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Associations between Arachidonic Acid Metabolism Gene Polymorphisms and Prostate Cancer Risk

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Abstract

BACKGROUND.—The arachidonic acid (AA) pathway is suspected to be involved in the development of various cancers, including prostate cancer. However, the role of single nucleotide polymorphisms (SNPs) of AA pathway genes remains unclear. The purpose of this case-control study was to evaluate the association between prostate cancer risk and 14 such SNPs in the *PTGS2*, *PTGES2*, *ALOX5*, *ALOX5AP*, and *LTA4H* genes.

METHODS.—Genotyping was conducted on 585 white prostate cancer cases and 585 healthy, age-matched controls. The best genetic model for each SNP was determined using Akaike's information criterion. Odds ratios for the association between each SNP and prostate cancer risk were calculated, both overall and stratified by obesity (BMI ≥ 30). Haplotype analysis was conducted for the *PTGES2* SNPs.

RESULTS.—*LTA4H*rs1978331 was inversely associated with prostate cancer risk overall (overdominant model OR=0.68, 95% CI: 0.51-0.91 for TC vs TT/CC). Among non-obese individuals, the GG genotype of *PTGES2*rs10987883 was associated with an increased risk for prostate cancer (recessive model OR=3.23, 95% CI: 1.27-8.23).

CONCLUSION.—Our results indicate that SNPs in certain AA metabolism genes may influence prostate cancer susceptibility. Furthermore, it is possible that obesity, which induces a chronic state of low-level inflammation in addition to several metabolic sequelae, may modify the impact of these SNPs. These findings should be confirmed in a larger study with power to detect differential effects by obesity. It would also be interesting to see if these effects differed by racial/ethnic group, especially since African-American men are at higher risk of prostate cancer.

Introduction

Over the course of the last 30 years, an enormous body of research on prostate cancer etiology has attempted to clarify the role of lipid metabolism, including that of the

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arachidonic acid (AA) pathway, in carcinogenesis (1-3). Arachidonic acid is an ω -6 polyunsaturated fatty acid that is obtained from dietary sources and stored as phospholipids in the bilayer of plasma membranes (4). The pathway regulating the release and metabolism of this essential fatty acid has been well-characterized, and some of the eicosanoid products of this pathway, such as prostaglandins, leukotrienes, and hydroxyeicosatetraenoic acids (HETEs), have been previously shown to be involved in cancer cell proliferation, cell motility, and angiogenesis (1,2,5-12). However, the impact of single-nucleotide polymorphisms (SNPs) of AA metabolism genes on prostate cancer risk is a newer area of research that merits further investigation.

The majority of recent studies that have examined SNPs of AA metabolism genes have only focused on a few selected genes, rather than taking a broader pathway-based approach (13-18). The results of these studies have been somewhat inconsistent, and the SNPs of several AA pathway genes have not yet been examined. The benefit of a pathway-based, exploratory analysis is that such an approach provides greater coverage of the genetic variation present in genes representing different parts of the pathway and thus, could direct future studies to the most influential candidate genes in the pathway. To date, a disproportionate number of studies on AA metabolism genes have examined SNPs in *PTGS2/COX2*, whereas analyses on SNPs in other genes of biosynthetic enzymes involved in the AA pathway, such as *ALOX5* and *PTGES2*, are uncommon in the literature.

Related to lipid metabolism, the effects of obesity on prostate cancer risk have been explored in several studies (19-23). While obesity appears to be associated with worse prostate cancer prognosis, some studies suggest that it may be inversely associated with overall prostate cancer risk (20,23). However, this hypothesis remains controversial, and few studies have addressed it directly. Nevertheless, it is clear that obesity and its related cascade of physiological effects (i.e., higher levels of lipids, adipokines, and other pro-inflammatory factors) may be relevant to the way in which SNPs in lipid metabolism genes influence prostate cancer risk.

The purpose of this pathway-based study was to examine the effects, both overall and stratified by obesity, of 14 SNPs in five AA metabolism pathway genes (*PTGS2*, *PTGES2*, *ALOX5*, *ALOX5AP*, and *LTA4H*) on prostate cancer risk in a population of 585 non-Hispanic white cases and 585 age-matched, non-Hispanic white controls.

Methods

Study Population.

DNA collected from prostate cancer cases and healthy control subjects was purchased from BioServe Biotechnologies, Ltd. (Beltsville, MD). BioServe's Global Biorepository contains DNA, RNA, and tissue samples from over 120,000 participants that are available to researchers. Tissue and blood samples of cases were obtained from patients recruited from hospitals and urology clinics. Controls were recruited from primary care and specialists' offices. We purchased DNA, collected in the United States, from 585 non-Hispanic white prostate cancer cases and 585 age-matched, white cancer-free controls. We also obtained self-reported information on demographics and other characteristics from the standard

questionnaires used by the biorepository. All subjects consented to have their specimens become part of BioServe's biorepository to be used for research.

Candidate Gene & SNP Selection.

Fourteen SNPs in five AA metabolism pathway genes, *PTGS2*, *PTGES2*, *ALOX5*, *ALOX5AP*, and *LTA4H*, were chosen for this analysis based on the previous literature (Table I). The gene products of these five candidate genes are representative of two different branches of the AA metabolism pathway (1,3,10). One branch of the pathway leads to the synthesis of prostaglandins and thromboxanes. *PTGS2* codes for cyclooxygenase-2 (COX2), also called prostaglandin-endoperoxide synthase 2, which is an inducible enzyme that has pro-inflammatory effects and is overexpressed in several cancers (11,24). COX2 catalyzes the conversion of prostaglandin G2 to prostaglandin H2; whereas prostaglandin E synthase-2, the product of the *PTGES2* gene, facilitates the next step in the pathway, the conversion of prostaglandin H2 to prostaglandin E2 (PGE2). Like COX2, PGE2 has also been shown to be dramatically overexpressed in malignant prostate tissue compared to benign surrounding tissue (2,9,25,26). As a result, COX2, PTGES2, PGE2, and related proteins that act further downstream have been considered to be possible therapeutic targets for cancer (9).

ALOX5, *ALOX5AP*, and *LTA4H* represent a different subpathway of AA metabolism. These genes encode arachidonate 5-lipoxygenase and arachidonate 5-lipoxygenase-activating protein (aka, FLAP), respectively, which are involved in synthesizing leukotrienes from arachidonic acid. Leukotrienes are important eicosanoid immunomodulators that are overexpressed in malignant cells of several cancers (8). Previous research has indicated that leukotrienes may have a role in increasing cell proliferation and decreasing apoptosis among cancer cells (8,27). *LTA4H* is the leukotriene A4 hydrolase gene. *LTA4H* acts downstream of *ALOX5* and converts leukotriene A4 into leukotriene B4, which is a chemotactic agent for neutrophils (28,29). Thus, these five genes were selected both on the basis of biological plausibility and the availability of previous research implying that polymorphisms in these genes may be relevant to prostate cancer susceptibility and development.

The 14 SNPs of the five candidate genes were chosen based on the following criteria: 1) commonality (>5% minor allele frequency in non-Hispanic whites); 2) potential biological significance (nonsynonymous SNPs or those shown to be associated with other cancers or inflammatory diseases); and 3) for the purpose of capturing more of the genetic variation within the candidate genes. More information about these SNPs is provided in Table I.

Genotyping.

DNA was originally extracted from peripheral blood lymphocytes using the QIAmp DNA Blood Mini Kit (Qiagen, Valencia, CA) according to the manufacturer's protocol and stored in the Global Biorepository. DNA quality was assessed by measuring the A260/A280 ratio on a spectrometer. Genotyping was done using the Sequenom MassARRAY iPLEX platform according to the manufacturer's instructions (<http://www.sequenom.com/sEq.genotyping.html>). The quality control analysis included the genotyping of internal positive control samples, the use of no template controls, and the use of replicates for 10%

of the samples. Genotyping plates were reviewed for results from positive / negative / DNA control wells that were organized in specific patterns to assist in the quality control process and to ensure correct plate orientations during processing and data review. Call rates ranged from 83-94% with only two being <90%.

Statistical Analysis.

For each SNP, deviations from Hardy-Weinberg equilibrium (HWE) were assessed among controls using the X^2 -test. Allele and genotype frequency distributions were compared between the cases and controls, and the best genetic model for each SNP was determined using Akaike's information criterion (AIC). Crude and age-adjusted odds ratios and 95% confidence intervals (CIs) for the association between each SNP and prostate cancer risk were calculated using logistic regression, both in the overall study population and stratified by obesity (BMI ≥ 30). As previously discussed, analyses were stratified by obesity because we hypothesized that the chronic low-level inflammation induced by obesity could potentially modify the effect of the polymorphisms on prostate cancer risk.

Pairwise linkage disequilibrium (LD) between the SNPs was examined, using LD coefficient r^2 and Lewontin's standardized coefficient D' . Haplo.stats (<http://www.mayo.edu/stagen>) was used to conduct haplotype analyses on the four *PTGES2* SNPs.

Results

Information on selected characteristics of prostate cancer cases and cancer-free controls is provided in Table II. Although controls were slightly younger and less likely to smoke, these differences were not statistically significant.

The genotype distributions of three (*ALOX5* rs2029253, *ALOX5* rs6593482, and *ALOX5AP* rs4076128) of the 14 SNPs of interest demonstrated deviations from HWE in the controls (Table I); these SNPs were excluded from further analysis. Logistic regression analyses of the remaining 11 SNPs revealed that two, *PTGES2* rs10987883 and *LTA4H* rs1978331, were associated with prostate cancer risk (Table III). An increased risk, of borderline statistical significance, was associated with the minor allele (G) of *PTGES2* rs10987883 among non-obese individuals, using a recessive model as determined by AIC (age-adjusted OR=2.53, 95% CI: 0.99-6.55 for GG vs. AA + AG). Among the overall study population, the heterozygous genotype (TC) of *LTA4H* rs1978331 was significantly protective against prostate cancer, compared to the homozygous genotypes (TT + CC) [age-adjusted OR=0.66, 95% CI: 0.50-0.86]. This inverse association remained present among both obese and non-obese subgroups. None of the other SNPs of interest were found to be significantly associated with prostate cancer risk in the overall study population or stratified by obesity.

Strong LD ($r^2 > 0.8$) was observed between the four SNPs in the *PTGES2* gene. Haplotype analyses were conducted on these SNPs, but no significant associations were found (Table IV). The global score test indicated that haplotype distributions were not significantly different between cases and controls ($p=0.12$).

Discussion

In this study, we conducted a pathway-based analysis to investigate the role of putative functional arachidonic acid metabolism gene polymorphisms in prostate cancer susceptibility. Our findings from the single-locus analyses indicate that one SNP from each of the two major branches of AA metabolism, the prostaglandin synthesis subpathway (*PTGES2* rs10987883) and the leukotriene synthesis subpathway (*LTA4H* rs1978331), may be relevant to prostate cancer risk. While the TC genotype of *LTA4H* rs1978331 was significantly protective against prostate cancer overall, the GG genotype of *PTGES2* rs10987883 was associated with an increased prostate cancer risk only among non-obese individuals. Interestingly, the divergent point estimates obtained from the stratified analyses imply that obesity, and its associated inflammatory and metabolic sequelae, may potentially be effect modifiers of the relationship between this *PTGES2* SNP and prostate cancer risk.

Previous studies examining the relationship between SNPs in AA metabolism genes and prostate cancer risk have largely focused on the genetic variation in the *PTGS2/COX2* gene, rather than taking a more inclusive pathway-based approach (13-16,18). Although these studies have provided some evidence that polymorphisms in the *PTGS2* gene may influence prostate cancer risk, their findings have been relatively inconsistent. For example, Cheng et al. reported that the heterozygous genotype of *PTGS2* rs2745557 (GA) was associated with a 36% lower odds of prostate cancer development in a hospital-based case-control study of 506 advanced prostate cancer cases and 506 healthy controls (GA vs. GG, OR=0.64, 95% CI: 0.49-0.84) (16). By contrast, an analysis conducted in a large, population-based Swedish case-control study of 1378 cases and 782 controls did not find an association between this SNP and localized or advanced disease (15). Like the latter study, we were also unable to detect an association between *PTGS2* rs2745557 and prostate cancer risk. Because our study is one of the first to take a pathway-based approach to identifying AA metabolism SNPs relevant to prostate cancer risk, we cannot corroborate the two significant associations reported here (*PTGES2* rs10987883 and *LTA4H* rs1978331) with the previous literature.

One reason why the previous literature on AA metabolism genes and prostate cancer has focused so heavily on *PTGS2* is the plethora of data suggesting that overexpression of COX2, which is considered to be the rate-limiting enzyme in prostaglandin synthesis, is involved in prostate carcinogenesis (11). However, several lines of evidence also support the potential importance of *PTGES2* and *LTA4H* expression in the carcinogenic process (2,9,25,26). The product of the *PTGES2* gene, prostaglandin E synthase-2, facilitates the conversion of prostaglandin H2 to PGE2, which is also significantly overexpressed in cancer tissues (2,25,26). Overproduction of PGE2 may play a role in immunosuppression, angiogenesis, and prostate cancer progression (2,9,25,26). In fact, it has been suggested that PGE2 should be a target for anti-angiogenic cancer therapy to provide an alternative to treatment with COX2 inhibitors, which have been associated with numerous side effects (9).

There is also strong biological plausibility behind a potential relationship between SNPs of the *LTA4H* gene and prostate cancer susceptibility. *LTA4H* codes for leukotriene A4 hydrolase, which facilitates the synthesis of leukotriene B4 (LB4) from leukotriene A4. LB4 is a strong chemotactic factor for mast cells and neutrophils and has been implicated in the

pathogenesis of several chronic inflammatory diseases, including asthma, rheumatoid arthritis, and inflammatory bowel disease (29,30). Leukotrienes, including LB4, are involved in a multitude of functions that are relevant to carcinogenesis, including increasing transcription of oncogenes, encouraging tumor cell proliferation, and improving tumor survival by decreasing apoptosis (8). It has, therefore, been suggested that disruption of leukotriene synthesis may prevent cancer growth (8,31). Here, we found that the TC genotype of *LTA4H* rs1978331 is associated with a significantly reduced risk of prostate cancer, compared to either homozygous genotype. Because the exact functional relevance of the heterozygous genotype of this intronic SNP is unknown, we cannot fully explain this seeming heterozygote advantage. However, given that the T allele of this SNP is associated with higher LB4 level (32,33), the TC genotype could be associated with somewhat lower LB4 levels than the TT genotype, but higher levels than the CC genotype, thus potentially reducing the tumorigenic activities of this leukotriene without completely diminishing its inflammatory effects. In fact, Tobin et al. arrived at a similar hypothesis with regard to the function of the heterozygous genotype of this SNP when they found that the TC genotype conferred protection against both tuberculosis and leprosy (33). They postulated that if the variant allele of this SNP resulted in lower LB4 expression levels, then the heterozygous genotype may confer an ideal balance of eicosanoids with pro- and anti-inflammatory effects during infection. Alternatively, we cannot discount the possibility that our finding may be either an artifact of our data or due to chance alone. Therefore, replication of this association in other larger study populations, as well as additional functional studies on this SNP, are warranted.

Perhaps the most interesting implication of our study is that obesity may be modifying the relationship between *PTGES2* rs10987883 and prostate cancer risk. We found that among the non-obese subgroup of our population, the GG genotype of *PTGES2* rs10987883 was associated with an increased prostate cancer risk, although this association was only of borderline statistical significance. Conversely, this genotype was associated with a non-significant protective effect among obese individuals. Obesity induces numerous metabolic, hormonal, and inflammatory consequences, including lower testosterone levels and increased secretion of certain cytokines (e.g. IL-12, IL-6, and VEGF), C-reactive protein, and other proinflammatory agents (20,22,23,34). Transcriptional activation of AA metabolism genes can be regulated by various cytokines. Thus, it is feasible that in the context of the chronic state of inflammation induced by obesity, the impact of this SNP on prostate cancer risk could differ, even by a mechanism as simple as a differential level of expression among obese individuals. Nevertheless, the vast inconsistencies in the results of studies that have attempted to clarify the relationship between obesity and prostate cancer point to the involvement of an extremely complicated network of physiological processes that cannot be easily discerned (20,21,23). We acknowledge that our stratified analysis on the effect of *PTGES2* rs10987883 was dependent on a very small number of individuals with the GG genotype and that the width of the confidence intervals reflects this fact. However, the possibility of an interaction between *PTGES2* rs10987883 and obesity in prostate cancer susceptibility poses an interesting hypothesis to explore in future research.

A limitation of our study is that body mass index was measured at study recruitment, which was after cancer diagnosis for the cases. However, the inability to assess exposures at the

etiologic time period of interest is a common problem in case-control studies, and the stratified analyses that we conducted were intended for the purposes of hypothesis generation. In addition, information on cancer stage and grade was unavailable for a substantial proportion of the cases (60.5% and 39.8%, respectively), precluding us from being able to examine associations specifically with advanced prostate cancer. Despite these limitations, our findings have provided some evidence that SNPs in the *PTGES2* and *LTA4H* genes may be relevant to prostate cancer etiology. It will be essential to validate these associations in future studies that include information on advanced disease and have greater statistical power for the detection of potential interactions. Furthermore, because our current study was restricted to non-Hispanic white participants, the effects of these SNPs should also be examined in African-American and Hispanic men, who tend to have higher risk and worse prognosis for prostate cancer, to see if these associations hold across racial and ethnic groups (35).

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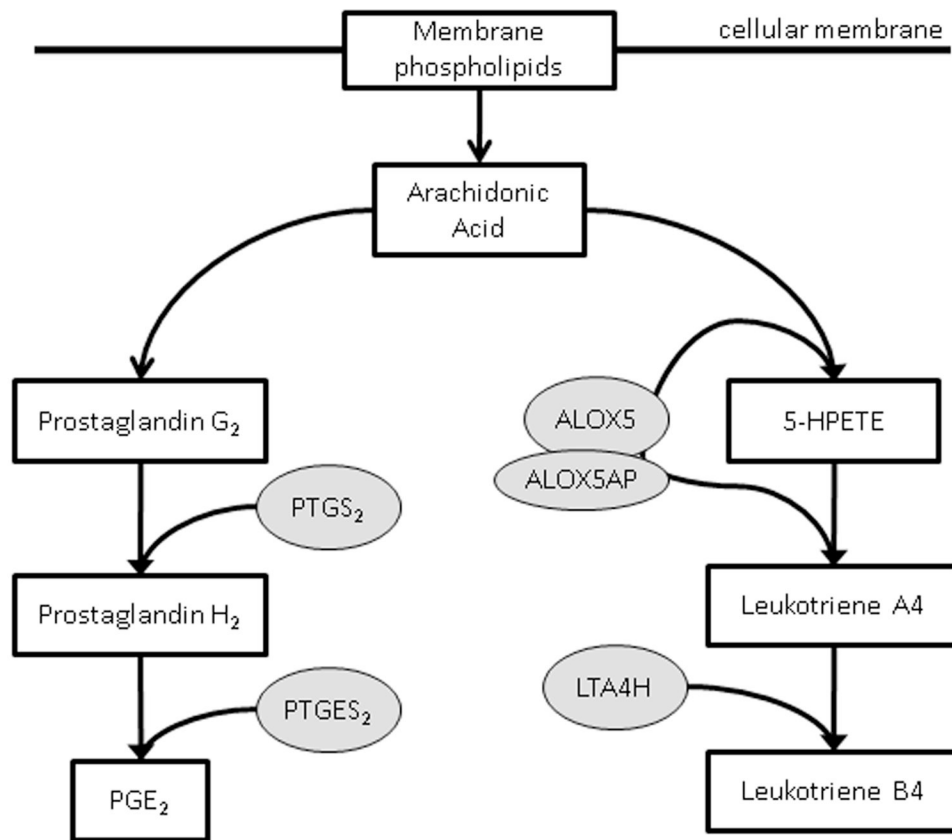


Figure 1. Simplified depiction of the arachidonic acid metabolism pathway, illustrating the roles of the protein products of genes of interest in the prostaglandin synthesis and leukotriene synthesis branches of the pathway.

Table 1.

Information about 14 genotyped SNPs in five arachidonic acid metabolism pathway genes

Genes	SNP ID	SNP region	Base change	P, HWE ^a	Details	Basis of selection		
						Reference ^b	Epidemiologic Associations	Reference
<i>Prostaglandin Synthesis Subpathway</i>								
<i>PTGS2/ COX2</i>	rs2745557	Intronic	G>A	0.40		(15)	Prostate cancer	(16)
<i>PTGES2</i>	rs884115	Intronic	C>T	0.72	Haplotype tagging SNP	(36)		
	rs10987883	Intronic	A>G	0.85	Haplotype tagging SNP	(36)		
	rs13283456	Intronic	C>T	0.25	Missense; Arg298His		Body mass index; Type 2 diabetes	(36,37)
	rs4837240	3' UTR	G>A	0.18	Haplotype tagging SNP	(36)		
<i>Leukotriene Synthesis Subpathway</i>								
<i>ALOX5</i>	rs2029253	Intronic	A>G	<.001			Acute coronary syndrome	(38)
	rs12762303	Promoter	T>C	0.41			Coronary artery disease	(39)
	rs2228065	Exon 6	G>T	1.0	Missense; Glu254Lys	(40)	Tuberculosis	(41)
	rs6593482	VNTR	G>T	<.001				
<i>ALOX5AP</i>	rs12721458	Intronic	C>G	0.84				
	rs4076128	5'UTR	A>G	<.001			Breast cancer	(10)
	rs4073259	5'UTR	G>A	0.13			Myocardial infarction	(42)
<i>LTA4H</i>	rs2660845	5'UTR	A>G	0.17			Asthma	(43)
	rs1978331	Intronic	T>C	0.65	Higher level of LB4 ^c associated with T allele	(33,42)	Asthma; leprosy; tuberculosis	(30,33)

^a P-values from Chi-squared test for Hardy-Weinberg equilibrium in controls. SNPs with significant p-values were excluded from further analysis.

^b Information from HapMap where not otherwise indicated.

^c LB4= leukotriene B4

Table II.

Characteristics of study population by case-control status

Characteristics	Cases (n=585) No. (%)	Controls (n=585) No. (%)	P-value
Mean age in years \pm SD	70.6 \pm 9.1	66.3 \pm 8.1	0.99
Smoking ^a			0.05
Never	221 (37.8)	254 (43.4)	
Ever	364 (62.2)	331 (56.6)	
Obesity ^b			0.63
BMI <30	448 (76.6)	455 (77.8)	
BMI \geq 30	137 (23.4)	130 (22.2)	
Gleason Score			
4	48 (8.3)		
5-7	468 (80.6)		
8	65 (11.2)		

^aIncludes cigarettes, cigars, and/or pipe use.^bObesity status at study recruitment. Cases and controls were matched on body mass index.

Genotype distributions and age-adjusted odds ratios for significant associations between arachidonic acid metabolism pathway SNPs and prostate cancer risk

Table III.

Arachidonic Acid Metabolism Subpathway	Gene & SNP ID	Model ^a	Genotype	No. of Cases/Controls	Overall Study Population (n= 585 cases; 585 controls)	Stratified by Obesity	
						Non-Obese OR (95% C.I.)	Obese OR (95% C.I.)
<i>Leukotriene Synthesis Subpathway</i>							
	<i>LTA4H</i> rs1978331	Overdominant	TT + CC	284/267	1.00	1.00	1.00
			TC	174/244	0.66 (0.50-0.86)	0.70 (0.52-0.95)	0.51 (0.29-0.90)
		Unrestricted	TT	183/175	1.00		
			TC	174/244	0.67 (0.50-0.90)		
			CC	101/193	1.05 (0.73-1.51)		
<i>Prostaglandin Synthesis Subpathway</i>							
	<i>PTGES2</i> rs10987883	Recessive	AA + AG	503/526	1.00	1.00	1.00
			GG	19/10	1.52 (0.69-3.35)	2.53 (0.99-6.55)	0.17 (0.02-1.57)
		Unrestricted	AA	384/404	1.00		
			AG	119/122	1.09 (0.81-1.46)		
			GG	19/10	1.55 (0.70-3.42)		

^aThe best genetic model for each SNP (bold), as determined by AIC, is provided along with the unrestricted model. Stratified analysis results are only provided for the best genetic models.

Table IV.Age-adjusted associations between *PTGES2* haplotypes and prostate cancer risk

Gene	Haplotype	Frequency ^a			OR (95% CI)	Global score test p-value ^b
		Total	Case	Control		
<i>PTGES2</i>						0.12
	C A C G	0.633	0.645	0.620	1.00	
	C A T G	0.187	0.185	0.188	1.05 (0.83-1.32)	
	T G C A	0.118	0.119	0.117	0.99 (0.76-1.30)	
	C A C A	0.032	0.033	0.032	1.11 (0.67-1.84)	

^aOnly haplotypes with frequencies>0.01 were examined; *PTGES2* rs884115, rs10987883, rs13283456, and rs4837240.

^bGenerated by permutation test (10,000x).

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