

- depletion of murine CD19 CAR T cells permanently reverses B cell aplasia. *J. Clin. Invest.* **126**, 4262–4272.
8. Weber, E.W., Lynn, R.C., Sotillo, E., Lattin, J., Xu, P., and Mackall, C.L. (2019). Pharmacologic control of CAR-T cell function using dasatinib. *Blood Adv.* **3**, 711–717.
  9. Cho, J.H., Collins, J.J., and Wong, W.W. (2018). Universal Chimeric Antigen Receptors for Multiplexed and Logical Control of T Cell Responses. *Cell* **173**, 1426–1438.e11.
  10. Mata, M., Gerken, C., Nguyen, P., Krenciute, G., Spencer, D.M., and Gottschalk, S. (2017). Inducible Activation of MyD88 and CD40 in CAR T Cells Results in Controllable and Potent Antitumor Activity in Preclinical Solid Tumor Models. *Cancer Discov.* **7**, 1306–1319.
  11. Wu, C.-Y., Roybal, K.T., Puchner, E.M., Onuffer, J., and Lim, W.A. (2015). Remote control of therapeutic T cells through a small molecule-gated chimeric receptor. *Science* **350**, aab4077.
  12. Leung, W.-H., Gay, J., Martin, U., Garrett, T.E., Horton, H.M., Certo, M.T., Blazar, B.R., Morgan, R.A., Gregory, P.D., Jarjour, J., and Astrakhan, A. (2019). Sensitive and adaptable pharmacological control of CAR T cells through extracellular receptor dimerization. *JCI Insight* **5**, e124430.
  13. Fedorov, V.D., Themeli, M., and Sadelain, M. (2013). PD-1- and CTLA-4-based inhibitory chimeric antigen receptors (iCARs) divert off-target immunotherapy responses. *Sci. Transl. Med.* **5**, 215ra172.
  14. Mayer, I.A., and Arteaga, C.L. (2016). The PI3K/AKT Pathway as a Target for Cancer Treatment. *Annu. Rev. Med.* **67**, 11–28.
  15. Sakemura, R., Terakura, S., Watanabe, K., Julamane, J., Takagi, E., Miyao, K., Koyama, D., Goto, T., Hanajiri, R., Nishida, T., et al. (2016). A Tet-On Inducible system for controlling CD19-Chimeric antigen receptor expression upon drug administration. *Cancer Immunol. Res.* **4**, 658–668.

## VEGF-B Gene Therapy for the Heart: Proceed with Caution

Mauro Giacca<sup>1,2</sup> and Fabio A. Recchia<sup>3,4,5</sup>

<https://doi.org/10.1016/j.ymthe.2020.06.014>

An unfortunate circumstance in biology is that the vascular endothelial growth factor type B (VEGF-B) was identified only after its homolog family member VEGF-A had already been discovered.<sup>1</sup> Should the opposite have occurred, VEGF-B would likely not have been named as such. Indeed, VEGF-B is a poor growth factor for endothelial cells and, unlike VEGF-A, it is not angiogenic in the strict sense, while its only known main receptor, VEGFR-1, is expressed in endothelial cells as well as in a vast series of other cell types. Little was known about the non-angiogenic functions of selective VEGFR-1 ligands until a study in 2008 showed marked anti-apoptotic effects of VEGF-B, both *in vitro* and *in vivo*.<sup>2</sup> Subsequent studies confirmed cell-protective functions of this factor in different cell types.

In this issue of *Molecular Therapy*, Lähteenvu et al.<sup>3</sup> add a new piece to the puzzle of VEGF-B activities. By using an adenoviral

vector to overexpress VEGF-B<sub>186</sub>, they show that this factor induces sympathetic nerve sprouting in mouse and pig hearts via a VEGFR-1 signaling-independent mechanism. The integrity of cardiac innervation is very important for electrical stability, contractility, myocardial metabolism, and coronary function.<sup>4</sup> Therefore, the induction of cardiac re-innervation by the VEGF-B<sub>186</sub> transgene should theoretically enhance post-ischemia cardiac recovery. Nonetheless, the authors report an association between a likely disordered nerve sprouting<sup>5</sup> and increased risk of ventricular arrhythmias and sudden cardiac death in pigs, at 6 days after infarction, unlike any other VEGFs tested in their previous studies. The authors should be commended for their rigorous approach based on retrospective analysis of 334 pigs and statistics, more usually utilized in the clinical studies realm. Such careful evaluation revealed an intrinsic risk in cardiac VEGF-B<sub>186</sub> gene therapy, whereas pre-

clinical studies, strictly prospective and necessarily based on smaller sample sizes, usually tend to overemphasize beneficial effects of experimental therapies, paying little attention to untoward events.

Does the observation by Lähteenvu et al.<sup>3</sup> raise a red flag on the therapeutic utilization of VEGF-B gene transfer in the heart? Not necessarily. If the goal is angiogenesis, VEGF-A or VEGF-D are more established therapeutic factors. Despite the fact that VEGF-A binds with 10-fold greater affinity to VEGFR-1 than to its angiogenic receptor VEGFR-2, it does not appear to be arrhythmogenic. However, if the goal is rather the direct protection of cardiomyocytes and other cell types in the ischemic or non-

<sup>1</sup>King's College London, British Heart Foundation Centre of Research Excellence, School of Cardiovascular Medicine & Sciences, Faculty of Life Sciences and Medicine, London, UK; <sup>2</sup>University of Trieste, Department of Medical, Surgical and Health Sciences, Trieste, Italy; <sup>3</sup>Institute of Life Sciences, Scuola Superiore Sant'Anna, Pisa, Italy; <sup>4</sup>Fondazione G. Monasterio, Pisa, Italy; <sup>5</sup>Cardiovascular Research Institute, Lewis Katz School of Medicine at Temple University, Philadelphia, PA, USA

**Correspondence:** Mauro Giacca, King's College London, British Heart Foundation Centre of Research Excellence, School of Cardiovascular Medicine & Sciences, Faculty of Life Sciences and Medicine, London, UK.

E-mail: [mauro.giacca@kcl.ac.uk](mailto:mauro.giacca@kcl.ac.uk)

**Correspondence:** Fabio A. Recchia, Institute of Life Sciences, Scuola Superiore Sant'Anna, Pisa, Italy.

E-mail: [frecchia@santannapisa.it](mailto:frecchia@santannapisa.it)



## Commentary

ischemic failing heart, then strong evidence suggests that the 167 aa isoform of VEGF-B (VEGF-B<sub>167</sub>) might be an equally effective and safer candidate. VEGF-B<sub>167</sub> is the most abundant VEGF-B isoform and is expressed in brown fat, myocardium, and skeletal muscle.<sup>6</sup> Mice knocked out for this factor develop normally but exhibit a mild cardiac phenotype, with dysfunctional coronary vasculature, impaired recovery from cardiac ischemia, and, most notably, decreased heart size.<sup>7</sup> Our own work using adeno-associated virus (AAV)-mediated gene delivery of VEGF-A<sub>165</sub> and VEGF-B<sub>167</sub> in infarcted rat hearts revealed that both factors produced a marked improvement in cardiac function by promoting cardiac contractility, preserving viable cardiac tissue, and preventing remodeling of the left ventricle over time.<sup>8</sup> However, VEGF-B<sub>167</sub> was more effective in preserving cardiac mass, even in the absence of a significant induction of angiogenesis. Two subsequent studies by our group in a dog model of tachypacing-induced dilated cardiomyopathy showed that AAV-VEGF-B<sub>167</sub>, injected directly in myocardium or infused in coronary arteries, nearly halted cardiac structural and functional abnormalities and delayed the onset of decompensated heart failure.<sup>9,10</sup> Lähteenluoto et al.<sup>3</sup> rightly observe that tachypacing might mask possible ventricular arrhythmias. However, our ongoing clinical trial in dogs with severe spontaneous heart failure is not revealing unexpected increases in sudden death after VEGF-B<sub>167</sub> gene delivery to the heart and, on the contrary, some canine patients monitored for months have displayed a surprising prolongation of life expectancy (unpublished data).

Over the last several years, there has been considerable interest in understanding the role of VEGF-B in the heart. Despite not being angiogenic in the strict sense, transgenically overexpressed VEGF-B was shown to induce overgrowth of the epicardial coronary vessels and of their main branches.<sup>6</sup> In addition, this factor was reported to regulate metabolism by promoting endothelial uptake and transcytosis of fatty acids from the bloodstream into various tissues, including myocardium,<sup>11</sup> and also to deplete endothelial plasma membrane cholesterol, with

consequent reduction in glucose uptake.<sup>12</sup> Of interest, under normal conditions, VEGF-B appears to be highly expressed by cardiomyocytes themselves, from which it is secreted and binds to the surrounding extracellular matrix. Endothelium-secreted heparinase can mobilize this extracellular pool of VEGF-B, thus enabling it to exert pro-survival and protective activities.<sup>13</sup> This protective mechanism appears to be severely attenuated in the setting of uncontrolled diabetes mellitus, which could explain the development of diabetic cardiomyopathy.<sup>13</sup>

How VEGF-B exerts its beneficial function on cardiomyocytes remains an open matter. VEGFR-1 is expressed by cardiomyocytes in a functional form at levels that are lower than those of endothelial cells but still sufficient to exert a protective effect.<sup>7,8</sup> In addition, this factor was also shown to influence myocardium through the activation of endothelial cells in the coronary vasculature.<sup>14</sup>

What are the molecular mechanisms underlying possible differences between the cardiac effects of VEGF-B<sub>186</sub> versus VEGF-B<sub>167</sub>? They are both specific ligands of VEGFR-1 and, similar to VEGF-A<sub>165</sub> and PIGF, bind neuropilin receptor-1 (Nrp-1),<sup>15</sup> the receptor that seems to mediate the neurotrophic effect in the heart in the Lähteenluoto et al.<sup>3</sup> study. However, VEGF-B<sub>167</sub> and VEGF-B<sub>186</sub> bind Nrp-1 with different modalities involving, respectively, a heparin-binding domain and proteolytic processing. Importantly, VEGF-A<sub>165</sub> and VEGF-B<sub>167</sub> tend to remain stably attached to heparin sulfates, while VEGF-B<sub>186</sub> and PIGF are more freely soluble and diffusible, probably functioning also as paracrine factors.

To what extent these subtle molecular differences might explain the different arrhythmogenic potential of the two VEGF-B isoforms in the absence of VEGFR-1 binding remains a matter for interesting future investigation. This appears to be an important field of research, since the marked beneficial effect of VEGF-B on the heart continues to be very attractive for the development of clinically effective gene therapy for heart failure.

## ACKNOWLEDGMENTS

This work was supported by the European Research Council (ERC; Advanced Grant 787971 “CuRE”), the British Heart Foundation (BHF; Programme Grant RG/19/11/34633), the European Commission Horizon 2020 programme (grant 874764 “REANIMA”), and the NIH (grant HL129120).

## REFERENCES

- Olofsson, B., Pajusola, K., Kaipainen, A., von Euler, G., Joukov, V., Saksela, O., Orpana, A., Pettersson, R.F., Alitalo, K., and Eriksson, U. (1996). Vascular endothelial growth factor B, a novel growth factor for endothelial cells. *Proc. Natl. Acad. Sci. USA* 93, 2576–2581.
- Li, Y., Zhang, F., Nagai, N., Tang, Z., Zhang, S., Scotney, P., Lennartsson, J., Zhu, C., Qu, Y., Fang, C., et al. (2008). VEGF-B inhibits apoptosis via VEGFR-1-mediated suppression of the expression of BH3-only protein genes in mice and rats. *J. Clin. Invest.* 118, 913–923.
- Lähteenluoto, J., Häntinen, O.P., Kuivanen, A., Huusko, J., Paanalanen, J., Lähteenluoto, M., Nurro, J., Hedman, M., Hartikainen, J., Laham-Karam, N., et al. (2020). Susceptibility to Cardiac Arrhythmias and Sympathetic Nerve Growth in VEGF-B Overexpressing Myocardium. *Mol. Ther.* 28, this issue, 1731–1740.
- Jamali, H.K., Waqar, F., and Gerson, M.C. (2017). Cardiac autonomic innervation. *J. Nucl. Cardiol.* 24, 1558–1570.
- Fukuda, K., Kanazawa, H., Aizawa, Y., Ardell, J.L., and Shivkumar, K. (2015). Cardiac innervation and sudden cardiac death. *Circ. Res.* 116, 2005–2019.
- Bry, M., Kivelä, R., Holopainen, T., Anisimov, A., Tammela, T., Soronen, J., Silvola, J., Saraste, A., Jeltsch, M., Korpisalo, P., et al. (2010). Vascular endothelial growth factor-B acts as a coronary growth factor in transgenic rats without inducing angiogenesis, vascular leak, or inflammation. *Circulation* 122, 1725–1733.
- Bellomo, D., Headrick, J.P., Silins, G.U., Paterson, C.A., Thomas, P.S., Gartside, M., Mould, A., Cahill, M.M., Tonks, I.D., Grimmond, S.M., et al. (2000). Mice lacking the vascular endothelial growth factor-B gene (*Vegfb*) have smaller hearts, dysfunctional coronary vasculature, and impaired recovery from cardiac ischemia. *Circ. Res.* 86, E29–E35.
- Zentilin, L., Puligadda, U., Lionetti, V., Zucchigna, S., Collesi, C., Patarini, L., Ruozzi, G., Camporesi, S., Sinagra, G., Pepe, M., et al. (2010). Cardiomyocyte VEGFR-1 activation by VEGF-B induces compensatory hypertrophy and preserves cardiac function after myocardial infarction. *FASEB J.* 24, 1467–1478.
- Pepe, M., Mamdani, M., Zentilin, L., Csizsar, A., Qanad, K., Zucchigna, S., Ungvari, Z., Puligadda, U., Moimas, S., Xu, X., et al. (2010). Intramyocardial VEGF-B167 gene delivery delays the progression towards congestive failure in dogs with pacing-induced dilated cardiomyopathy. *Circ. Res.* 106, 1893–1903.

## Commentary

10. Woitek, F., Zentilin, L., Hoffman, N.E., Powers, J.C., Ottiger, I., Parikh, S., Kulczycki, A.M., Hurst, M., Ring, N., Wang, T., et al. (2015). Intracoronary Cytoprotective Gene Therapy: A Study of VEGF-B167 in a Pre-Clinical Animal Model of Dilated Cardiomyopathy. *J. Am. Coll. Cardiol.* **66**, 139–153.
11. Hagberg, C.E., Falkevall, A., Wang, X., Larsson, E., Huusko, J., Nilsson, I., van Meeteren, L.A., Samen, E., Lu, L., Vanwildeveldsch, M., et al. (2010). Vascular endothelial growth factor B controls endothelial fatty acid uptake. *Nature* **464**, 917–921.
12. Moessinger, C., Nilsson, I., Muhl, L., Zeitelhofer, M., Heller Sahlgren, B., Skogsberg, J., and Eriksson, U. (2020). VEGF-B signaling impairs endothelial glucose transcytosis by decreasing membrane cholesterol content. *EMBO Rep.* **2020**, e49343.
13. Lal, N., Chiu, A.P., Wang, F., Zhang, D., Jia, J., Wan, A., Vlodavsky, I., Hussein, B., and Rodrigues, B. (2017). Loss of VEGFB and its signaling in the diabetic heart is associated with increased cell death signaling. *Am. J. Physiol. Heart Circ. Physiol.* **312**, H1163–H1175.
14. Kivelä, R., Hemanthakumar, K.A., Vaparanta, K., Robciuc, M., Izumiya, Y., Kidoya, H., Takakura, N., Peng, X., Sawyer, D.B., Elenius, K., et al. (2019). Endothelial Cells Regulate Physiological Cardiomyocyte Growth via VEGFR2-Mediated Paracrine Signaling. *Circulation* **139**, 2570–2584.
15. Mäkinen, T., Olofsson, B., Karpanen, T., Hellman, U., Soker, S., Klagsbrun, M., Eriksson, U., and Alitalo, K. (1999). Differential binding of vascular endothelial growth factor B splice and proteolytic isoforms to neuropilin-1. *J. Biol. Chem.* **274**, 21217–21222.