



Cell-Mediated Immune Responses to COVID-19 Infection

Amélie Guihot^{1,2}, Elena Litvinova², Brigitte Autran¹, Patrice Debré^{1,2} and Vincent Vieillard^{1*}

¹ Sorbonne Université, Inserm U1135, CNRS ERL 8255, Centre d'Immunologie et des Maladies Infectieuses (CIMI-Paris), Paris, France, ² Assistance Publique-Hôpitaux de Paris (AP-HP), Département d'Immunologie, Groupe Hospitalier Pitié-Salpêtrière, Paris, France

An unprecedented outbreak of pneumonia caused by a novel coronavirus (CoV), subsequently termed COVID-19 by the World Health Organization, emerged in Wuhan City (China) in December 2019. Despite rigorous containment and quarantine efforts, the incidence of COVID-19 continues to expand, causing explosive outbreaks in more than 160 countries with waves of morbidity and fatality, leading to significant public health problems. In the past 20 years, two additional epidemics caused by CoVs have occurred: severe acute respiratory syndrome-CoV, which has caused a large-scale epidemic in China and 24 other countries; and respiratory syndrome-CoV of the Middle East in Saudi Arabia, which continues to cause sporadic cases. All of these viruses affect the lower respiratory tract and manifest as pneumonia in humans, but the novel SARS-CoV-2 appears to be more contagious and has spread more rapidly worldwide. This mini-review focuses on the cellular immune response to COVID-19 in human subjects, compared to other clinically relevant coronaviruses to evaluate its role in the control of infection and pathogenesis and accelerate the development of a preventive vaccine or immune therapies.

Keywords: SARS - CoV-2, COVID-19, innate immunity, adaptive immunity, coronavirus

OPEN ACCESS

Edited by:

Denise L. Doolan,
James Cook University, Australia

Reviewed by:

Owen Kavanagh,
York St John University,
United Kingdom
Sofia A. Casares,
Naval Medical Research Center,
United States

*Correspondence:

Vincent Vieillard
vincent.vieillard@upmc.fr

Specialty section:

This article was submitted to
Vaccines and Molecular Therapeutics,
a section of the journal
Frontiers in Immunology

Received: 16 May 2020

Accepted: 22 June 2020

Published: 03 July 2020

Citation:

Guihot A, Litvinova E, Autran B,
Debré P and Vieillard V (2020)
Cell-Mediated Immune Responses to
COVID-19 Infection.
Front. Immunol. 11:1662.
doi: 10.3389/fimmu.2020.01662

INTRODUCTION

On December 31, 2019, a cluster of atypical pneumonia was reported in the Chinese city of Wuhan, mediated by a novel coronavirus (CoV) called SARS-CoV-2 (1, 2). The outbreak of this “coronavirus disease 2019” (COVID-19) has been declared a global pandemic by the World Health Organization (WHO), with more than 7 million cases in early June 2020 (3, 4) with a case-fatality rate of about 1%, as well as significant economic and social consequences. To date, no approved antiviral agents or efficient vaccines are available against the SARS-COV-2. For these reasons, necessary public health measures have been deployed, including worldwide quarantining of the populations and the use of barrier gestures to stop the progression of the SARS-COV-2.

CoVs are a class of positive-sense single-stranded RNA viruses found in a wide range of host species, including birds and mammals. Many of beta-CoV cause intestinal and respiratory infections in animals and humans. The zoonotic source of COVID-19 is not confirmed; however, sequencing of the SARS-CoV-2 reveals up to 80% identity with SARS-CoV and even more with several bat CoVs (5). This similarity suggests that bats could be the key reservoir, from which the virus was possibly directly transmitted to humans or through another unknown intermediate host.

A phylogenetic analysis of 160 genomes of patients with COVID-19 revealed three major variants, named A, B, and C; the A-type being the ancestral type, firstly detected in China. The A and C types are found in significant proportions in Europe and America, whereas the B type is the most common in East Asia (6).

In 2002–2003, a first “atypical pneumonia,” called severe acute respiratory syndrome (SARS) was reported in Guangdong Province in China. The disease then spread to 37 countries to cause more than 8,000 cases, with a case-fatality rate of ~10% (7). At that time, SARS had already posed a worldwide public health threat, with a major impact on the economy. More recently, the Middle East respiratory syndrome (MERS) spread to 27 countries, causing around 2,500 cases. Among the CoVs, MERS has the highest case fatality rate (about 30%), but it is rarely transmitted between humans, only via camel (8). Thus, for the third time in a few decades, a new CoV has crossed species to infect human populations. However, compared with the other two CoVs, SARS-CoV-2 is much more contagious. Until now, more than 7 million cases have been diagnosed globally, with over 400,000 fatalities worldwide through early June 2020, with a basic reproductive number estimated to be from 2.2 to 3.3 and a mortality rate of around 2.3% (3, 9).

Like the other CoVs, SARS-CoV-2 possesses a typical envelope structure with spike proteins at the surface; this characteristic certainly plays a major role in interspecies transmission. Based on similarities in spike structure characteristics between SARS-CoV-2 and SARS-CoV, several research groups have demonstrated that SARS-CoV-2 also utilizes the human angiotensin-converting enzyme 2 (ACE2) receptor as a cellular entry receptor (10, 11). ACE2 is mainly expressed in vascular endothelial cells and the renal tubular epithelium. PCR analysis revealed that ACE2 is also expressed in the lungs and gastrointestinal tract, which are tissues shown to harbor viruses (12). It was also suggested that CD147 (basigin or the EMMPRIN protein) could be another cell-surface receptor for SARS-CoV-2 (13). By co-immunoprecipitation, ELISA, and immuno-electron microscopy, they show that anti-CD147 antibody (Meplazumab) could competitively inhibit the binding of spike protein (SP) with CD147 and thus prevent infection of target cells. A phase II clinical trial entitled “Clinical study of anti-CD147 humanized Meplazumab for injection to treat with 2019-nCoV pneumonia” (ClinicalTrials.gov identifier: NCT04275245) is currently underway in China aiming to prevent SARS-CoV-2 SP binding and subsequent infection (14). CD209L (L-SGN) has been identified as another possible alternative receptor for SARS-CoV-2, as previously described for the SARS-CoV virus (15).

This review highlights some of the most recent advances in our understanding of the role of innate and adaptive cellular immunity in COVID-19 infection and discusses potential links to pathogenesis.

IMMUNOPATHOLOGY OF COVID-19

What of the Acute Infection?

The first symptoms associated with COVID-19 are mainly those of respiratory disease, although neurologic and digestive symptoms can also be observed. The primary mode of infection

is human-to-human transmission through close contact, via the spraying of droplets from infected individuals, primarily through the nasal and larynx mucosa, followed by entrance into the lungs through the respiratory tract. Next, in more severe cases, damage/oedema due to extracellular fluid may let the virus enter the peripheral blood from the lungs, causing viremia. COVID-19 has a probable asymptomatic incubation period between 2 and 14 days during which the virus can be transmitted (16), but importantly, the duration of SARS-CoV-2 RNA detection has not been well-characterized. Zhou et al. (12) found that viral titers in nasopharyngeal aspirates diminish 10–15 days after the onset of symptoms, but remains high when the clinical disease worsens. It is, however, noteworthy that the presence of viral RNA in specimens does not always correlate with viral transmissibility; a major limitation remains the inability to differentiate between infective and non-infective (dead or antibody-neutralized) viruses. For SARS and MERS, it had previously been shown that viral RNA persisted in the respiratory tract for at least 3 weeks after disease onset in a majority of patients (17).

What of the Severe Forms?

More than 80% of COVID-19 cases were asymptomatic or presented with mild symptoms, while the remaining cases were severe or critical (2, 18). It seems that the case-fatality rate of COVID-19 (about 1%) is lower than those of SARS (10%) and MERS (30%). Like other pathogenic CoVs, COVID-19 is associated with a typical influenza-like syndrome with fever, cough, fatigue and/or myalgia. Although diarrhea was reported in a foursome of patients with SARS and MERS, intestinal symptoms were rarely observed in patients with COVID-19 (2, 18, 19).

An early report in China found that 14% of COVID-19 patients were hospitalized, including 5% with ICU intervention (20). Similar proportions were observed later in Europe and the US (4). Among those who are seriously ill, acute hypoxemic respiratory failure due to acute respiratory distress syndrome (ARDS) is mainly observed (20, 21). At this stage, the need for mechanical ventilation is high, ranging from 40 to 100% (22); however, encephalitis and antiphospholipid syndrome are rare (23). Common complications of COVID-19 include acute kidney injury, elevated liver enzymes, and cardiac injury (23). The limited COVID-19 post mortem data show prominent alveolar edema, fibrin deposition, immune cell infiltration, and severe multi-organ damage, including renal, cardiac, and liver dysfunction (12, 24).

It was also reported that about 90% of COVID-19 hospitalized patients had at least one risk factor (www.cdc.gov/coronavirus/2019-ncov/index.html). Older age, in particular, as well as a higher sequential organ failure assessment (SOFA) score on admission, are associated with a higher probability of in-hospital death, whereas elevated levels of blood IL-6, high-sensitivity cardiac troponin I, and lymphopenia are more commonly seen in severe COVID-19 illness (12). It is still unknown why the cytokine storm may account for the severity of infection in elderly and immunocompromised (i.e., diabetics) but not in the young population who are mostly asymptomatic but have a fully

functional immune system. However, the variability of clinical cases observed during exposure and infection with SARS-CoV-2 likely suggests that human genetic factors can also influence the response to this virus. However, to date, very few studies have been conducted to determine its real impact.

Based on patients analyzed, SARS-CoV-2 infects all age groups equally, except perhaps children and adolescents. One unanswered question is why some patients develop severe disease, while others do not. Among the different parameters that can influence the severity of this infection, we will focus on the role of the cellular immune response.

RECENT PROGRESS IN IMMUNE CONTROL OF COVID-19 PATHOGENESIS

Usually, type I interferons (IFN- α/β) provide the first line of defense by generating cell-intrinsic antimicrobial states to limit virus replication. It seems, however, that pathogenic CoVs are particularly adapted to dampen responses mediated by IFN- α/β (25, 26). Several hypotheses were proposed to explain this early modulation of the immune response. It was shown that the Orf6 protein of SARS-CoV disrupts the karyopherin transport (27) and consequently inhibits the import of transcriptional factors, such as STAT1, into the nucleus, resulting in an inhibition of IFN response. Similarly, the Orf3b protein of SARS-CoV inhibits phosphorylation of interferon regulatory factor 3 (IRF3) (28), a protein involved in the activation of IFN- α/β . In China, the guidelines for the treatment of COVID-19 recommended administering IFN- α in combination with ribavirin (29), although no improvement was recorded. Interestingly, IFN- α effectively inhibited SARS-CoV replication but 50–90 times lower than IFN- β (30–32), suggesting that IFN- β could be a better antiviral component in patients' treatment. Thus, in the European DisCoVeRy trial, a combination of subcutaneous IFN- β with lopinavir/ritonavir is compared to hydroxychloroquine and remdesivir.

The loss of the “front line” antiviral defense mechanism mediated by IFN- α/β deficiency could be implicated in the induction of the cytokine storm leading to macrophage activation syndrome (MAS)-like pathology (33, 34). This cytokine storm is considered as the root cause of pathogenic inflammation in COVID-19. However, its initial trigger is not yet known, but it likely involves the immune system's detection of a large quantity of viral antigens released by dying cells. One in two fatal cases of COVID-19 experience a cytokine storm, 82% of whom are over the age of 60 (35). Interestingly, NLRP3, a major protein component of the inflammasome, could play a role. During aging, there is a steady increase in the abundance and activity of NLRP3 in immune cells in the lung, which contribute to pulmonary fibrosis (36). After age and hematological cancers, obesity is the next major risk factor for COVID-19 fatality, similar to type 2 diabetes. Obesity is well-known to increase the activity of NLRP3 and stimulate inflammation during viral infection (37).

The cytokine storm is mainly associated with a high production of pro-inflammatory cytokines (i.e. IL-1 β , IL-6, TNF- α) (**Figure 1**). For example, IL-6 production is about 3-fold

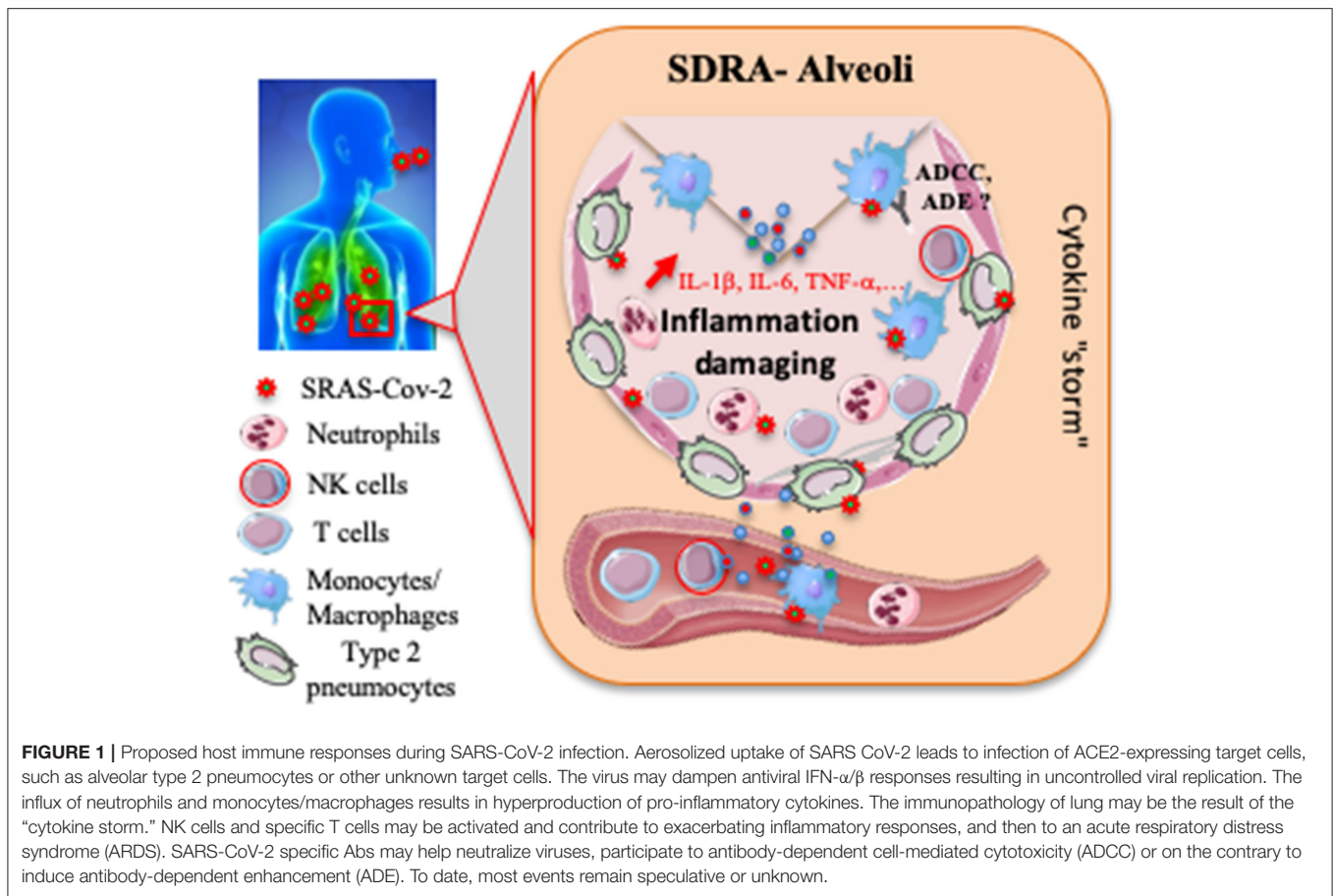
higher in patients with complicated COVID-19 compared to asymptomatic patients (38). Preliminary data with tocilizumab, a humanized anti-IL-6 monoclonal antibody, in patients with COVID-19 pneumonia reveal clinical improvement in a small number of patients (39). Similarly, interferon gamma-induced protein 10 (IP-10) is correlated with patient viral load, whereas monocyte-chemotactic protein 3 (MCP3) is associated with loss of lung function (PaO₂/FaO₂ ratio), lung injury (Murray Score) and fatal outcome (40). Systemic inflammation was also observed in fatal cases of H1N1, with high IL-6 and IP-10 concentrations in the lungs, associated with massive infiltration of immune cells in the lung (41), also reported in severe or fatal forms of avian H5N1 and H7N9 pulmonary infection (42, 43).

What of the Cell-Innate Immunity?

The epithelium of the lungs is the largest surface in the human body (>200 m²) in direct contact with the external environment. The lungs inhale daily about 10,000 l of air that contains various pathogenic particles, like the SARS-CoV-2 in fine droplets. Thus, this constant exposure to pathogens requires a very efficient immune system to sense the challenge and protect the host. To this end, the airways are endowed with physical barriers such as a layer of mucus, which is present over its entire surface to defend this tissue against pathogens, but also a vast network of cellular and humoral host defense mechanisms.

This network is mainly composed of epithelial cells of the respiratory tract, dendritic cells (DC) and alveolar macrophages. These cells trigger pro-inflammatory downstream immune responses in the presence of viral particles. Liao et al. (44) found that the depletion of tissue-resident alveolar macrophages and the accumulation of inflammatory macrophages in bronchoalveolar lavage cells were associated with disease severity. However, it would be necessary to finely test the infectivity of the monocyte/macrophage lineage with SARS-CoV-2 to determine better its impact on inflammatory responses. In this acute inflammatory reaction, neutrophils are also attracted and localized mainly in the bronchoalveolar space (45). Consistently, elevated neutrophil levels were reported in COVID-19⁺ patients (46, 47). Importantly, the lung constitutes the most important reservoir of neutrophils in the systemic circulation (~40% of total body neutrophils). It is plausible that elevated neutrophil level is associated with increased reactive oxygen species (ROS) and neutrophil extracellular traps (NETs), both considered as the most potent antimicrobial mechanisms used by neutrophils. Inappropriate levels of these neutrophil-derived products could contribute to the development of the “cytokine storm” initiated by the lung-infiltrating macrophages, and then to the partial destruction of lung tissues (**Figure 1**) (2, 48).

Mucosal-associated invariant T (MAIT) cells represent a population of innate T cells. They recognize metabolites that are presented by the major histocompatibility complex (MHC) class I-related protein MR1. Potential effectors of MAIT cell antimicrobial activity include the secretion of TNF- α , IFN- γ , IL-17A, and IL-22 as well as granzyme B and perforin (49, 50). Changes in MAIT cell frequencies have been reported in several viral infections; for example, higher cell numbers were found in survivors infected by H7N9 influenza, compared to



samples from fatalities (51). Consistently, *in vitro* coculture of primary peripheral blood mononuclear cells and H7N9-infected A549 airway epithelial cells was associated with increased intracellular IFN- γ and granzyme B levels in MAIT cells (51). Very recent preliminary data also suggested a very significant decrease of MAIT cells in COVID-19⁺ patients; expression of the CD69 activation marker on blood MAIT cells at inclusion was predictive of COVID-19 severity (52).

Natural killer (NK) cells are another key element of innate immunity (53). It was rapidly determined that in COVID-19 patients, the total number of NK cells is markedly decreased (54), as previously reported for the SARS (55). NK cells express a variety of receptors that transduce either activating or inhibitory signals. Integration of these signals regulates the effector functions of NK cells, including cytotoxic activity and cytokine secretion (53, 56). In patients infected with SARS-CoV-2, NKG2A expression was significantly increased on NK cells (54). The CD94/NK group 2 member A (NKG2A) heterodimeric receptor is one of the most prominent NK inhibitory receptors. It binds to a non-classical minimally polymorphic HLA class I molecule (HLA-E), which presents peptides derived from leader peptide sequences of other HLA class I molecules (57). Upon ligation by peptide-loaded HLA-E, NKG2A transduces inhibitory signaling through 2 inhibitory immune-receptor tyrosine-based inhibition motifs, thus suppressing NK cytokine secretion and

cytotoxicity (58). A clinical trial is ongoing in the presence of anti-NKG2A (Monalizumab) in Patients with advanced or metastatic cancer infected by SARS-CoV-2 (ClinicalTrials.gov Identifier: NCT04333914). However, more extensive phenotypic studies of NK cells will be necessary to determine the role of other cell markers and to measure their impact in disease evolution better. Consistent with increased NKG2A levels on NK cells from COVID-19 patients, low polyfunctional capacities were reported (54). Hence, SARS-CoV-2 may break down antiviral immunity mediated by NK cells at an early stage of infection, with putative consequences for the development of an efficient adaptive immunity. To increase NK-cell capability, a phase I clinical trial is ongoing to evaluate the safety and efficiency of allogenic NK-cell transfer in combination with standard therapy for 30 pneumonia patients infected with SARS-CoV-2 (ClinicalTrials.gov identifier: NCT04280224).

In other infectious situations, such as dengue virus infection, activation of NK cells by antibodies (Abs) can enhance controlled antibody-dependent enhancement (ADE) process (Figure 1), which occurs when Abs specific to a viral determinant facilitate secondary infection. Interestingly, it was shown previously that sera from SARS-CoV infected patients enhance viral entry into Fc receptor-expressing cells (59, 60). This mechanism should be extensively studied in a COVID-19 context to guide the development of future vaccine and antibody-based drug therapy.

Together, the preliminary data on COVID-19 patients suggest that SARS-CoV-2 could use different strategies to evade and/or antagonize different arms of the innate immune system.

What of the Cell-Adaptive Immunity?

Severe lymphopenia was observed until death in non-survivor patients with COVID-19 (12). Consistently, the acute phase of SARS in human patients was associated with marked leukopenia in up to 80% of hospitalized patients, associated with a dramatic loss of CD4 and CD8 T cells (61, 62). In SARS-CoV-infected patients, it was shown that infection of T lymphocytes directly contributes to lymphopenia and atrophy of the spleen and lymphoid tissue (63). Lymphopenia is also observed in MERS patients, albeit to a lesser degree than in SARS patients (64). Understanding the mechanism of lymphopenia could open the way to the development of a new strategy for the treatment of COVID-19. Several potential mechanisms could be considered: (i) The virus might directly infect lymphocytes, resulting in lymphocyte death, as recently reported by Wang et al. (65) for the SARS-CoV-1. (ii) The virus can damage different target organs, such as bone marrow and thymus, which can no longer function normally. (iii) Inflammatory cytokines are massively produced, perhaps leading to lymphocyte apoptosis. (iv) Lymphocytes are trapped in infected tissues (**Figure 1**). Further research is needed to confirm these hypotheses. Importantly, the loss of lymphocytes was transient; CD8⁺ T lymphocytes and memory CD4⁺ T cells of SARS patients returned to normal within 2–3 and 12 months after infection, whereas other CD4⁺ T cell subsets were still lower than in healthy controls (66).

The first study on patients with COVID-19 revealed that low levels of IFN- γ and TNF- α in CD4⁺ T cells are associated with severity. Consistently, in CD8⁺ T cells, the frequency of the exhausted (PD-1⁺CTLA-4⁺TIGIT⁺) subset was significantly higher in the severe group (67). Consequently, the low functionality of CD8⁺ T cells in severe patients could impact an efficient control of infection (67), as previously described in SARS-CoV infection (68). Furthermore, COVID-19 was associated with a significant decrease of T cell activation, determined by CD25, CD28, and CD69 expression on CD4⁺ and CD8⁺ T cell subsets (68). Despite a wave of information on the specific T cell responses to many other pathogens, less is known about respiratory CoV infections. CD8⁺ T cells are typically required for the control of influenza virus and other respiratory viruses (68). Furthermore, T resident memory cells (TRM) are critical in preventing re-infection from influenza virus (69). Their role in SARS-CoV-2 infection should be, however, more finely determined. In senescent mice infected by SARS-CoV, CD8⁺ CTLs alone are not sufficient to clear the virus in the absence of both CD4⁺ T cells and specific Abs (70).

On the other hand, depletion of CD4⁺ T cells in SARS-infected patients reduced production of neutralizing Abs and Th1 cytokines and induced lower recruitment of inflammatory monocytes in the lung. This mechanism can be bypassed by a passive transfer of neutralizing Abs against SARS-CoV, suggesting that the CD4-mediated control of infection

most likely operates through Ab- and/or cytokine-dependent mechanisms. In fatal human fulminant cases of H1N1 influenza pneumonia that required mechanical ventilation, a strong effector T-cell response in the lungs was also observed in conjunction with high production of IFN- γ and IP-10, suggesting a massive and effective translocation of specific T cells to the lungs (41).

Genetic differences in HLA haplotypes are also key parameters, known to contribute to individual sensitivity against pathogens as previously described for tuberculosis, leprosy, HIV, hepatitis B, and influenza (71). For example, HLA-A*11, HLA-B*35, and HLA-DRB1*10 confer susceptibility to H1N1 infection (72). For SARS-CoV-2, a preliminary *in silico* analysis of viral peptide-MHC class-I binding affinity suggests that individuals expressing HLA-B*46:01 may be particularly vulnerable to COVID-19, as previously shown for the SARS. At the same time, HLA-B*15:03 showed the greatest capacity to present highly conserved SARS-CoV-2 peptides shared among common human CoVs (73, 74). This observation suggests that the HLA distribution could affect the cellular immune response to SARS-CoV-2, and might explain the differences in COVID-19 susceptibility around the world. However, it seems crucial for the development of vaccine strategies to understand whether specific HLA haplotypes are associated with the development of anti-SARS-CoV-2 immunity. Interestingly, among the first 120 available SARS-CoV-2 sequences (as of February 21, 2020), several B cell and T cell epitopes specific to SARS-CoV-2 were identified for the spike and nucleocapsid proteins, that potentially induce protection against COVID-19 (75).

CONCLUDING COMMENTS

Current observations indicate that SARS-CoV-2 is particularly adapted to evade immune responses at the early stage of infection. Most mechanisms are linked to inappropriate type 1 IFN responses, massive inflammatory cytokine production, and possibly to a defect in NK-cell functions. Preliminary data also suggest adaptive immune evasion, as indicated by the exhaustion of T lymphocytes. However, current evidence strongly indicated that the Th1-type response is key to the successful control of human pathogenic CoVs, in the association with the presence of specific neutralizing Abs. Although there are clear relationships between the severity of the disease and immune responses, the role of protective immunity currently remains questionable.

Alarmingly, some patients remain viral positive, while others even relapse, after discharge from hospital, as recently stated by WHO (3), suggesting that complete control of the virus by the immune response could be difficult to induce at least in some patients. This could also have an impact on the development of the second wave of the epidemic, which is currently strongly envisaged. The vaccine remains the best way to counter this epidemic. However, to define the surrogate parameters of vaccine efficacy, it should be important to better monitor T/B cell responses of recovered patients and to better

understand the aging impact on the immune responses in COVID-19 patients, including the relative protection of younger individuals, excepted for some unexplained cases of Kawasaki-like syndrome. If overlapping epitopes among different human CoVs can be identified, this could help in the design of cross-reactive vaccines that protect against several pathogenic CoVs in the future.

DATA AVAILABILITY STATEMENT

All datasets presented in this study are included in the article.

REFERENCES

- Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med.* (2020) 382:727–33. doi: 10.1056/NEJMoa2001017
- Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet.* (2020) 395:497–506. doi: 10.1016/S0140-6736(20)30183-5
- World Health Organization. *Considerations in Adjusting Public Health and Social Measures in the Context of COVID-19: Interim Guidance*, 16 April 2020. (2020) Available online at: <https://apps.who.int/iris/handle/10665/331773>
- Lescure FX, Bouadma L, Nguyen D, Parisey M, Wicky PH, Behillil S, et al. Clinical and virological data of the first cases of COVID-19 in Europe: a case series. *Lancet Infect Dis.* (2020) 6:697–706. doi: 10.1016/S1473-3099(20)30200-0
- Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature.* (2020) 579:270–3. doi: 10.1038/s41586-020-2012-7
- Forster P, Forster L, Renfrew C, Forster M. Phylogenetic network analysis of SARS-CoV-2 genomes. *Proc Natl Acad Sci USA.* (2020) 117:9241–3. doi: 10.1073/pnas.2004999117
- Lipsitch M, Cohen T, Cooper B, Robins JM, Ma S, James L, et al. Transmission dynamics and control of severe acute respiratory syndrome. *Science.* (2003) 300:1966–70. doi: 10.1126/science.1086616
- Park JE, Jung S, Kim A, Park JE. MERS transmission and risk factors: a systematic review. *BMC Public Health.* (2018) 18:574. doi: 10.1186/s12889-018-5484-8
- Thompson R. Pandemic potential of 2019-nCoV. *Lancet Infect Dis.* (2020) 20:280. doi: 10.1016/S1473-3099(20)30068-2
- Letko M, Marzi A, Munster V. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses. *Nat Microbiol.* (2020) 5:562–9. doi: 10.1038/s41564-020-0688-y
- Jiang F, Yang J, Zhang Y, Dong M, Wang S, Zhang Q, et al. Angiotensin-converting enzyme 2 and angiotensin 1-7: novel therapeutic targets. *Nat Rev Cardiol.* (2014) 11:413–26. doi: 10.1038/nrcardio.2014.59
- Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet.* (2020) 395:1054–62. doi: 10.1016/S0140-6736(20)30566-3
- Wang K, Chen W, Zhou YS, Lian JQ, Zhang Z, Du P, et al. SARS-CoV-2 invades host cells via a novel route: CD147-spike protein. *bioRxiv.* (2020). doi: 10.1101/2020.03.14.988345
- Ulrich H, Pillat MM. CD147 as a target for COVID-19 treatment: suggested effects of azithromycin and stem cell engagement. *Stem Cell Rev Rep.* (2020) 16:434–40. doi: 10.1007/s12015-020-09976-7
- Jeffers SA, Tusell SM, Gillim-Ross L, Hemmila EM, Achenbach JE, Babcock GJ, et al. CD209L (L-SIGN) is a receptor for severe acute respiratory syndrome coronavirus. *Proc Natl Acad Sci USA.* (2004) 101:15748–53. doi: 10.1073/pnas.0403812101
- Wu F, Zhao S, Yu B, Chen YM, Wang W, Song ZG, et al. A new coronavirus associated with human respiratory disease in China. *Nature.* (2020) 579:265–9. doi: 10.1038/s41586-020-2008-3

AUTHOR CONTRIBUTIONS

All authors were involved in reading bibliography and writing the article. All co-authors reviewed the article.

FUNDING

This study was supported in part by the Agence Nationale de la Recherche (ANR, iCovid program), and Sorbonne Université. The funding organizations did not have any role in the design or conduct of this review.

- Oh MD, Park WB, Choe PG, Choi SJ, Kim JJ, Chae J, et al. Viral load kinetics of MERS coronavirus infection. *N Engl J Med.* (2016) 375:1303–5. doi: 10.1056/NEJMc1511695
- Chan JF, Yuan S, Kok KH, To KK, Chu H, Yang J, et al. A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster. *Lancet.* (2020) 395:514–23. doi: 10.1016/S0140-6736(20)30154-9
- Wu Z, McGoogan JM. Characteristics of and important lessons from the coronavirus disease 2019 (COVID-19) outbreak in china: summary of a report of 72 314 cases from the Chinese center for disease control and prevention. *JAMA.* (2020) 323:1239–42. doi: 10.1001/jama.2020.2648
- Arentz M, Yim E, Klaff L, Lokhandwala S, Riedo FX, Chong M, et al. Characteristics and outcomes of 21 critically ill patients with COVID-19 in Washington State. *JAMA.* (2020) 323:1612–4. doi: 10.1001/jama.2020.4326
- Grasselli G, Zangrillo A, Zanella A, Antonelli M, Cabrini L, Castelli A, et al. Baseline characteristics and outcomes of 1591 patients infected with SARS-CoV-2 admitted to ICUs of the Lombardy Region, Italy. *JAMA.* (2020) 323:1574–81. doi: 10.1001/jama.2020.5394
- Yang X, Yu Y, Xu J, Shu H, Xia J, Liu H, et al. Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. *Lancet Respir Med.* (2020) 8:475–81. doi: 10.1016/S2213-2600(20)30079-5
- Zhang Y, Xiao M, Zhang S, Xia P, Cao W, Jiang W, et al. Coagulopathy and antiphospholipid antibodies in patients with Covid-19. *N Engl J Med.* (2020) 382:e38. doi: 10.1056/NEJMc2007575
- Yao XH, Li TY, He ZC, Ping YF, Liu HW, Yu SC, et al. A pathological report of three COVID-19 cases by minimally invasive autopsies. *Zhonghua Bing Li Xue Za Zhi.* (2020) 49:E009. doi: 10.3760/cma.j.cn112151-20200312-00193
- Spiegel M, Pichlmair A, Martínez-Sobrido L, Cros J, García-Sastre A, Haller O, et al. Inhibition of beta interferon induction by severe acute respiratory syndrome coronavirus suggests a two-step model for activation of interferon regulatory factor 3. *J Virol.* (2005) 79:2079–86. doi: 10.1128/JVI.79.4.2079-2086.2005
- Kikkert M. Innate immune evasion by human respiratory RNA viruses. *J Innate Immun.* (2020) 12:4–20. doi: 10.1159/000503030
- Frieman M, Yount B, Heise M, Kopecky-Bromberg SA, Palese P, Baric RS. Severe acute respiratory syndrome coronavirus ORF6 antagonizes STAT1 function by sequestering nuclear import factors on the rough endoplasmic reticulum/Golgi membrane. *J Virol.* (2007) 81:9812–24. doi: 10.1128/JVI.01012-07
- Kopecky-Bromberg SA, Martínez-Sobrido L, Frieman M, Baric RA, Palese P. Severe acute respiratory syndrome coronavirus open reading frame (ORF) 3b, ORF 6, and nucleocapsid proteins function as interferon antagonists. *J Virol.* (2007) 81:548–57. doi: 10.1128/JVI.01782-06
- Lu H. Drug treatment options for the 2019-new coronavirus (2019-nCoV). *Biosci Trends.* (2020) 14:69–71. doi: 10.5582/bst.2020.01020
- Cinatl J, Morgenstern B, Bauer G, Chandra P, Rabenau H, Doerr HW. Treatment of SARS with human interferons. *Lancet.* (2003) 362:293–4. doi: 10.1016/S0140-6736(03)13973-6
- Lokugamage KG, Schindewolf C, Menachery VD. SARS-CoV-2 sensitive to type I interferon pretreatment. *bioRxiv. [Preprint].* (2020). doi: 10.1101/2020.03.07.982264

32. Sallard E, Lescure FX, Yazdanpanah Y, Mentre F, Peiffer-Smadja N. Type 1 interferons as a potential treatment against COVID-19. *Antiviral Res.* (2020) 178:104791. doi: 10.1016/j.antiviral.2020.104791
33. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ. HLH across speciality collaboration, UK. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet.* (2020) 395:1033–4. doi: 10.1016/S0140-6736(20)30628-0
34. McGonagle D, Sharif K, O'Regan A, Bridgewood C. The role of cytokines including interleukin-6 in COVID-19 induced pneumonia and macrophage activation syndrome-like disease. *Autoimmun Rev.* (2020) 3:102537. doi: 10.1016/j.autrev.2020.102537
35. Paranjpe I, Russak A, De Freitas JK, Lala A, Miotto R, Vaid A, et al. Clinical characteristics of hospitalized Covid-19 patients in New York City. *medRxiv. [Preprint].* (2020). doi: 10.1101/2020.04.19.20062117
36. Stout-Delgado HW, Cho SJ, Chu SG, Mitzel DN, Villalba J, El-Chemaly S, et al. Age-dependent susceptibility to pulmonary fibrosis is associated with NLRP3 inflammasome activation. *Am J Respir Cell Mol Biol.* (2016) 55:252–63. doi: 10.1165/rcmb.2015-0222OC
37. Guan WJ, Liang WH, Zhao Y, Liang HR, Chen ZS, Li YM, et al. Comorbidity and its impact on 1590 patients with COVID-19 in China: a nationwide analysis. *Eur Respir J.* (2020) 55:2000547. doi: 10.1183/13993003.01227-2020
38. Coomes A, Haghbayan H. Interleukin-6 in COVID-19: a systematic review and meta-analysis. *medRxiv. [Preprint].* (2020). doi: 10.1101/2020.03.30.20048058
39. Xu X, Han M, Li T, Sun W, Wang D, Fu B, et al. Effective treatment of severe COVID-19 patients with tocilizumab. *Proc Natl Acad Sci USA.* (2020) 117:10970–5. doi: 10.1073/pnas.2005615117
40. Xiong Y, Liu Y, Cao L, Wang D, Guo M, Jiang A, et al. Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients. *Emerg Microbes Infect.* (2020) 9:761–70. doi: 10.1080/22221751.2020.1747363
41. Guihot A, Luyt CE, Parrot A, Rousset D, Cavallion JM, Boutolleau D, et al. Low titers of serum antibodies inhibiting hemagglutination predict fatal fulminant influenza A(H1N1) 2009 infection. *Am J Respir Crit Care Med.* (2014) 189:1240–9. doi: 10.1164/rccm.201311-2071OC
42. de Jong MD, Simmons CP, Thanh TT, Hien VM, Smith GJ, Chau TN, et al. Fatal outcome of human influenza A(H5N1) is associated with high viral load and hypercytokinemia. *Nat Med.* (2006) 12:1203–7. doi: 10.1038/nm1477
43. Iwase S, Nakada TA, Hattori N, Takahashi W, Takahashi N, Aizimu T, et al. Interleukin-6 as a diagnostic marker for infection in critically ill patients: a systematic review and meta-analysis. *Am J Emerg Med.* (2019) 37:260–5. doi: 10.1016/j.ajem.2018.05.040
44. Liao M, Liu Y, Yuan J, Wen Y, Xu G, Zhao J, et al. The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing. *medRxiv. [Preprint].* (2020). doi: 10.1101/2020.02.23.20026690
45. Kruger P, Saffarzadeh M, Weber AN, Rieber N, Radsak M, von Bernuth H, et al. Neutrophils: between host defence, immune modulation, and tissue injury. *PLoS Pathog.* (2015) 11:e1004651. doi: 10.1371/journal.ppat.1004651
46. Mo P, Xing Y, Xiao Y, Deng L, Zhao Q, Wang H, et al. Clinical characteristics of refractory COVID-19 pneumonia in Wuhan, China. *Clin Infect Dis.* (2020). ciaa270. doi: 10.1093/cid/ciaa270
47. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with COVID-19 in Wuhan, China. *Clin Infect Dis.* (2020). ciaa248. doi: 10.1093/cid/ciaa248
48. Prompetchara E, Ketloy C, Palaga T. Immune responses in COVID-19 and potential vaccines: lessons learned from SARS and MERS epidemic. *Asian Pac J Allergy Immunol.* (2020) 38:1–9. doi: 10.12932/AP-200220-0772
49. Godfrey DI, Koay HF, McCluskey J, Gherardin NA. The biology and functional importance of MAIT cells. *Nat Immunol.* (2019) 20:1110–28. doi: 10.1038/s41590-019-0444-8
50. van Wilgenburg B, Scherwitzl I, Hutchinson EC, Leng T, Kurioka A, Kulicke C, et al. MAIT cells are activated during human viral infections. *Nat Commun.* (2016) 7:11653. doi: 10.1038/ncomms11653
51. Loh L, Wang Z, Sant S, Koutsakos M, Jegaskanda S, Corbett AJ, et al. Human mucosal-associated invariant T cells contribute to antiviral influenza immunity via IL-18-dependent activation. *Proc Natl Acad Sci USA.* (2016) 113:10133–8. doi: 10.1073/pnas.1610750113
52. Jouan Y, Guillon A, Gonzalez L, Perez Y, Ehrmann S, Ferreira M, et al. Functional alteration of innate T cells in critically ill Covid-19 patients. *medRxiv. [Preprint].* (2020). doi: 10.1101/2020.05.03.20089300
53. Lam VC, Lanier LL. NK cells in host responses to viral infections. *Curr Opin Immunol.* (2017) 44:43–51. doi: 10.1016/j.coi.2016.11.003
54. Zheng M, Gao Y, Wang G, Song G, Liu S, Sun D, et al. Functional exhaustion of antiviral lymphocytes in COVID-19 patients. *Cell Mol Immunol.* (2020) 17:533–5. doi: 10.1038/s41423-020-0402-2
55. National Research Project for SARS, Beijing Group. The involvement of natural killer cells in the pathogenesis of severe acute respiratory syndrome. *Am J Clin Pathol.* (2004) 121:507–11. doi: 10.1309/WPK7Y2XKNF4CBF3R
56. Hammer Q, Rückert T, Romagnani C. Natural killer cell specificity for viral infections. *Nat Immunol.* (2018) 19:800–8. doi: 10.1038/s41590-018-0163-6
57. Lazetic S, Chang C, Houchins JB, Lanier LL, Phillips JH. Human natural killer cell receptors involved in MHC class I recognition are disulfide-linked heterodimers of CD94 and NKG2 subunits. *J Immunol.* (1996) 157:4741–5.
58. Braud VM, Allan DS, O'Callaghan CA, Söderström K, D'Andrea A, Ogg GS, et al. HLA-E binds to natural killer cell receptors CD94/NKG2A, B and C. *Nature.* (1998) 391:795–9. doi: 10.1038/35869
59. Tetro JA. Is COVID-19 receiving ADE from other coronaviruses? *Microbes Infect.* (2020) 22:72–3. doi: 10.1016/j.micinf.2020.02.006
60. Wan Y, Shang J, Sun S, Tai W, Chen J, Geng Q, et al. Molecular mechanism for antibody-dependent enhancement of coronavirus entry. *J Virol.* (2020) 94:e02015-19. doi: 10.1128/JVI.02015-19
61. Li T, Qiu Z, Zhang L, Han Y, He W, Liu Z, et al. Significant changes of peripheral T lymphocyte subsets in patients with severe acute respiratory syndrome. *J Infect Dis.* (2004) 189:648–51. doi: 10.1086/381535
62. Channappanavar R, Zhao J, Perlman S. T cell-mediated immune response to respiratory coronaviruses. *Immunol Res.* (2014) 59:118–28. doi: 10.1007/s12026-014-8534-z
63. Gu J, Gong E, Zhang B, Zheng J, Gao Z, Zhong Y, et al. Multiple organ infection and the pathogenesis of SARS. *J Exp Med.* (2005) 202:415–24. doi: 10.1084/jem.20050828
64. Assiri A, Al-Tawfiq JA, Al-Rabeeh AA, Al-Rabiah FA, Al-Hajjar S, Al-Barrak A, et al. Epidemiological, demographic, and clinical characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia: a descriptive study. *Lancet Infect Dis.* (2013) 13:752–61. doi: 10.1016/S1473-3099(13)70204-4
65. Wang X, Xu W, Hu G, Xia S, Sun Z, Liu Z, et al. SARS-CoV-2 infects T lymphocytes through its spike protein-mediated membrane fusion. *Cell Mol Immunol.* (2020) 1–3. doi: 10.1038/s41423-020-0424-9
66. Lin L, Lu L, Cao W, Li T. Hypothesis for potential pathogenesis of SARS-CoV-2 infection—a review of immune changes in patients with viral pneumonia. *Emerg Microbes Infect.* (2020) 9:727–32. doi: 10.1080/22221751.2020.1746199
67. Zheng HY, Zhang M, Yang CX, Zhang N, Wang XC, Yang XP, et al. Elevated exhaustion levels and reduced functional diversity of T cells in peripheral blood may predict severe progression in COVID-19 patients. *Cell Mol Immunol.* (2020) 17:541–3. doi: 10.1038/s41423-020-0401-3
68. Schmidt ME, Varga SM. The CD8 T cell response to respiratory virus infections. *Front Immunol.* (2018) 9:678. doi: 10.3389/fimmu.2018.00678
69. Zens KD, Chen JK, Farber DL. Vaccine-generated lung tissue-resident memory T cells provide heterosubtypic protection to influenza infection. *JCI Insight.* (2016) 1:e85832. doi: 10.1172/jci.insight.85832
70. Chen J, Lau YF, Lamirande EW, Paddock CD, Bartlett JH, Zaki SR, Subbarao K. Cellular immune responses to severe acute respiratory syndrome coronavirus (SARS-CoV) infection in senescent BALB/c mice: CD4+ T cells are important in control of SARS-CoV infection. *J Virol.* (2010) 84:1289–301. doi: 10.1128/JVI.01281-09
71. Kenney AD, Dowdle JA, Boccazzo L, McMichael TM, St Gelais C, Panfil AR, et al. Human genetic determinants of viral diseases. *Annu Rev Genet.* (2017) 51:241–63. doi: 10.1146/annurev-genet-120116-023425
72. Dutta M, Dutta P, Medhi S, Borkakoty B, Biswas D. Polymorphism of HLA class I and class II alleles in influenza A (H1N1)pdm09 virus infected population of Assam, Northeast India. *J Med Virol.* (2018) 90:854–60. doi: 10.1002/jmv.25018

73. Nguyen A, David JK, Maden SK, Wood MA, Weeder BR, Nellore A, et al. Human leukocyte antigen susceptibility map for SARS-CoV-2. *J Virol.* (2020) 94:e00510-20. doi: 10.1101/2020.03.22.20040600
74. Lin M, Tseng HK, Trejaut JA, Lee HL, Loo JH, Chu CC, et al. Association of HLA class I with severe acute respiratory syndrome coronavirus infection. *BMC Med Genet.* (2003) 4:9. doi: 10.1186/1471-2350-4-9
75. Ahmed SF, Quadeer AA, McKay MR. Preliminary identification of potential vaccine targets for the COVID-19 coronavirus (SARS-CoV-2) based on SARS-CoV immunological studies. *Viruses.* (2020) 12:E254. doi: 10.3390/v12030254

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Guihot, Litvinova, Autran, Debré and Vieillard. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.