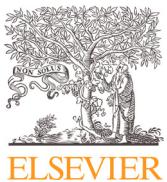




Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



## Recent advances in use of fresh frozen plasma, cryoprecipitate, immunoglobulins, and clotting factors for transfusion support in patients with hematologic disease

Prajeeda M. Nair<sup>a</sup>, Matthew J. Rendo<sup>b</sup>, Kristin M. Reddoch-Cardenas<sup>a</sup>, Jason K. Burris<sup>b</sup>, Michael A. Meledeo<sup>a</sup>, Andrew P. Cap<sup>a,c,\*</sup>

<sup>a</sup> United States Army Institute of Surgical Research, JBSA Fort Sam Houston, TX, USA

<sup>b</sup> San Antonio Military Medical Center, JBSA Fort Sam Houston, TX, USA

<sup>c</sup> Uniformed Services University, Bethesda, MD, USA



### ARTICLE INFO

#### Keywords:

Plasma transfusion  
Transfusion support  
Hematologic disease

### ABSTRACT

Hematologic diseases include a broad range of acquired and congenital disorders, many of which affect plasma proteins that control hemostasis and immune responses. Therapeutic interventions for these disorders include transfusion of plasma, cryoprecipitate, immunoglobulins, or convalescent plasma-containing therapeutic antibodies from patients recovering from infectious diseases, as well as concentrated pro- or anticoagulant factors. This review will focus on recent advances in the uses of plasma and its derivatives for patients with acquired and congenital hematologic disorders.

© 2020 Published by Elsevier Inc.

### Introduction

Hematologic diseases include a broad range of acquired and congenital disorders, spanning dysfunction of the bone marrow, red blood cells, leukocytes, and platelets. The plasma proteins of the coagulation system, the complement system as well as the immunoglobulins can also be affected. Finally, hematologic diseases affect the vascular endothelium from which blood arises during embryogenesis, and with which blood constantly interacts.

Unsurprisingly, a common component of treatment strategies for patients with hematologic disease involves replacement or enrichment of missing, dysfunctional or consumed constituents of blood through transfusion. This review will focus on the recent advances in the use of plasma and its derivatives, cryoprecipitate, immunoglobulin preparations, and individual clotting factors for patients with acquired and congenital hematologic disorders.

### The use of plasma transfusion for treating patients with hematologic disease

Plasma is the aqueous component of blood and is separated from blood cells by centrifugation of whole blood units or apheresis.

Plasma is a source of coagulation factors, albumin and immunoglobulins, as well as a large number of other proteins, lipids and other biological mediators. A variety of plasma products are currently available for transfusion including fresh frozen plasma (FFP), plasma frozen within 24 hours (PF24), thawed plasma (TP), liquid plasma (LP), and solvent-detergent plasma. FFP, PF24, TP, and LP have similar indications for use including in the management of preoperative or bleeding patients who require replacement of multiple factors (eg, liver disease, disseminated intravascular coagulation [DIC]); massive transfusion; urgent warfarin reversal; transfusion or plasma exchange in thrombotic thrombocytopenic purpura (TTP); congenital or acquired coagulation factor replacement when specific factor concentrates are unavailable; and rare specific plasma protein deficiencies [1]. In practice, FFP and PF24 are considered interchangeable, whereas TP and LP are not to be used to correct specific factor or plasma protein deficiencies when products containing higher concentrations of the required proteins are available. Solvent-detergent plasma is indicated in TTP and for replacement of multiple factors in acquired factor deficiency states including liver disease, liver transplantation, and cardiac surgery [2]. In addition, cryo-poor plasma is also a plasma-derived product wherein plasma is thawed at 1°C to 6°C to remove the precipitated fibrinogen, and it is indicated for transfusion or plasma exchange in patients with TTP or for providing limited clotting factors excluding fibrinogen, Factor VIII, Factor XIII, and vWF. These products differ in the content and activity of coagulation factors present in them and must be used within the stipulated shelf life.

\* Corresponding author. Andrew P. Cap, MD, PhD, United States Army Institute of Surgical Research, 3650 Chambers Pass, JBSA Fort Sam Houston, TX 78234. Tel.: +1-210-539-4858 (office), +1-210-323-6908 (mobile).

E-mail address: [andrew.p.cap.mil@mail.mil](mailto:andrew.p.cap.mil@mail.mil) (A.P. Cap).

With the recent advances in transfusion medicine, the use of plasma components is anticipated to drastically decline in the near future. The current guidelines call for plasma transfusions in patients with coagulopathy only when a specific therapy or factor concentrate is not appropriate or is unavailable. Plasma use has been discouraged as a treatment to improve international normalized ratio (INR) for low-risk procedures. However, the use of FFP to treat the acquired coagulopathies of DIC and liver diseases may still be relevant, as the replenishment of coagulation factors in these patients could be critical to treat the endothelial dysfunction associated with these conditions [3–7]. At basal conditions, endothelial cells are nonthrombogenic and are a main source of the tissue factor pathway inhibitor. Endothelial cells exert control of coagulation at critical steps of the clotting cascade [8,9]. Thus, endothelial dysfunction in these patients disturbs the finely tuned coagulation and fibrinolysis equilibrium causing blood failure [10–12], and can hence be classified as an acquired hematologic disease. For these patients, plasma or whole blood transfusion offers clear advantages over the clotting factor concentrates to treat endothelial dysfunction by supplying the adequate coagulation factors and fibrinolytic proteins to re-establish endothelial hemostasis [3]. Randomized clinical trials are warranted for developing evidence-based treatment recommendations in patients requiring multiple factor replacement in liver failure or DIC or in treating complex disorders like endothelial dysfunction arising from a variety of conditions.

Further, patients undergoing massive transfusion could potentially benefit from the clotting factors available in plasma transfusion, and a high FFP to RBC ratio (ie, 1:1) is advocated [13]. However, massive transfusion is a rare scenario in primary hematologic disease. In acquired coagulopathies arising from trauma and in other settings, an equal ratio of FFP, platelets, and RBCs (1:1:1) is used to mitigate platelet dysfunction and to reinstate hemostasis in these patients [14]. Conversely, when trauma patients who did not require a massive transfusion were transfused with FFP, a dose-related increase in adult respiratory distress syndrome, multi-organ failure, pneumonia, and sepsis was reported [15].

Another major indication for FFP is during warfarin or related vitamin K antagonists (VKA) treatment, for patients who are bleeding or undergoing urgent invasive procedures and need only transient reversal of warfarin effect. VKAs are routinely used for the primary and secondary prevention of arterial and venous thromboembolism, in patients with prosthetic heart valves, atrial fibrillation, peripheral arterial disease, and antiphospholipid syndrome [16,17]. The major concerns of plasma treatment in these patients are the varying levels of coagulation factors which may result in a partial or insufficient reversal of INR [18], and the large volume required to reverse the coagulation defect which can lead to cardiogenic pulmonary edema [19]. FFP transfusion is also specifically associated with a noncardiogenic pulmonary edema known as transfusion-related acute lung injury (TRALI), which is linked to formation of alloantibodies in prior transfusions or pregnancy. The incidence of TRALI has declined since the introduction of risk mitigation strategies such as collection of plasma from male or never pregnant female donors [19,20]. The more recently developed 4-factor prothrombin concentrate complex (4F-PCC) is currently considered optimal for treating warfarin reversal compared to plasma as it corrects INR more quickly with a lower volume of product infused [21–25]. As a result, plasma use in this setting is relegated to when 4F-PCC is not available.

Finally, TTP patients constitute an important cohort receiving FFP or cryo-poor plasma transfusion or plasma exchange to replace the VWF-cleaving protease, ADAMTS13. This enzyme prevents the formation of small-vessel platelet-rich thrombi, and the resulting thrombocytopenia, and the microangiopathic hemolytic anemia that characterizes TTP. TTP can be either an acquired syndrome

arising from an autoantibody against ADAMTS13 or a congenital syndrome, resulting from ADAMTS13 gene mutations. While acquired TTP still requires plasma exchange along with other treatments including rituximab, caplacizumab, and immunosuppressive agents to better manage the disease [26–28], a recombinant protein rADAMTS13 (BAX930) is currently underway in phase III clinical trials for use in congenital TTP patients [ClinicalTrials.gov Identifier: NCT03393975] [29]. This might be a significant breakthrough as these patients have a significant lifetime exposure to plasma that can cause severe complications such as allergic and anaphylactic reactions or volume overload, and infections from bloodborne pathogens [30].

In summary, FFP is recommended in patients with complex coagulopathies such as liver disease and DIC; however, there is little high-quality evidence to inform the optimal use of FFP in prophylaxis in these patients. Plasma has an established role in major hemorrhage and in TTP. Use of plasma is declining in warfarin reversal since the introduction of 4F-PCC. There are differing opinions about the efficacy of plasma transfusion to treat patients suffering from other hematologic diseases with no consistent evidence of significant benefit for prophylactic and therapeutic uses across a range of indications evaluated [31–35]. There is a pressing need for new clinical studies to evaluate the efficacy of plasma in non-bleeding patients to understand whether the risk is outweighed by the benefits.

#### *Use of convalescent plasma*

The plasma of patients recovering from acute viral diseases contains neutralizing antibodies that mediate immune clearance of viruses. Convalescent plasma has been safely used in a number of diseases including measles, influenza, Ebola, Severe acute respiratory syndrome (SARS), and Middle East respiratory syndrome, and may affect the clinical course of infection [36–40]. Most recently, it has been used in patients with COVID-19 (SARS-CoV-2 virus) [38,41–43]. Evidence of efficacy has been variable by disease, and in general very few adequately powered clinical trials have been performed. Convalescent plasma is considered by the US Food and Drug Administration to be a licensed plasma product used for investigational purposes [44]. Collection, storage, and transfusion of convalescent plasma are all performed as is standard for other plasma products. Dosing strategies have varied based on the availability of neutralizing antibody titer assays for various infections. Most protocols have stipulated the transfusion of 1 to 2 standard plasma units (200–600 mL total, or 10–20 mL/kg in pediatric patients). Published reports indicate that risks of convalescent plasma transfusion are similar to those of standard plasma, and adverse events are uncommon [40,45,46].

#### *The use of cryoprecipitate in treating patients with hematologic disease*

Cryoprecipitate was routinely used in the 1970s–1990s for hemophilia A and various factor deficiencies. However, its use has become increasingly confined to the treatment of hemorrhage with the development of individual factor concentrates. It derives its name from its own collection process, whereby FFP is thawed at 1°C to 6°C permitting precipitation of its cold-insoluble proteins. Centrifugation allows separation of these proteins: fibrinogen, factor VIII, factor XIII, von Willebrand factor, and fibronectin. A unit is typically stored at -18°C in 10 to 20 mL volumes of re-suspended plasma for up to 12 months [47]. After thawing, infusion is mandated within 4 hours [1,48]. While there are no definitive transfusion thresholds, general recommendations advise use when fibrinogen levels are less than 100 mg/dL in the setting of hemorrhage or DIC [47]. In the absence of bleeding or active

consumption, 1 unit of cryoprecipitate per 10 kg body weight typically raises the plasma fibrinogen concentration by approximately 50 mg/dL [49].

Its official name, cryoprecipitated antihemophilic factor, reflects its historical use to stop bleeding in patients with hemophilia A. Due to its high concentration of factor VIII, it significantly enhanced overall survival in patients with hemophilia A [47,50]. While the method for its acquisition was described in 1964, it was not approved for the use by the FDA until 1971 [51].

Until the early 1990s, most factor replacements were derived from human plasma. In 1992, the FDA-approved recombinant factor VIII [52]. Since then, individual factor concentrates have surpassed cryoprecipitate as front line for replacement therapies for hemophilia A, FXIII deficiency, hypofibrinogenemia and in von Willebrand disease. Moreover, clinical guidelines have recommended against cryoprecipitate for these conditions unless specific factor replacement products are unavailable [50]. This is because individual factor concentrates generally are associated with fewer transfusion reactions and episodes of TRALI, and lower infection risk compared to cryoprecipitate [50]. However, fibrinogen concentrate in the United States remains licensed only for congenital deficiencies and not acquired fibrinogen deficiencies [53]. As a result, cryoprecipitate is primarily used as a concentrated source of fibrinogen in the setting of acquired fibrinogen deficiencies: massive blood loss from trauma, hemorrhagic obstetric complications, liver transplant, and DIC [54]. Fibrinogen is the most abundant coagulation factor in plasma. However, it is highly susceptible to hemodilution from fluid resuscitation and blood loss, both of which are common in the setting of massive transfusion. It is the earliest clotting factor to become depleted in hemorrhage and thus is targeted for replacement in such patients [55]. Cryoprecipitate maintains a place in the setting of major hemorrhage from trauma. Trauma-induced coagulopathy is a phenomenon resulting in accelerated fibrinolysis, induced hypofibrinogenemia, and subsequent dysfibrinogenemia [56,57]. It heralds increasing transfusion requirements and mortality as acquired hypofibrinogenemia is associated with coagulopathy and inferior outcomes in hemorrhage control [55,58]. As a result, cryoprecipitate is still often integral in massive transfusion protocols as a fibrinogen source. It can be utilized when plasma fibrinogen is found to be less than 150 to 200 mg/dL or viscoelastic test values indicate a functional fibrinogen deficit [55,59]. A retrospective review of US Army combat soldiers who received massive transfusions (10 or more packed red blood cell transfusions in 24 hours) showed improved mortality with a higher ratio of cryoprecipitate to red blood cells [60]. While a limiting factor may be time-to-administration after thawing the product, early supplementation has been shown to be feasible in trauma patients [61].

Cryoprecipitate still has a role in obstetric hemorrhage when fibrinogen is found to be less than 100 mg/dL. However, many institutions are moving toward use of fibrinogen concentrate, given ease of use and limited infection risks with this product. Fibrinogen concentrate has been shown to be as efficacious as cryoprecipitate in correcting hypofibrinogenemia identified in major obstetric hemorrhage [62]. Given current guidelines indicating focus on single factor replacement, cryoprecipitate's role in obstetric hemorrhage may soon become historical or limited to resource-constrained settings.

Dysfibrinogenemia has been observed in various states of liver disease, ranging from cirrhosis, biliary obstruction, or acute and chronic liver failure [63]. Furthermore, when synthetic liver function is compromised in such disease states, fibrinogen synthesis is reduced. During liver transplant, the ischemic liver graft releases tissue plasminogen activator that disseminates into circulation after reperfusion. The ensuing fibrinolysis diminishes fibrinogen levels which may promote intraoperative hemorrhage. Thus,

cryoprecipitate is advised in the setting of clinically significant bleeding after liver transplant with a target fibrinogen level of 150 to 200 mg/dL [64].

The consumptive coagulopathy that occurs from DIC is among the most common causes of acquired hypofibrinogenemia. DIC often results secondary to an underlining disorder such as malignancy (including due to treatment of acute lymphoblastic leukemia with asparaginase), infection, trauma, or complication of pregnancy. Cryoprecipitate is often provided to patients with fibrinogen levels below 100 to 150 mg/dL to mitigate hemorrhagic complications.

There are various genetic defects that result in undetectable circulating fibrinogen and a state of afibrinogenemia. Congenital hypofibrinogenemia is typically seen in heterozygous carriers of afibrinogenemia mutations [65]. International guidelines advise use of cryoprecipitate only if fibrinogen concentrates are unavailable [66,67].

The current FDA and AABB practice guidelines mandate that cryoprecipitate be transfused within 4 hours of thawing (or 6 hours if prepoled in a closed system prior to freezing) [48]. These current guidelines were established in the 1970s to ensure adequate FVIII levels for treatment of hemophilia A and reduced risk of bacterial growth. However, given that cryoprecipitate is now mostly limited to use in hemorrhage, recent studies have sought to expand this shelf life and expand availability in emergencies. It has been recently shown that refrigerated cryoprecipitate retains hemostatic function for 14 days after thawing. Moreover, the fibrinogen concentration was not significantly changed with storage at 4 weeks in room temperature or in a refrigerator for 5 weeks after thawing [68]. Even longer shelf life may be possible through use of lyophilization and pathogen reduction technologies.

#### *The uses of immunoglobulins in hematologic diseases*

Immunoglobulin therapy, commonly referred to as IVIG, requires extensive manipulation of plasma and is one of the most processed transfusion products used for the treatment of hematologic diseases. From collection to packaging, this product requires months of complex manufacturing steps to prepare it from the pooled plasma of many donors [69]. Additionally, the increased demand from both on- and off-label uses has contributed to the ongoing national shortage of this transfusion product [70].

The mechanism of action for this class of products depends on the hematologic condition for which it is used and the specific formulation. It can be used as a replacement therapy for primary or secondary immunodeficiencies or as a specific therapy for certain pathogens (ie, rabies, hepatitis B, etc.) when collected and used as "hyperimmune" globulin. Furthermore, immunoglobulins can occupy and saturate Fc receptors of the reticuloendothelial system, thereby reducing destruction of antibody-covered neutrophils, RBCs, or platelets, explaining their use in conditions like immune thrombocytopenic purpura (ITP). However, there is also evidence that infused immunoglobulins can affect the overall function of the immune system through its direct interaction with T- and B-cells [71], interference in the maturation and differentiation of dendritic cells [72], reduction in proinflammatory monocytes and their secreted cytokines [73], or the inhibition of the expression of the IgG receptor itself, and thus exert broad immunomodulatory effects [71].

Primary humoral immunodeficiency is a congenital loss of B-cell function resulting in lack of antibody production and loss of humoral immunity. These congenital diseases include congenital agammaglobulinemia, common variable immunodeficiency, X-linked agammaglobulinemia, Wiskott-Aldrich syndrome, and severe combined immunodeficiencies [74].

Secondary humoral immunodeficiency is an acquired loss of B-cell function secondary to autoimmune destruction and dysfunction or hematological malignancies. IVIG therapy can decrease the frequency of bacterial infections in hematological malignancies, like chronic lymphocytic leukemia and multiple myeloma, as well as cytomegalovirus, interstitial pneumonia, and graft-versus-host disease in allogeneic bone marrow transplant patients [69]. However, the frequency of severe bacterial infections, fungal, and viral infections, and mortality were not significantly reduced in chronic lymphocytic leukemia patients. Further analysis reveals that IVIG therapy increases treatment costs for relatively minor decreases in infection-free days. Moreover, the development of very effective anticytomegalovirus medications, more sophisticated graft-versus-host disease prophylaxis, and lack of survival advantage has reduced the use of this treatment for bone marrow transplant patients [75].

Use of IVIG in autoimmunity continues to generate mixed results. Immune thrombocytopenia (ITP) is the primary FDA indication for use of IVIG. This treatment combined with high-dose corticosteroids can rapidly increase platelet counts in hours to days when a rapid rise in platelets are needed. Moreover, the anti-Rh(D) immunoglobulin can be used in Rh (+) patients that do not have a concomitant hemolytic anemia. As for other autoimmune cytopenias (ie, refractory disease) associated with either pediatric, adult, or perinatal conditions, the evidence is not as strong and suggested use is limited. IVIG is also indicated for treatment of chronic inflammatory demyelinating polyneuropathy (CIDP) [76]. As for other systemic autoimmunity conditions, only dermatomyositis and severe, refractory juvenile idiopathic arthritis have convincing evidence supporting use. This is due to lack of trials with this treatment as well as breakthrough developments in biologic therapies [69].

Thrombosis is the major complication associated with use of IVIG and has resulted in an FDA “boxed warning.” The risk factors for developing this adverse event include advanced age, prolonged immobilization, hypercoagulable conditions, history of arterial or venous thrombosis, estrogen use, indwelling central venous catheters, hyperviscosity, and cardiovascular risk factors [77]. Also, the rate of infusion and underlying hematological malignancy may affect this risk as well [78,79]. Furthermore, in 2010, an investigation into this risk showed that certain lots of IVIG were contaminated with kallikrein and Factor Xla, with Factor Xla deduced to be the major cause of impurity-driven thrombotic events [80]. Investigation into specific products associated with thrombosis revealed Octagam to be the offending agent, and it was removed from the market. Once the manufacturing process was improved, however, it was re-introduced with no increased rate of drug-induced thrombosis [81].

Renal dysfunction and acute kidney injury are also associated with IVIG use. Those at increased risk include those with preexisting chronic kidney disease, diabetes mellitus, age greater than 65, volume depletion, sepsis, paraproteinemia, or patients receiving nephrotoxic drugs [82]. Sucrose-containing products seem to be the most closely associated with this adverse event. The mechanism is likely osmotic nephrosis, where tubular cells absorb the large sugar load, become vacuolated, swell, and develop tubular nephropathy [83]. This can be minimized through the selection of sucrose-free IVIG if available and if not, infusion of the product at  $\leq 3$  mg sucrose/kg/min or use of lower concentrated product (ie, 5%), dividing the dose to 500 mg/kg/day [84]. Case reports suggest that IVIG-induced hemolysis from a sucrose-free IVIG product led to pigment-induced nephropathy and may be an unrecognized cause of acute kidney injury as well [85].

The most common hematologic adverse event in use of IVIG is hemolytic anemia. The mechanism of action is the presence of anti-A and anti-B antibodies (isohemagglutinins) in the IVIG

product. The major risk factors for developing this are high-dose infusions ( $>1$  g/kg/day, doses  $>100$  g), female sex, and non-O blood group [86]. Manufacturers have identified methods for reducing isohemagglutinins which may reduce these risks [87]. Other strategies available to clinicians to reduce hemolysis risk include switching brands or lots if possible as well as subcutaneous administration [88].

Recent data suggest potential new trends in use of IVIG. As indicated on the website clinicaltrials.gov, recent efforts in IVIG research are focused on exploring use of these products in infectious or autoimmune processes. Evidence of activity in mixtures of antibodies then stimulate efforts to isolate the specific therapeutic antibodies and targets and use these to manufacture disease-specific immune globulin or the development of monoclonal antibodies to specific targets. Regrettably, there have been no major developments in the recent past for the routine use of IVIG, but certain areas continue to be a focus for improving its utility. One such area is the use of subcutaneous immunoglobulins for maintenance therapy in CIDP as well as other neuromuscular disorders like multifocal motor neuropathy [89]. Recent advances in Kawasaki’s disease in children show that the addition of steroids to IVIG decreases development of coronary artery anomalies [90]. Promising data also exist for the use of IVIG to treat toxic epidermal necrolysis (TEN, or Stevens-Johnson Syndrome), but further randomized controlled trial data are needed [91].

## **The uses of clotting factor concentrate in hematologic diseases**

### *Replacement of clotting factors*

#### *Hemophilia*

A discussion of clotting factor therapies inevitably begins with hemophilia A (congenital Factor VIII deficiency) and hemophilia B (congenital Factor IX deficiency) as the classic cases of bleeding disorder mutations. These have been observed for centuries (if not millennia) [Reference: PMID 24513149], but identification of the responsible proteins did not occur until characterization of the coagulation cascade in the 1950s and 1960s [92]. Prior to that, factor replacement therapies were only achieved inefficiently by blood or plasma transfusions, but in the late 1960s and 1970s, cryoprecipitate, delivering a more concentrated dose of plasma factors, and Factor VIII and IX concentrates were available [93]. By the end of the 1990s, recombinant Factor VIII and IX were widely used, and further improvements were made by mitigating the immune response to transfused factors (which was also critical for acquired hemophiliacs) [93]. Multiple entries at clinicaltrials.gov demonstrate that new formulations of recombinant Factor VIII and IX are constantly being evaluated.

While the gains made over the past century have greatly improved longevity and quality of life for hemophiliacs, more recent advancements have focused on treating the diseases at their roots by gene therapy [94,95]. The methods and modalities by which gene therapy have been conducted have varied, ranging from viral vector delivery [96], stem cell transplant [97,98], CRISPR/Cas editing [99,100], and synthetic transgene development (all primarily in animal models to date) [101]. There are more than 30 active gene therapy trials for Factor VIII and IX replacement currently listed at clinicaltrials.gov.

Additionally, a unique alternative to factor replacement has emerged for Factor VIII: emicizumab, a bispecific chimeric monoclonal antibody, will mimic the cofactor activity of Factor VIII by co-locating Factor IXa and Factor X to catalyze clot formation [102-104]. This is a major advancement in the field since it is not affected by alloantibodies that commonly appear with repeated lifelong usage of factor concentrates and avoids usage of “bypass agents” like prothrombin complex concentrates or recombinant

Factor VIIa (each discussed below), both of which can result in complications for hemophilia A patients.

#### Fibrinogen

Fibrinogen plays a central role in hemostasis as the substrate by which clots are formed. Deficiencies of a congenital or acquired nature have significant morbidity and mortality effects, and therefore supplementation is required in these patients [105,106]. Plasma and cryoprecipitate have both been used as replacement therapies for decades, but neither of these is ideal, requiring large volumes (less so in cryoprecipitate), risking pathogen infection, and requiring lengthy thawing [107,108].

Alternatively, fibrinogen concentrate products are currently on the market with still others in development, although these are more expensive than plasma or cryoprecipitate [109]. These are currently indicated for congenital fibrinogen deficiency, including afibrinogenemia and hypofibrinogenemia. Studies have shown that FC usage reduces bleeding and transfusion requirements [110].

However, fibrinogen concentrates are not indicated for acquired hypofibrinogenemia, bleeding associated with aortic reconstruction and deep hypothermic circulatory arrest, dysfibrinogenemia, obstetric hemorrhage including postpartum hemorrhage in persons without congenital fibrinogen deficiency, perioperative or trauma-associated hemorrhage (without congenital fibrinogen deficiency). But several studies have been completed for the above off-label usages indications [111-113]; there are also currently multiple trials on clinicaltrials.gov recruiting for usage in pediatric cardiac surgery [NCT03884725, NCT04376762], traumatic brain injury [NCT03304899], severe traumatic hemorrhage (including pediatric and prehospital usage) [NCT04149171, NCT03508141], and acquired hypofibrinogenemia [NCT03444324]. Additional approved usages may appear in the near future. For purposes of severe bleeding, an interesting approach has been the implementation of liquid fibrin glue or stiff fibrin patches containing freeze-dried concentrates of fibrinogen, FXIII, fibronectin, and thrombin [114].

#### Prothrombin

Prothrombin is available in the form of a concentrate of multiple factors, referred to as prothrombin complex concentrate, and it exists in 2 forms: 3-factor PCC (3F-PCC) consisting of Factors II (prothrombin), IX, and X, as well as Protein C and S, and 4-factor PCC (4F-PCC) that also contains Factor VII [115]. Its indication is currently only for urgent reversal of the VKA warfarin in adults with acute major bleeding; however, multiple off-label uses exist including reversal of direct oral anticoagulants in major bleeding or surgery [116,117], treatment of bleeding in congenital deficiencies of any of the coagulation factors found in PCC (the vitamin K-dependent factors), prophylactic usage to reduce perioperative bleeding and reduce transfusion requirements [118-120], and in traumatic bleeding alongside FFP to correct coagulopathy [121-123]. Clinicaltrials.gov lists multiple ongoing studies investigating PCC for usage in reducing perioperative bleeding [NCT02740335, NCT04244981, NCT03341156] as well as treatment of coagulopathy and bleeding including the prehospital setting [NCT03981484, NCT04019015].

#### Factor V

Factor V is a component of the prothrombinase complex that increases the rate of conversion of prothrombin to thrombin, rapidly resulting in clot formation upon activation. Factor V deficiency (Owren's disease) is rare and acquired Factor V deficiency is even rarer, resulting in increased bleeding risk [124]. Treatment of these patients is through supplementation with plasma, as no Factor V concentrates exist currently. Alternatively, platelet transfusion has been shown to be effective in treating Factor V deficiency due

to the presence of activated Factor V in platelets. A new plasma-derived Factor V concentrate is currently under investigation but not yet in clinical trials [125,126]. More common are mutations such as Factor V Leiden which result in resistance to degradation and increased risk of thrombosis, but these patients are treated with anticoagulants rather than additional clotting factors [127]. A mutant Factor V, resistant to cleavage by Protein C is under investigation to treat severe bleeding [128-131].

#### Factor VII

Factor VII is activated when blood is exposed to tissue factor found in the extravascular space, resulting in downstream cascade activation of the tissue factor-activated pathway (AKA extrinsic pathway). A recombinant activated Factor VII (rFVIIa) has been available for several years, with FDA-approved indications of usage for treatment of bleeding episodes and perioperative management in adults and children with hemophilia A or B with inhibitors (or acquired hemophilia) [132,133], congenital Factor VII deficiency [134], and Glanzmann's thrombasthenia with refractoriness to platelet transfusions [135]. Due to several severe adverse thrombotic events associated with rFVIIa shown in trials, off-label usage of the drug (such as in traumatic hemorrhage) has greatly diminished from its initially perceived potential [136]. However, warnings and precautions have been issued by the FDA to prevent or reduce the risk of thrombosis for vulnerable populations, especially considering interactions with other procoagulant drugs. Multiple reports have demonstrated the safety of rFVIIa when used appropriately [137-140], and clinicaltrials.gov lists at least one study currently evaluating rFVIIa in hemorrhagic stroke [NCT03496883], so rFVII clinical development is likely to continue.

#### Factor X

Factor X is critical for efficient conversion of prothrombin to thrombin in the clotting cascade, and supplementation with Factor X concentrates or PCCs as described above is indicated in FX-deficient patients for prophylactic reduction of spontaneous bleeding, control of bleeding episodes, and perioperative management of bleeding in mild or moderate FX deficiency. Clinicaltrials.gov indicates that a study of FX concentrates in severe Factor X deficient patients undergoing surgery will be completed by 2021 [NCT03161626].

#### Factor XI

Factor XI is responsible for downstream activation of Factor IX in the contact activation pathway, and deficiency in this protein is rare (known as hemophilia C), sometimes resulting in abnormal bleeding. A plasma-derived concentrate has existed for over 2 decades now with a good safety profile for its limited patient population [141]. Expansion of extracorporeal life support system usage has demonstrated that Factor XI concentrate exacerbates the already prevalent clot formation in the bypass circuit [142].

#### Factor XII

Factor XII deficiencies are rare and have little impact on the patient [142]. No treatments are necessary.

#### Factor XIII

Factor XIII is responsible for crosslinking and stabilizing clot formation, and deficiency in this protein, while rare, results in bleeding disorders that require replacement. Both plasma-derived and recombinant Factor XIII are prescribed for routine prophylactic treatment and perioperative management of bleeding. Factor XIII is difficult to measure by current methods [143], but at least one study has been planned on clinicaltrials.gov to evaluate the level of Factor XIII in trauma to determine the need for acute supplementation [NCT03634215].

### ADAMTS-13

ADAMTS-13 is a metalloproteinase responsible for maintaining von Willebrand factor at appropriate multimeric lengths. Severe deficiency in this enzyme's function results in TTP [144], an emergent condition requiring plasma exchange in both acquired TTP (to remove ADAMTS-13 autoantibodies) or congenital TTP (to provide limited but sufficient levels of ADAMTS-13) [145]. Plasma exchange is an invasive procedure that has many adverse effects, particularly for pediatric patients, but newly developed recombinant ADAMTS-13 has provided a safe and sustainable alternative [29,146]. Prophylactic and therapeutic trials are ongoing.

### Replacement of anticoagulant factors

#### Antithrombin

Produced by the liver, antithrombin (AT) is a vitamin K-independent glycoprotein that blocks coagulation by irreversibly inactivating various enzymes of the coagulation cascade, most notably Factor Xa and thrombin. AT deficiency can be acquired or congenital and results in insufficient endogenous anticoagulation and subsequent predisposition to increased thrombosis [147]. Acquired AT deficiency can occur as a result of increased consumption (eg, DIC, AML), decreased synthesis (eg, liver disease), or can be drug-induced (eg, L-asparaginase, heparin). In the United States, 2 AT concentrates exist—Thrombate III (Grifols/Talecris), a pooled lyophilized product derived from human plasma and ATryn (Ovation), a recombinant AT product derived from transgenic goats [148]. AT concentrates have been proven safe and effective in patients with AT deficiency and acute VTE [149,150]. Thrombate III is FDA-approved to treat hereditary AT deficiency and for the treatment and prevention of thromboembolism that arises from surgical or obstetrical procedures [151], whereas ATryn is used in hereditary AT-deficient patients for the prevention of perioperative and peripartum thromboembolic events [152].

AT concentrates have also found success off-label for treatment and prevention of veno-occlusive disease [153,154] and for extracorporeal membrane oxygenation patients [155,156].

#### Recombinant human-soluble thrombomodulin

Akin to thrombomodulin, recombinant human-soluble thrombomodulin (rhTM) binds thrombin via a functional extracellular domain, activates protein C, and thwarts excessive coagulation [157]. rhTM has been assessed as a novel therapeutic agent for management of DIC. Several studies in Japan have evaluated the usefulness of rhTM in sepsis-induced DIC patients [158–161], but results have been conflicting. One multicenter retrospective study evaluating rhTM administration in DIC patients showed no reduction in mortality, and a meta-analysis of RCTs generally supported this conclusion [159,161]. Contrastingly, results from a multicenter retrospective study showed improvement of survival outcomes in sepsis-induced DIC patients who received rhTM [160]. In the United States, a randomized, double-blind placebo-controlled phase 3 clinical trial evaluating high-risk patients (ClinicalTrials.gov Identifier: NCT01598831) recently reported no significant reduction in 38-day mortality of sepsis-associated coagulopathic patients who received rhTM (ART-123) over placebo [162]. Studies evaluating sepsis-induced DIC patients with severe respiratory failure (eg, ARDS) have also been performed with both positive and negative correlations in reducing mortality [161,163]. Further study of this product is required to determine its role in treatment of coagulopathy.

#### Protein C

Produced by hepatocytes, protein C is a vitamin K-dependent anticoagulation factor that becomes activated by thrombin [164]. Activated protein C along with its cofactor, protein S, targets

Factors Va and VIIIa for inhibition to downregulate thrombin generation. The recombinant human activated protein C, Xigris (Eli Lilly), was FDA-approved for treatment of severe sepsis but later withdrawn from the market after clinical trials failed to confirm survival benefit for patients with severe sepsis and septic shock [165–167]. The human Protein C Concentrate, cepritin (Baxter Healthcare Corp.), is currently FDA-indicated for use in patients with severe congenital Protein C deficiency to prevent and treat venous thrombosis and purpura fulminans [168].

#### Protein S

Protein S has multiple functions including as a cofactor for protein C inactivation of Factors Va and VIIIa. Protein S deficiency is a rare disorder resulting in an increased risk of thrombosis [169,170]. Acute treatment with heparin and prophylactic treatment with warfarin are common after the first thrombotic incident, but lifelong usage of warfarin is not without risks [171]. There are currently no protein S concentrates available. Direct oral anticoagulants have been used successfully in cases where warfarin is ineffective or side effects become too burdensome [172,173], but no randomized controlled trials have been conducted to validate their true effectiveness [174].

#### C1 inhibitor

Heredity angioedema is a congenital deficiency or dysfunction of C1 esterase inhibitor (C1-INH). While C1-INH is a complement factor rather than a coagulation factor, its loss directly affects high-level elements of the contact-activated (intrinsic) pathway, including Factor XII, kallikrein, and high molecular weight kininogen (whose cleavage product is bradykinin). Prophylactic and acute treatment of the disease has included the prominent antifibrinolytic tranexamic acid, although this is an off-label use of the drug reserved for those who cannot tolerate the androgens dana-zol and stanozolol which are effective treatments with a number of significant side effects [175,176]. In acute flare ups, plasma has been given to restore C1-INH, but this is less than ideal as it contains the panoply of proteases that can serve to exacerbate the attack [177,178]. Therefore, more recently developed therapies such as icatibant (a bradykinin receptor antagonist), ecallantide (a kallikrein inhibitor), and plasma-derived or recombinant C1-INH are currently used with safer profiles for hereditary angioedema patients [179–181]. New alternatives include the monoclonal antibody lanadelumab which functions by directly blocking bradykinin [182]. A multitude of other inhibitors, antibodies, and other drugs are currently under investigation.

#### Alpha-2 antiplasmin

Initiation of fibrinolysis is orchestrated by the generation of plasmin from plasminogen. The fibrinolysis process must be properly regulated to prevent excess bleeding and tissue damage. In combination with plasminogen activator inhibitor and thrombin activatable fibrinolysis inhibitor,  $\alpha_2$ -antiplasmin ( $\alpha_2$ -AP) serves as a major regulator of fibrinolysis by acting on plasminogen [183].  $\alpha_2$ -AP has most recently been utilized for reduction of bleeding secondary to thrombolytic therapy without affecting thrombolysis [184]. Additional efforts to produce variants of  $\alpha_2$ -AP for therapeutic benefit are also under evaluation [183,184].

### Conclusions

Plasma and its derivatives are widely used in a variety of congenital and acquired hematologic disorders. Major trends in use of these products can be identified. Plasma and cryoprecipitate use are becoming most frequent in the management of bleeding or complex coagulopathies like DIC, or less frequently, liver disease, since these disorders affect a large number of plasma proteins,

and specific therapies are lacking. Plasma use is decreasing for the purpose of warfarin reversal, prophylaxis prior to procedures and in replacement of coagulation factors or other plasma proteins as concentrates become available. Plasma use in TTP also may eventually decrease if ADAMTS13 products are successfully developed. Immunoglobulin therapy remains a bedrock of treatment of humoral immunodeficiency states, ITP and CIDP, and off-label use is common in other autoimmune disorders. Use of IVIG may decrease as more targeted immunotherapies are developed. Significant advances have been made to replace many pro- and anticoagulant factors as well as other plasma proteins. As development efforts continue in these areas, recombinant products or engineered proteins such as bi-specific antibodies will continue to displace plasma concentrates. Further early stage work is needed to optimize therapies and inform randomized trials for complex conditions like DIC and trauma-induced coagulopathy.

## Conflicts of interest

The authors declare no conflicts of interest.

## References

- [1] American Association of Blood Banks (AABB). Circular of information for the use of human blood and blood components. 2017.
- [2] Octapharma. Pooled plasma (human), solvent/detergent treated solution for intravenous infusion. Octapharma; 2013.
- [3] Mueller MM, Bomke B, Seifried E. Fresh frozen plasma in patients with disseminated intravascular coagulation or in patients with liver diseases. *Thromb Res* 2002;107(Suppl 1):S9–17.
- [4] Rockey DC. Endothelial dysfunction in advanced liver disease. *Am J Med Sci* 2015;349(1):6–16.
- [5] Vairappan B. Endothelial dysfunction in cirrhosis: role of inflammation and oxidative stress. *World J Hepatol* 2015;7(3):443–59.
- [6] Poisson J, Lemoine S, Boulanger C, et al. Liver sinusoidal endothelial cells: physiology and role in liver diseases. *J Hepatol* 2017;66(1):212–27.
- [7] Walborn A, Rondina M, Mosier M, Fareed J, Hoppensteadt D. Endothelial dysfunction is associated with mortality and severity of coagulopathy in patients with sepsis and disseminated intravascular coagulation. *Clin Appl Thromb Hemost* 2019;25:1076029619852163.
- [8] Broze GJ Jr. Tissue factor pathway inhibitor. *Thromb Haemostas* 1995;74(1):90–3.
- [9] Bombeli T, Mueller M, Haeberli A. Anticoagulant properties of the vascular endothelium. *Thromb Haemostas* 1997;77(3):408–23.
- [10] Bjerkvig CK, Strandenes G, Eliasson HS, et al. "Blood failure" time to view blood as an organ: how oxygen debt contributes to blood failure and its implications for remote damage control resuscitation. *Transfusion* 2016;56(Suppl 2):S182–9.
- [11] White NJ, Ward KR, Pati S, Strandenes G, Cap AP. Hemorrhagic blood failure: oxygen debt, coagulopathy, and endothelial damage. *J Trauma Acute Care Surg* 2017;82(6S Suppl 1):S41–9.
- [12] Cap AP, Pidcock HF, Spinella P, et al. Damage control resuscitation. *Military Med* 2018;183(Suppl 2):36–43.
- [13] Borgman MA, Spinella PC, Perkins JG, et al. The ratio of blood products transfused affects mortality in patients receiving massive transfusions at a combat support hospital. *J Trauma* 2007;63(4):805–13.
- [14] Holcomb JB, Tilley BC, Baraniuk S, et al. Transfusion of plasma, platelets, and red blood cells in a 1:1:1 vs a 1:1:2 ratio and mortality in patients with severe trauma: the proper randomized clinical trial. *JAMA* 2015;313(5):471–82.
- [15] Inaba K, Branco BC, Rhee P, et al. Impact of plasma transfusion in trauma patients who do not require massive transfusion. *J Am Coll Surg* 2010;210(6):957–65.
- [16] Schulman S. Clinical practice. Care of patients receiving long-term anticoagulant therapy. *New Engl J Med* 2003;349(7):675–83.
- [17] Baglin TP, Keeling DM, Watson HG. Guidelines on oral anticoagulation (warfarin): third edition—2005 update. *Br J Haematol* 2006;132(3):277–85.
- [18] Makris M, Watson HG. The management of coumarin-induced over-anticoagulation annotation. *Br J Haematol* 2001;114(2):271–80.
- [19] Bux J. Transfusion-related acute lung injury (TRALI): a serious adverse event of blood transfusion. *Vox Sanguin* 2005;89(1):1–10.
- [20] Khan H, Belsher J, Yilmaz M, et al. Fresh-frozen plasma and platelet transfusions are associated with development of acute lung injury in critically ill medical patients. *Chest* 2007;131(5):1308–14.
- [21] Tao J, Bukanova EN, Akhtar S. Safety of 4-factor prothrombin complex concentrate (4f-pcc) for emergent reversal of factor xa inhibitors. *J Intensive Care* 2018;6:34.
- [22] Levy JH, Douketis J, Steiner T, Goldstein JN, Milling TJ. Prothrombin complex concentrates for perioperative vitamin k antagonist and non-vitamin k anti-coagulant reversal. *Anesthesiology* 2018;129(6):1171–84.
- [23] Brekelmans MPA, Ginkel KV, Daams JG, et al. Benefits and harms of 4-factor prothrombin complex concentrate for reversal of vitamin k antagonist associated bleeding: a systematic review and meta-analysis. *J Thrombos Thrombol* 2017;44(1):118–29.
- [24] Keeling D, Baglin T, Tait C, et al. Guidelines on oral anticoagulation with warfarin – fourth edition. *Br J Haematol* 2011;154(3):311–24.
- [25] Makris M, Greaves M, Phillips WS, et al. Emergency oral anticoagulant reversal: the relative efficacy of infusions of fresh frozen plasma and clotting factor concentrate on correction of the coagulopathy. *Thromb Haemostas* 1997;77(3):477–80.
- [26] Scully M, Cataland SR, Peyvandi F, et al. Caplacizumab treatment for acquired thrombotic thrombocytopenic purpura. *New Engl J Med* 2019;380(4):335–46.
- [27] Chander DP, Loch MM, Cataland SR, George JN. Caplacizumab therapy without plasma exchange for acquired thrombotic thrombocytopenic purpura. *New Engl J Med* 2019;381(1):92–4.
- [28] Blombery P, Scully M. Management of thrombotic thrombocytopenic purpura: current perspectives. *J Blood Med* 2014;5:15–23.
- [29] Scully M, Knobl P, Kentouche K, et al. Recombinant adams-13: first-in-human pharmacokinetics and safety in congenital thrombotic thrombocytopenic purpura. *Blood* 2017;130(19):2055–63.
- [30] Engelhardt DL, Sarnoski J. Variations in the cell-free translating apparatus of cultured animal cells as a function of time during cell growth. *J Cell Physiol* 1975;86(1):15–29.
- [31] Nascimento B, Callum J, Rubenfeld G, et al. Clinical review: Fresh frozen plasma in massive bleedings - more questions than answers. *Crit Care (Lond, Engl)* 2010;14(1):202.
- [32] Bernal W, Caldwell SH, Lisman T. Nails in the coffin of fresh frozen plasma to prevent or treat bleeding in cirrhosis? *J Hepatol* 2020;72(1):12–13.
- [33] Desborough MJ, Stanworth SJ, Curry NS. Uses and abuses of fresh frozen plasma for the treatment of bleeding. *Clin Med (Lond, Engl)* 2013;13(2):200–2.
- [34] Stanworth SJ. The evidence-based use of ffp and cryoprecipitate for abnormalities of coagulation tests and clinical coagulopathy. *Hematology. American Society of Hematology Education Program*; 2007. p. 179–86.
- [35] Stanworth SJ, Brunskill SJ, Hyde CJ, McClelland DB, Murphy MF. Is fresh frozen plasma clinically effective? A systematic review of randomized controlled trials. *Br J Haematol* 2004;126(1):139–52.
- [36] Arabi YM, Hajee AH, Luke T, et al. Feasibility of using convalescent plasma immunotherapy for Mers-CoV infection, Saudi Arabia. *Emerg Infect Dis* 2016;22(9):1554–61.
- [37] Cheng Y, Wong R, Soo YOY, et al. Use of convalescent plasma therapy in SARS patients in Hong Kong. *Eur J Clin Microbiol Infect Dis* 2005;24(1):44–6.
- [38] Ahn JY, Sohn Y, Lee SH, et al. Use of convalescent plasma therapy in two COVID-19 patients with acute respiratory distress syndrome in Korea. *J Korean Med Sci* 2020;35(14):e149.
- [39] van Griensven J, Edwards T, de Lamballerie X, et al. Evaluation of convalescent plasma for Ebola virus disease in Guinea. *New Engl J Med* 2016;374(1):33–42.
- [40] Mair-Jenkins J, Saavedra-Campos M, Baillie JK, et al. The effectiveness of convalescent plasma and hyperimmune immunoglobulin for the treatment of severe acute respiratory infections of viral etiology: a systematic review and exploratory meta-analysis. *J Infect Dis* 2015;211(1):80–90.
- [41] Duan K, Liu B, Li C, et al. Effectiveness of convalescent plasma therapy in severe COVID-19 patients. *Proc Natl Acad Sci U S A* 2020;117(17):9490–6.
- [42] Shen C, Wang Z, Zhao F, et al. Treatment of 5 critically ill patients with COVID-19 with convalescent plasma. *JAMA* 2020;323(16):1582–9.
- [43] Ye M, Fu D, Ren Y, et al. Treatment with convalescent plasma for COVID-19 patients in Wuhan, China. *J Med Virol* 2020. doi:10.1002/jmv.25882. [Online ahead of print].
- [44] US Food and Drug Administration. Investigational COVID-19 convalescent plasma – emergency inds. 2020. Available from: <https://www.fda.gov/vaccines-blood-biologics/investigational-new-drug-ind-or-device-exemption-ide-process-cber/recommendations-investigational-covid-19-convalescent-plasma>.
- [45] Yeh KM, Chiueh TS, Siu LK, et al. Experience of using convalescent plasma for severe acute respiratory syndrome among healthcare workers in a Taiwan hospital. *J Antimicrob Chemother* 2005;56(5):919–22.
- [46] Ko JH, Seok H, Cho SY, et al. Challenges of convalescent plasma infusion therapy in middle east respiratory coronavirus infection: a single centre experience. *Antivir Ther* 2018;23(7):617–22.
- [47] Callum JL, Karkouti K, Lin Y. Cryoprecipitate: the current state of knowledge. *Transfus Med Rev* 2009;23(3):177–88.
- [48] American Association of Blood Banks (AABB). Circular of information for the use of human blood and blood components. 2013.
- [49] Arya RC, Wander GS, Gupta P. Blood component therapy: which, when and how much. *J Anaesthesiol Clin Pharmacol* 2011;27:278–84.
- [50] Sørensen B, Bevan D. A critical evaluation of cryoprecipitate for replacement of fibrinogen. *Br J Haematol* 2010;149(6):934–43.
- [51] Pool JG, Hershgold Ej, Pappenhaben AR. High-potency antihæmophilic factor concentrate prepared from cryoglobulin precipitate. *Nature* 1964;203:312.
- [52] Prevention Center for Disease Control and PreventionCDC. Treatment of hemophilia. 2020. Available from: <https://www.cdc.gov/ncbddd/hemophilia/treatment.html>.
- [53] Costa-Filho R, Hochleitner G, Wendt M, Teruya A, Spahn DR. Over 50 years of fibrinogen concentrate. *Clin Appl Thromb Hemostas* 2016;22(2):109–14.

- [54] Yang L, Stanworth S, Baglin T. Cryoprecipitate: an outmoded treatment? *Transfus Med* 2012;22:315–20.
- [55] Rourke C, Curry N, Khan S. Fibrinogen levels during trauma hemorrhage, response to replacement therapy, and association with patient outcomes. *J Thromb Haemost* 2012;10(7):1342–51.
- [56] Kattula S, Byrnes JR, Wolberg AS. Fibrinogen and fibrin in hemostasis and thrombosis. *Arterioscler Thromb Vasc Biol* 2017;37:e13–21.
- [57] Winearls J, Campbell D, Hurn C, et al. Fibrinogen in traumatic haemorrhage: a narrative review. *Injury* 2017;48:230–42.
- [58] Montgomery HR, Butler FK, Kerr W. TCCC guidelines comprehensive review and update: TCCC guidelines change 16-03. *J Spec Oper Med* 2017;17:21–98.
- [59] Spahn DR, Bouillon B, Cerny V. Management of bleeding and coagulopathy following major trauma: an updated European guideline. *Crit Care (Lond, Engl)* 2013;17(2):R76.
- [60] Stinger HK, Spinella PC, Perkins JG. The ratio of fibrinogen to red cells transfused affects survival in casualties receiving massive transfusions at an army combat support hospital. *J Trauma* 2008;64(2):S79.
- [61] Curry N, Rourke C, Davenport R. Early cryoprecipitate for major haemorrhage in trauma: a randomised controlled feasibility trial. *Br J Anaesth* 2015;115:76–83.
- [62] Ahmed S, Harrity C, Johnson S, et al. The efficacy of fibrinogen concentrate compared with cryoprecipitate in major obstetric haemorrhage—an observational study. *Transfus med* 2012;22(5):344–9.
- [63] Francis JL, Armstrong DJ. Acquired dysfibrinogenaemia in liver disease. *J Clin Pathol* 1982;35(3):667.
- [64] Bezinover D, Dirkmann D, Findlay J, et al. Perioperative coagulation management in liver transplant recipients. *Transplantation* 2018;102(4):578–92.
- [65] Asselta R, Plate M, Robusto M, et al. Clinical and molecular characteristics of 21 patients affected by quantitative fibrinogen deficiency. *Thromb Haemost* 2015;113:567.
- [66] Peyvandi F, Palla R, Menegatti M, et al. Rare bleeding disorders: general aspects of clinical features, diagnosis, and management. *Semin Thromb Hemost* 2009;35(4):349–55.
- [67] Palla R, Peyvandi F, Shapior AD. Rare bleeding disorders: diagnosis and treatment. *Blood* 2015;125(13):2052–61.
- [68] Fenderson JL, Meledeo MA, Rendo MJ, et al. Hemostatic characteristics of thawed, pooled cryoprecipitate stored for 35 days at refrigerated and room temperatures. *Transfusion* 2019;59(S2):1560–7.
- [69] Perez EE, Orange JS, Bonilla F, et al. Update on the use of immunoglobulin in human disease: a review of evidence. *J Allergy Clin Immunol* 2017;139(3S):S1–S46.
- [70] US Food and Drug Administration. Information about immune globulin (human) product shortage. 2020; (04.30.20). Available from: <https://www.fda.gov/vaccines-blood-biologics/safety-availability-biologics/information-about-immune-globulin-human-product-shortage>.
- [71] Savage W, Chou ST. Transfusion medicine. In: Cuker A, Altman JK, Gerds AT, Wun T, editors. American Society of Hematology self-assessment program. 7th ed Washington, DC: American Society of Hematology; 2019. p. 350–86.
- [72] Kaveri SV, Lacroix-Desmazes S, Bayry J. The antiinflammatory IGG. *New Engl J Med* 2008;359(3):307–9.
- [73] Siedlar M, Strach M, Bukowska-Strakova K, et al. Preparations of intravenous immunoglobulins diminish the number and proinflammatory response of CD14+ CD16++ monocytes in common variable immunodeficiency (CVID) patients. *Clin Immunol* 2011;139(2):122–32.
- [74] Privigen. (Immune Globulin Intravenous [human], 10% liquid.). [package insert]. Kankakee, IL: CSL Behring; 2017.
- [75] Darabi K, Abdel-Wahab O, Dzik WH. Current usage of intravenous immune globulin and the rationale behind it: the Massachusetts General Hospital data and a review of the literature. *Transfusion* 2006;46(5):741–53.
- [76] Privigen. (Immune Globulin Intravenous [human], 10% liquid.) [package insert]. Kankakee, IL: CSL Behring; 2007.
- [77] Gamunex-C (Immune Globulin Injection [human]. 10% caprylate/chromatography purified) [package insert]. Research Triangle Park, NC: Grifols Therapeutics Inc; 2016.
- [78] Ammann EM, Jones MP, Link BK, et al. Intravenous immune globulin and thromboembolic adverse events in patients with hematologic malignancy. *Blood* 2016;127(2):200–7.
- [79] Caress JB, Hobson-Webb L, Passmore LV, Finkbiner AP, Cartwright MS. Case-control study of thromboembolic events associated with iv immunoglobulin. *J Neurol* 2009;256(3):339–42.
- [80] Etscheid M, Breitner-Ruddock S, Gross S, et al. Identification of Kallikrein and Fxia as impurities in therapeutic immunoglobulins: implications for the safety and control of intravenous blood products. *Vox Sanguin* 2012;102(1):40–6.
- [81] Funk MB, Gross N, Gross S, et al. Thromboembolic events associated with immunoglobulin treatment. *Vox Sanguin* 2013;105(1):54–64.
- [82] Gammagard liquid. (Immune Globulin Intravenous [human] 10%) [package insert]. Westlake Village, CA: Baxter International Inc; 2005.
- [83] Dickenmann M, Oettl T, Mihatsch MJ. Osmotic nephrosis: acute kidney injury with accumulation of proximal tubular lysosomes due to administration of exogenous solutes. *Am J Kidney Dis* 2008;51(3):491–503.
- [84] Lozerton P, Not A, Theaudin M, et al. Safety of intravenous immunoglobulin in the elderly treated for a dysimmune neuromuscular disease. *Muscle Nerve* 2016;53(5):683–9.
- [85] Welles CC, Tamma S, Lafayette RA. Hemoglobinuria and acute kidney injury requiring hemodialysis following intravenous immunoglobulin infusion. *Am J Kidney Dis* 2010;55(1):148–51.
- [86] Daw Z, Padmore R, Neurath D, et al. Hemolytic transfusion reactions after administration of intravenous immune (gamma) globulin: a case series analysis. *Transfusion* 2008;48(8):1598–601.
- [87] Hoefferer L, Glauser I, Gaida A, et al. Isoagglutinin reduction by a dedicated immunoaffinity chromatography step in the manufacturing process of human immunoglobulin products. *Transfusion* 2015;55(Suppl 2):S117–21.
- [88] Quinti I, Pulvirenti F, Milito C, et al. Hemolysis in patients with antibody deficiencies on immunoglobulin replacement treatment. *Transfusion* 2015;55(5):1067–74.
- [89] Dacci P, Riva N, Scarlato M, et al. Subcutaneous immunoglobulin therapy for the treatment of multifocal motor neuropathy: a case report. *Neurol Sci* 2010;31(6):829–31.
- [90] Kobayashi T, Saji T, Otani T, et al. Efficacy of immunoglobulin plus prednisolone for prevention of coronary artery abnormalities in severe Kawasaki disease (raise study): a randomised, open-label, blinded-endpoints trial. *Lancet* 2012;379(9826):1613–20.
- [91] Huang YC, Li YC, Chen TJ. The efficacy of intravenous immunoglobulin for the treatment of toxic epidermal necrolysis: a systematic review and meta-analysis. *Br J Dermatol* 2012;167(2):424–32.
- [92] Franchini M, Mannucci PM. The history of hemophilia. *Semin Thromb Hemost* 2014;40(5):571–6.
- [93] Page D. Comprehensive care for hemophilia and other inherited bleeding disorders. *Transfus Apher Sci* 2019;58(5):565–8.
- [94] VandenDriessche T, Chuah MK. Hemophilia gene therapy: ready for prime time? *Hum Gene Ther* 2017;28(11):1013–23.
- [95] Rodriguez-Merchan EC. What's new in gene therapy of hemophilia. *Curr Gene Ther* 2018;18(2):107–14.
- [96] Nienhuis AW, Nathwani AC, Davidoff AM. Gene therapy for hemophilia. *Mol Ther* 2017;25(5):1163–7.
- [97] Couto DL, Cuqueris J, El Ayoubi R, et al. Hierarchical scaffold design for mesenchymal stem cell-based gene therapy of hemophilia b. *Biomaterials* 2011;32(1):295–305.
- [98] Porada CD, Sanada C, Kuo CJ, et al. Phenotypic correction of hemophilia a in sheep by postnatal intraperitoneal transplantation of FVIII-expressing MSC. *Exp Hematol* 2011;39(12):1124–35.e4.
- [99] Wang L, Yang Y, Breton CA, et al. Crispr/cas9-mediated in vivo gene targeting corrects hemostasis in newborn and adult factor IX-knockout mice. *Blood* 2019;133(26):2745–52.
- [100] Stephens CJ, Lauron Ej, Kashentseva E, et al. Long-term correction of hemophilia b using adenoviral delivery of CRISPR/CAS9. *J Control Release* 2019;298:128–41.
- [101] Brown HC, Zakas PM, George SN, et al. Target-cell-directed bioengineering approaches for gene therapy of hemophilia a. *Mol Ther Methods Clin Dev* 2018;9:57–69.
- [102] Shima M, Hanabusa H, Taki M, et al. Factor VIII-mimetic function of humanized bispecific antibody in hemophilia a. *New Engl J Med* 2016;374(21):2044–53.
- [103] Mahlangu J, Oldenburg J, Paz-Priel I, et al. Emicizumab prophylaxis in patients who have hemophilia a without inhibitors. *New Engl J Med* 2018;379(9):811–22.
- [104] Shima M, Nogami K, Nagami S, et al. A multicentre, open-label study of emicizumab given every 2 or 4 weeks in children with severe haemophilia a without inhibitors. *Haemophilia* 2019;25(6):979–87.
- [105] Kaur J, Jain A. Fibrinogen. Treasure Island, FL: StatPearls Publishing; 2020. [Updated 2020 Jan 16]. In: StatPearls [Internet]. Treasure Island (FL); [Updated 2020 Mar 25]. In: StatPearls [Internet]Jan-. Available from <https://www.ncbi.nlm.nih.gov/books/NBK537184>.
- [106] Grottkau O, Mallaiah S, Karkouti K, Saner F, Haas T. Fibrinogen supplementation and its indications. *Semin Thromb Hemost* 2020;46(1):38–49.
- [107] Tziomalos K, Vakalopoulou S, Perifanis V, Garipidou V. Treatment of congenital fibrinogen deficiency: overview and recent findings. *Vasc Health Risk Manag* 2009;5:843–8.
- [108] Bevan DH. Cryoprecipitate: no longer the best therapeutic choice in congenital fibrinogen disorders? *Thromb Res* 2009;124(Suppl 2):S12–16.
- [109] Okerberg CK, Williams LA 3rd, Kilgore ML, et al. Cryoprecipitate AHF vs. fibrinogen concentrates for fibrinogen replacement in acquired bleeding patients – an economic evaluation. *Vox Sanguin* 2016;111(3):292–8.
- [110] Gollop ND, Chilcott J, Benton A, et al. National audit of the use of fibrinogen concentrate to correct hypofibrinogenaemia. *Transfus Med* 2012;22(5):350–5.
- [111] Matsunaga S, Takai Y, Nakamura E, et al. The clinical efficacy of fibrinogen concentrate in massive obstetric haemorrhage with hypofibrinogenaemia. *Sci Rep* 2017;7:46749.
- [112] Bell SF, Rayment R, Collins PW, Collis RE. The use of fibrinogen concentrate to correct hypofibrinogenaemia rapidly during obstetric haemorrhage. *Int J Obstet Anesth* 2010;19(2):218–23.
- [113] Yamamoto K, Yamaguchi A, Sawano M, et al. Pre-emptive administration of fibrinogen concentrate contributes to improved prognosis in patients with severe trauma. *Trauma Surg Acute Care Open* 2016;1(1):e000037.
- [114] Chiara O, Cimbanassi S, Bellanova G, et al. A systematic review on the use of topical hemostats in trauma and emergency surgery. *BMC Surg* 2018;18(1):68.
- [115] Baskaran J, Lopez RA, Cassagnol M. Prothrombin Complex Concentrate. StatPearls [Internet] [Updated 2020 Mar 25]. Treasure Island, FL: StatPearls Publishing; 2020. Jan-. Available from <https://www.ncbi.nlm.nih.gov/books/NBK539716>.

- [116] Moia M, Squizzato A. Reversal agents for oral anticoagulant-associated major or life-threatening bleeding. *Intern Emerg Med* 2019;14(8):1233–9.
- [117] Desai NR, Cornutt D. Reversal agents for direct oral anticoagulants: considerations for hospital physicians and intensivists. *Hosp Pract* (1995) 1999;47(3):113–22.
- [118] Hashmi NK, Ghadimi K, Srinivasan AJ, et al. Three-factor prothrombin complex concentrates for refractory bleeding after cardiovascular surgery within an algorithmic approach to haemostasis. *Vox Sanguin* 2019;114(4):374–85.
- [119] Quick JA, Meyer JM, Coughenour JP, Barnes SL. Less is more: low-dose prothrombin complex concentrate effective in acute care surgery patients. *Am Surg* 2015;81(6):646–50.
- [120] Lin DM, Murphy LS, Tran MH. Use of prothrombin complex concentrates and fibrinogen concentrates in the perioperative setting: a systematic review. *Transfus Med Rev* 2013;27(2):91–104.
- [121] Matsushima K, Benjamin E, Demetriaides D. Prothrombin complex concentrate in trauma patients. *Am J Surg* 2015;209(2):413–17.
- [122] Jehan F, Aziz H, O'Keefe T, et al. The role of four-factor prothrombin complex concentrate in coagulopathy of trauma: a propensity matched analysis. *J Trauma Acute Care Surg* 2018;85(1):18–24.
- [123] Zeeshan M, Hamidi M, Feinstein AJ, et al. Four-factor prothrombin complex concentrate is associated with improved survival in trauma-related hemorrhage: a nationwide propensity-matched analysis. *J Trauma Acute Care Surg* 2019;87(2):274–81.
- [124] Vetri D, Lumera G, Tarasco S, et al. A case of acquired factor V deficiency in patient with bleeding. *TH Open* 2020;4(2):e77–e9.
- [125] Tabibian S, Shiravand Y, Shams M, et al. A comprehensive overview of coagulation factor V and congenital factor V deficiency. *Semin Thromb Hemost* 2019;45(5):523–43.
- [126] Bulato C, Novembrino C, Anzoletti MB, et al. "In vitro" correction of the severe factor V deficiency-related coagulopathy by a novel plasma-derived factor V concentrate. *Haemophilia* 2018;24(4):648–56.
- [127] Campello E, Spiezia L, Simioni P. Diagnosis and management of factor V Leiden. *Expert Rev Hematol* 2016;9(12):1139–49.
- [128] Gale AJ, Bhat V, Pellequer JL, et al. Safety, stability and pharmacokinetic properties of (super)factor VA, a novel engineered coagulation factor V for treatment of severe bleeding. *Pharm Res* 2016;33(6):1517–26.
- [129] Bhat V, von Drygalski A, Gale AJ, Griffin JH, Mosnier LO. Improved coagulation and haemostasis in haemophilia with inhibitors by combinations of superfactor VA and factor VIIA. *Thromb Haemostas* 2016;115(3):551–61.
- [130] von Drygalski A, Cramer TJ, Bhat V, et al. Improved hemostasis in hemophiliac mice by means of an engineered factor VA mutant. *J Thromb Haemost* 2014;12(3):363–72.
- [131] von Drygalski A, Bhat V, Gale AJ, et al. An engineered factor VA prevents bleeding induced by anticoagulant WT activated protein C. *PLoS One* 2014;9(8):e104304.
- [132] Tiede A, Worster A. Lessons from a systematic literature review of the effectiveness of recombinant factor VIIA in acquired haemophilia. *Ann Hematol* 2018;97(10):1889–901.
- [133] Kessler CM, Benchikhi El Fegouen S, Worster A. Methodologies for data collection in congenital haemophilia with inhibitors (CHWI): critical assessment of the literature and lessons learned from recombinant factor VIIA. *Haemophilia* 2018;24(4):536–47.
- [134] Hunault M, Bauer KA. Recombinant factor VIIA for the treatment of congenital factor VII deficiency. *Semin Thromb Hemost* 2000;26(4):401–5.
- [135] Franchini M, Lippi G. Novoseven (recombinant factor viia) for the treatment of bleeding episodes and perioperative management in patients with Glanzmann's thrombasthenia. *Expert Rev Hematol* 2014;7(6):733–40.
- [136] Goodnough LT, Levy JH. The judicious use of recombinant factor VIIA. *Semin Thromb Hemost* 2016;42(2):125–32.
- [137] Budnik I, Shenkman B, Morozova O, Einav Y. In-vitro assessment of the effects of fibrinogen, recombinant factor VIIA and factor XIII on trauma-induced coagulopathy. *Blood Coagul Fibrinolysis* 2020;31(4):253–7.
- [138] Vincent JL, Artigas A, Petersen LC, Meyer C. A multicenter, randomized, double-blind, placebo-controlled, dose-escalation trial assessing safety and efficacy of active site inactivated recombinant factor VIIA in subjects with acute lung injury or acute respiratory distress syndrome. *Crit Care Med* 2009;37(6):1874–80.
- [139] Stein DM, Dutton RP, Kramer ME, Scalea TM. Reversal of coagulopathy in critically ill patients with traumatic brain injury: recombinant factor VIIA is more cost-effective than plasma. *J Trauma* 2009;66(1):63–72.
- [140] Kluger Y, Riou B, Rossaint R, et al. Safety of RVIIA in hemodynamically unstable polytrauma patients with traumatic brain injury: post hoc analysis of 30 patients from a prospective, randomized, placebo-controlled, double-blind clinical trial. *Crit Care (Lond, Engl)* 2007;11(4):R85.
- [141] Ling G, Kagdi H, Subel B, Chowdary P, Gomez K. Safety and efficacy of factor XI (FXI) concentrate use in patients with FXI deficiency: a single-centre experience of 19 years. *Haemophilia* 2016;22(3):411–18.
- [142] Hulsman N, van der Meulen J, Talacua H, Essoussi B, Hermanns H. Clot formation in cardiopulmonary bypass circuit after application of factor XI concentrate. *J Cardiothorac Vasc Anesth* 2020;34(8):2178–80.
- [143] Lassila R. Clinical use of factor xiii concentrates. *Semin Thromb Hemost* 2016;42(4):440–4.
- [144] Zhou Z, Nguyen TC, Guchhait P, Dong JF, Von willebrand factor, adamts-13, and thrombotic thrombocytopenic purpura. *Semin Thromb Hemost* 2010;36(1):71–81.
- [145] Crawley JT, Scully MA. Thrombotic thrombocytopenic purpura: basic pathophysiology and therapeutic strategies. *Hematol Am Soc Hematol Educ Prog* 2013;2013:292–9.
- [146] Kopic A, Benamara K, Piskernik C, et al. Preclinical assessment of a new recombinant adamts-13 drug product (bax930) for the treatment of thrombotic thrombocytopenic purpura. *J Thromb Haemost* 2016;14(7):1410–19.
- [147] Salas CM, Miyares MA. Antithrombin III utilization in a large teaching hospital. *P T* 2013;38(12):764–79.
- [148] Adiguzel C, Iqbal O, Demir M, Fareed J. European community and US-FDA approval of recombinant human antithrombin produced in genetically altered goats. *Clin Appl Thromb/Hemostas* 2009;15(6):645–51.
- [149] Menache D, O'Malley JP, Schorr JB, et al. Evaluation of the safety, recovery, half-life, and clinical efficacy of antithrombin III (human) in patients with hereditary antithrombin III deficiency. Cooperative study group. *Blood* 1990;75(1):33–9.
- [150] Schwartz RS, Bauer KA, Rosenberg RD, et al. Clinical experience with antithrombin III concentrate in treatment of congenital and acquired deficiency of antithrombin. The antithrombin III study group. *Am J Med* 1989;87(3b):53s–60s.
- [151] Thrombate III, prescribing information. Triangle park, NC: Grifols/Talecris; 2009.
- [152] Atryn, prescribing information. Deerfield, IL: Ovation Pharmaceuticals, Inc.; 2009.
- [153] Senzolo M, Germani G, Cholongitas E, Burra P, Burroughs AK. Veno occlusive disease: update on clinical management. *World J Gastroenterol* 2007;13(29):3918–24.
- [154] Pegram AA, Kennedy LD. Prevention and treatment of veno-occlusive disease. *Ann Pharmacother* 2001;35(7–8):935–42.
- [155] Niebler RA, Christensen M, Berens R, et al. Antithrombin replacement during extracorporeal membrane oxygenation. *Artif Organs* 2011;35(11):1024–8.
- [156] Agati S, Ciccarello G, Salvo D, et al. Use of a novel anticoagulation strategy during ECMO in a pediatric population: single-center experience. *ASAIO J* 2006;52(5):513–16.
- [157] Mohri M, Sugimoto E, Sata M, Asano T. The inhibitory effect of recombinant human soluble thrombomodulin on initiation and extension of coagulation—a comparison with other anticoagulants. *Thromb Haemostas* 1999;82(6):1687–93.
- [158] Yoshimura J, Yamakawa K, Ogura H, et al. Benefit profile of recombinant human soluble thrombomodulin in sepsis-induced disseminated intravascular coagulation: a multicenter propensity score analysis. *Crit Care (Lond, Engl)* 2015;19:78.
- [159] Yamakawa K, Aihara M, Ogura H, et al. Recombinant human soluble thrombomodulin in severe sepsis: a systematic review and meta-analysis. *J Thromb Haemost* 2015;13(4):508–19.
- [160] Hayakawa M, Yamakawa K, Saito S, et al. Recombinant human soluble thrombomodulin and mortality in sepsis-induced disseminated intravascular coagulation. A multicentre retrospective study. *Thromb Haemostas* 2016;115(6):1157–66.
- [161] Tagami T, Matsui H, Horiguchi H, Fushimi K, Yasunaga H. Recombinant human soluble thrombomodulin and mortality in severe pneumonia patients with sepsis-associated disseminated intravascular coagulation: an observational nationwide study. *J Thromb Haemost* 2015;13(1):31–40.
- [162] Vincent JL, Francois B, Zabolotskikh I, et al. Effect of a recombinant human soluble thrombomodulin on mortality in patients with sepsis-associated coagulopathy: the scarlet randomized clinical trial. *JAMA* 2019;321(20):1993–2002.
- [163] Uba T, Nishi K, Umegaki T, et al. The influence of human soluble recombinant thrombomodulin on in-hospital mortality in patients with acute respiratory distress syndrome and disseminated intravascular coagulation: a retrospective multicenter study. *J Intensive Crit Care* 2017;3(4):40.
- [164] Kroiss S, Albisetti M. Use of human protein C concentrates in the treatment of patients with severe congenital protein C deficiency. *Biologics* 2010;4:51–60.
- [165] Alaniz C. An update on activated protein C (Xigris) in the management of sepsis. *P t* 2010;35(9):504–29.
- [166] Nadel S, Goldstein B, Williams MD, et al. Drotrecogin alfa (activated) in children with severe sepsis: a multicentre phase III randomised controlled trial. *Lancet* 2007;369(9564):836–43.
- [167] Abraham E, Laterre PF, Garg R, et al. Drotrecogin alfa (activated) for adults with severe sepsis and a low risk of death. *New Engl J Med* 2005;353(13):1332–41.
- [168] Colombo S, Crivellari M, Mucci M, Plumari V, Silvani P, Della Valle P. Human protein C concentrate in a patient with meningitis and bleeding as a complication of treatment with recombinant activated protein C. *Signa Vitae* 2013;8(1):62–4.
- [169] Fearon A, Pearcey P, Venkataraman S, Shah P. Protein S deficiency and arterial thromboembolism: a case report and review of the literature. *J Hematol* 2019;8(1):37–9.
- [170] ten Kate MK, van der Meer J. Protein S deficiency: a clinical perspective. *Haemophilia* 2008;14(6):1222–8.
- [171] Fraga R, Diniz LM, Lucas EA, Emerich PS. Warfarin-induced skin necrosis in a patient with protein s deficiency. *An Bras Dermatol* 2018;93(4):612–13.
- [172] Ohashi I, Wada S, Yoshino F, et al. Ischemic stroke with protein s deficiency treated by apixaban. *J Stroke Cerebrovasc Dis* 2020;29(4):104608.
- [173] Ameku K, Higa M. Rivaroxaban treatment for warfarin-refractory thrombosis in a patient with hereditary protein s deficiency. *Case Rep Hematol* 2018;2018:5217301.
- [174] Yagi S, Kagawa K, Fujimoto E, Sata M. Recurrent venous thrombosis during

- direct oral anticoagulant therapy in a patient with protein S deficiency. *J Med Invest* 2019;66(1.2):182–4.
- [175] Zanichelli A, Wu MA, Andreoli A, Mansi M, Cicardi M. The safety of treatments for angioedema with hereditary c1 inhibitor deficiency. *Expert Opin Drug Saf* 2015;14(11):1725–36.
- [176] Longhurst H, Zinser E. Prophylactic therapy for hereditary angioedema. *Immunol Allergy Clin North Am* 2017;37(3):557–70.
- [177] Tang R, Chen S, Zhang HY. Fresh frozen plasma for the treatment of hereditary angioedema acute attacks. *Chin Med Sci J= Chung-kuo i Hsueh k'o Hsueh tsa Chih* 2012;27(2):92–5.
- [178] Prematta M, Gibbs JG, Pratt EL, Stoughton TR, Craig TJ. Fresh frozen plasma for the treatment of hereditary angioedema. *Ann Allergy Asthma Immunol* 2007;98(4):383–8.
- [179] Lumry W, Soteres D, Gower R, et al. Safety and efficacy of c1 esterase inhibitor for acute attacks in children with hereditary angioedema. *Pediatric Allergy Immunol* 2015;26(7):674–80.
- [180] Sabharwal G, Craig T. Recombinant human c1 esterase inhibitor for the treatment of hereditary angioedema due to c1 inhibitor deficiency (c1-Inh-Hae). *Expert Rev Clin Immunol* 2015;11(3):319–27.
- [181] Henry Li H, Riedl M, Kashkin J. Update on the use of c1-esterase inhibitor replacement therapy in the acute and prophylactic treatment of hereditary angioedema. *Clin Rev Allergy Immunol* 2019;56(2):207–18.
- [182] Bova M, Valerieva A, Wu MA, Senter R, Perego F. Lanadelumab injection treatment for the prevention of hereditary angioedema (HAE): design, development and place in therapy. *Drug Des Dev Ther* 2019;13:3635–46.
- [183] Carpenter SL, Matheu P. A2-antiplasmin and its deficiency: fibrinolysis out of balance. *Haemophilia* 2008;14(6):1250–4.
- [184] Lee KN, Jackson KW, Christiansen VJ, Chung KH, McKee PA. Alpha2-antiplasmin: potential therapeutic roles in fibrin survival and removal. *Curr Med Chem Cardiovasc Hematol Agents* 2004;2(4):303–10.