance and preserves muscle capillary density in high-fat diet-fed rats. *Am J Physiol Endocrinol Metab*. 2016;**311**(3):E640-E648.
40. Simoneau JA, Kelley DE. Altered glycolytic and oxidative capacities of skeletal muscle contribute to insulin resistance in NIDDM. *J Appl Physiol (1985)*. 1997;**83**(1):166-171.
41. Petersen KF, Dufour S, Befroy D, Garcia R, Shulman GI. Impaired mitochondrial activity in the insulin-resistant offspring of patients with type 2 diabetes. *N Engl J Med*. 2004;**350**(7):664-671.
42. Patti ME. Gene expression in humans with diabetes and prediabetes: what have we learned about diabetes pathophysiology? *Curr Opin Clin Nutr Metab Care*. 2004;**7**(4):383-390.
43. Patti ME, Butte AJ, Crunkhorn S, et al. Coordinated reduction of genes of oxidative metabolism in humans with insulin resistance and diabetes: potential role of PGC1 and NRF1. *Proc Natl Acad Sci U S A*. 2003;**100**(14):8466-8471.
44. Mootha VK, Lindgren CM, Eriksson KF, et al. PGC-1alpha-responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nat Genet*. 2003;**34**(3):267-273.
45. Dresner A, Laurent D, Marcucci M, et al. Effects of free fatty acids on glucose transport and IRS-1-associated phosphatidylinositol 3-kinase activity. *J Clin Invest*. 1999;**103**(2):253-259.
46. Kelley DE, He J, Menshikova EV, Ritov VB. Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. *Diabetes*. 2002;**51**(10):2944-2950.
47. Cree-Green M, Gupta A, Coe GV, et al. Insulin resistance in type 2 diabetes youth relates to serum free fatty acids and muscle mitochondrial dysfunction. *J Diabetes Complications*. 2017;**31**(1):141-148.
48. Barrett EJ, Eggleston EM, Inyard AC, et al. The vascular actions of insulin control its delivery to muscle and regulate the rate-limiting step in skeletal muscle insulin action. *Diabetologia*. 2009;**52**(5):752-764.
49. Bugger H, Abel ED. Mitochondria in the diabetic heart. *Cardiovasc Res*. 2010;**88**(2):229-240.
50. Diamant M, Lamb HJ, Groeneveld Y, et al. Diastolic dysfunction is associated with altered myocardial metabolism in asymptomatic normotensive patients with well-controlled type 2 diabetes mellitus. *J Am Coll Cardiol*. 2003;**42**(2):328-335.
51. Casademont J, Miró O. Electron transport chain defects in heart failure. *Heart Fail Rev*. 2002;**7**(2):131-139.
52. Kadkhodayan A, Lin CH, Coggan AR, et al. Sex affects myocardial blood flow and fatty acid substrate metabolism in humans with nonischemic heart failure. *J Nucl Cardiol*. 2017;**24**(4):1226-1235.
53. Peterson LR, Herrero P, Coggan AR, et al. Type 2 diabetes, obesity, and sex difference affect the fate of glucose in the human heart. *Am J Physiol Heart Circ Physiol*. 2015;**308**(12):H1510-H1516.
54. Neubauer S, Horn M, Cramer M, et al. Myocardial phosphocreatine-to-ATP ratio is a predictor of mortality in patients with dilated cardiomyopathy. *Circulation*. 1997;**96**(7):2190-2196.
55. Vega RB, Konhilas JP, Kelly DP, Leinwand LA. Molecular mechanisms underlying cardiac adaptation to exercise. *Cell Metab*. 2017;**25**(5):1012-1026.
56. Wang H, Wang AX, Liu Z, Chai W, Barrett EJ. The trafficking/interaction of eNOS and caveolin-1 induced by insulin modulates endothelial nitric oxide production. *Mol Endocrinol*. 2009;**23**(10):1613-1623.
57. Davidson SM. Endothelial mitochondria and heart disease. *Cardiovasc Res*. 2010;**88**(1):58-66.
58. Scherrer U, Randin D, Vollenweider P, Vollenweider L, Nicod P. Nitric oxide release accounts for insulin's vascular effects in humans. *J Clin Invest*. 1994;**94**(6):2511-2515.
59. Montagnani M, Chen H, Barr VA, Quon MJ. Insulin-stimulated activation of eNOS is independent of Ca²⁺ but requires phosphorylation by Akt at Ser(1179). *J Biol Chem*. 2001;**276**(32):30392-30398.

60. Xing W, Li Y, Zhang H, et al. Improvement of vascular insulin sensitivity by downregulation of GRK2 mediates exercise-induced alleviation of hypertension in spontaneously hypertensive rats. *Am J Physiol Heart Circ Physiol*. 2013;**305**(8):H1111-H1119.
61. Lim HS, MacFadyen RJ, Lip GY. Diabetes mellitus, the renin-angiotensin-aldosterone system, and the heart. *Arch Intern Med*. 2004;**164**(16):1737-1748.
62. Vlassara H, Uribarri J. Advanced glycation end products (AGE) and diabetes: cause, effect, or both? *Curr Diab Rep*. 2014;**14**(1):453.
63. Yan SF, Ramasamy R, Schmidt AM. The RAGE axis: a fundamental mechanism signaling danger to the vulnerable vasculature. *Circ Res*. 2010;**106**(5):842-853.
64. Huebschmann AG, Regensteiner JG, Vlassara H, Reusch JE. Diabetes and advanced glycoxidation end products. *Diabetes Care*. 2006;**29**(6):1420-1432.
65. Avogaro A, Fadini GP, Gallo A, Pagnin E, de Kreutzenberg S. Endothelial dysfunction in type 2 diabetes mellitus. *Nutr Metab Cardiovasc Dis*. 2006;**16**(Suppl 1):S39-S45.
66. Joseph JJ, Echouffo Tcheugui JB, Effoe VS, Hsueh WA, Allison MA, Golden SH. Renin-angiotensin-aldosterone system, glucose metabolism and incident type 2 diabetes mellitus: MESA. *J Am Heart Assoc*. 2018;**7**(17):e009890.
67. Miyata T, van Ypersele de Strihou C, Ueda Y, et al. Angiotensin II receptor antagonists and angiotensin-converting enzyme inhibitors lower in vitro the formation of advanced glycation end products: biochemical mechanisms. *J Am Soc Nephrol*. 2002;**13**(10):2478-2487.
68. Matsui T, Nishino Y, Maeda S, Takeuchi M, Yamagishi S. Irbesartan inhibits advanced glycation end product (AGE)-induced up-regulation of vascular cell adhesion molecule-1 (VCAM-1) mRNA levels in glomerular endothelial cells. *Microvasc Res*. 2011;**81**(3):269-273.
69. Yamagishi S, Matsui T, Nakamura K, et al. Olmesartan blocks inflammatory reactions in endothelial cells evoked by advanced glycation end products by suppressing generation of reactive oxygen species. *Ophthalmic Res*. 2008;**40**(1):10-15.
70. Heart Outcomes Prevention Evaluation (HOPE) Study Investigators. Effects of ramipril on cardiovascular and microvascular outcomes in people with diabetes mellitus: results of the HOPE study and MICRO-HOPE substudy. *Lancet*. 2000;**355**(9200):253-259.
71. Parving HH, Lehnert H, Bröchner-Mortensen J, Gomis R, Andersen S, Arner P; Irbesartan in Patients with Type 2 Diabetes and Microalbuminuria Study Group. The effect of irbesartan on the development of diabetic nephropathy in patients with type 2 diabetes. *N Engl J Med*. 2001;**345**(12):870-878.
72. Persson F, Rossing P, Hovind P, et al. Endothelial dysfunction and inflammation predict development of diabetic nephropathy in the Irbesartan in Patients with Type 2 Diabetes and Microalbuminuria (IRMA 2) study. *Scand J Clin Lab Invest*. 2008;**68**(8):731-738.
73. Persson F, Rossing P, Hovind P, et al. Irbesartan treatment reduces biomarkers of inflammatory activity in patients with type 2 diabetes and microalbuminuria: an IRMA 2 substudy. *Diabetes*. 2006;**55**(12):3550-3555.
74. Shah AS, El Ghormli L, Gidding SS, et al. Prevalence of arterial stiffness in adolescents with type 2 diabetes in the TODAY cohort: relationships to glycemic control and other risk factors. *J Diabetes Complications*. 2018;**32**(8):740-745.
75. Starritt EC, Angus D, Hargreaves M. Effect of short-term training on mitochondrial ATP production rate in human skeletal muscle. *J Appl Physiol (1985)*. 1999;**86**(2):450-454.
76. Spina RJ, Chi MM, Hopkins MG, Nemeth PM, Lowry OH, Holloszy JO. Mitochondrial enzymes increase in muscle in response to 7-10 days of cycle exercise. *J Appl Physiol (1985)*. 1996;**80**(6):2250-2254.
77. Bishop DJ, Granata C, Eynon N. Can we optimise the exercise training prescription to maximise improvements in mitochondria function and content? *Biochim Biophys Acta*. 2014;**1840**(4):1266-1275.
78. Knaub LA, McCune S, Chicco AJ, et al. Impaired response to exercise intervention in the vasculature in metabolic syndrome. *Diab Vasc Dis Res*. 2013;**10**(3):222-238.
79. Nisoli E, Clementi E, Paolucci C, et al. Mitochondrial biogenesis in mammals: the role of endogenous nitric oxide. *Science*. 2003;**299**(5608):896-899.
80. Nisoli E, Carruba MO. Nitric oxide and mitochondrial biogenesis. *J Cell Sci*. 2006;**119**(Pt 14):2855-2862.
81. Nisoli E, Tonello C, Cardile A, et al. Calorie restriction promotes mitochondrial biogenesis by inducing the expression of eNOS. *Science*. 2005;**310**(5746):314-317.
82. Abudukadier A, Fujita Y, Obara A, et al. Tetrahydrobiopterin has a glucose-lowering effect by suppressing hepatic gluconeogenesis in an endothelial nitric oxide synthase-dependent manner in diabetic mice. *Diabetes*. 2013;**62**(9):3033-3043.
83. Keller AC, Knaub LA, Miller MW, Birdsey N, Klemm DJ, Reusch JE. Saxagliptin restores vascular mitochondrial exercise response in the Goto-Kakizaki rat. *J Cardiovasc Pharmacol*. 2015;**65**(2):137-147.

84. Miller MW, Knaub LA, Olivera-Fragoso LF, et al. Nitric oxide regulates vascular adaptive mitochondrial dynamics. *Am J Physiol Heart Circ Physiol*. 2013;**304**(12):H1624-H1633.
85. Scalzo RL, Knaub LA, Hull SE, et al. Glucagon-like peptide-1 receptor antagonism impairs basal exercise capacity and vascular adaptation to aerobic exercise training in rats. *Physiol Rep*. 2018;**6**(13):e13754.
86. Wadley GD, Choate J, McConell GK. NOS isoform-specific regulation of basal but not exercise-induced mitochondrial biogenesis in mouse skeletal muscle. *J Physiol*. 2007;**585**(Pt 1):253-262.
87. Keller AC, Knaub LA, Scalzo RL, et al. Sepiapterin improves vascular reactivity and insulin-stimulated glucose in Wistar rats. *Oxid Med Cell Longev*. 2018;**2018**:7363485.
88. Ban K, Noyan-Ashraf MH, Hoefler J, Bolz SS, Drucker DJ, Husain M. Cardioprotective and vasodilatory actions of glucagon-like peptide 1 receptor are mediated through both glucagon-like peptide 1 receptor-dependent and -independent pathways. *Circulation*. 2008;**117**(18):2340-2350.
89. Scalzo RL, Rafferty D, Schauer I, et al. Sitagliptin improves diastolic cardiac function but not cardiorespiratory fitness in adults with type 2 diabetes. *J Diabetes Complications*. 2019;**33**(8):561-566.
90. Scalzo RL, Moreau KL, Ozemek C, et al. Exenatide improves diastolic function and attenuates arterial stiffness but does not alter exercise capacity in individuals with type 2 diabetes. *J Diabetes Complications*. 2017;**31**(2):449-455.
91. Wascher TC, Graier WF, Dittrich P, et al. Effects of low-dose L-arginine on insulin-mediated vasodilatation and insulin sensitivity. *Eur J Clin Invest*. 1997;**27**(8):690-695.
92. Regensteiner JG, Popylisen S, Bauer TA, et al. Oral L-arginine and vitamins E and C improve endothelial function in women with type 2 diabetes. *Vasc Med*. 2003;**8**(3):169-175.
93. Crabtree MJ, Channon KM. Synthesis and recycling of tetrahydrobiopterin in endothelial function and vascular disease. *Nitric Oxide*. 2011;**25**(2):81-88.
94. Sjøberg KA, Frøsig C, Kjøbsted R, et al. Exercise increases human skeletal muscle insulin sensitivity via coordinated increases in microvascular perfusion and molecular signaling. *Diabetes*. 2017;**66**(6):1501-1510.
95. Bauer TA, Reusch JE, Levi M, Regensteiner JG. Skeletal muscle deoxygenation after the onset of moderate exercise suggests slowed microvascular blood flow kinetics in type 2 diabetes. *Diabetes Care*. 2007;**30**(11):2880-2885.
96. Mason McClatchey P, Wu F, Olfert IM, et al. Impaired tissue oxygenation in metabolic syndrome requires increased microvascular perfusion heterogeneity. *J Cardiovasc Transl Res*. 2017;**10**(1):69-81.
97. Cree-Green M, Scalzo RL, Harrall K, et al. Supplemental oxygen improves in vivo mitochondrial oxidative phosphorylation flux in sedentary obese adults with type 2 diabetes. *Diabetes*. 2018;**67**(7):1369-1379.
98. Reitsma S, Slaaf DW, Vink H, van Zandvoort MA, oude Egbrink MG. The endothelial glycocalyx: composition, functions, and visualization. *Pflugers Arch*. 2007;**454**(3):345-359.
99. Kearney K, Tomlinson D, Smith K, Ajjan R. Hypofibrinolysis in diabetes: a therapeutic target for the reduction of cardiovascular risk. *Cardiovasc Diabetol*. 2017;**16**(1):34.
100. Brandenburg SL, Reusch JE, Felder KK, et al. Impaired fibrinolysis in premenopausal women and age-matched men with Type 2 diabetes mellitus: a pilot study. *J Investig Med*. 2002;**50**(2):110-115.
101. Bryk AH, Prior SM, Plens K, et al. Predictors of neutrophil extracellular traps markers in type 2 diabetes mellitus: associations with a prothrombotic state and hypofibrinolysis. *Cardiovasc Diabetol*. 2019;**18**(1):49.
102. Prior SJ, Goldberg AP, Ortmeier HK, et al. Increased skeletal muscle capillarization independently enhances insulin sensitivity in older adults after exercise training and detraining. *Diabetes*. 2015;**64**(10):3386-3395.
103. Prior SJ, Blumenthal JB, Katzell LI, Goldberg AP, Ryan AS. Increased skeletal muscle capillarization after aerobic exercise training and weight loss improves insulin sensitivity in adults with IGT. *Diabetes Care*. 2014;**37**(5):1469-1475.
104. McClatchey PM, Schafer M, Hunter KS, Reusch JE. The endothelial glycocalyx promotes homogenous blood flow distribution within the microvasculature. *Am J Physiol Heart Circ Physiol*. 2016;**311**(1):H168-H176.
105. Stout R. Diabetes, atherosclerosis and aging. *Diabetes Care*. 1990;**13**:20-23.
106. Regensteiner JG, Bauer TA, Huebschmann AG, et al. Sex differences in the effects of type 2 diabetes on exercise performance. *Med Sci Sports Exerc*. 2015;**47**(1):58-65.
107. Kannel WB, McGee DL. Diabetes and cardiovascular disease. The Framingham study. *Jama*. 1979;**241**(19):2035-2038.
108. Regensteiner JG, Golden S, Huebschmann AG, et al.; American Heart Association Diabetes Committee of the Council on Lifestyle and Cardiometabolic Health, Council on Epidemiology and

- Prevention, Council on Functional Genomics and Translational Biology, and Council on Hypertension. Sex differences in the cardiovascular consequences of diabetes mellitus: a scientific statement from the American Heart Association. *Circulation*. 2015;**132**(25):2424-2447.
109. Prospective Studies Collaboration, Asia Pacific Cohort Studies Collaboration. Sex-specific relevance of diabetes to occlusive vascular and other mortality: a collaborative meta-analysis of individual data from 980 793 adults from 68 prospective studies. *Lancet Diabetes Endocrinol*. 2018;**6**(7):538-546.
 110. Kalyani RR, Lazo M, Ouyang P, et al. Sex differences in diabetes and risk of incident coronary artery disease in healthy young and middle-aged adults. *Diabetes Care*. 2014;**37**(3):830-838.
 111. Huebschmann AG, Huxley RR, Kohrt WM, Zeitler P, Regensteiner JG, Reusch JEB. Sex differences in the burden of type 2 diabetes and cardiovascular risk across the life course. *Diabetologia*. 2019;**62**(10):1761-1772.
 112. Kanaya AM, Grady D, Barrett-Connor E. Explaining the sex difference in coronary heart disease mortality among patients with type 2 diabetes mellitus: a meta-analysis. *Arch Intern Med*. 2002;**162**(15):1737-1745.
 113. Wenger NK. Coronary heart disease in women: highlights of the past 2 years—stepping stones, milestones and obstructing boulders. *Nat Clin Pract Cardiovasc Med*. 2006;**3**(4):194-202.
 114. Sattar N. Gender aspects in type 2 diabetes mellitus and cardiometabolic risk. *Best Pract Res Clin Endocrinol Metab*. 2013;**27**(4):501-507.
 115. Fang ZY, Sharman J, Prins JB, Marwick TH. Determinants of exercise capacity in patients with type 2 diabetes. *Diabetes Care*. 2005;**28**(7):1643-1648.
 116. Scholes S, Bann D. Education-related disparities in reported physical activity during leisure-time, active transportation, and work among US adults: repeated cross-sectional analysis from the National Health and Nutrition Examination Surveys, 2007 to 2016. *BMC Public Health*. 2018;**18**(1):926.
 117. Sisson SB, Camhi SM, Church TS, et al. Leisure time sedentary behavior, occupational/domestic physical activity, and metabolic syndrome in U.S. men and women. *Metab Syndr Relat Disord*. 2009;**7**(6):529-536.
 118. Barrett JE, Plotnikoff RC, Courneya KS, Raine KD. Physical activity and type 2 diabetes: exploring the role of gender and income. *Diabetes Educ*. 2007;**33**(1):128-143.
 119. Edwards ES, Sackett SC. Psychosocial variables related to why women are less active than men and related health implications. *Clin Med Insights Womens Health*. 2016;**9**(Suppl 1):47-56.
 120. Huebschmann AG, Kohrt WM, Herlache L, et al. Type 2 diabetes exaggerates exercise effort and impairs exercise performance in older women. *BMJ Open Diabetes Res Care*. 2015;**3**(1):e000124.
 121. Ha JW, Lee HC, Park S, et al. Gender-related difference in left ventricular diastolic elastance during exercise in patients with diabetes mellitus. *Circ J*. 2008;**72**(9):1443-1448.
 122. Boulé NG, Weisnagel SJ, Lakka TA, et al.; HERITAGE Family Study. Effects of exercise training on glucose homeostasis: the HERITAGE Family Study. *Diabetes Care*. 2005;**28**(1):108-114.
 123. Ross R, Goodpaster BH, Koch LG, et al. Precision exercise medicine: understanding exercise response variability. *Br J Sports Med*. 2019;**53**(18):1141-1153.
 124. Bouchard C, An P, Rice T, et al. Familial aggregation of VO₂(max) response to exercise training: results from the HERITAGE Family Study. *J Appl Physiol (1985)*. 1999;**87**(3):1003-1008.
 125. Pandey A, Johnson JL, Slentz CA, et al. Short-term changes in cardiorespiratory fitness in response to exercise training and the association with long-term cardiorespiratory fitness decline: the STRRIDE Reunion Study. *J Am Heart Assoc*. 2019;**8**(20):e012876.