



Clinical impact of the lung tissue transcriptome in a teenager with multifocal invasive mucinous adenocarcinoma – a case report

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Abstract: The transcriptional profiling of cancer and normal tissues harboring cancer can be a clinical and discovery tool, especially for the study of rare tumors. Invasive mucinous adenocarcinoma (IMA) is a rare lung cancer histotype, which mostly affects the elderly and commonly has a poor prognosis. We investigated the exceptional case of a teenager, exposed to passive smoke and chemical carcinogens, who developed a multifocal IMA with bilateral involvement. The malignancy was asymptomatic and was diagnosed occasionally during hospitalization for acute abdominal pain due to adnexitis. The young patient underwent video-assisted thoracoscopic surgery and lung samples were analysed by RNA-Sequencing. The transcriptome of patient’s normal and neoplastic lung tissues was compared with matched healthy controls and IMA signature cases, using Gene Set Enrichment Analyses, Gene Ontology and Genotype Tissue Expression database. Compared to healthy controls, the patient’s lung tissue lacked the expression of lymphocyte and humoral-mediated immune response genes, whereas genes driving the response to stimulus, chemical and organic substances, primarily, *CXCL8*, *ACKR1*, *RAB7B*, *HOXC9*, *HOXD9*, *KLF5* and *NKX2-8* were overexpressed. Genes driving extracellular structure organization, cell adhesion, cell movement, metabolic and apoptotic processes were down-modulated in patient’s lung tissue. When compared to IMA signature cases, the patient’s IMA revealed a prevalent expression of genes regulating the response to stimulus, myeloid and neutrophil activation and immune system processes, primarily *CD1a* and *CXCL13/BCA1*, whereas stemness genes and proto-oncogenes, such as *SOX4*, *HES1*, *IER3* and *SERPINH1* were downmodulated. These transcriptional signature associated with a favorable clinical course, since the patient was healthy five years after initial diagnosis. The transcriptome of the normal tissues bearing tumor provides meaningful information on the gene pathways driving tumor histogenesis, with a prospective impact on early diagnosis. Unlike the tumor histotype-related transcriptional signature, the individual patient’s signature enables tailored treatment and accurate prognosis.

Keywords: Invasive mucinous adenocarcinoma (IMA); lung tissue transcriptome; RNA sequencing; molecular signature; immune-response genes

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Introduction

Lung cancer is the leading cause of death worldwide and the most common cancer, excluding skin cancers, after the prostate and breast cancers (1). The American Cancer Society’s estimates for 2019 about 228,150 new cases of

lung cancer and 142,670 deaths in the United States with considerable Public Health impact.

The discovery of biomarkers for tumor prevention, early diagnosis and accurate prognosis, which involves a deeper understanding of the tumor histogenesis, would improve the

management of lung cancer patient and “not yet patient” at risk to develop cancer.

To achieve this goal, genetic and molecular investigation must extend beyond the tumor specimen and include the histologically normal tissue adjacent to and distant from cancer.

Histologically normal-appearing tissues adjacent to cancers display cancer-associated molecular abnormalities known as “field effect”, that can either be due to “first hit” genetic alterations in a clone of cells from which the tumor ultimately develops, or to effects of the tumor on the local microenvironment (2-4). On the other hand, the prolonged exposure to genotoxic agents induces widespread molecular changes, defined as “field cancerization”, which can lead to the onset of multiple primary tumors (5). Decipher the transcriptional aberrations underlying these histologically normal-appearing conditions is proving to be fundamental in understanding the pathogenesis of tumor and, in particular, of the rare tumor histotypes in a singular clinical context, and in developing a personalized prognostic and therapeutic approach.

Invasive mucinous adenocarcinoma (IMA) is a distinct lung cancer histotype, which represents 5% of lung-adenocarcinomas, and mostly affects adults over 65 (6). Patients frequently present with pneumonia-like symptoms and multifocal disease (7). The prognostic implication is still a matter of debate, since some studies report a recurrence-free survival comparable to stage-matched non mucinous adenocarcinomas, while others describe a worse clinical behavior, with high recurrence rate (76% in 5 years) (8,9).

Here, we report the unprecedented case of a teenager with multifocal IMA and investigate the transcriptional signature of both the patient’s tumor and histologically normal lung tissues, distant from cancer. Integrated and comparative transcriptome analyses highlight the potential of the transcriptional profiles of normal and neoplastic lung tissues to unmask gene pathways driving oncogenesis with important clinical implications. We present the following case in accordance with the CARE guideline checklist.

Case presentation

A 16 years old girl was hospitalized due to acute adnexitis. Unexpectedly, the chest X-ray and subsequent spiral computed tomography (CT) imaging revealed, in the left-lower lobe, a nodule of 15 mm and, in the right middle lobe, a nodule of 7 mm with irregular margins (*Figure 1A*).

Signs and symptoms of lung cancer were absent.

The girl had never smoked, but had a history of passive smoking in early life, and exposure to dichloroethylene and hexavalent chromium released by a tannery near her home. The paternal uncle died at the age of 46 due to metastatic lung cancer. No histopathological and molecular data available. The patient underwent a wide wedge resection of the left inferior lobe and lymph node sampling by video assisted thoracoscopic surgery. The pathological diagnosis was IMA with lepidic growth pattern areas and intratumoral lymphoid follicle-like structures (*Figure 1B*). The tumor stained negative for Transcription Termination Factor 1 (TTF-1), Caudal Type Homeobox 2 (CDX2), ALK Receptor Tyrosine Kinase (ALK) and ROS Proto-Oncogene 1 (ROS1). Molecular analyses revealed no *Epidermal Growth Factor Receptor (EGFR)* gene mutations, whereas a G12D mutation was found in the *KRAS* gene. Three months later, the smaller nodule was also removed by a wide wedge resection of the right upper and middle lobes, with lymph node sampling. Histopathological and molecular reports were consistent with previous findings. No therapy was given. Five years after initial diagnosis, a total body CT confirmed that the patient was tumor free and healthy (*Figure 1C*).

Histology and immunohistochemistry

The investigations were carried out following the rules of the Declaration of Helsinki of 1975, revised in 2013. The study was approved by the ethical committee of the “G. d’Annunzio” University (COET 19.07.2012) and written consent was obtained from the patient and parents.

For histology, lung tissue samples were formalin fixed and paraffin embedded, sectioned at 4 µm, and stained with hematoxylin and eosin (H&E). For immunohistochemistry, paraffin embedded sections, after antigen retrieval, were stained with the following antibodies (Abs), TTF-1 (clone 8G7G3/1, Agilent Technologies), CDX2 (clone DAK-CDX2, Agilent Technologies), ALK (clone D5F3, Roche), ROS1 (clone CST D4D6, Cell Signaling), C-X-C Motif Chemokine Ligand 13 (CXCL13; R&D Systems).

qBiomarker somatic mutation PCR Arrays

To assess *KRAS* and *EGFR* mutations, the DNA was extracted from 8 formalin fixed, paraffin embedded sections (10 µm). *EGFR* and *KRAS* mutations were analyzed with the qBiomarker™ Somatic Mutation PCR Arrays (Qiagen). Data were analysed with the *Data analysis software for*

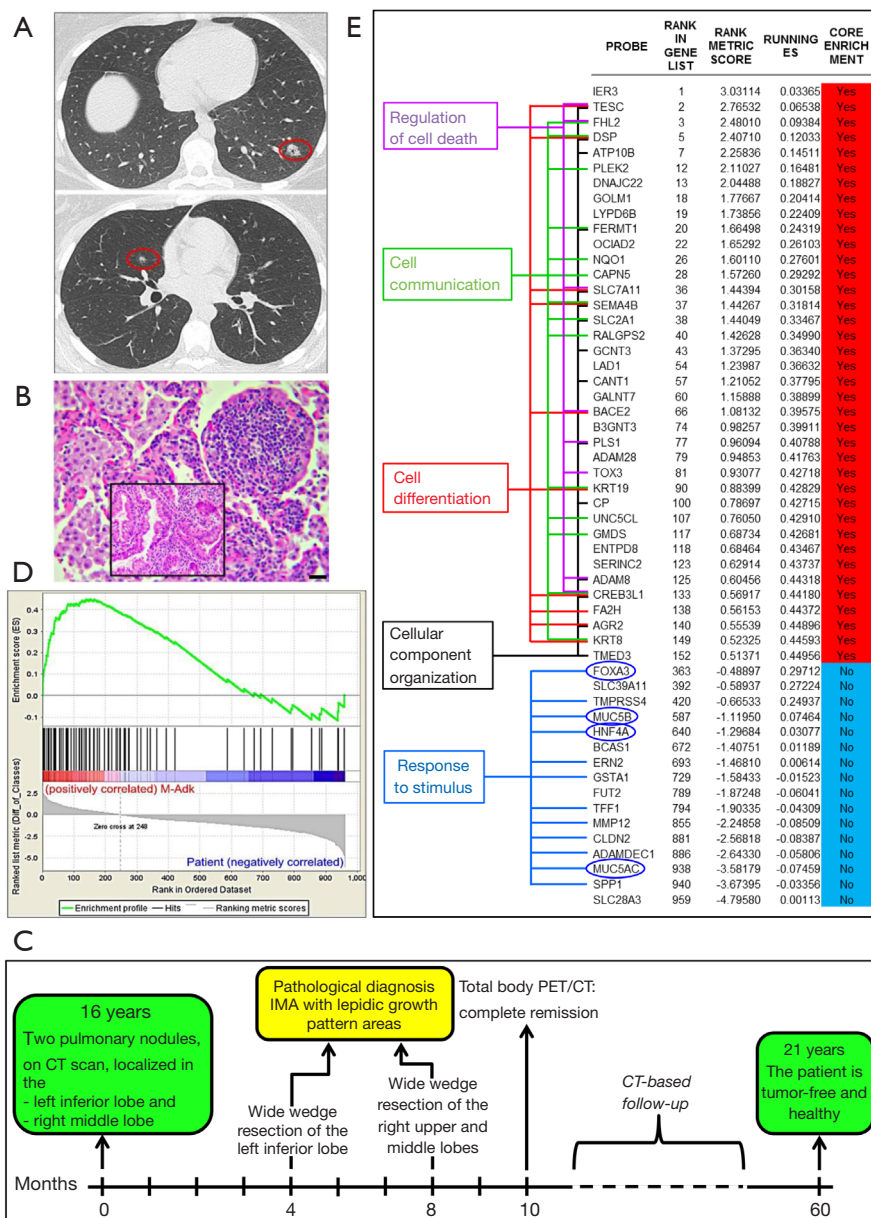


Figure 1 Radiological, histological and transcriptomic analyses of patient's IMA. (A) Spiral CT scans demonstrated, in the left lower lobe, a nodular consolidation area, with irregular margins, of 15 mm in size (red circle) and, in the right middle lobe, an excavated nodule of 7 mm in size (red circle). (B) Hematoxylin and eosin staining of one of the lung neoplastic nodules revealed the histologic feature of IMA, with an intratumoral lymphoid follicle-like structure. Magnification: x200. Scale bar: 30 μm. The inset shows the aspects of lepidic growth and a considerable lymphomononuclear cell infiltrate in the tumor stroma. Magnification: x400. Scale bar: 40 μm. (C) Timeline of the patient. (D) Gene Set Enrichment Analysis (GSEA) (<http://software.broadinstitute.org/gsea/index.jsp>) shows that the IMA Signature (143 genes) developed by Guo *et al.* in 2017, through the analysis of six IMA cases (Enrichment Score =0.4495621; Nominal P value <1%; False Discovery Rate <25%), is not enriched in our patient (Enrichment score <0.1). (E) The table shows the genes, included in the IMA signature, ordered by the normalized enrichment score. Genes with a “Yes value” (in red), contribute to the leading edge subset within the gene set (i.e., genes that contributes most to the enrichment result). Genes with a “No value” (in blue) contribute less to the enrichment result. Genes that regulate cell death, communication, differentiation and cellular component organization prevail in the IMA signature, whereas expression of genes regulating the response to stimulus, including typical IMA-associated genes, such as *HNF4A*, *FOXA3*, *MUC5AC* and *MUC5B* (circled in dark blue), are prevalent in the patient's IMA. IMA, invasive mucinous adenocarcinoma.

qBiomarker Somatic Mutation PCR Arrays (www.SABiosciences.com/somaticmutationdataanalysis.php).

Next generation sequencing (NGS)

RNA was extracted from fresh frozen neoplastic samples and histologically normal lung samples. According to The Cancer Genome Atlas (TCGA) Research Network protocols (<http://cancergenome.nih.gov/>), histologically normal tissues adjacent to tumor samples have been collected more than 2 cm distant from cancer, to avoid the field effect (10).

RNA was extracted from 10 frozen tissue sections (10 µm) and the NGS was performed by Lexogen GmbH, using the Illumina HiSeq 2500 V4 System. The reads alignment was done with STAR aligner and differential expression analysis was performed with DESeq software package.

Gene set enrichment analysis (GSEA)

For the GSEA, we used the GSEA computational method (<http://software.broadinstitute.org/gsea/index.jsp>), setting the IMA signature, identified by Guo *et al.* (11), as gene set.

Functional classification of differentially expressed genes

The functional classification of genes was performed using the Gene Ontology Consortium website (<http://geneontology.org>), with the PANTHER Overrepresentation Test (GO Ontology database, Released 2018-12-01), followed by Fisher's Exact test with Bonferroni correction. Subsequently, the functional categories were also screened using the DAVID (12,13) and the UniProt database (<https://www.uniprot.org/>).

Comparative transcriptome analyses

The cohort of patients, identified by Guo *et al.* to determine the IMA signature, was used for the comparative transcriptome analyses with the IMA of the patient.

For comparative transcriptome analysis of normal lung tissues, the data of the reference cohort (consisting of women died for accidental causes and matched for age and smoking history) were obtained from the GTEx portal (<https://gtexportal.org/home/>) (V6p release).

The IMA signature is not enriched in the patient's IMA

To assess whether, this unusual clinical case, was linked to

a particular gene signature, we analyzed, through RNA-sequencing, the transcriptome of the patient's IMA that was compared, through the GSEA method (14), to the recently defined IMA signature (11). The signature was not enriched in our patient (*Figure 1D*). Genes that regulate cell death, communication, differentiation and cellular component organization prevailed in IMA signature, whereas expression of genes regulating the response to stimulus, including typical IMA-associated genes *Hepatocyte Nuclear Factor 4 Alpha (HNF4A)*, *Forkhead Box A3 (FOXA3)*, *Mucin 5AC (MUC5A)* and *Mucin 5B (MUC5B)*, was prominent in the patient's IMA (*Figure 1E*).

The transcriptome of the patient's normal lung reveals the down-regulation of genes driving cell adhesion and apoptosis and up-regulation of RAS family members and genes driving the response to chemical stimulus

Next, we wondered whether these alterations could also be found in the patient's histologically normal lung tissue, far from cancer (10). Comparative transcriptome analyses with lung tissue of healthy age matched never smoker women revealed that genes driving the response to stimulus, chemical and organic substances, primarily *CXCL8/IL-8*, *Atypical Chemokine Receptor 1 (ACKR1/DARC)*, and the member of the RAS oncogene family *RAB7B* were expressed in the patient's normal lung, but absent in the lung of healthy women (*Figure 2A*).

Additional genes driving the response to chemical, primarily *Homeobox C9 (HOXC9)*, *Homeobox D9 (HOXD9)*, *Kruppel Like Factor 5 (KLF5)*, *NK2 Homeobox 8 (NKX2-8)*, together with regulators of cell signaling, electron transport chain and mitochondrial organization, gene transcription and expression were upregulated in the patient's normal lung (*Figure 2B*). Interestingly, genes driving cell adhesion, cell movement, metabolic and apoptotic processes were downregulated in patient's normal lung (*Figure 2C*), whereas genes driving immune system processes, humoral and lymphocyte immune responses were expressed in healthy-woman's lung, but lacked in the patient's normal lung (*Figure 2D*).

The transcriptome of the patient's IMA revealed overexpression of genes driving the immune system process and the response to chemical stimulus and down-regulation of specific proto-oncogenes

Comparative analyses, by functional categories, between

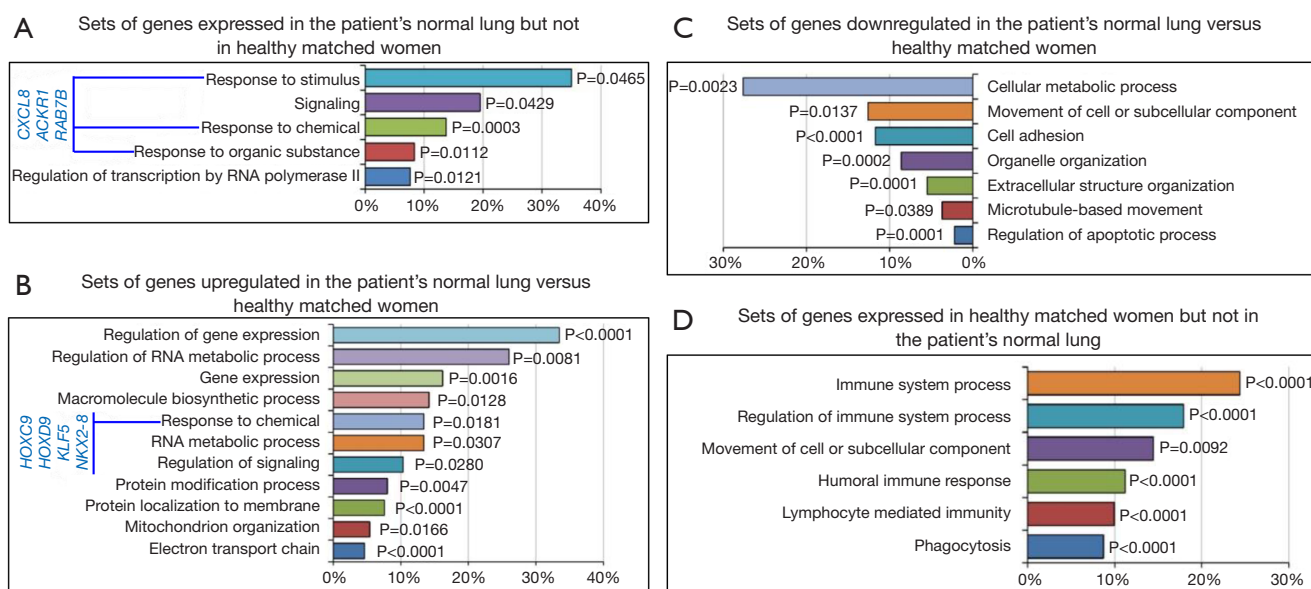


Figure 2 Comparative transcriptomic analysis of patient's normal lung versus healthy matched women, performed using the Gene Ontology Consortium website (<http://geneontology.org>). (A) Sets of genes expressed in the patient's normal lung tissue, but lacking in the normal lung of healthy age-matched women. The functional categories of the response to stimulus, chemicals and organic substance were also screened using the DAVID and UniProt database (<https://www.uniprot.org/>). The RAS oncogene family member *RAB7B*, *ACKR1/DARC* and *CXCL8/IL-8* genes are the most represented in the functional categories of genes driving the *response stimulus*, *chemical* and *organic substance*. (B) Sets of genes upregulated in the patient's normal lung versus normal lung of healthy age-matched women. The functional category of the response to chemicals was also screened using the DAVID and UniProt database (<https://www.uniprot.org/>). *HOXC9*, *HOXD9*, *KLF5*, *NKX2-8* genes are the most represented in the functional category of genes driving the response to chemical. (C) Sets of genes downregulated in the patient's normal lung versus the normal lung of healthy age-matched women. Genes driving *cell adhesion*, *cell movement*, *metabolic* and *apoptotic processes* are downregulated in the patient's normal lung tissue. (D) Sets of genes expressed in the normal lung of healthy age matched women, but unexpressed in the patient's normal lung. Genes driving *immune system process*, *humoral* and *lymphocyte immune responses* are expressed in healthy women's lung, but not in the patient's lung tissue. Lung tissue transcriptomes from healthy age-matched never smoker women, died for accidental causes, were obtained from the GTEx portal (<https://gtexportal.org/home/>) (V6p release). P = Bonferroni corrected P value.

patient's IMA and IMA-signature cases revealed that genes driving the response to stimulus and immune system process, primarily *CD1a* and *CXCL13/BCA-1*, and genes involved in leukocyte, myeloid cell and neutrophil activation were expressed in the patient's IMA, but lacked in the reference cohort (Figure 3A,B,C). Thirty seven genes revealed opposite expression. Specifically, in contrast to IMA signature cases, in the patient's IMA 4/37 genes, *Serpins Family H Member 1 (SERPINH1)*, *SRY-Box 4 (SOX4)*, *Hes Family BHLH Transcription Factor 1 (HES1)* and *Immediate Early Response 3 (IER3)* were downmodulated, whereas 33/37 genes, including *HLA-DOA*, *CD74* and *Class II*

Major Histocompatibility Complex Transactivator (CIITA) were upregulated (Figure 3D).

Discussion

The present clinical case stands for the very young age of the patient who developed a rare histotype of lung cancer with multifocal onset and bilateral lung involvement.

According to the 8th TNM staging system of lung cancer, the matching of morphological, immunohistochemical and molecular profiles between the two neoplastic nodules was consistent with the diagnosis of intrapulmonary metastasis

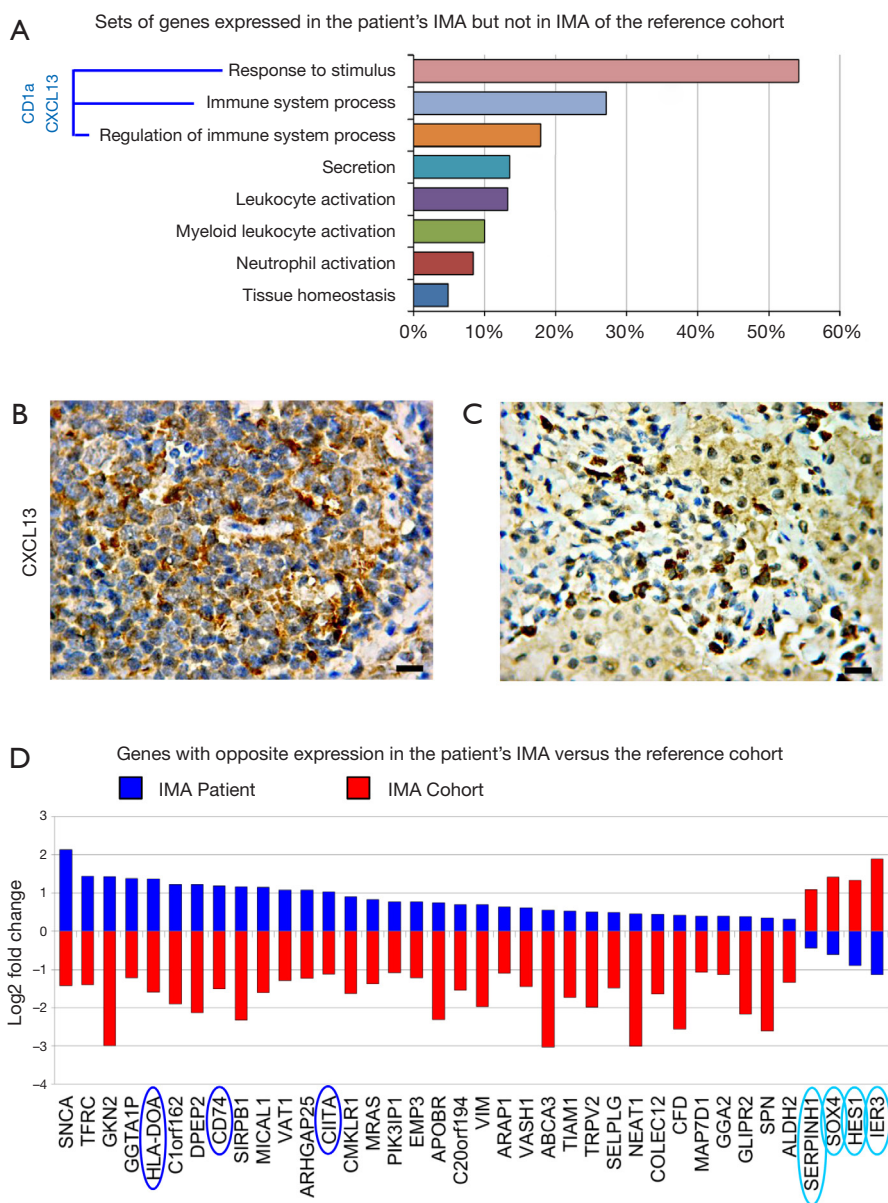


Figure 3 Comparative transcriptomic analysis of patient's IMA versus IMA of the reference cohort, performed using the Gene Ontology Consortium website (<http://geneontology.org>), and immunopathological analysis of patient's IMA. (A) Sets of genes expressed in the patient's IMA and unexpressed in IMA of the reference cohort. The functional categories of the *response to stimulus*, *immune system process* and *regulation of immune system process* were also screened using the DAVID and UniProt database (<https://www.uniprot.org/>). Gene transcripts of *CD1a* and *CXCL13/BCA-1* are the most represented in the functional categories of genes driving the *response to stimulus*, *immune system process* and *regulation of immune system process*. P = Bonferroni corrected P value. (B,C) Immunohistochemistry reveals a prominent expression of *CXCL13/BCA-1* both (B) in the context of lymphoid follicle-like structures (magnification: $\times 630$. Scale bar: 20 μm) and (C) in inflammatory cells infiltrating the patient's IMA (magnification: $\times 400$. Scale bar: 30 μm). (D) Identification of 37 genes showing opposite expression in the patient's IMA (blue) in comparison with IMA of the reference cohort (red). Thirty-three out of thirty-seven genes, including *HLA-DOA*, *CD74* and *CIITA* (circled in dark blue) were upregulated in the patient's IMA and downmodulated in IMA of the reference cohort. On the contrary 4/37 genes, *SERPINH1*, *SOX4*, *HES1* and *IER3* (circled in sky blue) were downmodulated in the patient's IMA and upregulated in IMA of the reference cohort.

and classified the tumor as pathologic stage T1bN0M1a, i.e., stage IVA (15). However, the possibility of multiple primary cancers cannot be excluded (16).

Genes driving the response to stimulus, which include the typical IMA markers *HNF4A*, *FOXA3*, *MUC5A* and *MUC5B* (11,17), and the response to chemicals were overexpressed in both the patient's tumor compared to the IMA signature, and the patient's normal lung compared to control lungs. This molecular trait may have been triggered by long term exposure to passive smoking (18-20) and chemical carcinogens (21,22), although an inherited predisposition to early onset lung cancer cannot be ruled out (23).

Among the functional categories of genes driving the response to stimulus, chemical and organic substances, the genes coding for *CXCL8/IL-8*, *ACKR1/DARC*, and the member of the RAS oncogene family *RAB7B* were the most represented in the normal lung tissue of our young patient.

The multifunctional cytokine *CXCL8/IL-8*, by interacting with its cognate receptors, C-X-C chemokine receptor 1 (*CXCR1*) and *CXCR2* mediates the initiation and development of various cancers, including lung cancer (24). Circulating *CXCL8/IL-8* may predict the risk of lung cancer, since it is upregulated prior to clinical diagnosis (25,26). The Lung cancer screening trial has shown that high expression of *CXCL8/IL-8* can raise the risk of lung cancer by 45–86% (25,26). *CXCL8/CXCR1-2* axis promotes the transactivation of *EGFR*, which results in Ras GTPase activation and NSCLC cell proliferation (27).

ACKR1 binds over 20 inflammatory chemokines belonging to the CC and CXC families (28). It is expressed by erythrocytes and endothelial cells lining small veins and venules. Recent evidences suggest that *ACKR1* expressed by erythrocytes acts as a chemokine buffer and can limit excessive leukocyte extravasation, whereas endothelial *ACKR1* increases neutrophil and monocyte extravasation and promotes acute and chronic inflammation (29).

RAB7B is a lysosome-associated small GTPase, that regulates vesicular traffic from early to late endosomal stages of the endocytic pathway (30). It has been reported that increased expression of *RAB7B*, caused by growth factor withdrawal in mammalian cells, induces the downregulation of nutrient transporters, which activate stress response pathways and trigger apoptosis (31). However, its function and role in cancer remain to be defined.

Among the genes driving the response to chemicals, *HOXC9*, *HOXD9*, *KLF5* and *NKX2-8* were the most prevalent, and were upregulated in the patient's normal lung when compared with age-matched healthy controls.

Both Homeobox transcription factors, *HOXC9* and *HOXD9*, have been associated with increased risk to develop cancer and have been described as proto-oncogenes (32), but deciphering the cancer context specific role of these transcription factor remains a challenge.

KLF5 is a member of the zinc finger class of DNA-binding transcriptional regulators that regulates cell growth, proliferation, differentiation, apoptosis and tumorigenesis in a number of systems. It has been reported to act as a proto-oncogene in the lung, by promoting lung tumorigenesis via *SOX4* up-regulation (33).

The function of *NKX2-8* in lung cancer is still unclear. Indeed, knockout of *NKX2-8* leads to proliferation of lung progenitor cells and widespread dysplasia in the large airways of mice (34), yet overexpression of this protein seems to enhance the tumorigenicity of malignant cell lines (35).

Co-activation of the *TTF-1* and *NKX2-8* pathways was discovered to confer resistance to cisplatin in lung cancer cell lines and was found to be associated with poor survival in NSCLC patients (36), however the meaning of *NKX2-8* overexpression in the normal lung tissue is to be defined.

Notably, genes driving immune responses were found unexpressed in the patient's normal lung versus healthy control tissues, whereas genes governing cell adhesion, subcellular organization and apoptosis were dramatically downregulated, suggesting a favorable soil for the loss of cell-cell interactions and aberrant cell proliferation out of immunological control.

Molecular analyses of the neoplastic tissue revealed mutation of the *KRAS* gene, which is a common driver aberration in IMA (6) and occurs in our patient as *KRAS codon 12 mutation (G12D)*. *KRAS* positive status significantly associates with IMA and *MUC5B* and *MUC5AC* expression (37). It has been demonstrated that a single endogenous mutant *KRAS* allele is sufficient to promote lung tumor formation, meanwhile malignant progression requires additional genetic alterations (38). *KRAS G12D* mutation, which induces a high-level of resistance to apoptosis and predisposition to anchorage-independent growth (39), may have synergized with the transcriptional alterations found in the patient's normal lung tissue resulting in tumor progression. The immunomodulatory potential of *KRAS* mutant cancer cells, which promotes the recruitment of M1 macrophages, neutrophils and myeloid-derived suppressor cells (40), likely predominates in the patient's IMA microenvironment compared to the reference cohort, as demonstrated by the overexpression of myeloid lineage-related genes. Genes

that regulate immune system processes and leukocyte activation, such as the dendritic cell marker *CD1a* and the chemokine *CXCL13/BCA-1* were also overexpressed. *CD1a* is a marker for myeloid DC, which are a major source of IL-12 and Th1 CD4⁺ T cell polarization (41). Structurally related to the major histocompatibility complex (MHC) proteins, *CD1a* appears to be crucial for the presentation of non peptide tumour antigens to the immune system (42). Moreover, the density of CD1a⁺DCs within human tumors has been associated with survival (43).

CXCL13/BCA-1 has a controversial role in lung cancer. It may be induced by carcinogens and favor cancer cell proliferation (44), but it has also been proven to be fundamental to the development of tumor-associated lymphoid structures, which are detected in the patient's tumor, and have been associated with a favorable prognosis in NSCLC (45).

Comparative transcriptomic analyses between the patient's IMA and the IMA signature cases also revealed a limited range of genes with opposite expression pattern, mostly with an unclear function. Notably, *HLA-DOA*, *CD74* and *CIITA*, encoding for MHCII molecules, which influence tumor antigen specific immune responses (46-48) were overexpressed in the patient's IMA and downregulated in the IMA signature cases, whereas stemness and pro-tumoral genes, such as *SOX4*, *HES1*, *IER3* and *SERPINH1* (49-52) were downregulated in the patient's IMA. This peculiar transcriptional profile is likely behind the favorable clinical course of the patient that, despite the 20.9 months of median survival for stage IV IMA (53).

The limits of this work lie in a transcriptional analysis that is confined to a single, albeit rare and unprecedented, clinical case. However, highlighting the value of both normal-like and cancer tissues in deciphering patient-specific tumor histogenesis, this study emphasizes the need for a personalized diagnostic and clinical management of the lung cancer patient.

Assessing the transcriptional profiles of both normal and neoplastic lung tissues provides fundamental information: (I) to define the case specific molecular pathways driving the natural history of a particular lung cancer histotype, and (II) to develop an accurate prognostic nomogram and patient-tailored treatment approach.

Beyond the long recognized clinical and prognostic value of the specific tumor histotype and stage, assessing the transcriptional signature of both normal and neoplastic lung tissues paves the way to personalized clinical treatment and outcome prediction.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tlcr-20-177>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). A written informed consent was obtained from the patient for the publication of this case report and for any images.

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