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Absence of the c.169+50delTAAACAG mutation of *SOD1* gene in a sample of keratoconus patients in Brazilian population

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Abstract

Objective: To determine the presence of the 7-bp deletion c.169+50delTAAACAG in intron 2 of *Superoxide Dismutase-1* gene in keratoconic patients from the State of São Paulo, Brazil, which promotes splicing variations, resulting in non-functional Superoxide Dismutase-1 antioxidant proteins, which may damage the corneal structure.

Results: A group of 35 keratoconic patients, from whom 35 peripheral blood samples and 58 samples of corneal fragments were evaluated, and a control group of 89 individuals, from whom 41 blood samples and 149 samples of corneal fragments were collected. After the amplification of DNA fragments by polymerase chain reaction, mutational screening analysis was performed by enzymatic digestion, followed by direct sequencing. The absence of the 7-bp c.169+50delTAAACAG mutation in intron 2 of Superoxide Dismutase-1 gene was detected in the analyzed subjects of the 2 groups, both in the cornea and peripheral blood samples. Then, according to our results, there is no involvement of c.169+50delTAAACAG deletion in the pathogenesis of keratoconus in this population, once it was not detected. But we emphasize that studies involving this deletion must be continued in an attempt to elucidate this issue

Keywords: Keratoconus, Cornea, Superoxide Dismutase-1 gene, Corneal Diseases, Polymorphism, Genetic

Introduction

Keratoconus (KC) is an asymmetrically bilateral corneal dystrophy characterized by progressive thinning of the cornea, which becomes cone-shaped in a process called ectasia, with a prevalence of approximately 1 in 2000 people [1–7]. It is currently known that KC is a multifactorial disease, involving genetic, environmental,

and behavioral factors and etiology remains poorly understood [3, 7–14].

In the literature, several studies define KC as a noninflammatory dystrophy; however, some recently published studies demonstrate an increase in inflammatory mediators (cytokines), such as interleukin-6, in tear samples of affected patients [15–18].

At first, the disease manifests asymptomatically; however, with its progression, visual acuity decreases, culminating in the clinical picture of progressive myopia and irregular astigmatism which may last until the fourth decade of life. Severe cases of KC require corneal transplantation [1, 3, 7, 12, 14, 19–21].

The genetic factors involved, the following are the candidate genes in the etiology of KC: *SOD1, VSX1, LOX, DOCK9,* among others. However, the role of these

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Lopes et al. BMC Res Notes (2020) 13:328 Page 2 of 7

genes in the disease is still considered complex and controversial [13, 22–25].

Genome-Wide Association Study (GWAS) and other isolated gene association studies have already identified a lot of polymorphisms in more than 60 keratoconusrelated loci. But these studies show that the existence of a relevant genetic alteration to the onset of the disease is still unknown, suggesting that further genetic research should be carried out [26–29].

The antioxidant enzyme superoxide dismutase 1 (SOD1) plays a crucial role in corneal physiology; it neutralized superoxide radicals formed owing to exposure to light, which can damage the corneal structure. Interestingly, this enzyme is differently distributed in keratoconic corneas in comparison with that in normal corneas [23, 24, 30, 31].

To date, the most relevant mutation found in *SOD1* and related to KC is a 7-bp deletion, c.169+50delTAAACA G, located in the intron 2 of *SOD1*, which may interfere with the splicing process, generating 2 transcript variants which result in non-functional SOD1 proteins [25, 32, 33].

It is also important to notice that the vast majority of studies on KC have only analyzed blood samples and not cornea, the latter being the target tissue of the disease. Then, to contribute to the genetic knowledge of this important and complex eye anomaly, in this study genetic screening of the intron 2 of *SOD1* was performed to detect the c.169+50delTAAACAG mutation in keratoconic patients from the northwestern region of São Paulo State, Brazil. The search for indexers was found to be unprecedented in the Brazilian population. Another relevant aspect was the analysis of 2 tissues, blood and cornea, in an attempt to detect if the mutation could be acquired.

Main text

Experimental population

The participants were divided into 2 groups: the group of keratoconic patients (35 patients: 35 blood samples and 58 corneal samples) and the control group

comprising healthy individuals (89 subjects: 41 blood samples and 149 corneal samples) (Table 1), and the corneal fragments and peripheral blood were analyzed in a paired manner.

The samples were collected from the Ophthalmology Outpatient Clinic Hospital de Base (HB-FUNFARME) and of the Visum Eye Center in São José do Rio Preto, SP and were obtained through keratorefractive procedures and corneal crosslinking.

the patients were subjected to clinical examination and topographic and tomographic evaluations. The clinical check comprised dynamic and static refractions, slit-lamp biomicroscopy, retinoscopy, tonometry, and fundus examination. The diagnosis of KC was based on axial placido disc topography with Easygraph/Topolyzer (Oculus-Wetzlar, Germany), with elevation derived from the anterior corneal curvature map, along with the tomographic examination with Pentacam HR (Oculus-Wetzlar, Germany), based on criteria used in the "Collaborative Longitudinal Evaluation of Keratoconus, CLEK" [34]. In the Pentacam, 3 examinations were performed with optimal fixation and quality, and average values for the following parameters were considered: axial curvature, anterior elevation, posterior elevation, thickness, and BAD III. Elevation data were taken from an 8-mm fixed zone using as reference the Best Fit Sphere manually adjusted.

The following factors were considered as inclusion criteria: absence of previous eye surgery; discontinued use of rigid contact lens (CL) for 4 weeks and gelatinous CL for 2 weeks; absence of previous eye trauma, and absence of primary nasal pterygium. Exclusion criteria were as follows: patients with keratoconjunctivitis sicca, acne rosacea, and severe dysfunction of the meibomian glands; patients using systemic or topical immunosuppressive drugs or those with autoimmune disease; chronic use of eye medication; vulnerable patient owing to physical or mental illness; corneal scar owing to KC; corneal hydrops; infectious keratitis; pregnancy, and lactation.

Table 1 Phenotypic characteristics of subjects included in mutational screening of SOD1 gene

Subjects	Gender	Age (mean/years)	SD	CL(% of individuals)	ER(% of individuals)
KC individuals ^a (35)	Male (16)	25.1	±6.4	25.7	62.8
	Female (19)	25.2	±5.3		
Control individuals (89)	Male (31)	36.6	±11.1	26.9	22.4
	Female (58)	37.8	±10.7		

^a KC individuals belonged to isolated cases

SD, standard deviation; CL, contact lens wear; ER, Act of eye rubbing

Lopes et al. BMC Res Notes (2020) 13:328 Page 3 of 7

DNA extraction, PCR-Random Fragment Length Polymorphism (RFLP) conditions and sequencing

Genomic DNA from peripheral blood (5 ml) was extracted with the QIAamp® DNA Blood QIAGEN kit (Hilden, Germany) according to the manufacturer's instructions. The genomic DNA from corneal epithelium samples was extracted using the standard phenol–chloroform extraction protocol.

The primer oligonucleotides which were used for PCR amplification of the fragment (approximately 218 bp) comprising part of the exon 2 and part of the intronic region of *SOD1* were as follows: F-CAGAAA CTCTCTCCAACTTGC and R-GAGGGGTTTACT GTTTAGGG [22].

PCR reaction mixture (25 μ l) containing 100 ng of genomic DNA, 12.5 μ l GoTaq® Green Master Promega (WI, USA), 7.5 μ l nuclease-free water Promega (WI, USA), and 0.2 pM of each primer (forward and reverse). The cycling conditions comprised an initial denaturation at 95 °C for 3 min, followed by 35 cycles at 95 °C for 30 s, annealing temperature at 59 °C for 30 s, 72 °C for 30 s, and a final extension at 72 °C for 10 min.

RFLP was performed with the restriction enzyme $HpyCH_4$ III, which has a restriction site in samples harboring the deletion.

After digestion, the fragments were applied to 2.5% agarose gel and stained with 8 μ l Sybr® Safe gel stain (Invitrogen-Thermo Scientific). The electrophoresis lasted 60 min at 100 V in TBE buffer (tris-borate-EDTA), and then the gels were photographed.

Subsequently, samples from the keratoconic group and random samples from the control group were sequenced to confirm the results. The nucleotide sequences were initially analyzed using the Sequence Scanner 2 software (Applied Biosystems) and aligned and edited with the BioEdit 7.0.5 software (Tom Hall, Ibis Therapeutics, Carlsbad, CA).

For reference analyses, the sequences were checked against the National Center for Biotechnology Information GenBank database. The reference sequence used is registered under the accession number NG_008689.1.

Results

PCR–RFLP of the fragment comprising part of the exon 2 and part of the intronic region of *SOD1* showed no restriction polymorphism in the samples analyzed.

Literature data confirm the presence of the c.169+50delTAAACAG deletion in the splicing region and is associated with KC [22, 24, 31, 33]. When present, this mutation promotes alternative splicing and generates 2 mRNA variant molecules, with the loss of exon 2 and/or exons 2 and 3, which results in structural damage and decreased function of the SOD1 protein. None of the samples from this study, either blood or cornea, had a restriction site for the enzyme HpyCH₄ III, which is capable of cleaving DNA at a specific site promoted by the 7 bp deletion, i.e., the presence of the restriction site would indicate the presence of the deletion, which was not found in our analyses (Fig. 1).

Direct genotyping and nucleotide sequence analyses of the samples following sequencing did not show any polymorphism and/or mutation in the studied fragment, which is consistent with the results obtained in the gel (Fig. 2).

Discussion

SOD1 comprises 9320 bp, encoding a 645-bp transcript comprising 5 exons, resulting in a protein composed of 154 amino acids [33].

Owing to its antioxidant property in corneal physiology, this gene appears to be one of the primary genes related to KC pathogenesis. Several studies of genetic screening found in the literature have investigated this gene as well as other such as VSX1, LOX, TIMP3, DOCK9, and

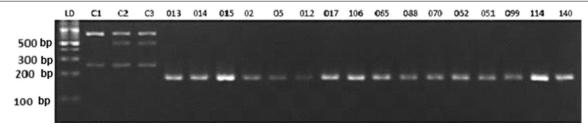


Fig. 1 Agarose gel electrophoresis (2.5%) of the fragment comprising part of the intron 2 region and part of the exon 2 region of *SOD1*. The bands indicate fragments of approximately 218 bp that were previously treated with the restriction enzyme HpyCH₄ III, and which showed no restriction polymorphisms. LD: DNA ladder (100 bp). Keratoconic blood samples (013, 014, 015, 02, 05, 012, 017, and 106) and cornea (065, 088, 070, 052, 051, 099, 114, and 140). C1, C2, and C3: positive reaction controls (ABO blood group gene fragment, which contains the restriction sites for the HpyCH₄ III enzyme)

Lopes et al. BMC Res Notes (2020) 13:328 Page 4 of 7

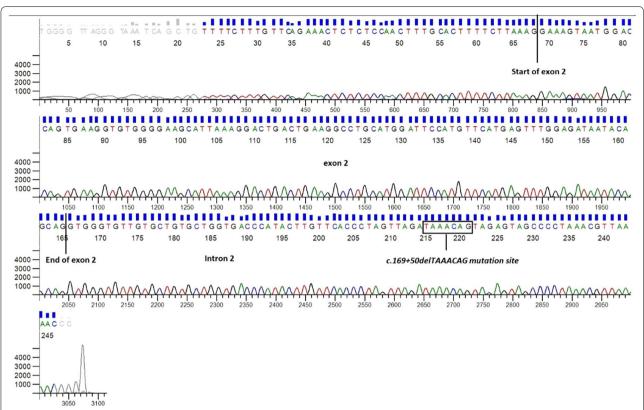


Fig. 2 Ectropherogram obtained by direct DNA sequencing of KC samples, showing the absence of the intronic 7-base deletion (c.169+50delTAA ACAG) in intron 2 of *SOD1* gene. The data reveal the exon 2 fragment and part of intron 2 in which we can observe the presence of the 7 nucleotides (TAAACAG) corresponding to the mutation site. The sequenced fragments exhibited 100% DNA identity to the *SOD1* gene reference sequence from GenBank database (accession number NG_008689.1)

collagen genes and identified genetic changes that may be related to the disease. Nevertheless, these results alone are not consistent enough to consider these genetic agents as disease-triggering [23, 24, 35–39].

The genetic variations found in these genes in different populations worldwide are predominantly single nucleotide polymorphisms, which in most cases have a low association with the disease and are considered controversial by several researchers [3, 8, 9, 11, 12, 24, 37, 40].

Specifically, concerning *SOD1*, the most relevant mutation associated with the disease is a intronic 7-bp deletion called c.169+50delTAAACAG, first described in 2006 by Udar et al. [33], through a genetic screening study involving 15 keratoconic patients, of whom 2 presented the mutation. According to the authors, this mutation occurs in the intron 2 of the gene which promotes alternative splicing, resulting in 2 variants of transcripts, which are mRNA molecules with the absence of exon 2 and/or exons 2 and 3, respectively. They generate non-functional SOD1 proteins owing to the loss of their active sites.

De Bonis et al. [22] performed a mutational screening of *SOD1* in 302 Italian keratoconic patients and also identified the presence of the mutation c.169+50delTAA ACAG in the intron 2 of the gene in 2 patients. The authors concluded that despite its low frequency (0.9%), this mutation needs to be investigated.

In this study, in a literature review considering some of the most relevant studies on this 7-base deletion published so far [22, 32, 33, 41], we reported a total of 13 KC patients who presented the mutation out in a total of 483 KC patients. Although this study also reported a low frequency of this mutation (2.6%), we emphasize the need for further research on the role of the *SOD1* gene in the KC etiology, due to its great importance in the corneal tissue physiology, as mentioned above.

Moschos et al. [32] also conducted a genetic study involving *SOD1* in a Greek population comprising 33 keratoconus cases and 78 control participants. After DNA sequencing analyses, they identified the presence of 7 bp deletion in 9 patients with sporadic KC. Besides, the mutation appeared in 4 control subjects. Thus, the

Lopes et al. BMC Res Notes (2020) 13:328 Page 5 of 7

authors emphasize that the role of this mutation in the KC pathogenesis should be further investigated.

In this literature review, we highlight 2 studies that investigated this deletion in keratoconic individuals from other populations: Al-Muammar et al. [42] and Nejabat et al. [41] investigated 55 Saudi patients and 20 patients from southern Iran, respectively. In contrast to the deletion-positive studies mentioned above, these authors did not detect the mutation.

In the present study involving blood and corneal samples from 35 keratoconic patients, where screening of the intron 2 of *SOD1* was performed, no intronic 7-bp deletion was identified, unlike in other populations. Nevertheless, we emphasize that our result is in agreement with some studies discussed above, which also have reported negative results.

We highlight that this study involving genetic screening of *SOD1* is a pioneer in a Brazilian population. It also stands out for studying rare corneal samples which were collected through surgical procedures.

Finally, we conclude that the genetic factors involved in KC pathogenesis, despite all efforts to date, are poorly defined and controversial. About the presence of the c.169+50delTAAACAG mutation in KC patients, despite the low frequency and weak association with the origin of the disease, it must be considered as, when present, it may trigger the disease or not, depending also on the association of other susceptibility factors, since KC is a multifactorial disease and that environmental and behavioral aspects are involved in its etiology.

Limitations

- Despite all efforts to date, the genetic factors involved in the pathogenesis of KC are poorly defined and controversial.
- Regarding the presence of the c.169+50delTAA
 ACAG mutation in keratoconic patients, it has an
 extremely low frequency and when present, may
 trigger the disease depending on the association
 with other susceptibility factors because KC is a
 multifactorial disease and the environmental and
 behavioral aspects are strongly related to its origin.
- There is no involvement of c.169+50delTAAACAG deletion in the pathogenesis of keratoconus in this population.

Abbreviations

KC: Keratoconus; CL: Contact lens; SOD1: Superoxide Dismutase-1; VSX1: Visual System Homeobox 1; LOX: Lysyl Oxidase; DOCK9: Dedicator of Cytokinesis 9; TIMP3: Tissue Inhibitor of Metalloproteinase 3.

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Authors' contributions

Conceived and designed the experiments: AGL, GCAJr, LC. Performed the experiments: AGL, RMT. Performed the inclusion of patients, sample collection, and developed the clinical evaluation and clinical analyses: GCAJr. Analyzed the data: AGL, LCM, CCBM and LC. Wrote the paper: AGL, GCAJr, LCM and LC. All authors read and approved the final manuscript.

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Availability of data and materials

This manuscript is part of the PhD Degree of Alessandro Garcia Lopes, as must be view in UNESP Thesis repository: https://repositorio.unesp.br/bitstream/handle/11449/153175/lopes_ag_dr_sjrp_int.pdf?isAllowed=y&sequence=6. The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request. The reference sequence of *SOD 1* gene used in this study is registered under the accession number NG_008689.1 in the National Center for Biotechnology Information GenBank database.

Ethics approval and consent to participate

This study was conducted according to the principles of the Declaration of Helsinki and was approved by the Research Ethics Committee of the São José do Rio Preto Medical School, FAMERP (CAAE 44071315.7.0000.5415). All patients were informed about the objectives of the study and signed a consent form.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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References

 Mas Tur V, MacGregor C, Jayaswal R, O'Brart D, Maycock N. A review of keratoconus: diagnosis, pathophysiology, and genetics. Surv Ophthalmol. 2017;62:770–83. https://doi.org/10.1016/j.survophthal.2017.06.009.

- Ben-Eli H, Erdinest N, Solomon A. Pathogenesis and complications of chronic eye rubbing in ocular allergy. Philadelphia: Lippincott Williams and Wilkins; 2019. https://doi.org/10.1097/aci.00000000000000571.
- Hashemi H, Heydarian S, Hooshmand E, Saatchi M, Yekta A, Aghamirsalim M, et al. The prevalence and risk factors for keratoconus: a systematic review and meta-analysis. Cornea. 2020;39:263–70. https://doi. org/10.1097/ICO.00000000000002150.
- Ghassembaglou N, Djalilian AR. Keratoconus; A true corneal disease. J Ophthalmic Vis Res. 2016;11:1–2.
- Masiwa LE, Moodley V. A review of corneal imaging methods for the early diagnosis of pre-clinical Keratoconus. J Optom. 2020. https://doi. org/10.1016/j.optom.2019.11.001.
- Mounir A, El Saman IS, Anbar M. The Correlation between Corneal Topographic Indices and Corneal High Order Aberrations in Keratoconus. Med hypothesis, Discov Innov Ophthalmol J. 2019;8:1–6. https://pubmed.ncbi. nlm.nih.gov/30923715/. Accessed 24 Feb 2020.
- Koh S, Ambrósio R, Inoue R, Maeda N, Miki A, Nishida K. Detection of subclinical corneal ectasia using corneal tomographic and biomechanical assessments in a Japanese population. J Refract Surg. 2019;35:383–90. https://doi.org/10.3928/1081597X-20190417-01.
- Loukovitis E, Sfakianakis K, Syrmakesi P, Tsotridou E, Orfanidou M, Bakaloudi DR, et al. Genetic aspects of keratoconus: a literature review exploring potential genetic contributions and possible genetic relationships with comorbidities. Ophthalmol Ther. 2018;7:263–92. https://doi.org/10.1007/ s40123-018-0144-8.
- Volatier TLA, Figueiredo FC, Connon CJ. Keratoconus at a molecular level: a review. Anat Rec. 2019. https://doi.org/10.1002/ar.24090.
- di Martino E, Ali M, Inglehearn CF. Matrix metalloproteinases in keratoconus—too much of a good thing? Exp Eye Res. 2019;182:137–43. https://doi. org/10.1016/j.exer.2019.03.016.
- Shinde V, Hu N, Renuse S, Mahale A, Pandey A, Eberhart C, et al. Mapping keratoconus molecular substrates by multiplexed high-resolution proteomics of unpooled corneas. Omi A J Integr Biol. 2019;23:583–97. https://doi. org/10.1089/omi.2019.0143.
- Barrientez B, Nicholas SE, Whelchel A, Sharif R, Hjortdal J, Karamichos D. Corneal injury: clinical and molecular aspects. Exp Eye Res. 2019;186:107709. https://doi.org/10.1016/j.exer.2019.107709.
- Bykhovskaya Y, Margines B, Rabinowitz YS. Genetics in keratoconus: where are we? Eye Vis. 2016;3:1–10. https://doi.org/10.1186/s40662-016-0047-5.
- Song P, Yang K, Li P, Liu Y, Liang D, Ren S, et al. Assessment of corneal pachymetry distribution and morphologic changes in subclinical keratoconus with normal biomechanics. Biomed Res Int. 2019. https://doi. org/10.1155/2019/1748579.
- Jun AS, Cope L, Speck C, Feng X, Lee S, Meng H, et al. Subnormal cytokine profile in the tear fluid of keratoconus patients. PloS one. 2011;6(1):e16437. https://doi.org/10.1371/journal.pone.0016437.
- Soiberman US, Shehata AEM, Lu MX, Young T, Daoud YJ, Chakravarti S, et al. Small molecule modulation of the integrated stress response governs the keratoconic phenotype in vitro. Invest Ophthalmol Vis Sci. 2019;60:3422–31. https://doi.org/10.1167/iovs.19-27151.
- Wisse RPL, Kuiper JJW, Gans R, Imhof S, Radstake TRDJ, Van Der Lelij A. Cytokine expression in keratoconus and its corneal microenvironment: a systematic review. Ocular Surf. 2015;13:272–83. https://doi.org/10.1016/j. jtos.2015.04.006.
- Sacchetti M, Bruscolini A, Lambiase A. Neurotrophic factors and nerve growth factor in ocular allergy. Curr Opin Allergy Clin Immunol. 2019;19:510–6. https://doi.org/10.1097/ACI.000000000000555.
- Lechner J, Porter LF, Rice A, Vitart V, Armstrong DJ, Schorderet DF, et al. Enrichment of pathogenic alleles in the brittle cornea gene, ZNF469, in keratoconus. Hum Mol Genet. 2014;23:5527–35. https://doi.org/10.1093/ hmg/ddu253.
- Mohammadpour M, Heidari Z, Hashemi H. Updates on managements for keratoconus. J Curr Ophthalmol. 2018;30:110–24. https://doi.org/10.1016/j. joco.2017.11.002.
- Loukovitis E, Kozeis N, Gatzioufas Z, Kozei A, Tsotridou E, Stoila M, et al. The
 proteins of keratoconus: a literature review exploring their contribution to
 the pathophysiology of the disease. Adv Ther. 2019;36:2205–22. https://doi.
 org/10.1007/s12325-019-01026-0.
- de Bonis P, Laborante A, Pizzicoli C, Stallone R, Barbano R, Longo C, et al. Mutational screening of VSX1, SPARC, SOD1, LOX, and TIMP3 in keratoconus. Mol Vis. 2011;17:2482–94.

- Valgaeren H, Koppen C, Van Camp G. A new perspective on the genetics of keratoconus: why have we not been more successful? Ophthalmic Genet. 2018;39:158–74. https://doi.org/10.1080/13816810.2017.1393831.
- Skorodumova LO, Belodedova AV, Sharova EI, Malyugin BE. Search for genetic markers for precise diagnostics of keratoconus. Biochem Suppl Ser B Biomed Chem. 2019;13:214–27. https://doi.org/10.1134/S1990750819030090.
- Al-Raddadi HS, Al-Barry MA, Al-Harbi E, Samman MI, Albalawi AM, Basit S. Sequence analysis of the VSX1 and SOD1 genes in families with Keratoconus and a review of the literature. J Taibah Univ Med Sci. 2016;11:115– 20. https://doi.org/10.1016/j.jtumed.2015.08.004.
- Li X, Bykhovskaya Y, Haritunians T, Siscovick D, Aldave A, Szczotka-Flynn L, et al. A genome-wide association study identifies a potential novel gene locus for keratoconus, one of the commonest causes for corneal transplantation in developed countries. Hum Mol Genet. 2012;21:421–9. https://doi. org/10.1093/hmg/ddr460.
- Lu Y, Vitart V, Burdon KP, Khor CC, Bykhovskaya Y, Mirshahi A, et al. Genomewide association analyses identify multiple loci associated with central corneal thickness and keratoconus. Nat Genet. 2013;45:155–63. https://doi. org/10.1038/ng.2506.
- Abu-Amero KK, Helwa I, Al-Muammar A, Strickland S, Hauser MA, Allingham RR, et al. Case-control association between CCT-associated variants and keratoconus in a Saudi Arabian population. J Negat Results Biomed. 2015;14:10. https://doi.org/10.1186/s12952-015-0029-5.
- Moussa S, Grabner G, Ruckhofer J, Dietrich M, Reitsamer H. Genetics in keratoconus—what is new? Open Ophthalmol J. 2017;11:201–10. https:// doi.org/10.2174/1874364101711010201.
- Saee-Rad S, Raoofian R, Mahbod M, Miraftab M, Mojarrad M, Asgari S, et al. Analysis of superoxide dismutase 1, dual-specificity phosphatase 1, and transforming growth factor, beta 1 genes expression in keratoconic and non-keratoconic corneas. Mol Vis. 2013;19:2501–7.
- Lucas SEM, Zhou T, Blackburn NB, Mills RA, Ellis J, Leo P, et al. Rare, potentially pathogenic variants in 21 keratoconus candidate genes are not enriched in cases in a large Australian cohort of European descent. PLoS One. 2018;13:e0199178. https://doi.org/10.1371/journal.pone.0199178.
- Moschos MM, Kokolakis N, Gazouli M, Chatziralli IP, Droutsas D, Anagnou NP, et al. Polymorphism analysis of VSX1 and SOD1 genes in greek patients with keratoconus. Ophthalmic Genet. 2015;36:213–7. https://doi.org/10.3109/13816810.2013.843712.
- Udar N, Atilano SR, Brown DJ, Holguin B, Small K, Nesburn AB, et al. SOD1: a candidate gene for keratoconus. Investig Ophthalmol Vis Sci. 2006;47:3345– 51. https://doi.org/10.1167/jovs.05-1500.
- Zadnik K, Barr JT, Edrington TB, Everett DF, Jameson M, McMahon TT, et al. Baseline findings in the collaborative longitudinal evaluation of keratoconus (CLEK) study. Investig Ophthalmol Vis Sci. 1998;39:2537–46.
- van Trier DC, van der Burgt I, Draaijer RW, Cruysberg JRM, Noordam C, Draaisma JM. Ocular findings in Noonan syndrome: a retrospective cohort study of 105 patients. Eur J Pediatr. 2018;177:1293–8. https://doi.org/10.1007/ s00431-018-3183-1.
- Ionescu C, Corbu CG, Tanase C, Jonescu-Cuypers C, Nicula C, Dascalescu D, et al. Inflammatory Biomarkers Profile as Microenvironmental Expression in Keratoconus. Dis Markers. 2016;2016:1243819. https://doi.org/10.1155/2016/1243819.
- Khawaja AP, Rojas Lopez KE, Hardcastle AJ, Hammond CJ, Liskova P, Davidson AE, et al. Genetic variants associated with corneal biomechanical properties and potentially conferring susceptibility to keratoconus in a genome-wide association study. JAMA Ophthalmol. 2019;137:1005–12. https://doi.org/10.1001/jamaophthalmol.2019.2058.
- Atilano S, Lee D, Fukuhara P, Chwa M, Nesburn A, Udar N, et al. Corneal oxidative damage in keratoconus cells due to decreased oxidant elimination from modified expression levels of SOD enzymes, PRDX6, SCARA3, CPSF3, and FOXM1. J Ophthalmic Vis Res. 2019;14:62–70. https://doi.org/10.4103/jovr.jovr. 80 18.
- Cao K, Sahebjada S, Richardson AJ, Baird PN. Do age-related macular degeneration genes show association with keratoconus? Eye Vis. 2019. https://doi.org/10.1186/s40662-019-0164-z.
- Wang YM, Kam KW, Chan TCY, Young AL, Jhanji V, Chen GLJ, et al. Keratoconus genes in chinese advances in vision research, vol. II. Singapore: Springer; 2019. p. 435–47. https://doi.org/10.1007/978-981-13-0884-0_35.
- 41. Nejabat M, Naghash P, Dastsooz H, Mohammadi S, Alipour M, Fardaei M. VSX1 and SOD1 mutation screening in patients with keratoconus in the

Lopes et al. BMC Res Notes (2020) 13:328 Page 7 of 7

- South of Iran. J Ophthalmic Vis Res. 2017;12:135–40. https://doi.org/10.4103/jovr.jovr_97_16.
- 42. Al-Muammar AM, Kalantan H, Azad TA, Sultan T, Abu-Amero KK. Analysis of the *SOD1* gene in keratoconus patients from Saudi Arabia. Ophthalmic Genet. 2015;36:373–5. https://doi.org/10.3109/13816810.2014.889173.

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