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Interpretation and Validation of Maximum Absorbance Data Obtained from Turbidimetry Analysis of Plasma Clots

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Abstract

Turbidimetry is used to characterize fibrin clot properties. In purified systems, maximum absorbance (MA) directly relates to fibrin fiber cross-sectional area. However, in plasma samples there are discrepancies in the relationships between MA and fibrinogen concentration, fiber diameter, other clot properties, and cardiovascular disease outcomes, which complicate data interpretation. This study aims to advance understanding of MA of plasma clots through testing how well it relates to fundamental dependence on fibrinogen concentration and fiber diameter as predicted by light scattering theory, other clot properties and lifestyle, and biochemical variables. Plasma samples from 30 apparently healthy individuals with a fibrinogen concentration from 2.4 to 6.4 g/L were included. We performed turbidimetry, permeability, scanning electron microscopy, and rheometry on *in vitro* formed plasma clots. MA correlated more strongly with fibrinogen concentration (r = 0.65; p < 0.001) than with fiber diameter (r = 0.47; p = 0.01), which combined explained only 46% of the MA variance. Of additional variables measured, only low-density lipoprotein cholesterol correlated with MA (r = 0.46; p = 0.01) and clot lysis (r = 0.62; p < 0.0001) but not with fiber diameter or fibrinogen concentration. MA correlated with clot lysis time (r =0.59; p = 0.001, storage modulus (r = 0.61; p = 0.001), and loss modulus (r = 0.59; p = 0.001), and negatively with clot permeability (r = -0.60; p = 0.001) also after adjustment for fibrinogen concentration and fiber diameter. Increased MA is indicative of a prothrombotic clot phenotype irrespective of fibrinogen concentration. MA is more indicative of overall clot density than of fiber diameter. Other plasma components can alter internal fiber density without altering fiber diameter and should be considered when interpreting MA of plasma samples.

Keywords

turbidimetry; fibrinogen; fibrin structure; fibrin viscoelastic properties; blood plasma samples

Conflict of Interest None declared.

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M.P. and Z.d.L. designed the research; Z.d.L. and C.N. performed the experiments and analyzed the data; M.P., M.G., and Z.d.L. critically evaluated the results and wrote the manuscript. All authors approved the final manuscript and figures.

Introduction

Turbidimetry is a high-throughput, highly sensitive technique, providing information about fibrin clot formation and structure, that can be used in large-scale clinical and epidemiological studies with good reproducibility.¹ A turbidity curve is recorded by plotting light absorbance against time during *in vitro* clot formation.² Three main variables are typically obtained from this curve, namely lag time (time required for protofibrils to grow to sufficient length to allow lateral aggregation), slope (rate of lateral aggregation), and maximum absorbance, which depends on fibrinogen concentration and clot structural parameters, such as fiber diameter and internal fiber density, defined as the internal mass density of the fiber measured in g/cm³.^{3–5} More recently, this method has been modified to also include clot lysis time (CLT) by the addition of tissue plasminogen activator (tPA), resulting in the breakdown of the formed clot.⁶ The interpretation of maximum absorbance in terms of clot structural properties started with the early observation by Ferry and Morrison⁷ that fibrin gels with varying maximum absorbance contain fibers with varying fiber diameters. In 1978, Carr and Hermans³ derived an equation that relates absorbance to fibrinogen concentration and specific structural properties of a clot. In this model, the fibrin gel is treated as a dilute solution of randomly oriented cylindrical, long, thin rods that attenuate transmitted light by scattering (absorption and other light-attenuating processes are ignored). According to this model, the turbidity, τ , which equals $\ln(10)$ ·absorbance under common experimental conditions, see Supplementary Material (available in the online version), is given by

$$\tau = \alpha \mu \left(\frac{1}{\lambda^3} - \frac{\beta D^2}{4\lambda^5} \right) \tag{1}$$

where λ is the wavelength of the incident and transmitted light, $\alpha = \frac{88\pi^3}{15N_A} n_S \left(\frac{dn}{dc}\right)^2 \cdot c$, and $\beta = \frac{184\pi^2 \cdot n_s^2}{231} \cdot N_A. N_A \text{ is Avogadro's number; } n_s \text{ is the refractive index of the solvent (1.33)}$ for water); $\left(\frac{dn}{dc}\right)$ is the specific refractive index increment for fibrin (= 0.17594 cm³/g)—these values are typically constant for a given experiment; c is the concentration of fibrinogen in g/mL; μ is the molecular mass-to-length ratio of the fiber, measured in Daltons/cm; D is the diameter of the fiber. It is important to note that μ depends on the diameter of the fiber and on the internal fiber density, ρ_f , since it measures molecular mass per fiber length. For a homogenous, cylindrical fiber, $\mu = \rho_f \cdot N_A \frac{\pi \cdot D^2}{4}$, where ρ_f is the internal fiber density, measured in g/cm³. A thicker fiber will have a larger μ than a thinner fiber (at constant internal fiber density), and a denser fiber will have a larger μ than a less dense fiber (at constant diameter). Furthermore, it should be noted that the fibrinogen concentration, c, which appears in the prefactor, a, also has an effect on fiber diameter; higher fibrinogen concentrations have been shown to result in larger fiber diameters.^{5,8,9} Thus, according to this model (Eq. 1), the absorbance of a clot explicitly depends on the fibrinogen concentration, c, the fiber molecular mass per length, μ , and the fiber diameter, D, with the added complication that c, D, and μ are interrelated since c affects D, and D affects μ . Eq.

(1), or modified versions of this equation, have also been used to determine fiber diameter, D, and average fiber mass-to-length ratio, μ , from turbidity versus wavelength measurements of fully formed clots.^{4,10–13}

The model resulting in Eq. (1) was initially developed for purified fibrinogen solutions; however, turbidimetry is also used in plasma samples, typically when comparing fibrin clot properties of healthy control individuals with cardiovascular disease (CVD) patients since altered fibrin clot structure is considered a risk factor for arterial and venous thrombotic events.¹⁴ The correlation between τ and c, and between τ and D, as predicted by this equation, have, however, not been validated in plasma samples. Moreover, there are some apparent discrepancies in plasma studies when correlating maximum absorbance with c, D, other clot properties, and CVD outcomes. For example, in two case–control studies using plasma samples, maximum absorbance was increased in thrombotic patients.^{15,16} However, Undas et al¹⁶ reported these fibers to be thicker, whereas Mills et al¹⁵ reported thinner fibers when measured directly with scanning electron microscopy (SEM). Differences in maximum absorbance were also reported between healthy control and CVD patients despite having similar fibrinogen concentrations.^{17–22}

Regarding other clot properties, not included in Eq. (1), some studies show increased maximum absorbance to be associated with higher clot permeability,^{17,19,23} whereas in others, it was associated with denser clots with decreased permeability.^{15,16,18,20–22} In terms of disease outcome itself, both decreased^{19,24} and increased^{15,20–22} maximum absorbance have been reported in CVD patients, as compared with healthy controls.

These inconsistencies indicate a need to better understand turbidity measurements in plasma samples. This study therefore aims to advance understanding of maximum absorbance in plasma samples through a series of association studies. Specifically, we tested how well maximum absorbance relates to (i) the predicted fundamental dependence on fibrinogen concentration and fiber diameter (Eq. 1), (ii) other clot properties (clot formation, permeability, lysis, and mechanical properties), and (iii) lifestyle and biochemical variables of the study participants. As expected (Eq. 1), maximum absorbance correlated with both, fiber diameter and fibrinogen concentration, with the latter association being the strongest. However, fibrinogen concentration and fiber diameter did not fully explain the variance in maximum absorbance, since, when keeping the fibrinogen concentration constant, a twofold difference in maximum absorbance was still found. This suggests that in plasma, there are additional parameters that affect maximum absorbance, most likely by altering the internal fiber density, which is very challenging to measure directly with a method independent from turbidity. Of additional lifestyle and biochemical variables measured, we found that lowdensity lipoprotein cholesterol (LDL-C) strongly and positively correlated with maximum absorbance and clot lysis but not with fiber diameter or fibrinogen concentration. This finding suggests that LDL-C increases the internal fiber density, which, in turn, increases the maximum absorbance. Finally, we also found positive correlations between maximum absorbance and CLT, storage modulus, and loss modulus, and a negative correlation with clot permeability.

Methods

Study Population and Design

Data for this study were obtained from apparently healthy, black South Africans, older than 30 years (not using chronic medication for noncommunicable diseases) enrolled in the South African arm of the International Prospective Urban and Rural Epidemiology study.²⁵ Baseline data were collected in 2005 (n = 2,010) and follow-up data in 2010 (n = 1,288) and 2015 (n = 926). For this study, a subsample of 30 participants was systematically selected across the total fibrinogen concentration range (1.5–7.5 g/L) of the participants who partook in the 2015 data collection. All participants gave voluntary written informed consent and data collection complied with the Declaration of Helsinki of 1975 (as revised in 2013). The study was approved by the Health Research Ethics Committee of the North-West University, South Africa (NWU-00016–10-A1).

Questionnaire Data

Information regarding self-reported smoking, alcohol consumption, and contraceptive use was recorded using standardized, validated questionnaires.

Blood Sample Collection and Preparation

Fasting blood samples were collected between 07:00 a.m. and 11:00 a.m. from the antecubital vein using sterile winged infusion sets and syringes. For total and γ ' fibrinogen concentration, clot properties, and plasminogen activator inhibitor-1 (PAI-1) activity (PAI-1_{act}), samples were collected in 3.2% sodium citrate tubes. For serum total cholesterol (TC), high-density lipoprotein cholesterol (HDL-C) and LDL-C, triglycerides (TG), C-reactive protein (CRP), albumin, and creatinine, blood samples were collected in tubes without anticoagulants, and for plasma glucose, sodium fluoride tubes were used. Samples were centrifuged within 30 minutes of collection at 2,000 × *g* for 15 minutes and stored at -80°C until analyses.

Biochemical Analysis

Serum high-sensitivity CRP, TG, TC, HDL-C, albumin, creatinine, and plasma glucose were determined using the Cobas Integra 400 analyzer (Roche Diagnostics, Indianapolis, Indiana, United States). LDL-C was calculated according to the formula described by Sathiyakumar et al.²⁶ PAI-1_{act} was determined by an indirect enzymatic method (Spectrolyze PAI-1, Trinity Biotech, Bray, Ireland). Fibrinogen was analyzed with a modified Clauss method using an automated coagulation laboratory (Instrumentation Laboratory, Milan, Italy). Fibrinogen γ ' was determined with an enzyme-linked immunosorbent assay²⁷ and reported as percentage of total fibrinogen.

Fibrin Clot Properties

Turbidity—Plasma fibrinolytic potential was determined using a turbidity assay.²⁸ tPA (80 ng/mL tPA; Actilyse, Boehringer Ingelheim, Ingelheim, Germany) was added to plasma containing 17 mmol/L CaCl₂, tissue factor (TF) (1,750 × diluted TF; Dade Innovin, Siemens Healthcare Diagnostics Inc., Marburg, Germany) and 10 mmol/L phospholipid vesicles

(Rossix, Mölndal, Sweden). The tPA and TF concentrations were selected to obtain CLTs between 60 and 100 minutes. To determine clotting and lysis times from the resultant curves, sigmoidal curve fitting was used (Origin software version 8.5 [Origin lab, 2010]). Lag time (minutes), slope ($\times 10^{-3}$ au/s), maximum absorbance (*au*), and CLT (minutes) were calculated as previously described.⁶

Permeability—Clot permeability (K_s), an indication of the intrinsic pore size of the network, was measured as described previously.²⁹ Essentially, plasma clots were prepared in triplicate in 3-cm sections of 1 mL plastic serological pipettes. Plasma was clotted with the addition of 1 U/mL human α -thrombin (Merck, Darmstadt, Germany) and 20 mM CaCl₂. Buffer was permeated through at a pressure height of h = 4 cm and K_s calculated from $K_s = \frac{\Delta Q L \eta}{\Delta T \Delta \Delta P} \Delta Q / \Delta T$ is the flow rate (flow through volume/time), η is the viscosity ($\eta_{water} = 1.0$ cP at 20°C), L is the clot length, A is the cross-sectional area of the clot, and P is pressure drop. $P = \rho_{water} \cdot g \cdot h$, where the density of water, $\rho_{water} = 1$ g/cm³, the acceleration due to gravity, g = 980 cm/second², and h = 4 cm is the height of the buffer

above the clot.

SEM and Fiber Diameter Measurement—After completion of permeation, each clot container was rinsed with cacodylate buffer; thereafter, clots were fixed overnight in 2% glutaraldehyde (Merck) and recovered from the clot containers. Samples were then prepared for SEM imaging by dehydration with ethanol in successive dose increases and chemically dried with hexamethyldisilazane (Merck). Dried clots were mounted onto stubs and sputter coated with gold-palladium before being viewed and photographed with a FEI Corporation Quanta 200ESEM (Hillsboro, Oregon, United States). ImageJ (v 1.48, National Institutes of Health, Bethesda, Maryland, United States) was used to measure the fiber diameter of 100 systematically selected fibers in each of five micrographs per individual.

Rheometry—Plasma clots were prepared with 1 U/mL human α -thrombin and 20 mM CaCl₂ and viscoelastic properties determined during plasma clot formation, by performing oscillatory shear measurements at 37°C on an ARES-G2 Rheometer (TA Instruments, New Castle, Delaware, United States). A time sweep test was performed using a 40-mm stainless steel cone, under an oscillation procedure of 3% strain, at an angular frequency of 5 rad/s (10 half sampling cycles) with a sampling interval of 3 points per second. The geometry (truncation) gap and loading gap were set at 0.045 and 15.0 mm, respectively. After lowering the stainless steel cone, immersion oil (100–120 mPa·s) (Merck) was placed around the plates to prevent the clot from drying out. The test was performed for 40 minutes, measuring the storage modulus (G') and loss modulus (G''), that is, elastic and viscous properties, respectively, for each plasma sample at 3-second intervals. A summary of the methods used in this study and the data obtained from these methods, are presented in \blacktriangleright Fig. 1. The coefficient of variation for all methods was < 10%.

Statistical Analysis

Statistical analyses were performed using the Statistica software version 13. Significance was set at p = 0.05. Normality was determined using histograms and the Shapiro–Wilks test.

Normally distributed data (fibrinogen concentration, turbidimetry data, K_s , and fiber diameter) are reported as mean standard deviation. Nonnormally distributed data (% γ ' fibrinogen and rheometry data) were log transformed to improve normality for further statistical analyses but are reported as median (25th-75th percentiles). Multiple regression analysis was used to determine the contribution of fibrinogen concentration and fiber diameter to maximum absorbance variance. Pearson's correlations and partial correlations (when adjusting for fibrinogen concentration or fiber diameter) were used to determine the correlation between maximum absorbance, fibrinogen concentration, and other fibrin clot properties. Scatterplots were inspected for outliers to ensure the correlations were not driven by single outlier data points. To determine whether there are other factors that are associated with maximum absorbance in plasma, Mann–Whitney U tests (because of the small sample size of the subdivisions) were used to compare maximum absorbance and fiber diameter between categorical variables (alcohol consumption, smoking, and oral contraceptive use) with analysis of covariance when adjusting for differences in fibrinogen concentration. Pearson's correlations and partial correlations (adjusting for fibrinogen concentration) with visual inspection of scatterplots were used to determine associations with continuous variables (age, body mass index [BMI], glucose, CRP, blood lipids, albumin, and creatinine).

Results

The descriptive characteristics of the participants are presented in \blacktriangleright Table 1. Twenty-three of the 30 participants were women, 5 of whom used oral contraceptives. Eleven participants smoked and eight consumed alcohol. The mean age of the study group was 58.6 (53.5–66.7) years and the mean BMI was 26.2 (21.8–33.8) kg/m². The mean fibrinogen concentration was 4.16 ± 0.85 g/L.

Association between Maximum Absorbance, Fibrinogen Concentration, and Fiber Diameter

In a first series of experiments, we investigated the correlation between maximum absorbance and fundamental clot parameters as predicted from light scattering theory (Eq. 1). Maximum absorbance correlated with both fibrinogen concentration and fiber diameter; however, the correlation with fibrinogen concentration was stronger than that with fiber diameter (r = 0.65; p < 0.001 compared with r = 0.47; p = 0.01) (\blacktriangleright Table 2). Fiber diameter on the other hand demonstrated similar correlations with maximum absorbance and fibringen concentration (r = 0.47; p = 0.01 and r = 0.45; p = 0.01). After adjusting for differences in fibrinogen concentration between samples, maximum absorbance no longer correlated with fiber diameter (r = 0.26; p = 0.18) (\blacktriangleright Table 3) indicating that this association is largely dependent on the fibrinogen concentration. In a separate analysis, when adjusting for fiber diameter, the association between maximum absorbance and fibrinogen concentration was only moderately reduced (r = 0.56; p = 0.002 vs. r = 0.65; p < 0.001). This suggests that this relationship is only partly attributable to fiber size with the remaining relationship likely reflecting clot density (volume occupied by fibers per total clot volume). ► Fig. 2 depicts scanning electron images of clots prepared from plasma samples with varying fibrinogen concentrations. Based on visual comparison, clots prepared from plasma

with higher fibrinogen concentrations had thicker fibers and had a higher fiber density than clots prepared from plasma samples with a lower fibrinogen concentration.

Despite the strong relationship between maximum absorbance and fibrinogen concentration, samples with similar fibrinogen concentrations, had maximum absorbance values that differed up to twofold (►Supplementary Table S1, available in the online version). In addition, fibrinogen concentration and fiber diameter combined, explained only 46% of the variance in maximum absorbance with fibrinogen concentration contributing 42% and fiber diameter 4%.

Association of Maximum Absorbance with Lifestyle and Other Measured Biochemical Variables

To identify other factors that may explain the remaining variance in maximum absorbance, maximum absorbance, fiber diameter, and fibrinogen concentration were correlated with other biochemical and lifestyle variables (Supplementary Table S2, available in the online version). ► Table 4 presents the variables that correlated significantly with at least one of the three. Of these measured variables, LDL-C was the only variable that correlated significantly with maximum absorbance (r = 0.46; p = 0.01) but not with fiber diameter (r = 0.17; p =0.36) or fibrinogen concentration (r = 0.18; p = 0.34). Referring to Eq. (1) (absorbance depends on variables c, D, and μ), this finding suggests that increased LDL-C is associated with an increased internal fiber density without increasing fiber diameter. LDL-C furthermore correlated significantly with CLT (r = 0.62 and r = 0.61 after adjusting for fibrinogen concentration; p < 0.0001 for both) suggesting a link between increased LDL-C, increased internal fiber density, and longer lysis time. LDL-C did not differ between smokers and nonsmokers, alcohol users and nonusers, or oral contraceptive users and nonusers. Maximum absorbance also correlated significantly with CRP (r = 0.47; p = 0.008) and fiber diameter with HDL-C (r = -0.38; p = 0.04), but significance disappeared after adjustment for fibrinogen concentration. There was no correlation with age, blood glucose, TC, TG, albumin, creatinine, or BMI (>Supplementary Material, available in the online version).

In addition, higher maximum absorbance values were observed for women using oral contraceptives compared with those who did not. However, after adjustment for the higher fibrinogen concentration in this group, the difference in maximum absorbance was no longer significant. Fiber diameter did not differ significantly between the two groups. There was no difference in either maximum absorbance or fiber diameter between smokers and nonsmokers (►Fig. 3). Although maximum absorbance tended to be higher in alcohol consumers, this difference disappeared after adjustment for fibrinogen concentration.

Association between Maximum Absorbance and Other Clot Properties

In a third series of experiments, we determined the correlation between maximum absorbance and kinetic and bio-physical clot properties. Maximum absorbance had strong significant positive correlations with lag time (r = 0.39; p = 0.03), slope (r = 0.69; p < 0.001), CLT (r = 0.63; p < 0.001), stiffness (storage modulus, G') (r = 0.67; p < 0.001), and plasticity (loss modulus, G") (r = 0.68; p < 0.001), and a negative correlation with

permeability (r = -0.67; p < 0.001) (\blacktriangleright Table 2). These associations remained essentially unchanged after adjustment for fibrinogen concentration and fiber diameter, individually and combined (\blacktriangleright Table 3). In contrast, aside from its significant correlation with maximum absorbance, fiber diameter correlated significantly with slope (r = 0.38; p = 0.04) only. After adjusting for fibrinogen concentration this association disappeared, indicating that this

Discussion

In purified systems, maximum absorbance is considered to be directly related to the fibrin fiber cross-sectional area.³ This relationship was established through light scattering theory, which modeled fibrin fibers as diluted, randomly arranged, long, thin cylinders. It predicts that turbidity (maximum absorbance) depends on fibrinogen concentration, fiber diameter,

and internal fiber density (as part of the molecular mass-to-length ratio, $\mu = \rho_f \cdot N_A \frac{\pi \cdot D^2}{4}$),

association is largely mediated through the fibrinogen concentration.

as can be seen from Eq. (1). Since maximum absorbance depends on all three, the direct association between maximum absorbance and fiber diameter is contingent on fibrinogen concentration and internal fibrin density being constant. The use of the turbidity assay has also been extended to plasma samples to characterize clot properties of mainly CVD patients, compared with healthy individuals. Superficially, plasma clots have a very similar appearance to fibrin clots prepared from purified fibrinogen. Thus, it may seem reasonable to assume that Eq. (1) also applies to plasma clots. However, this assumption needed to be tested as the plasma environment differs from purified systems in that the fibrinogen concentrations in the samples vary and plasma contains additional components, not present in purified fibrinogen systems. These components may modify internal fiber structure, which our data in fact suggest to be the case. In addition, in plasma studies, maximum absorbance is often interpreted to reflect fiber size, as is done in purified studies, without considering the influence of the varying fibrinogen concentrations or the additional plasma components. Validation studies in plasma samples are therefore required to advance our understanding of how to interpret maximum absorbance in plasma samples.

Traditionally, fibrin clot networks are defined as either fine or coarse networks.^{30,31} Clots composed of thinner fibers are typically denser and more rigid, with a stiffer network arrangement, reduced permeability (smaller pore sizes), and enhanced resistance to fibrinolysis (prothrombotic); whereas clots consisting of thicker fibers generally contain looser and less rigid networks that are more susceptible to fibrinolysis (antithrombotic).^{32–38} As mentioned in the introduction, in plasma (case–control) studies, these relationships are less clear, with inconsistencies existing regarding the association between other clot properties and maximum absorbance, which is often used as a proxy marker/indirect measurement of fiber diameter. Our data showed that clots with increased maximum absorbance, have an increased rate of lateral aggregation, have thicker fibers, increased clot stiffness, and decreased permeability and lysis. After adjustment for fibrinogen concentration, maximum absorbance no longer correlated with fiber diameter but all other associations remained. This suggests that increased maximum absorbance is indicative of a prothrombotic clot phenotype that is more dense, with increased stiffness and decreased lysis rate, irrespective of the fibrinogen concentration. These associations were further more

not dependent on fiber diameter as fiber diameter did not associate significantly with any of the other clot properties nor did adjustment for fiber diameter alter the associations. In agreement with these associations, many studies in the literature found maximum absorbance to be increased in CVD patients regardless of whether fibrinogen concentration was increased or not.^{15,16,18,20–22,39,40}

As predicted by Eq. (1), both fiber diameter and fibrinogen concentration associated positively with maximum absorbance. However, after adjusting for fibrinogen concentration, fiber diameter and maximum absorbance no longer correlated, indicating that this relationship is largely driven by the fibrinogen concentration. The relationship between maximum absorbance and fibrinogen concentration, however, remained after adjusting for fiber diameter. This indicates that fiber size only partly contributes to this relationship with the remainder likely reflecting the fiber density of the clot (i.e., volume occupied by fibers per total clot volume). Based on the SEM images, it is clear that clots with higher fibrinogen concentration formed both more and thicker fibers, increasing the total density of the clot. Maximum absorbance furthermore correlated more strongly with fibrinogen concentration than fiber diameter, further supporting the concept that it is a marker of overall clot density rather than simply reflecting fiber diameter.

Fibrinogen concentration and fiber diameter, however, explained only about half of the variance in maximum absorbance, and when keeping the fibrinogen concentration constant, a twofold difference in maximum absorbance still existed. As maximum absorbance is influenced not only by fibrinogen concentration and fiber diameter, but also by internal fiber density (Eq. 1), this suggests that the internal fiber density, ρ_f which enters Eq. (1) through the mass-to-length ratio, μ , $\left(\mu = \rho_f \cdot N_A \frac{\pi \cdot D^2}{4}\right)$, is an additional important parameter that affects absorbance in plasma and which may be influenced by additional components

present in plasma, but absent in the purified system. Several lifestyle and biochemical variables, for example, CRP and oral contraceptive use (with borderline significance for alcohol consumption), positively associated with maximum absorbance and not fiber diameter, but after adjustment for fibrinogen concentration, these associations disappeared indicating that the relationship was driven by the associated increased fibrinogen concentration. Only LDL-C was positively associated with maximum absorbance independent of fiber diameter and/or fibrinogen concentration. This suggests that LDL-C potentially increases internal fiber density by binding to fibrin fibers and in so doing increases maximum absorbance. It was furthermore also positively associated with CLT. Increased LDL-C has been reported in the literature to be associated with stiffer clots,⁴¹ slower lag time, ⁴² lower clot permeability, ^{43,44} and enhanced resistance to fibrinolysis. ^{41,45} These associations, together with our data, suggest that this inhibited fibrinolysis may be the result of LDL-C-associated apolipoproteins binding to fibrin clots and thereby hindering permeation and action of lytic enzymes.^{41,46} In addition, surface-bound LDL has been found to bind to tPA, thus preventing the formation of plasmin.⁴⁷ The oxidation of lipids may furthermore influence the effect of lipids on clot structure. It has been suggested that fibrin may be directly affected by highly oxidized LDL altering the ensuing clot network by enhancing hypercoagulation and fibrinolytic resistance in comparison to nonoxidized LDL.

⁴⁸ These results suggest a potential, novel role for LDL-C in thrombotic disease, which deserves further investigation. Intrafibrillar fibrin structure has recently been demonstrated to be modifiable and to significantly influence clot mechanical properties and resistance to lysis.⁴⁹ Domingues et al⁵⁰ demonstrated that thrombin concentration and γ ' fibrinogen altered protofibril content and protein density within fibers, altering the overall strength of the fibrin network. In support of this concept, in our study, maximum absorbance also correlated positively with fibrin clot stiffness even after adjusting for fiber diameter and fibrinogen concentration, although the exact mechanism behind this association remains to be determined.

A potential limitation of this study was that native plasma clots were used to obtain turbidimetric, permeability, and rheometry data, while fibrin clots were dehydrated for SEM analysis. The turbidity assay furthermore made use of TF as a clotting agent, while thrombin was used in the other methods. However, these assays were used with the intention of being complementary to one another, providing supporting data obtained from different methods. Furthermore, maximum absorbance demonstrated strong significant associations not only with the turbidimetry data, but also with the other assays, supporting internal consistency of the data. We could not investigate molecular mass-to-length ratio (μ), as an indirect marker of internal fiber density due to a lack of sample. Nor could we directly measure clot/protein density. We recommend the development and/or validation of such direct measures, to further assess the association between maximum absorbance and internal fiber density and overall clot protein density. Although we identified environmental and plasma components that contributed to altered clot properties, the study was not designed to identify an exhaustive list of fibrin clot structure determinants and follow-up experimentation is required to better characterize additional plasma components that may influence the internal fiber density of plasma clots. Being a cross-sectional study, we could furthermore not determine causality, which limits the conclusions that can be drawn from the data. It furthermore remains to be determined whether these results, obtained in apparently healthy participants, are also valid in patients with thrombosis.

Conclusion

In this study increased maximum absorbance was associated with a prothrombotic clot phenotype characterized by increased rate of lateral aggregation, increased clot stiffness, and decreased clot permeability and lysis irrespective of fiber diameter and fibrinogen concentration. Although maximum absorbance correlates positively with fiber diameter, in plasma samples it is more strongly associated with fibrinogen concentration and more indicative of overall clot density than of fiber diameter. Other components in plasma, such as LDL-C, can alter internal fiber density without altering fiber diameter, likely by binding to fibrin fibers and in so doing influence maximum absorbance. Our data show that while Eq. (1) is valid to use in plasma samples, the difference in fibrinogen concentration and potential influence of plasma constituents on internal fiber density should be considered when interpreting maximum absorbance.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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What is known about this topic?

- According to light scattering theory, the absorbance of a clot depends on the fibrinogen concentration, the internal fiber density (as part of the fiber molecular mass per length ratio), and the fiber diameter.
- In purified fibrinogen systems, absorbance is indicative of fiber diameter at fixed fibrinogen concentrations but these associations remain to be validated in plasma samples.
- There are discrepancies in published data from plasma samples when relating maximum absorbance to fibrinogen concentration, fiber diameter, other clot properties, and CVD outcomes.

What does this paper add?

- In this study, increased maximum absorbance was indicative of a prothrombotic clot phenotype that was denser, with increased stiffness and decreased lysis, irrespective of fibrinogen concentration or fiber diameter.
- Maximum absorbance correlated more strongly with fibrinogen concentration than fiber diameter, suggesting it to be a marker of overall clot density (volume occupied by fibers per total clot volume) rather than simply fiber diameter.
- Fibrinogen concentration and fiber diameter explained only about half of the variance in maximum absorbance.
- We demonstrated that other plasma components, such as LDL-C, can alter maximum absorbance by altering internal fiber density.

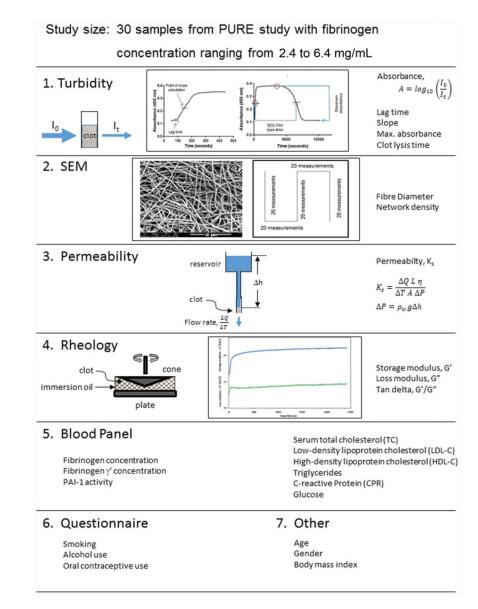


Fig. 1.

Summary of the methods used in this article. The turbidity curves of this figure are included with permission from Pieters et al.⁶

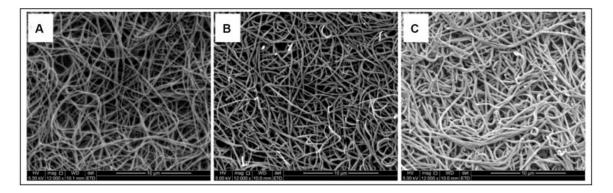


Fig. 2.

Scanning electron microscopy (SEM) images, at $12,000 \times$ magnification, of plasma clots with varying total fibrinogen concentration and increasing fiber diameters as follows: (A) 2.4 g/L, 162 nm; (B) 4.17 g/L, 196 nm; and (C) 6.34 g/L, 218 nm.

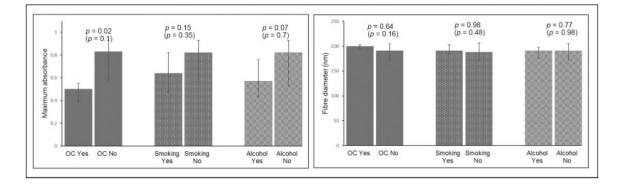


Fig. 3.

Maximum absorbance and fiber diameter in relation to oral contraceptive use, smoking, and alcohol consumption. Bars denote median and interquartile range. *p*-Values indicate significance between users and nonusers. *p*-Values in brackets were obtained after adjustment for fibrinogen concentration. OC, oral contraceptive users n=5; oral contraceptive nonusers n = 18; smokers n = 10; nonsmokers n = 19; alcohol consumers n = 8; alcohol nonusers n = 22.

Table 1

Descriptive characteristics of the participants

Variable	Participants $(n = 30)$
Sex (male / female) n (%)	7 (23.3) / 23 (76.7)
Age (years)	58.6 (53.5–66.7)
Tobacco users, $n(\%)$	11 (36.6)
Alcohol users, $n(\%)$	8 (26.7)
Oral contraceptive users, $n(\%)$	5 (21.7)
BMI (kg/m ²)	26.2 (21.8–33.8)
Glucose (mmol/L)	5.63 [1.97]
PAI-1 (U/mL)	1.91 (0-8.45)
CRP (mg/L)	5.09 (2.64–10.5)
TC (mmol/L)	4.59 [1.43]
HDL-C (mmol/L)	1.36 [0.42]
LDL-C (mmol/L)	2.87 [1.38]
TG (mmol/L)	1.18 [0.56]
Albumin (g/L)	39.3 [6.88]
Creatinine (µmol/L)	59.8 [17.3]
Fibrinogen (g/L)	4.16 [0.85]
Fibrinogen y' (%)	9.18 (8.36–12.9)
Lag time (min)	4.18 [0.66]
Slope (au/s)	10.8 [5.00]
Maximum absorbance (au)	0.71 [0.24]
Clot lysis time (min)	64.0 [9.47]
Fiber diameter (nm)	191 [18.5]
Permeability (cm ² × 10 ⁻⁹)	9.50 [3.20]
Storage modulus (G') (Pa)	56.2 (35.1–102)
Loss modulus (G") (Pa)	2.55 (1.69-3.94)
Tan (delta) (G'/G")	0.05 (0.04–0.06)

Abbreviations: au/s, absorbance unit per second; BMI, body mass index; CRP, C-reactive protein; HDL-C, high-density lipoprotein cholesterol; PAI-1, plasminogen activator inhibitor-1; TC, total cholesterol; TG, triglycerides.

Note: au indicates change in absorbance units. Normally distributed data are reported as mean [standard deviation] and nonnormally distributed data as median (25th-75th percentile).

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Correlation

Clot property	Max abs	Fiber diameter	Fibrinogen	Fibrinogen $\gamma^{*}(\%)$	Lag time	Slope	CLT	Permeability	Storage modulus	Loss modulus	Tan (delta)
Max abs (au)		0.47^b	$0.65^{\mathcal{C}}$	-0.12	0.39 ^a	$0.69^{\mathcal{C}}$	$0.63^{\mathcal{C}}$	-0.67^{c}	0.67 ^c	$0.68^{\mathcal{C}}$	-0.19
Fiber diameter (nm)	0.47^{b}		0.45^{b}	-0.21	0.04	0.38 ^a	0.30	-0.05	0.24	0.26	-0.09
Fibrinogen (g/L)	0.65 ^C	0.45^{b}		-0.13	0.11	$0.64^{\mathcal{C}}$	0.29	-0.54^{b}	0.37^{a}	0.41 ^{<i>a</i>}	0.04
Fibrinogen γ' (%)	-0.12	-0.21	-0.13		-0.19	0.17	0.03	-0.04	-0.15	-0.10	0.26
Lag time (min)	0.39 ^a	0.04	0.11	-0.19		-0.02	0.006	-0.12	0.30	0.27	-0.12
Slope (au/s)	0.69 ^C	0.38 ^a	$0.64^{\mathcal{C}}$	0.17	-0.02		$0.56^{\mathcal{C}}$	$-0.63^{\mathcal{C}}$	0.27	0.31	0.05
CLT (min)	$0.63^{\mathcal{C}}$	0.30	0.29	0.03	0.006	$0.56^{\mathcal{C}}$		-0.36^{a}	0.49^{b}	0.51^{b}	-0.14
Permeability (cm ²)	-0.67^{c}	-0.05	-0.54^{b}	-0.04	-0.12	$-0.63^{\mathcal{C}}$	-0.36^{a}		0.41 ^{<i>a</i>}	-0.46^{b}	-0.18
Storage modulus (Pa)	$0.67^{\mathcal{C}}$	0.24	0.37^{a}	-0.15	0.30	0.27	0.49^{b}	-0.41^{a}		$0.98^{\mathcal{C}}$	-0.49^{b}
Loss modulus (Pa)	$0.68^{\mathcal{C}}$	0.26	0.41^{a}	-0.10	0.27	0.31	0.51^{b}	-0.46^{b}	0.98 ^c		-0.33
Tan (delta) (G'/G")	-0.19	-0.09	0.04	0.26	-0.12	0.05	-0.14	-0.18	-0.49^{b}	-0.33	

Abbreviations: CLT, clot lysis time; Max abs, maximum absorbance; au, change in absorbance units.

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a p < 0.05.

 $b \\ p < 0.01.$ $c \\ p < 0.001.$

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Table 3

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Clot property	Unadjusted	Adjusted - fibrinogen	Adjusted - fiber diameter	Adjusted - fibrinogen and fiber diameter
	r	r	r	ľ
Fiber diameter (nm)	0.47^{b}	0.26	1	-
Lag time (min)	0.39 ^a	0.43 ^a	0.42 ^a	0.45 ^a
Slope (au/s)	$0.69^{\mathcal{C}}$	$_{0.47}^{b}$	$0.64^{\mathcal{C}}$	0.47^{b}
Clot lysis time (min)	$0.63^{\mathcal{C}}$	$0.61^{\mathcal{C}}$	q65.0	9 ⁶⁵⁰
Permeability (cm ²)	$-0.67^{\mathcal{C}}$	-0.49^{b}	$-0.73^{\mathcal{C}}$	-0.60 b
Storage modulus (Pa)	$0.67^{\mathcal{C}}$	$0.62^{\mathcal{C}}$	0.65 ^C	0.61^{b}
Loss modulus (Pa)	$0.68^{\mathcal{C}}$	$0.60^{\mathcal{C}}$	0.65 ^C	0.59 ^b
Tan (delta) (G'/G")	-0.19	-0.29	-0.18	-0.28
Fibrinogen γ' (%)	-0.12	-0.06	-0.04	-0.01

Abbreviations: au/s, absorbance unit per second; Max abs, maximum absorbance.

a p < 0.05.

 $^{b}_{p < 0.01.}$

 $\overset{\mathcal{C}}{p} < 0.001.$

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Variable	Maximum	Maximum absorbance Fiber diameter	Fiber d	iameter	Fibrinogen	Fibrinogen concentration
	r	<i>p</i> -Value	r	<i>p</i> -Value	r	<i>p</i> -Value
LDL-C - unadjusted - adjusted for fbg 0.46	0.46	0.01	0.17 0.36		0.18	0.34
	0.46	0.01	0.11 0.59	0.59	-	
HDL-C - unadjusted - adjusted for fbg -0.32	-0.32	0.08	-0.38 0.04	0.04	-0.41	0.02
	-0.08	0.69	-0.24 0.22	0.22		
CRP - unadjusted - adjusted for fbg	0.47	0.008	0.15 0.44	0.44	0.56	0.001
	0.17	0.40	-0.14 0.48	0.48	-	

Abbreviations: CRP, C-reactive protein; fbg, fibrinogen; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol.