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COVID-19: Emergence of Infectious Diseases, Nanotechnology Aspects, Challenges, and Future Perspectives

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Wuhan, a city of China, is the epicenter for the pandemic outbreak of coronavirus disease-2019 (COVID-19). It has become a severe public health challenge to the world and established a public health emergency of international worry. This infectious disease has pulled down the economy of almost all top developed nations. The coronaviruses (CoVs) known for various epidemics caused time to time. Infectious diseases such as severe acute respiratory syndrome (SARS) and middle east respiratory syndrome (MERS), followed by COVID-19, are all coronaviruses led outbreaks that scourged the history of mankind. CoVs evolved themselves to more infectious, transmissible, and more pandemic with time. To prevent the spread

Introduction

In last two decades, entire world faced three major outbreaks of coronaviruses like Severe Acute respiratory syndrome (SARS), middle east respiratory syndrome (MERS) and novel coronavirus disease i.e., COVID-19. The first case of recent outbreak of COVID-19 was recognized at Wuhan, Hubei, China five months ago. Later, it spread to the whole world and affects health as well economy globally in a very short span. Although, the world is well equipped with technological advances and well aware of the complete structural details to deal with the

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of the SARS-CoV-2, many countries have ordered the complete lockdown to combat the outbreak. This paper briefly discussed the historical background of CoVs and the evolution of human coronaviruses (HCoVs), the case studies and the development of their antiviral medications. The viral infection encountered with present-day challenges and futuristic approaches with the help of nanotechnology to minimize the spread of infectious viruses. The antiviral drugs and their clinical advances, along with herbal medicines for viral inhibition and immunity boosters, are described. Elaboration of tables related to CoVs for the compilation of the literature has been adopted for the better understanding.

outbreak of coronavirus (CoVs), still all are struggling to find out its cure. Since ancient times, appearance or reappearance of several diseases have affected the human life significantly.^[1] From the early 1900s to date, around 250 viruses species have been evolved, and these keep on increases in the coming years (Figure 1).^[2]

Historical background

Starting from the tobacco mosaic virus (1892), foot-mouth disease virus (1898), yellow fever virus (1901) followed by a deadly virus created influenza epidemic during World War-I and took around 50 million lives. It is an all-time high in the mankind history^[2] and this virus probably one of the influenza virus species.^[3]

Coronavirus disease (COVID) was first reported in 1931, and the first coronavirus (HCoV-229E) isolated from humans in 1965. These infect various birds, animals, and mammals, including humans. The isolation of the prototype murine coronavirus strain of coronavirus was reported in 1949.^[4] The pathogenesis and replication mechanisms of various coronaviruses have been elaborated in details since the 1970s. Human coronaviruses (HCoVs) were first identified in 1960 and caused acute upper respiratory infection (URI).^[5] CoVs affected the humans worldwide and are the pathogenic agents for both mammals and avian. These viruses can infect respiratory, hepatic, central nervous, and gastrointestinal systems of humans, bats, birds, mice, and some wild animals.^[6] These viruses are highly pathogenic to humans. Previously, CoV causes an epidemic of SARS in humans and infected thousands



of people in Guangdong province of China, in 2002-2003, followed by MERS in Saudi Arabia in 2012.

CoVs are pleiomorphic, enveloped, with an average diameter of \sim 120 nm, and club-shaped surface projections 20 nm (glycosylated spike glycoprotein) as shown in Figure 2.^[7] These



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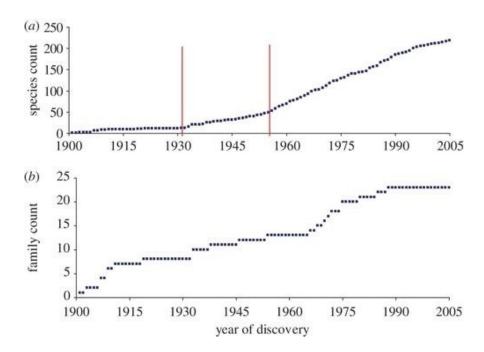


Figure 1. Discovery curves for human viruses. (a) Virus discovery curve by species. A cumulative number of species reported to infect humans. Statistically significant upward breakpoints are shown (vertical lines). (b) Virus discovery curve by family. A cumulative number of families containing species reported to infect humans. (Copyright permission © 2012 The Royal Society).^[2]

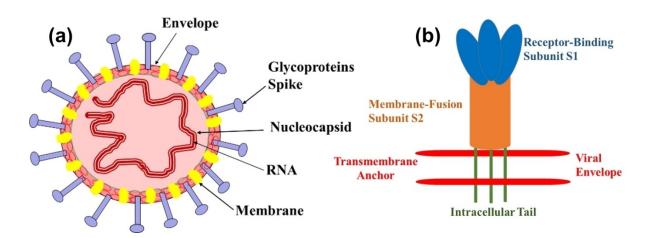


Figure 2. (a) General structure of CoVs and (b) Representation of the general structure of prefusion CoVs spikes.

viruses belong to family Coronaviridae, which shows crown-like appearances under an electron microscope. The CoVs contain large, positively charged strands of RNA associated with nucleocapsid phospholipid protein and the genetic material is surrounded by viral glycoproteins. The CoVs have three types of membrane proteins, which include the spike glycoprotein, membrane protein, and small hydrophobic membrane protein. In addition to these proteins, other proteins like hemagglutinin esterase (HE) has been isolated from a different group of coronaviruses which are not present in the SARS-CoV genome.^[8] The spikes of coronavirus cause infection in host cells. The structure of spike is clove shaped and has trimeric S1 head and S2 stalk, respectively (Figure 2b). During infection, S1 (head) of virus spike binds to host cell receptor for viral attachment and S2 (stalk) allow viral genome to enter into host cell through fusion of viral and host membrane surface.^[9]

Apart from these four principle structural proteins, various coronavirus also encodes some special structural and accessory proteins like 3a/b protein, 4a/b protein and hemagglutininesterase (HE) protein.^[10] The subgenomic RNAs (sgRNAs) of coronaviruses encodes all the structural and accessory proteins.^[11] The translation of coronavirus genomic RNA is controlled by the replicase-transcriptase protein genes. These replicase-transcriptase proteins encoded by two open-reading frames (ORF) namely, ORF1a and ORF1b, respectively. These two ORFs are synthesized two large polyproteins like pp1a and pp1ab. The first ORF is 2/3rd of the entire genome length from the 5'-terminal and they encode a pp1ab polyprotein.^[7b] The



pp1ab polyprotein further divide into 16 nonstructural proteins (1-16 nsps) except in the gamma-coronavirus lacks nsp1. Most of the nonstructural proteins are very important and plays critical role in coronavirus viral RNA synthesis. The functions of some nonstructural proteins are not reported so far. There are 16 nsps of coronavirus form Double-Membrane Vesicles (DMVs). Some nsps have hydrophobic transmembrane domains which acts as achor to the pp1a/pp1ab proteins with replication-transcription membranes during complexes formation.^[12] Interestingly, the nsps or viral replication processes are found to be potential antiviral drug targets which might be helpful in the development of drugs against coronavirus.^[13] These nsps regulate genome transcription and replication processes.^[7b,10] For example, nsp10 has role in viral replication/transcription complexes formation whereas the viral RNA replication and transcription are controlled by nsp12.^[14] It is essential to understand the function of all nsps for the development of promising drugs/vaccines against the COVID-19 pandemic. The function of 16 nsps of coronaviruses are shown in the Table 1 that helps to interpret CoVs for effective design of drugs.

Human coronavirus is spreading rapidly among people via viral respiratory infections like cough and sneeze. Coronavirus disease is most contagious and lethal. Various stains of coronaviruses have been reported so far (Table 2). The incubation periods and clinical symptoms of each coronaviruses are varying significantly. The symptoms and incubation periods of human coronaviruses play very important role in the identification and diagnosis of coronavirus diseases, shown in the Table 2.

The viral spike glycoprotein is dimer or trimer and has two functions: attaching the virus to receptor sites located on the host surface and activate the virion membrane to fuse with host membrane so that RNA genome released inside the host cell.^[38] It may be noted that coronaviruses attack inside the cell membrane rather than acting on the surface of host cells. CoVs genome is non-segmental, and the two-third portion consists of overlapping ORFs responsible for translating the polypro-

Table 1. Functions of non-structural proteins (nsps) of CoVs.					
nsps	Functions	References			
nsp1	Promotes degradation of cellular mRNA as well as blocks translation of host cell, obstructive distinctive immunity reaction, inhibit interferon (IFN) signals	[15]			
nsp2	Some functions are not known, holds to prohibitin proteins	[16]			
nsp3	Multi-domain large transmembrane protein, functions include: • N protein interact with Ac and Ub11 domains	[17]			
	 Cytokine expression promote due to ADRP activity Viral polyprotein cleaved by PLPro/Deubiquitinase domain which blocks host innate immune response Unknown function of NAB, SUD Ubl2, G2M, and Y domains 				
nsp4	Potential transmembrane scaffold protein, Role in (DMVs formation	[18]			
nsp5	chymotrypsin-like protease (3CLpro), main protease (Mpro), cleaves viral polypeptides, inhibit interferon (IFN) signals	[19]			
nsp6	Restrict expansion of autophagosome, Potential transmembrane scaffold protein, double-membrane vesicle (DMV) formation	[20]			
nsp7	Formation of hexadecameric complex (cofactor) with nsp8 and nsp12, role as a processivity clamp and primase for RNA polymerase	[21]			
nsp8	Forms hexadecameric complex (cofactor) with nsp7 and 12, may act as primase as well as processivity clamp for RNA polymerase	[21a,c, 22]			
nsp9	RNA binding protein, Dimerization	[23]			
nsp10	Stimulates 2-O-MT and ExoN activities, Scaffold protein for nsp14 and nsp16,	[24]			
nsp11	unknown functions	[14,15d]			
nsp12	Primer and RNA-dependent, RNA polymerase	[25]			
nsp13	RNA helicase, 5' triphosphatase	[26]			
nsp14	Exoribonuclease activity for viral genome proofreading, N7-Mtase activity adds 5' cap to viral RNAs, 3'-5' exoribonuclease,	[24b,27]			
nsp15	nsp15 endoribonuclease, evasion of dsRNA sensors	[28]			
nsp16	2'-O-Methyltransferase (2'-O-Mtase); avoiding MDA5 recognition, negative regulation of innate immunity	[24a,c, 29]			

Table 2. HCoVs and their clinical symptoms ^[30]						
HCoVs	Clinical Symptoms	Incubation Period	References			
HKU1	Fever, Cough, Respiratory Tract Illness (RTI), Sneezing, Dyspnea, Pneumonia	2–4 days	[31]			
229E	Fever, Chills, Cough, Acute Rhinorrhea, Malaise, Nasal Congestion and Discharge, Headache, Throat Sore	2–5 days	[32]			
OC43	Fever, Cold, Cough, Sputum, Dyspnea, Headache, Nasal Congestion and Discharge, Sneezing, Throat Sore	2–5 days	[33]			
NL63	Fever, Cold, Cough, Respiratory Distress, Wheeze, Rales, Rhinorrhea, Tachypnea, Hypoxia	2–4 days	[34]			
MERS-CoV	Fever, Cough, Cold, Shortness of Breath, Gastrointestinal Symptoms, Sore Throat, Arthralgia, Diarrhea and Vomiting, Pneumonia, Acute Renal Impairment, Multiple Organ Failures, Rapid Kidney Failure	2–13 days	[35]			
SARS-CoV	Fever, Cough, Cold, Rigor, Shortness Of Breath, Gastrointestinal Symptoms, Myalgias, 2–11 days Headache, Malaise, Dyspnea, Respiratory Distress, Diarrhea, Pneumonia		[35a,36]			
SARS-CoV-2 (COVID-19)	Fever, Coughing, Cold, Sore Throat, Nasal Congestion and Rhinorrhea, Diarrhea, Asympto- matic, Organ Function Damage, Acute Kidney And Cardiac Infection, Liver Dysfunction, Pneumothorax	2–14 days but in some cases extended to 24 days	[10,37]			



teins. Remaining ORFs of genomes contained for structural proteins. Depending upon the protein sequences CoVs are subcategorised into four genera (α -, β -, γ -, and δ -CoVs).^[39] The two genera α - and β - were primarily associated with rodents and bats for their genes source, although birds are the key reservoir for γ - and δ -CoVs. So far, there are seven human coronaviruses (HCoVs) reported in literature where two of the coronaviruses namely, SARS, MERS, OC43, HKU1, and SARS-CoV-2 are β -CoVs.^[40] However, past few years, studies are underway to explore these viruses and develop potential drugs to cure the people from the infection.^[7b,30, 32c, 33a, 41]

There are various natural hosts of coronaviruses have been reported so far. Natural hosts are mainly mammals, birds, dog, man, pig, cat, mouse, bat, mouse, tree sparrow, chicken, etc. (Table 3). These hosts are very important in the identification of specific coronavirus strains. Several coronaviruses are classified as species in the Coronaviridae.^[7a,42] A total of 37 coronaviruses and their natural hosts are shown in Table 3, which provides information about specific genera like α , β , δ and γ and acronym of each CoVs. Woo et al. analyzed 3306 birds species

in which only 1.1% of species found positive for CoV as mentioned in Table 3. $^{\rm [42b]}$

Bats and birds are reservoirs of distinct types of CoVs. The evolution model of HCoVs is shown in Figure 3.^[30] According to the evolutionary model (Figure 3), the origin of first CoVs might be in the bat and transfer to birds or vice-versa. Based on the Figure 3, the bat coronavirus family transfer the virus to other bat species and then, further transfers to mammals, including humans. Similarly, the bird CoVs transfer to another bird species and further transfer in birds and mammals like pigs and whale.^[15d]

Human coronaviruses

The human coronavirus first emerged in 1965 and was obtained from human embryonic tracheal culture from adults suffering from the common cold by Tyrell and Bynoe.^[43] Similar findings were carried out by Hamre and Procknow^[44] on common cold specimens and isolated a new category of virus known as 229E from WI-38 lung cell line culture. These two human respiratory viruses were sensitive to ethers and get multiplied in the presence of inhibitors of DNA synthesis.^[45]

	Table 3. Coronaviridae with their natural hosts, acronyms, and genere ^[7a,42] .							
S.No.	Natural Host	Virus	Acronym	Genere				
1	Dog	Canine coronavirus	CCV	α				
2	Dog	Canine enteric coronavirus	CECV	α				
3	Man	Human coronavirus 229E	HCoV-229E	α				
4	Man	Human coronavirus NL63	HCoV-NL63	α				
5	Cat	Feline infectious peritonitis virus	FIPV	α				
6	Pig, Swine	Porcine epidemic diarrhea coronavirus	PEDV	α				
7	Pig, Swine	Porcine hemagglutinating encephalomyelitis virus	HEV	α				
8	Pig	Porcine transmissible gastroenteritis virus	TGEV	α				
9	Pig	Porcine coronavirus	PorCoV-HKU15	α				
10	Bat	Rhinolophus bat coronavirus	HKU2	α				
11	Bat	Myotis bat coronavirus	HKU6	α				
12	Bat	Miniopterus bat coronavirus	HKU7	α				
13	Bat	Miniopterus bat coronavirus	HKU8	α				
14	Bat	Hipposideros and Rousettus bat coronavirus	HKU10	α				
15	Cattle	Bovine coronavirus	BCV	β				
16	Man	Human coronavirus OC43	HCV OC43	β				
17	Man	Human coronavirus HKU1	HCoV-HKU1	β				
18	Man	Middle East respiratory syndrome coronavirus	MERS-CoV	β				
19	Mouse	Murine hepatisis virus	MHV	β				
20	Puffin	Puffin coronavirus	PuCoV	β				
21	Bat	SARS-related Rhinolophus bat coronaviruse	SARSr–Rh-Bat HKU3	β				
22	Bat	Tylonycteris bat coronavirus	HKU4	β				
23	Bat	Pipistrellus bat coronavirus	HKU5	β				
24	Bat	Rousettus bat coronavirus	HKU9	β				
25	Man	Severe acute respiratory syndrome coronavirus	SARS-CoV	ß				
26	Chinese bulbul, Red-whiskered bulbul, Sooty-headed bulbul	Bulbul coronavirus	BuCoV HKU11	δ				
27	Grey-backed thrush	Thrush coronavirus	ThCoV HKU12	δ				
28	White-rumped munia, Chestnut munia	Munia coronavirus	MunCoV HKU13	δ				
29	Japanese white eye	White-eye coronavirus	WECoV HKU16	δ				
30	Tree sparrow	Sparrow coronavirus	SpCoV HKU17	δ				
31	Oriental magpie robin	Magpie robin coronavirus	MrCoV HKU18	δ				
32	Black-crowned night heron	Night heron coronavirus	NHCoV HKU19	δ				
33	Eurasian wigeon	Wigeon coronavirus	WiCoV HKU20	δ				
34	Common moorhen	Common moorhen coronavirus	CMCoV HKU21	δ				
35	Chicken, Fowl	Infectious bronchitis virus	IBV	γ				
36	Pheasant	Pheasant coronavirus	PhCoV	Ϋ́				
37	Turkey	Turkey coronavirus	TCV	Ý				

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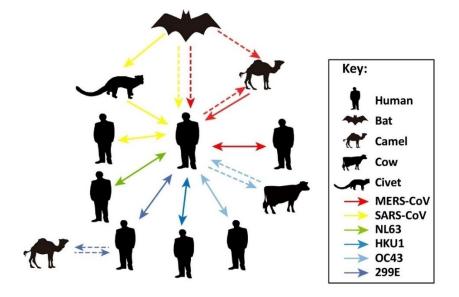


Figure 3. Intra- and Inter-Species Transmission of Human Coronaviruses. Red, yellow, green, blue, brown, and purple arrows represent the transmission of MERS-CoV, SARS-CoV, NL63, HKU1, OC43, and 229E, respectively, between bats, camels, cows, humans, and masked palm civets (shown in a legend on the side of the figure). Unbroken arrows represent confirmed transmission between the two species in question, and broken arrows represent suspected transmission (Copyright permission @ 2016 Elsevier Ltd.^[30]).

Later, McIntosh and his group at the NIH, Bethesda isolated six morphologically related viruses grown in organ culture, OC43 and OC38. The term "coronaviruses" described in 1968 based on crown-like projections on the surface for the new generation of viruses.^[46]

Human and animal coronaviruses were classified under three main categories, category-I has 229E and similar type of other viruses, category-II has OC43 and similar viruses, while in category-III, avian infectious bronchitis viruses (IBVs) were included.^[47] Both 229E and OC43 coronaviruses are impacting globally in the winter season, and incubation time is less than one weak.^[48] Another epidemic was caused in 2002 emerged from Guangdong Province, China caused by SARS-HCoV. The virus spread in almost 30 countries, and more than 7900 patients were reported to suffer from this viral infection. The SARS-HCoV was isolated from the Himalayan palm civets and was believed to originated from a live animal market in Guangdong Province, China.^[49] The worldwide spread of the virus was accompanied by the incubation period of 4-7 days and the highest viral load on the 10th day.^[50] Patients infected with this virus show myalgia, headache, fever, and respiratory difficulties. Severe conditions present with lymphopenia, liver disfunction, diffused alveolar damage, an increased number of macrophages.^[42e,51] The virus was re-emerged in late 2003 because of favourable condition available for amplification of virus and it was isolated from horseshoe bats.^[50]

The fourth human coronavirus strain, HCoV-NL63 was identified by Hoek and co-workers^[52] in Netherland and was isolated from seven months old baby in 2004. It was observed that about 4.7% of common respiratory problems in children due to HCoV-NL63.^[30] In 2004 again, fifth human coronavirus strain, HCoV-HKU1 was isolated by Patrick and co-workers^[53] from 71 years old man suffering from pneumonia and

bronchiolitis in Hong Kong. The old person was returned from Shenzhen, China. The virus causes acute asthmatic exacerbation and it was different from common pneumonia.^[22]

In Saudi Arabia 2012, MERS-HCoV (6th HCoV) reported in an old aged patient who suffered from acute pneumonia and renal failure. MERS-HCoV sequenced and resemble with SARS-CoV group and placed in the same category with a new lineage. MERS-HCoV, unlike SARS-CoV, observed with severe symptoms along with more epidemic and sporadic nature.^[54] Recently, emergence of 7th HCoV reported in late December 2019 from Wuhan of China. The pandemic outbreak of the novel coronavirus is most devastated to date resulting in 6.27 million infected and 379 thousands deaths so far until this paper was written up to 3rd June 2020. SARS-CoV-2, which shares 82% nucleotides similarities with SARS-HcoV..^[55] The novel coronavirus shows an incubation period of 2-14 days but in some cases extended to 24 days and spread rapidly through inanimate surfaces (metal, plastic, and glass) where it persists up to 9 days. The pandemic outspread can only be controlled through surface disinfestation procedures by applying ethanol (62-71%), 1-propanol (70%), 2-propanol (70%), H₂O₂ (0.5%), and sodium hypochlorite (0.1%).^[56]

Genome replication and transcription

Coronavirus enters into host cells through viral spike (S) glycoproteins.^[9b,57] Spike (S) contains two subunits, namely, S1 and S2. The S1 subunit contains bind to host cell receptors via the receptor-binding domain. Receptor binding is a crucial step to determine the host range for a coronavirus. Some human coronavirus has receptors like angiotensin-converting enzyme 2 (ACE2) for SARS-CoV and HCoV-NL63, aminopeptidase N (APN) for HCoV-229E, 9-O-acetylated sialic acid HCoV-HKU1,



and HCoV-OC43, and dipeptidyl peptidase 4 (DPP4) for MERS-CoV.^[58] In the case of novel COVID-19, the receptor-binding domain (RBD) of S protein has a strong affinity to angiotensinconverting enzyme 2 (ACE2) receptors of the bat and human.^[59] While S2 subunit comprises specialized fusion machinery that mediates fusion of viral and host cell membranes.^[9a,b, 57, 60] Later, S is cleaved at S2 of fusion peptide by proteases activities of the host in all coronaviruses.^[61] After entry, viruses get uncoated and start cap-dependent translation of ORF1a from viral genomic RNA to produce polyprotein pp1a, and further translation of ORF1b encodes longer polyprotein pp1ab.^[62] The pp1a and pp1ab produce 15-16 nonstructural proteins (nsps) through autoproteolytic cleavage. Significantly, nsp12 encodes the RNA-dependent RNA polymerase (RdRP) activity,^[25] whereas nsp3 and nsp5 encode papain-like protease (PLPro) and main protease (Mpro) activities, respectively.^[63] The nsp3, nsp4, and nsp6 from DMVs or spherules via also induce the rearrangement of the cellular membrane.^[20a,64] Further, the coronavirus replication transcription complex (RTC) is anchored and assembled at DMVs. The entire life cycle, replication and pathogenicity of SARS-CoV-2 is shown in Figure 4.^[58a,65]

Case studies

HCoVs have different symptoms for various host, and accordingly dissimilar disease caused as mentioned in Table 2. Dromedary camels (Saudi Arabia) were responsible for the spread of different HCoVs in the middle east, including South Korea (2015).^[30] Several countries such as the United States of America, the United Kingdom, Saudi Arabia, Kenya, France, China, and other countries are profoundly affected by different types of HCoVs before Wuhan 2019 outbreak (Table 4). In 2012, a novel β -CoV MERS was found in an old aged patient in Jeddah, Saudi Arabia suffered from pneumonia, renal failure, and respiratory infection who linked to Jordon. Later on, doctors and nurses were examined for the CoV, but they found negative, which proposed that it was not scattered easily. The MERS-CoV caused lower respiratory tract illness (RTI) in almost all over the globe, predominantly in the Middle East, and took

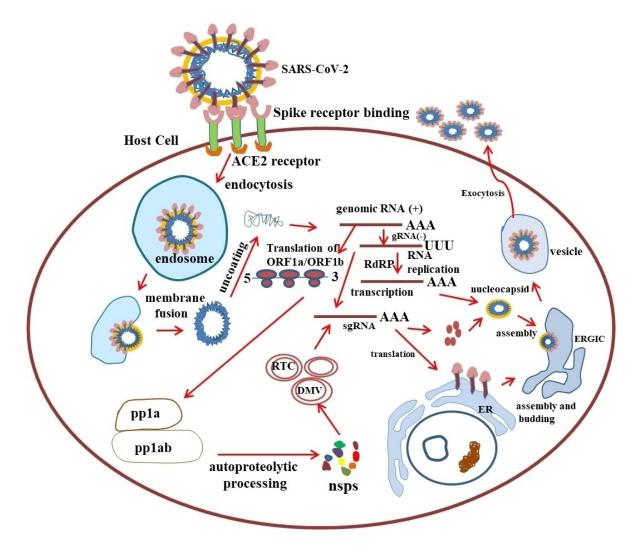


Figure 4. Modified schematic overview of SARS-CoV-2 life cycle in host cells. gRNA-genomic RNA; ERGIC-ER-Golgi intermediate complex; sgRNA- subgenomic RNA; RTC-replication transcription complexes; RdRP- RNA-dependent RNA polymerase; nsps-non-structural proteins.



	Table 4. Case study of HCoVs in several countries.							
S.No.	Country	Coronaviruses strains	Year	Effect	Ref.			
1	Chicago, USA	HCoV 229E	1962	Respiratory infections among medical students at the University of Chicago	[5a]			
2	Netherlands	HCoV-NL63	2002- 2003	Replicated faster in monkey kidney cells as compared to HCoV 229E and OC43, matched with porcine epidemic diarrhea virus and HCoV 229E	[42c,52]			
3	Frankfurt, Germany	SARS-CoV	2003	The patient traveled Singapore, Hong Kong, New York and During a stopover in Frankfurt, Germany on day 7	[67]			
4	Hong-Kong	MERS-CoV, SARC-CoV	2003	SARS-CoV behind the outbreak of severe respiratory infection in China in this period	[51,68]			
5	Shenzhen, China	SARS-CoV, NL63, HKU1	2003- 2004	Isolation of SARS-CoV from civet cats, the study revealed that it might be originated from animals	[69]			
6	Kenya	HCoV 229E, NL63, OC43, HKU1	2009– 2012	Out of 417 samples, 2.4% HCoV-NL63, ~ 3% OC43, ~ 2% HKU1, 1% 29E. CoVs are spreaded globally	[41a]			
7	Jordan, Sau- di Arabia	MERS-CoV	2012	Relatives are bat coronaviruses HKU4 and HKU5, 60 years old man died due to renal failure and respiratory infection linked to Jordon and SA	[42d,66]			
8	UK	The origin of this novel coronavirus is unknown	2012	Severe respiratory infection, the patient traveled from Qatar and Saudi Arabia	[70]			
9	France	HCoV-OC43	2001- 2013	Genomic studies revealed to understand the dynamics of evolution of CoVs	[71]			
10	USA	HCoVs OC43, NL63, 229E, HKU1	2014- 2017	Out of 854575 cases; 2.2% OC43, 1.0% NL63, 0.8% 229E, 0.6% HKU1. Cases inceased in between December and March	[40a]			

356 lives by February 2015.^[42d,66] Sipilwa et al. have carried out the study to find and describe HCoV strains spreading all regions of Kenya and explored the epidemiological significance of human coronaviruses.^[41a]

Drugs and vaccines in the current scenario

Presently, the entire world is fighting againt COVID-19 pandemic which is caused by SARS-CoV-2. This disease was first reported in the Hubai, China, in December 2019 and spreaded throughout the globe in a very short time.^[1,37a] According to "WHO" as of 3rd June 2020, the total number of confirmed COVID-19 cases is approximately 6.27 million and 379 thousand deaths throughout the 216 countries in the world. At present, there is no effective drug available in the market to control the spread of COVID-19. The researchers are working for the development of therapeutic drugs to treat infections.

In silico methods offer a way to methodically and rapidly yield additional repurposing candidates. Three-dimensional structure for the main protease of SARS-CoV-2 in complex with N3 (PDB ID: 6LU7) available on the RCSB protein databank is being use in insilico study. Bioinformatics analysis is used to virtually screen the molecules to get the promising candidate against the main protease of SARS-CoV-2. A recent study published, relied on this approach, using the predicted structure of all SARS-CoV-2 proteins based on their homology with other known coronavirus protein structures, and identified several compounds with potential antiviral activity.^[41d,72] Since the outbreak of SARS-CoV in 2003 and MERS-CoV during 2005, evoked scientists to explore and elaborate the HCoVs.^[7b,40b] Complete information at molecular level of any disease causing agents (virus/bacteria) may helpful in the development of the medicine. There are many approved drugs to cure the patients suffering from different diseases. These durgs are approved based on the efficacy and safety (pharmacology, formulation and toxicity) of medicines to humans. Subsequently, then many therapeutic and defensive agents have been discovered. More than 500 patents have been filed to combat these viruses in various authorities worldwide, so far.^[73] The treatment regimen of this epidemic appearance hard as the world is eyeing for the finding of effective drugs and vaccines. The pioneer of the pathophysiology of COVID-19 sacked the world to discover the drug/antiviral.^[37a,41d, 74] Many of the broad-spectrum antiviral drugs are in the trial stage at different levels to combat the effect of COVID-19 by blocking the RNA polymerase, RdRp.^[73,75] Some of the drugs are in trial stages such as quinine, chloroquine, lopinavir, ritonavir, alferon LDO, poly-ICLC, streptokinase, arbidol and glucocorticoid alone or in a combination of hydroxychloroquine, ritonavir/ribavirin, ASC09/ritonavir, daunavir/lopinavir, interferon α -2b/ribavirin, camrelizumab/thymosin, etc.^[73,75a, 76] Till date, none of the compounds is approved drugs for curing from COVID-19 infection in satisfactory way. In most of the countries on the trial basis, hydroxychlroquine with azithromycin have been used to cure the infected people. Further, it is reported that hydroxychlroquine is less effective when administered alone than in combination with the azithromycin. Although, hydroxychlroquine and azithromycin have toxicity to heart and hydroxychloroquine showed toxic effect on eye.[39,77]

A biological preparation provides active acquired immunity against particular infectious disease like COVID-19. It contains an agent that is similar to virus or microorganism caused disease. There are many vaccines are under preparation and at developmental stages throughout the world. Many vaccines are at first stage and few are waiting for human trial very soon. Till date, eight vaccines are under clinical trials throughout the world. These are Adenovirus Type 5 Vector, ChAdOx1, DNA plasmid vaccine with electropolation,^[78] Inactivated, Inactivated + alum, mRNA and LNP-encapsulated mRNA and these are developed by CanSino Biological Inc./Beijing Institute of Biotechnology, University of Oxford, Inovio Pharmaceuticals, Wuhan Institute of Biological Products/Sinopharm, Beijing



Institute of Biological Products/Sinopharm, Sinovac, BioNTech/ Fosun Pharma/Pfizer and Moderna/NIAID, respectively.^[79]

Nanotechnology: viral inhibition and recognition with potent nanomedicines

Still, we are strugling to find out rapid detection kit, effective antiviral and drug to make potent theranostics against SARS-CoV-2. The peripheral protein sequence adaptation and genome modification of virus lead to the resistance toward conventional therapies. With the advent of nanotechnology in recent years, nanomedicine have emerged to be an alternative for conventional therapeutics.^[80] Professor Chan emphasis on the development of nanotechnological tools as best weapon to deal with COVID-19 and focussed on rapid point-of-care diagnostics, surveillance and monitoring, therapeutics and vaccine development. Identification, treatment and prevention using nanotechnology tools would escalate the development to fight against COVID-19 as frontline tools.^[81] Kerry et al. explored the nano-based technique to combat NIPAH virus and a similar method may be explored to combat HCoVs where nanosystems could be developed and used to treat viral diseases.^[80,82] In the last few years, nanomaterials have also been investigated against various viral strains, and promising results were attained. A prominent approach, development of iron-based magnetic nanoparticles (IMNPs) as nanomedicine would help immensely to deal with COVID-19.^[80] The movement of IMNPs can be controlled by the external magnetic field and targeted to affected organs invaded by COVID-19, i.e., lung, throat, etc. without going to other organelles, as shown in Figure 5.^[80] IMNPs alone could interact with the nanovirus, COVID-19^[83] as of similar size regime and destroy or inhibit it. Additionally, in combination with photodynamic therapy, it deforms the structure of nanovirus and inhibit the growth.^[83] Except magnetic NPs, other nanomedicine can be utilized as antiviral agents such as Ti-based NPs known for influenza virus,^[84] carbon-based NPs elaborated for ebola, respiratory syncytial virus, etc.^[85] Inhibition of norovirus and pseudorabies virus through viral replications or by attacking histo-blood group antigens using Carbon dots,^[86] graphene oxides based nanomaterials preventing viral spread (coronavirus, respiratory syncytial virus) on the damaging virus by interacting with negative charge present over it^[87] and similarly other NPs such as Se NPs, solid lipid NPs, Ag NPs, Au NPs, etc. are also active in fighting against viruses.^[88] Huy et al. reported the electrochemical synthesis of AgNPs and explored their activity against non-enveloped viruses. The concentration of AgNPs up to 100 ppm or less was not cytotoxic, while antiviral activities were found in 3.13 ppm for 30 minutes of incubation with poliovirus.^[89] Viruses inactivation were carried out with different nanoparticles such as TiO₂ NPs,^[84b] Au NPs,^[88d] and Silica NPs.^[90] The composite of various nanomaterials also exhibits promising results that may comprise the delivery of target species through nanocarrier. The nucleic acid-based antiviral drugs face problems of cell penetration. The fragments of nucleic acid, therefore, introduced with the help of nanomaterials. Such an attempt was made by Levina et al., where they prepared titanium oxide nanoparticles and polylysine containing oligonucleotides (TiO₂-PL-DNA) and used to target the influenza A virus (IAV).^[84c] The authors reported (-) RNA strands as more sensitive for IAV segment 5. However, IMNPs have great advantageous here over other nanomedicine and drug because of less toxicity.

Another strategy could be the development of nanobiosensor to recognize and neutralize the epidemic of virus based on earlier development and detection of coronavirus. Nanosensor would open new possibilities to stop the outbreak of HCoVs by identifying the molecular species in real-time. These sensors, based on semiconductors or hybrid nanomaterials, show rapid identification of the virus even at ultra-

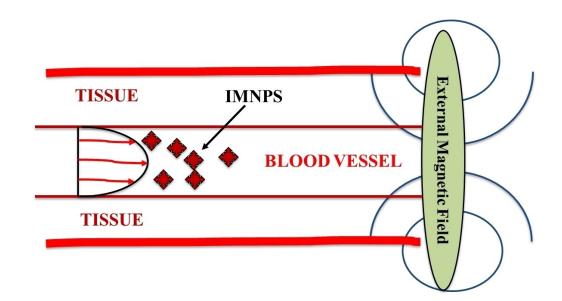


Figure 5. Diagrammatic illustration of IMNPs based drug delivery system.



low concentration.^[82,91] CdTe quantum dots (CdTe QDs) mixed with sandwiched complexes, NH₂-reporter, and biotin receptor and target DNA accelerating the research for developing a sensor for human T-lymphotropic virus-1 (HTLV-1) identification.^[92] Au NPs based materials are well known for the detection of various viruses such as dengue virus, influenza virus, bovine viral diarrhea coronavirus, etc.^[93] Recently, Seo et al.^[94] reported the field-effect transistor (FET) based biosensor for SARS-CoV-2. The novel biosensor was fabricated by immobilizing the SARS-CoV-2 spike antibody on the graphene surface through 1-pyrenebutyric acid N-hydroxysuccinimide ester (PBASE) as probe linker. The FET based biosensor exhibited a good response for clinical samples obtained from nasopharyngeal swabs and cultured SARS-CoV-2 with a limit of detection of 1 fg/mL. Further, thiol modified antisense oligonucleotides capped AuNPs showed better response and sensitivity against RNA sequence of SARS-CoV-2 (0.18 ng/µL virus load) by modification in surface plasmon resonance behavior.^[95]

Role of medicinal plants and herbs in the treatment of COVID-19

Till today, there is no single vaccine developed for effective treatment of deadly coronavirus disease COVID-19. On exposure to the deadly virus, the recovery rate of the patient mainly depends upon the strength of the immune system. According to WHO, 80% population of the developing country make use of herbal plants for successful treatment of various disease as well as for boosting up the immunity of individuals. The best preventive measures under practice to minimize the rate of infection includes primarily controlling the source of infection along with maintaining proper hygiene, avoiding crowded places, isolation, early diagnosis, and supportive treatments (boosting the immune system using herbal medicine). The herbal drugs are designed using specific parts of plants (roots, stem, bark, flowers, seeds, and fruits) to increase the resistance against the emerging and re-emerging viruses and bacteria (used in Ayurveda, as mentioned in Charaka Samhita and Susruta Samhita). Various plants produce beneficial ingredients such as oregano oil, fennel, garlic, peppermint, echinacea, sambucus, licorice, astragalus, ginger, tulsi (ocimum tenuiflorum), turmeric, etc. which contain different flavones, alkaloids, polyphenols, etc. These substances release interferons and antibodies which play an essential role to fight against various viral disease. It also increases the quantity of phagocytosis that protects the body from harmful particles. One of the herbal drugs 'fifatrol', a natural antibiotic, is a combination of vital phytoconstituents, immunomodulatory and antioxidants. It is advantagenous against viruses, allergens, and bacteria, thereby providing multi symptoms relief.

The essential oil of garlic is a source of organosulfur compounds and allicin is a reactive sulfur species found in it. The organosulfur in the essential garlic oil inhibit the ACE2 (host-receptor site of the virus) and main protease of the virus as well as to treat the infection due to SARS-CoV-2. Docking simulation and synergistic interactions show that the essential garlic oil has the ability to inhibit ACE2 and resist CoV-19.^[96] This oil is also beneficial in treatment of antithrombotic, hypoglycemia, hypotension, and immunomodulation.

Natural products present in common vegetables are explored due to its tendency to effectively bind the active sites of COVID-19 protease and hinder the viral replication. Turmeric (Curcuma longa), coriandrin, apple peels (ursolic acid), cucurbit vegetables (hederagenin), olive oil (oleanolic acid), rosemary, mint family plants (sageone), red pepper (apigenin), glycyrrhiza glabra (glabridin) are some of the natural compounds that have better insight into the mode of inhibition of viruses and have the potential for further research.^[97] Further, Anthocyanins represent a class of glycosylated polyphenol, that can act as a potential therapeutic antiviral compounds. It produces anthocyanidins whose nanoformulation are used in drug delivery systems and can reveal antiviral effects through a number of ways, i.e., by binding to host cells, inhibiting viral life cycle, or stimulating host immunity.^[98] Arundina gramnifolia belongs to Orchid family and produces gramniphenol (orange gum) and others phenolic compounds that can unveil anti-tobacco mosaic virus activity, in addition to anti-HIV-1 activity.^[99] Codiaeum peltatum bark extract is used as an anti viral, attributed to its structural novelty and antiviral potential against zika, chikungunya and dengue.^[100] Similarly, zingiber officinale (ginger) was used as an antiviral for the treatment of chikungunya virus.^[101]

The anti-infectives is highly dependent on natural products and their structures. Combinatorial chemistry techniques have emerged as a successful approach for optimizing structures and have been used effectively in the optimization of various drugs.^[102] Few natural products that can act as starting materials for host-targeting antiviral drugs show a broad spectrum in treating antiviral problems. Tannins that are normally found in plants acts as antimicrobial secondary metabolites. Hydrolysable tannins i.e. chebulagic acid and punicalagin possess inhibitory mechanism against bacteria, viruses, and eukaryotic microorganisms.^[103] Mycalamide is used for treatment of polio, HSV-1 Influenza and CoV-A59 (Mice survival 14 days after A59 CoV infection under 0.1 mg/kg of 2% mycalamide A) and Griffithsin exhibits inhibitory effects against SARS-CoV virus.^[103] Finally, there is a urgent need to develop natural antiviral medicines which can possess high chemical diversity and biochemical specificity to control coronavirus pandemics in future.

Future challenges and perspectives

 Controlling and development of vaccines to prevent the outbreak of infectious diseases such as COVID-19, needs fast action. Till date, no efficient vaccine/drug for its treatment is developed. Clinically patients treatment is mainly supported by emphasizing on the strength of their immune systems. In case of respiratory failure, patients are kept in the intensive care unit. Various nonspecific drugs such as hydroxychloroquine, ribavirin, lopinavir etc. are used for treating COVID-19; however, their actual effectiveness is still not apparent.



- Since nanotechnology has improvised therapeutic advancements in recent years and is an advanced combating tool in drug designing as well as drug targeting. Scientists could be encouraged towards the use of nanomaterials for targeting viral structures and depriving the impact of such novel viral infections.
- The traditional diagnostic methods for viral disease sometimes are bulky, expensive and require skilled staff to operate, as it requires cell culture and sample preparation and thus, they are limited to centralized laboratories. But, the application of nanosensors, new tools offers massive advantages in the diagnosis of viral diseases; it requires only suitable nanoparticles. The development of nanosensors will be beneficial due to low cost, rapid detection tool and can help in the epidemics such as COVID-19. Moreover, the existing toxicity evaluations of the NPs, though, cannot eliminate health risks.
- To encounter the pandemic situation in the current scenario is herculean. However, it could be possible by undergoing strong preventive major like social distancing, avoiding crowded places, wearing a good quality mask, maintaining personal hygiene like washing hands regularly, and early diagnosis of infection is of utmost crucial if symptoms of disease appear.
- The herbal drugs designed using specific parts of plants (roots, stem, bark, flowers, seeds, and fruits) increase the resistance against the emerging and re-emerging viruses and bacteria (used in Ayurveda, as mentioned in Charaka Samhita and Susruta Samhita). Oregano oil, fennel, garlic, peppermint, echinacea, sambucus, licorice, astragalus, ginger, tulsi, turmeric, etc. have flavinoids, alkaloids, polyphenol's plays an essential role against various viral infections. These herbs release interferons and antibodies against viruses. It also increases the quantity of phagocytosis that protects the body by harmful particles. One of the herbal drugs 'fifatrol' natural antibiotic combination of vital phytoconstituents, immunomodulatory and antioxidants is helpful against viruses, allergens, and bacteria, thereby providing multi symptoms relief.^[104]
- Since COVID-19 outbreak is a global concern, therefore, confronting similar future situations needs uncompromising regulations. Zoonotic origin and genetically crossed species barriers need to be seriously encountered through the scientific database. Since 2019-nCoV probably originated from bats and there are many future chances to get more viruses like this, which may prove more deadly and transmissible. Therefore, scientists must take fast action to recognize the origin of this deadly virus, discover active and safe drug and therapeutics to prevent its escalation and to combat any upcoming pandemic.

Conflict of Interest

The authors declare no conflict of interest.

Keywords: SARS-CoV-2 · Antiviral agents · Coronavirus · Herbal · Biosensors

- L. Zhang, D. Lin, X. Sun, U. Curth, C. Drosten, L. Sauerhering, S. Becker, K. Rox, R. Hilgenfeld, *Science* 2020, *368*, 409–412.
- [2] M. Woolhouse, F. Scott, Z. Hudson, R. Howey, M. Chase-Topping, Philos. Trans. R. Soc. London Ser. B 2012, 367, 2864–2871.
- [3] a) M. Billings, Web page, June 1997; b) S. Riley, Proc. Natl. Acad. Sci. USA 2014, 111, 7892–7893.
- [4] a) F. S. Cheever, J. B. Daniels, A. M. Pappenheimer, O. T. Bailey, J. Exp. Med. 1949, 90, 181; b) O. T. Bailey, A. M. Pappenheimer, F. S. Cheever, J. B. Daniels, J. Exp. Med. 1949, 90, 195–212.
- [5] a) D. Hamre, J. J. Procknow, Proc. Soc. Exp. Biol. Med. 1966, 121, 190– 193; b) E. Kendall, M. Bynoe, D. Tyrrell, Br. Med. J. 1962, 2, 82.
- [6] a) L. F. Wang, Z. Shi, S. Zhang, H. Field, P. Daszak, B. T. Eaton, *Emerging Infect. Dis.* **2006**, *12*, 1834; b) X. Y. Ge, J. L. Li, X. L. Yang, A. A. Chmura, G. Zhu, J. H. Epstein, J. K. Mazet, B. Hu, W. Zhang, C. Peng, *Nature* **2013**, *503*, 535–538; c) Y. Chen, D. Guo, *Virol. Sin.* **2016**, *31*, 3–11.
- [7] a) D. Cavanagh, Coronaviruses with Special Emphasis on First Insights Concerning SARS, Springer, 2005, pp. 1–54; b) J. Cui, F. Li, Z. L. Shi, Nat. Rev. Microbiol. 2019, 17, 181–192; c) F. Li, Annu. Rev. Virol. 2016, 3, 237– 261.
- [8] D. A. Brian, B. G. Hogue, T. E. Kienzle, *The Coronaviridae*, Springer, 1995, pp. 165–179.
- [9] a) R. N. Kirchdoerfer, C. A. Cottrell, N. Wang, J. Pallesen, H. M. Yassine, H. L. Turner, K. S. Corbett, B. S. Graham, J. S. McLellan, A. B. Ward, *Nature* 2016, *531*, 118–121; b) A. C. Walls, M. A. Tortorici, B.-J. Bosch, B. Frenz, P. J. Rottier, F. DiMaio, F. A. Rey, D. Veesler, *Nature* 2016, *531*, 114–117; c) D. R. Beniac, A. Andonov, E. Grudeski, T. F. Booth, *Nat. Struct. Mol. Biol.* 2006, *13*, 751–752.
- [10] N. Chen, M. Zhou, X. Dong, J. Qu, F. Gong, Y. Han, Y. Qiu, J. Wang, Y. Liu, Y. Wei, *The Lancet* 2020, 395, 507–513.
- [11] S. Hussain, Y. Chen, Y. Yang, J. Xu, Y. Peng, Y. Wu, Z. Li, Y. Zhu, P. Tien, D. Guo, *J. Virol.* **2005**, *79*, 5288–5295.
- [12] S. G. Sawicki, D. L. Sawicki, S. G. Siddell, J. Virol. 2007, 81, 20–29.
- [13] A. Zumla, J. F. Chan, E. I. Azhar, D. S. Hui, K.-Y. Yuen, Nat. Rev. Drug Discovery 2016, 15, 327.
- [14] S. G. Fang, H. Shen, J. Wang, F. P. Tay, D. X. Liu, Virology 2008, 379, 175–180.
- [15] a) C. Huang, K. G. Lokugamage, J. M. Rozovics, K. Narayanan, B. L. Semler, S. Makino, *J. Virol.* 2011, *85*, 638–643; b) W. Kamitani, C. Huang, K. Narayanan, K. G. Lokugamage, S. Makino, *Nat. Struct. Mol. Biol.* 2009, *16*, 1134; c) T. Tanaka, W. Kamitani, M. L. DeDiego, L. Enjuanes, Y. Matsuura, *J. Virol.* 2012, *86*, 11128–11137; d) A. R. Fehr, S. Perlman, *Coronaviruses*, Springer, 2015, pp. 1–23.
- [16] a) R. L. Graham, A. C. Sims, S. M. Brockway, R. S. Baric, M. R. Denison, J. Virol. 2005, 79, 13399–13411; b) C. T. Cornillez-Ty, L. Liao, J. R. Yates, P. Kuhn, M. J. Buchmeier, J. Virol. 2009, 83, 10314–10318; c) M. J. Gadlage, R. L. Graham, M. R. Denison, J. Virol. 2008, 82, 11964–11969.
- [17] a) J. Lei, Y. Kusov, R. Hilgenfeld, *Antiviral Res.* 2018, *149*, 58–74; b) P. Serrano, M. A. Johnson, A. Chatterjee, B. W. Neuman, J. S. Joseph, M. J. Buchmeier, P. Kuhn, K. Wüthrich, *J. Virol.* 2009, *83*, 12998–13008; c) B. W. Neuman, J. S. Joseph, K. S. Saikatendu, P. Serrano, A. Chatterjee, M. A. Johnson, L. Liao, J. P. Klaus, J. R. Yates, K. Wüthrich, *J. Virol.* 2008, *82*, 5279–5294.
- [18] a) M. A. Clementz, A. Kanjanahaluethai, T. E. O'Brien, S. C. Baker, Virology 2008, 375, 118–129; b) M. J. Gadlage, J. S. Sparks, D. C. Beachboard, R. G. Cox, J. D. Doyle, C. C. Stobart, M. R. Denison, J. Virol. 2010, 84, 280–290; c) D. C. Beachboard, J. M. Anderson-Daniels, M. R. Denison, J. Virol. 2015, 89, 2080–2089.
- [19] a) Y. Lu, X. Lu, M. R. Denison, J. Virol. 1995, 69, 3554–3559; b) C. C. Stobart, N. R. Sexton, H. Munjal, X. Lu, K. L. Molland, S. Tomar, A. D. Mesecar, M. R. Denison, J. Virol. 2013, 87, 12611–12618; c) X. Zhu, L. Fang, D. Wang, Y. Yang, J. Chen, X. Ye, M. F. Foda, S. Xiao, Virology 2017, 502, 33–38; d) X. Zhu, D. Wang, J. Zhou, T. Pan, J. Chen, Y. Yang, M. Lv, X. Ye, G. Peng, L. Fang, J. Virol. 2017, 91, e00003–00017.
- [20] a) M. M. Angelini, M. Akhlaghpour, B. W. Neuman, M. J. Buchmeier, *mBio* 2013, 4, e00524–00513; b) M. Oostra, M. C. Hagemeijer, M. van Gent, C. P. Bekker, E. G. te Lintelo, P. J. Rottier, C. A. de Haan, J.

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Virol. **2008**, *82*, 12392–12405; c) E. M. Cottam, M. C. Whelband, T. Wileman, *Autophagy* **2014**, *10*, 1426–1441.

- [21] a) Y. Zhai, F. Sun, X. Li, H. Pang, X. Xu, M. Bartlam, Z. Rao, *Nat. Struct. Mol. Biol.* 2005, *12*, 980–986; b) R. N. Kirchdoerfer, A. B. Ward, *Nat. Commun.* 2019, *10*, 1–9; c) A. J. Te Velthuis, S. H. van den Worm, E. J. Snijder, *Nucleic Acids Res.* 2012, *40*, 1737–1747.
- [22] I. Imbert, J. C. Guillemot, J. M. Bourhis, C. Bussetta, B. Coutard, M. P. Egloff, F. Ferron, A. E. Gorbalenya, B. Canard, *EMBO J.* 2006, 25, 4933–4942.
- [23] a) M. P. Egloff, F. Ferron, V. Campanacci, S. Longhi, C. Rancurel, H. Dutartre, E. J. Snijder, A. E. Gorbalenya, C. Cambillau, B. Canard, *Proc. Natl. Acad. Sci. USA* 2004, *101*, 3792–3796; b) Z. Zeng, F. Deng, K. Shi, G. Ye, G. Wang, L. Fang, S. Xiao, Z. Fu, G. Peng, *J. Virol.* 2018, *92*.
- [24] a) E. Decroly, C. Debarnot, F. Ferron, M. Bouvet, B. Coutard, I. Imbert, L. Gluais, N. Papageorgiou, A. Sharff, G. Bricogne, *PLoS Pathog.* 2011, *7*; b) Y. Ma, L. Wu, N. Shaw, Y. Gao, J. Wang, Y. Sun, Z. Lou, L. Yan, R. Zhang, Z. Rao, *Proc. Natl. Acad. Sci. USA* 2015, *112*, 9436–9441; c) Y. Chen, C. Su, M. Ke, X. Jin, L. Xu, Z. Zhang, A. Wu, Y. Sun, Z. Yang, P. Tien, *PLoS Pathog.* 2011, *7*.
- [25] X. Xu, Y. Liu, S. Weiss, E. Arnold, S. G. Sarafianos, J. Ding, Nucleic Acids Res. 2003, 31, 7117–7130.
- [26] a) K. A. Ivanov, V. Thiel, J. C. Dobbe, Y. van der Meer, E. J. Snijder, J. Ziebuhr, *J. Virol.* 2004, *78*, 5619–5632; b) K. A. Ivanov, J. Ziebuhr, *J. Virol.* 2004, *78*, 7833–7838; c) W. Hao, J. A. Wojdyla, R. Zhao, R. Han, R. Das, I. Zlatev, M. Manoharan, M. Wang, S. Cui, *PLoS Pathog.* 2017, *13*, e1006474; d) A. O. Adedeji, H. Lazarus, *mSphere* 2016, *1*, e00235–00216.
- [27] a) L. D. Eckerle, M. M. Becker, R. A. Halpin, K. Li, E. Venter, X. Lu, S. Scherbakova, R. L. Graham, R. S. Baric, T. B. Stockwell, *PLoS Pathog.* 2010, 6; b) Y. Chen, H. Cai, N. Xiang, P. Tien, T. Ahola, D. Guo, *Proc. Natl. Acad. Sci. USA* 2009, *106*, 3484–3489.
- [28] a) K. Bhardwaj, J. Sun, A. Holzenburg, L. A. Guarino, C. C. Kao, J. Mol. Biol. 2006, 361, 243–256; b) X. Deng, M. Hackbart, R. C. Mettelman, A. O'Brien, A. M. Mielech, G. Yi, C. C. Kao, S. C. Baker, Proc. Natl. Acad. Sci. USA 2017, 114, e4251-E4260; c) L. Zhang, L. Li, L. Yan, Z. Ming, Z. Jia, Z. Lou, Z. Rao, J. Virol. 2018, 92, e00893–00818.
- [29] P. Shi, Y. Su, R. Li, Z. Liang, S. Dong, J. Huang, Virus Res. 2019, 265, 57– 66.
- [30] S. Su, G. Wong, W. Shi, J. Liu, A. C. Lai, J. Zhou, W. Liu, Y. Bi, G. F. Gao, *Trends Microbiol.* 2016, 24, 490–502.
- [31] a) T. P. Sloots, P. McErlean, D. J. Speicher, K. E. Arden, M. D. Nissen, I. M. Mackay, *J. Clin. Virol.* **2006**, *35*, 99–102; b) F. Esper, C. Weibel, D. Ferguson, M. L. Landry, J. S. Kahn, *Emerging Infect. Dis.* **2006**, *12*, 775.
- [32] a) J. O. Hendley, H. B. Fishburne, J. M. Gwaltney Jr, Am. Rev. Respir. Dis. 1972, 105, 805–811; b) A. Bradburne, M. Bynoe, D. Tyrrell, Br. Med. J. 1967, 3, 767; c) M. J. Mäkelä, T. Puhakka, O. Ruuskanen, M. Leinonen, P. Saikku, M. Kimpimäki, S. Blomqvist, T. Hyypiä, P. Arstila, J. Clin. Microbiol. 1998, 36, 539–542.
- [33] a) E. R. Gaunt, A. Hardie, E. C. Claas, P. Simmonds, K. E. Templeton, J. Clin. Microbiol. 2010, 48, 2940–2947; b) S. f Zhang, J. I Tuo, X. b Huang, X. Zhu, D.-m. Zhang, K. Zhou, L. Yuan, H.-j. Luo, B.-j. Zheng, K. Y. Yuen, PLoS One 2018, 13.
- [34] a) K. E. Arden, M. D. Nissen, T. P. Sloots, I. M. Mackay, J. Med. Virol. 2005, 75, 455–462; b) H. Hofmann, K. Pyrc, L. van der Hoek, M. Geier, B. Berkhout, S. Pöhlmann. Proc. Natl. Acad. Sci. USA 2005, 102, 7988–7993.
- [35] a) H. Momattin, K. Mohammed, A. Zumla, Z. A. Memish, J. A. Al-Tawfiq, *Int. J. Infect. Dis.* **2013**, *17*, e792–798; b) G. K. M. Goh, A. K. Dunker, V. Uversky, *PLoS Curr.* **2013**, *5*.
- [36] a) M. D. Christian, M. Loutfy, L. C. McDonald, K. F. Martinez, M. Ofner, T. Wong, T. Wallington, W. L. Gold, B. Mederski, K. Green, *Emerging Infect. Dis.* 2004, *10*, 287; b) M. J. Cameron, J. F. Bermejo-Martin, A. Danesh, M. P. Muller, D. J. Kelvin, *Virus Res.* 2008, *133*, 13–19.
- [37] a) X. Yang, Y. Yu, J. Xu, H. Shu, H. Liu, Y. Wu, L. Zhang, Z. Yu, M. Fang, T. Yu, *Lancet Respir. Med.* **2020**; b) Y. Xu, X. Li, B. Zhu, H. Liang, C. Fang, Y. Gong, Q. Guo, X. Sun, D. Zhao, J. Shen, *Nat. Med.* **2020**, 1–4.
- [38] G. Simmons, J. D. Reeves, A. J. Rennekamp, S. M. Amberg, A. J. Piefer, P. Bates, Proc. Natl. Acad. Sci. USA 2004, 101, 4240–4245.
- [39] L. S. Nguyen, C. Dolladille, M. D. Drici, C. Fenioux, J. Alexandre, J. P. Mira, J. J. Moslehi, D. M. Roden, C. Funck-Brentano, J. E. Salem, *Circulation* 2020.

- [40] a) M. E. Killerby, H. M. Biggs, A. Haynes, R. M. Dahl, D. Mustaquim, S. I. Gerber, J. T. Watson, *J. Clin. Virol.* **2018**, *101*, 52–56; b) M. E. Killerby, H. M. Biggs, A. Haynes, **2018**.
- [41] a) L. A. Sipulwa, J. R. Ongus, R. L. Coldren, W. D. Bulimo, *Virology journal* 2016, *13*, 18; b) S. R. Dominguez, C. C. Robinson, K. V. Holmes, *J. Med. Virol.* 2009, *81*, 1597–1604; c) H. K. Talbot, B. E. Shepherd, J. E. Crowe Jr, M. R. Griffin, K. M. Edwards, A. B. Podsiad, S. J. Tollefson, P. F. Wright, J. V. Williams, *Pediatr. Infect. Dis. J.* 2009, *28*, 682; d) L. Dong, S. Hu, J. Gao, *Drug Discoveries Ther.* 2020, *14*, 58–60.
- [42] a) D. Cavanagh, D. Brian, M. Brinton, L. Enjuanes, K. Holmes, M. Horzinek, M. Lai, H. Laude, P. Plagemann, S. Siddell, in Coronaviruses, Springer, 1994, pp. 255–257; b) P. C. Woo, S. K. Lau, C. S. Lam, C. C. Lau, A. K. Tsang, J. H. Lau, R. Bai, J. L. Teng, C. C. Tsang, M. Wang, J. Virol. 2012, 86, 3995–4008; c) R. A. Fouchier, N. G. Hartwig, T. M. Bestebroer, B. Niemeyer, J. C. de Jong, J. H. Simon, A. D. Osterhaus, Proc. Natl. Acad. Sci. USA 2004, 101, 6212-6216; d) A. M. Zaki, S. Van Boheemen, T. M. Bestebroer, A. D. Osterhaus, R. A. Fouchier, N. Engl. J. Med. 2012, 367, 1814–1820; e) K. K. To, I. F. Hung, J. F. Chan, K. Y. Yuen, J. Thorac. Dis. 2013, 5, S103; f) S. K. Lau, P. C. Woo, K. S. Li, Y. Huang, M. Wang, C. S. Lam, H. Xu, R. Guo, K. h Chan, B. j Zheng, Virology 2007, 367, 428-439; g) P. C. Woo, S. K. Lau, K. S. Li, R. W. Poon, B. H. Wong, H. w Tsoi, B. C. Yip, Y. Huang, K.-h. Chan, K. y Yuen, Virology 2006, 351, 180-187; h) P. C. Woo, Y. Huang, S. K. Lau, K.-Y. Yuen, Viruses 2010, 2, 1804–1820; i) S. K. Lau, R. W. Poon, B. H. Wong, M. Wang, Y. Huang, H. Xu, R. Guo, K. S. Li, K. Gao, K.-H. Chan, J. Virol. 2010, 84, 11385-11394.
- [43] D. A. Tyrrell, M. L. Bynoe, *Lancet* **1966**, *1*, 76–77.
- [44] D. Hamre, J. J. Procknow, Proc. Soc. Exp. Biol. Med.. 1966, 121, 190-193.
- [45] J. D. Almeida, D. A. Tyrrell, J. Gen. Virol. 1967, 1, 175–178.
- [46] D. A. Tyrrell, J. D. Almeida, C. H. Cunningham, W. R. Dowdle, M. S. Hofstad, K. McIntosh, M. Tajima, L. Y. Zakstelskaya, B. C. Easterday, A. Kapikian, R. W. Bingham, *Intervirology* **1975**, *5*, 76–82.
- [47] K. V. Holmes, M. M. C. Lai, Fields Virol. 2001, 3, 1075-1093.
- [48] K. McIntosh, J. H. Dees, W. B. Becker, A. Z. Kapikian, R. M. Chanock, Proc. Natl. Acad. Sci. USA 1967, 57, 933.
- [49] Y. Guan, B. Zheng, Y. He, X. Liu, Z. Zhuang, C. Cheung, S. Luo, P. Li, L. Zhang, Y. Guan, *Science* 2003, *302*, 276–278.
- [50] V. C. C. Cheng, S. K. P. Lau, P. C. Y. Woo, K. Y. Yuen, *Clin. Microbiol. Rev.* 2007, 20, 660–694.
- [51] J. Peiris, S. Lai, L. Poon, Y. Guan, L. Yam, W. Lim, J. Nicholls, W. Yee, W. Yan, M. Cheung, *Lancet* 2003, *361*, 1319–1325.
- [52] L. van der Hoek, K. Pyrc, M. F. Jebbink, W. Vermeulen-Oost, R. J. Berkhout, K. C. Wolthers, P. M. Wertheim-van Dillen, J. Kaandorp, J. Spaargaren, B. Berkhout, *Nat. Med.* **2004**, *10*, 368–373.
- [53] P. C. Y. Woo, S. K. P. Lau, C.-M. Chu, K.-h. Chan, H.-W. Tsoi, Y. Huang, B. H. L. Wong, R. W. S. Poon, J. J. Cai, W.-k. Luk, L. L. M. Poon, S. S. Y. Wong, Y. Guan, J. S. M. Peiris, K. y Yuen, *J. Virol.* **2005**, *79*, 884.
- [54] S. S. Lee, N. S. Wong, Int. J. Infect. Dis. 2015, 38, 65-67.
- [55] a) J. F. W. Chan, K. H. Kok, Z. Zhu, H. Chu, K. K. W. To, S. Yuan, K. Y. Yuen, *Emerg. Microbes Infect.* **2020**, *9*, 221–236; b) V. Surveillances, *China CDC Weekly* **2020**, *2*, 113–122.
- [56] G. Kampf, D. Todt, S. Pfaender, E. Steinmann, J. Hosp. Infect. 2020, 104, 246–251.
- [57] A. C. Walls, M. A. Tortorici, J. Snijder, X. Xiong, B. J. Bosch, F. A. Rey, D. Veesler, Proc. Natl. Acad. Sci. USA 2017, 114, 11157–11162.
- [58] a) T. S. Fung, D. X. Liu, Annu. Rev. Microbiol. 2019, 73, 529–557; b) Y. X. Lim, Y. L. Ng, J. P. Tam, D. X. Liu, Diseases 2016, 4, 26 (1-28).
- [59] W. Tai, L. He, X. Zhang, J. Pu, D. Voronin, S. Jiang, Y. Zhou, L. Du, Cell. Mol. Immunol. 2020, 1–8.
- [60] W. Song, M. Gui, X. Wang, Y. Xiang, PLoS Pathog. 2018, 14, e1007236.
- [61] a) J. K. Millet, G. R. Whittaker, Virus Res. 2015, 202, 120–134; b) I. G. Madu, S. L. Roth, S. Belouzard, G. R. Whittaker, J. Virol. 2009, 83, 7411–7421.
- [62] P. S. Masters, Adv. Virus Res. 2006, 66, 193-292.
- [63] J. Ziebuhr, E. J. Snijder, A. E. Gorbalenya, J. Gen. Virol. 2000, 81, 853– 879.
- [64] H. J. Maier, P. C. Hawes, E. M. Cottam, J. Mantell, P. Verkade, P. Monaghan, T. Wileman, P. Britton, *mBio* 2013, 4, e00801–00813.
- [65] M. A. Shereen, S. Khan, A. Kazmi, N. Bashir, R. Siddique, J. Adv. Res. 2020, 24, 91–98.



- [66] B. Hijawi, M. Abdallat, A. Sayaydeh, S. Alqasrawi, A. Haddadin, N. Jaarour, S. El Sheikh, T. Alsanouri, *East Mediterr Health J.* 2013, 19, S12-S18.
- [67] C. Drosten, S. Günther, W. Preiser, S. Van Der Werf, H.-R. Brodt, S. Becker, H. Rabenau, M. Panning, L. Kolesnikova, R. A. Fouchier, *N. Engl. J. Med.* 2003, 348, 1967–1976.
- [68] S. S. Chiu, K. Hung Chan, K. Wing Chu, S. W. Kwan, Y. Guan, L. L. Man Poon, J. Peiris, *Clin. Infect. Dis.* 2005, 40, 1721–1729.
- [69] P. C. Woo, S. K. Lau, C. M. Chu, K. H. Chan, H. W. Tsoi, Y. Huang, B. H. Wong, R. W. Poon, J. J. Cai, W. K. Luk, J. Virol. 2005, 79, 884–895.
- [70] A. Bermingham, M. Chand, C. Brown, E. Aarons, C. Tong, C. Langrish, K. Hoschler, K. Brown, M. Galiano, R. Myers, *Eurosurveillance* 2012, *17*, 20290.
- [71] N. Kin, F. Miszczak, W. Lin, M. A. Gouilh, A. Vabret, Viruses 2015, 7, 2358–2377.
- [72] a) Y. Zhou, Y. Hou, J. Shen, Y. Huang, W. Martin, F. Cheng, *Cell Discovery* 2020, 6, 1–18; b) C. Harrison, *Nat. Biotechnol.* 2020.
- [73] C. Liu, Q. Zhou, Y. Li, L. V. Garner, S. P. Watkins, L. J. Carter, J. Smoot, A. C. Gregg, A. D. Daniels, S. Jervey, ACS Cent. Sci. 2020, 6, 315–331.
- [74] D. Kumar, K. Kumari, A. Jayaraj, V. Kumar, R. V. Kumar, S. K. Dass, R. Chandra, P. Singh, J. Biomol. Struct. Dyn. 2020, 1–14 (https://doi.org/10.1080/07391102.2020.1752310); b) D. Kumar, K. Kumari, V. Kumar, Vishvakarma, A. Jayaraj, D. Kumar, V. K. Ramappa, R. Patel, V. Kumar, S. K. Dass, R. Chandra, P. Singh, J. Biomol. Struct. Dyn. 2020 (https://doi.org/10.1080/07391102.2020.1779131).
- [75] a) G. Li, E. De Clercq, *Nat. Rev. Drug Discovery* **2020**; b) R. Qiu, X. Wei, M. Zhao, C. Zhong, C. Zhao, J. Hu, M. Li, Y. Huang, S. Han, T. He, *MedRxiv* **2020**, https://doi.org/10.1101/2020.03.04.20031401.
- [76] a) S. C. Wu, Biotechnol. J. 2020, 15, 2000147; b) S. Caddy, British Medical Journal Publishing Group, 2020; c) T. T. Le, Z. Andreadakis, A. Kumar, R. G. Roman, S. Tollefsen, M. Saville, S. Mayhew, Nat. Rev. Drug Discovery 2020, 19, 305–306; d) L. Peeples, Proc. Natl. Acad. Sci. USA 2020, 117, 8218–8221; e) G. Yamey, M. Schäferhoff, R. Hatchett, M. Pate, F. Zhao, K. K. McDade, Lancet 2020, 395, 1405–1406; f) M. P. Lythgoe, P. Middleton, Trends Pharmacol. Sci. 2020, 41, 363–382; g) W. H. Organization, World 2020 (https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines); h) A. Talevi, C. L. Bellera, Expert Opin. Drug Discovery 2020, 15, 397–401.
- [77] T. J. Kamp, M. H. Hamdan, C. T. January, J. Am. Heart Assoc. 2020, 9, e016887.
- [78] a) Y. C. Kim, B. Dema, A. Reyes-Sandoval, *NPJ Vaccines* 2020, *5*, 1–3;
 b) M. S. Diamond, T. C. Pierson, *Cell Host Microbe* 2020, *27*, 699–703.
- [79] a) A. S. Espeseth, P. J. Cejas, M. P. Citron, D. Wang, D. J. DiStefano, C. Callahan, G. O'Donnell, J. D. Galli, R. Swoyer, S. Touch, *NPJ vaccines* **2020**, *5*, 1–14; b) K. M. Ella, V. K. Mohan, *Indian Pediatr.* **2020**, *57*, 407–410.
- [80] V. Kumar, A. K. Choudhary, P. Kumar, S. Sharma, Nanoscience & Nanotechnology-Asia 2019, 9, 64–78.
- [81] a) W. C. Chan, ACS Nano 2020, 14, 4, 3719–3720; b) B. Fadeel, K. Kostarelos, Nat. Nanotechnol. 2020, 15, 164; c) A. Gupta, S. Kumar, V. Kumar, Challenges for Assessing Toxicity of Nanomaterials in Biochemical Toxicology-Heavy Metals and Nanomaterials, IntechOpen, 2019.
- [82] R. G. Kerry, S. Malik, Y. T. Redda, S. Sahoo, J. K. Patra, S. Majhi, Nanomedicine: Nanotechnology, Biology and Medicine 2019, 18, 196– 220.
- [83] T. Y. Hu, M. Frieman, J. Wolfram, Nat. Nanotechnol. 2020, 15, 1–3.
- [84] a) N. Mazurkova, Y. E. Spitsyna, N. Shikina, Z. Ismagilov, S. Zagrebel'nyi, E. Ryabchikova, *Nanotechnol. Russ.* 2010, *5*, 417–420; b) V. I. Syngouna, C. V. Chrysikopoulos, *J. Colloid Interface Sci.* 2017, 497, 117–125; c) A. S. Levina, M. N. Repkova, E. V. Bessudnova, E. I. Filippova, N. A. Mazurkova, V. F. Zarytova, *Beilstein J. Nanotechnol.* 2016, *7*, 1166–1173.
- [85] a) J. Luczkowiak, A. Muñoz, M. Sánchez-Navarro, R. Ribeiro-Viana, A. Ginieis, B. M. Illescas, N. Martín, R. Delgado, J. Rojo, *Biomacromolecules*

2013, 14, 431–437; b) S. Dostalova, A. Moulick, V. Milosavljevic, R. Guran, M. Kominkova, K. Cihalova, Z. Heger, L. Blazkova, P. Kopel, D. Hynek, *Monatsh. Chem.* 2016, 147, 905–918.

- [86] a) T. Du, J. Liang, N. Dong, L. Liu, L. Fang, S. Xiao, H. Han, *Carbon* 2016, 110, 278–285; b) X. Dong, M. M. Moyer, F. Yang, Y. P. Sun, L. Yang, *Sci. Rep.* 2017, *7*, 1–10.
- [87] a) S. Ye, K. Shao, Z. Li, N. Guo, Y. Zuo, Q. Li, Z. Lu, L. Chen, Q. He, H. Han, *ACS Appl. Mater. Interfaces* 2015, *7*, 21571–21579; b) X. X. Yang, C. M. Li, Y. F. Li, J. Wang, C. Z. Huang, *Nanoscale* 2017, *9*, 16086–16092; c) Y. N. Chen, Y. H. Hsueh, C. T. Hsieh, D. Y. Tzou, P. L. Chang, *Int. J. Environ. Res. Public Health* 2016, *13*, 430.
- [88] a) Y. Li, Z. Lin, M. Guo, Y. Xia, M. Zhao, C. Wang, T. Xu, T. Chen, B. Zhu, Int. J. Nanomed. 2017, 12, 5733; b) J. Torrecilla, A. D. P. Rodríguez, M. Y. Solinís, P. S. Apaolaza, B. B. Herranz, C. R. López, A. B. Herranz, A. R. Gascón, Colloids Surf B Biointerfaces 2016, 146, 808–817; c) S. Gaikwad, A. Ingle, A. Gade, M. Rai, A. Falanga, N. Incoronato, L. Russo, S. Galdiero, M. Galdiero, Int. J. Nanomed. 2013, 8, 4303; d) A. M. Paul, Y. Shi, D. Acharya, J. R. Douglas, A. Cooley, J. F. Anderson, F. Huang, F. Bai, J. Gen. Virol. 2014, 95, 1712.
- [89] T. Q. Huy, N. T. Hien Thanh, N. T. Thuy, P. V. Chung, P. N. Hung, A.-T. Le, N. T. Hong Hanh, J. Virol. Methods 2017, 241, 52–57.
- [90] E. C. Lee, N. Davis-Poynter, C. T. Nguyen, A. A. Peters, G. R. Monteith, E. Strounina, A. Popat, B. P. Ross, *Nanoscale* 2016, 8, 16192–16196.
- [91] a) H. Zhu, Z. Fohlerová, J. Pekárek, E. Basova, P. Neužil, *Biosens. Bioelectron.* 2020, 112041; b) A. Mokhtarzadeh, R. Eivazzadeh-Keihan, P. Pashazadeh, M. Hejazi, N. Gharaatifar, M. Hasanzadeh, B. Baradaran, M. de la Guardia, *TrAC Trends Anal. Chem.* 2017, *97*, 445–457.
- [92] M. Norouzi, M. Zarei Ghobadi, M. Golmimi, S.-H. Mozhgani, H. Ghourchian, S. A. Rezaee, Anal. Lett. 2017, 50, 2402–2411.
- [93] a) J. R. Carter, V. Balaraman, C. A. Kucharski, T. S. Fraser, M. J. Fraser, Virol. J. 2013, 10, 201; b) S. C. Gopinath, K. Awazu, M. Fujimaki, K. Shimizu, T. Shima, PLoS One 2013, 8, e69121; c) M. Askaravi, S. E. Rezatofighi, S. Rastegarzadeh, M. R. S. A. Shapouri, AMB Express 2017, 7, 137.
- [94] G. Seo, G. Lee, M. J. Kim, S. H. Baek, M. Choi, K. B. Ku, C. S. Lee, S. Jun, D. Park, H. G. Kim, S. J. Kim, J. O. Lee, B. T. Kim, E. C. Park, S. I. Kim, ACS Nano 2020, 14(4), 5135–5142.
- [95] P. Moitra, M. Alafeef, K. Dighe, M. Frieman, D. Pan, ACS Nano 2020.
- [96] B. T. P. Thuy, T. T. A. My, N. T. T. Hai, L. T. Hieu, T. T. Hoa, H. Thi Phuong Loan, N. T. Triet, T. T. V. Anh, P. T. Quy, P. V. Tat, ACS Omega 2020, 5(14), 8312–8320.
- [97] a) S. Khaerunnisa, H. Kurniawan, R. Awaluddin, S. Suhartati, S. Soetjipto, *Prepr. doi10. 20944/preprints202003. 0226. v1* **2020**, 1–14; b) M. H. Sampangi-Ramaiah, R. Vishwakarma, R. U. Shaanker, *Curr. Sci.* **2020**, *118*, 1087.
- [98] P. M. Pour, S. Fakhri, S. Asgary, M. H. Farzaei, J. Echeverria, Front. Pharmacol. 2019, 10.
- [99] Q. F. Hu, B. Zhou, J. M. Huang, X. M. Gao, L. D. Shu, G. Y. Yang, C. T. Che, J. Nat. Prod. 2013, 76, 292–296.
- [100] F. Olivon, S. Remy, G. Grelier, C. C. Apel, C. C. Eydoux, J. C. Guillemot, J. Neyts, L. Delang, D. Touboul, F. Roussi, J. Nat. Prod. 2019, 82, 330–340.
- [101] S. Kaushik, G. Jangra, V. Kundu, J. P. Yadav, S. Kaushik, VirusDis. 2020 (https://doi.org/10.1007/s13337-020-00584-0).
- [102] D. J. Newman, G. M. Cragg, J. Nat. Prod. 2020, 83, 770–803.
- [103] J. Martinez, F. Sasse, M. Brönstrup, J. Diez, A. Meyerhans, Nat. Prod. Rep. 2015, 32, 29–48.
- [104] R. K. Ganjhu, P. P. Mudgal, H. Maity, D. Dowarha, S. Devadiga, S. Nag, G. Arunkumar, *VirusDis.* 2015, 26, 225–236.

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