

# *In silico* assessment of the impact of 2019 novel coronavirus genomic variation on the efficiency of published real-time quantitative polymerase chain reaction detection assays

Hang Fan, Xiang-Li-Lan Zhang, Ya-Wei Zhang, Yong Huang, Yue Teng, Yan Guo, Zhi-Qiang Mi, Rui-Fu Yang, Ya-Jun Song, Yu-Jun Cui

State Key Laboratory of Pathogen and Biosecurity, Beijing Institute of Microbiology and Epidemiology, Beijing 100071, China.

*To the Editor:* In December 2019, coronavirus disease 2019 outbreak caused by the 2019 novel coronavirus (2019-nCoV) happened in Wuhan, China. Now, it has posed a worldwide public health threat. Real-time quantitative polymerase chain reaction (RT-qPCR) was recommended as an effective pathogen detection method and has played an important role in prevention and control of the current outbreak. Many research institutions have released their primer sets for RT-qPCR. If the variant sites were located in the primer regions, the efficiency of RT-qPCR would be reduced, thus possibly causing false negative results, and leading to unpredictable impact on the diagnosis of patients and the control of this outbreak. Therefore, a comprehensive investigation on 2019-nCoV genome variation is necessary to evaluate the effectiveness of current released RT-qPCR methods.

Here, we analyzed 77 public full-length genome sequences of 2019-nCoV from the GISAID website [Supplementary Table 1, <http://links.lww.com/CM9/A222>]. All the sequences were aligned by using MAFFT v7.450.<sup>[1]</sup> A total of 85 variant sites were found, all of which are single nucleotide variants. Among the 85 variant sites, seven were shared by two 2019-nCoV sequences and nine were found in three or more sequences [Supplementary Table 2, <http://links.lww.com/CM9/A223>].

We investigated the published 2019-nCoV RT-qPCR detection assays and found a total number of 13 RT-qPCR primer sets designed by eight institutions [Table 1, Supplementary Table 3, <http://links.lww.com/CM9/A224>]. These primers were designed to amplify genes of ORF1ab, Spike (S), Envelope (E), and Nucleocapsid (N). The reverse primers of primer sets 3 and 6 had one mismatch against all of the

released 2019-nCoV sequences. Among all the observed variation sites, three (positions on the reference genome IVDC-HB-01: 28291, 28688, and 29200) are separately located on the region of forward primers of the primer sets 7 and 9, and the probe of the primer set 8. It is noted that the above three variations was found in the sequences BetaCoV/Shenzhen/SZTH-003/2020, BetaCoV/Shandong/IVDC-SD-001/2020|EPI\_ISL\_408482, and BetaCoV/Chongqing/YC01/2020|EPI\_ISL\_408478, respectively. These variants may affect the RT-qPCR detection efficiency. In particular, variations on the probe region of the primer set 8 may have largely negative effects on detection efficiency according to the previous research.<sup>[2]</sup>

In conclusion, using any of the five RT-qPCR primer sets mentioned above to detect 2019-nCoV may potentially cause false-negative results. Among the five, two have mismatches and three contain some 2019-nCoV genome variants which occurred during the outbreak. It is worth noting that the three primer sets containing variants are all located on the N gene. Therefore, it is suggested that conservative regions, such as nsp12 (*RdRp*) gene, would be preferable primer targets. Although the multiple-targets designation of RT-qPCR protocol would reduce the false-negative results caused by genome variation, more careful performance evaluation of the currently used primers is needed. Moreover, it is necessary to keep continuous surveillance on the genome variants and their effects on the RT-qPCR assays during the whole outbreak.

## Acknowledgements

We gratefully acknowledge the Authors, the Originating and Submitting Laboratories for their sequences and metadata shared through GISAID, on which this research

### Access this article online

Quick Response Code:



Website:  
[www.cmj.org](http://www.cmj.org)

DOI:  
10.1097/CM9.0000000000000817

**Correspondence to:** Dr. Yu-Jun Cui, State Key Laboratory of Pathogen and Biosecurity, Beijing Institute of Microbiology and Epidemiology, Beijing 100071, China  
E-Mail: [cuiyujun.new@gmail.com](mailto:cuiyujun.new@gmail.com)

Copyright © 2020 The Chinese Medical Association, produced by Wolters Kluwer, Inc. under the CC-BY-NC-ND license. This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Chinese Medical Journal 2020;133(13)

Received: 26-02-2020 Edited by: Pei-Fang Wei

**Table 1: The detailed information of publicly released 2019-nCoV RT-qPCR primers.**

Primer No.	Target gene	Primer type	Primer name	Primer sequence (5'-3')	Pos. on Ref.	Variation site
1	ORF1ab	Forward	HKU-ORF1b-nsp14F	TGGGGYTTTACRGGTAAACCT	18778..18797	
		Reverse	HKU- ORF1b-nsp14R	AACRCGCTTAACAAAGCACTC	18889..18909	
		Probe	HKU-ORF1b-nsp141P	FAM-TAGTTGTGATGCWATCATGACTAG-TAMRA	18849..18872	
2	N	Forward	HKU-NF	TAATCAGACAAGGAACTGATTA	29145..29166	
		Reverse	HKU-NR	CGAAGGTGTGACTTCCATG	29236..29254	
		Probe	HKU-NP	FAM-GCAAATTGTGCAATTTGCCGG-TAMRA	29177..29196	
3	ORF1ab	Forward	RdRP_SARsR-F2	GTGARATGGTCATGTGTGGCGG	15431..15452	
		Reverse	RdRP_SARsR-R1	CARATGTTAAAS <sup>u</sup> ACACTATTAGCATA	15505..15530	15519
		Probe	RdRP_SARsR-P2	FAM-CAGGTGGAACCTCATCAGGAGATGC-BBQ	15470..15494	
4	E	Forward	E_Sarbeco_F1	ACAGGTACGTTAATAGTTAATAGCGT	26269..26294	
		Reverse	E_Sarbeco_R2	ATATTGCAGCAGTACGGCACACA	26360..26381	
		Probe	E_Sarbeco_P1	FAM-ACACTAGCCATCCTTACTGCGCTTCG-BBQ	26332..26357	
5	N	Forward	NbatCoV_F1	TTTGGTGGACCCCTCAGATTC	28322..28341	
6	N	Reverse	NbatCoV_R1	GGGTGCCAATGTGATCTTTT	28699..28718	
		Forward	NIID_2019-nCoV_N_F2	AAATTTGGGGACCAGGAAC	29125..29144	
		Reverse	NIID_2019-nCoV_N_R2	TGGCA <sup>u</sup> CTGTGTAGGTCAAC	29263..29282	29277
7	N	Probe	NIID_2019-nCoV_N_P2	FAM-ATGTCGCGCATTGGCATGGA-BHQ	29222..29241	
		Forward	2019-nCoV_N1-F	GACCCCAAATCAGCGAAAT	28287..28306	28291
		Reverse	2019-nCoV_N1-R	TCTGGTACTGCCAGTTGAATCTG	28335..28358	
8	N	Probe	2019-nCoV_N1-P	FAM-ACCCCGCATTACGTTTGGTGGACC-BHQ1	28309..28332	
		Forward	2019-nCoV_N2-F	TTACAAACATTGGCCGCAAA	29164..29183	
		Reverse	2019-nCoV_N2-R	GCGCGACATTCGGAAGAA	29213..29230	
9	N	Probe	2019-nCoV_N2-P	FAM-ACAATTTGCCCCAGCGCTTCAG-BHQ1	29188..29210	29200
		Forward	2019-nCoV_N3-F	GGGAGCC <sup>u</sup> TGAATACACCAAAA	28681..28702	28688
		Reverse	2019-nCoV_N3-R	TGTAGCACGATTGCAGCATTG	28732..28752	
10	ORF1ab	Probe	2019-nCoV_N3-P	FAM-AYCACATTGGCACCCGCAATCCTG-BHQ1	28704..28727	
		Forward	F	CCCTGTGGGTTTTACTTAA	13342..13362	
		Reverse	R	ACGATTGTGCATCAGCTGA	13442..13460	
11	N	Probe	P	FAM-CCGTCTGCGGTATGTGGAAAGGTTATGG-BHQ1	13377..13404	
		Forward	F	GGGGAAC <sup>u</sup> TCTCCTGCTAGAAT	28881..28902	
		Reverse	R	CAGACATTTTGCTCTCAAGCTG	28958..28979	
12	E	Probe	P	FAM-TTGCTGCTGCTTGACAGATT-TAMRA	28934..28953	
		Forward	F	ACTTCTTTTTCTTGCTTTCGTGGT	26295..26318	
		Reverse	R	GCAGCAGTACGCACACAATC	26357..26376	
13	N	Probe	P	CY5-CTAGTACTAGCCATCCTTACTGC-BHQ1	26326..26351	
		Forward	WH-NIC N-F	CGTTTGGTGGACCCCTCAGAT	28320..28339	
		Reverse	WH-NIC N-R	CCCCACTGCGTTCTCCATT	28358..28376	
		Probe	WH-NIC N-P	FAM-CAACTGGCAGTAACCA- BQH1	28341..28356	

The reference genome is IVDC-HB-01. The underlined letters in the column of primer sequence represent the variant sites. 2019-nCoV: 2019 Novel coronavirus; RT-qPCR: Real-time quantitative polymerase chain reaction; Pos: Position; Ref: Reference genome.

is based. All submitters of data may be contacted directly via <https://www.gisaid.org>.

### Funding

This work was supported by the National Key Research & Development Program of China (No. 2020YFC0840900) and the Beijing Municipal Science and Technology Project (No. Z201100001020004).

### Conflicts of interest

None.

### References

- Katoh K, Standley DM. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol* 2013;30:772–780. doi: 10.1093/molbev/mst010.
- Süss B, Flekna G, Wagner M, Hein I. Studying the effect of single mismatches in primer and probe binding regions on amplification curves and quantification in real-time PCR. *J Microbiol Methods* 2009;76:316–319. doi: 10.1016/j.mimet.2008.12.003.

**How to cite this article:** Fan H, Zhang XLL, Zhang YW, Huang Y, Teng Y, Guo Y, Mi ZQ, Yang RF, Song YJ, Cui YJ. *In silico* assessment of the impact of 2019 novel coronavirus genomic variation on the efficiency of published real-time quantitative polymerase chain reaction detection assays. *Chin Med J* 2020;133:1612–1613. doi: 10.1097/CMJ.0000000000000817