



REVIEW

# Optimizing G6PD testing for *Plasmodium vivax* case management: why sex, counseling, and community engagement matter [version 1; peer review: 2 approved]

Cindy S Chu <sup>1,2</sup>, Germana Bancone <sup>1,2</sup>, Maureen Kelley<sup>3</sup>, Nicole Advani<sup>4</sup>, Gonzalo J Domingo <sup>4</sup>, Eva M Cutiongo-de la Paz<sup>5,6</sup>, Nicole van der Merwe<sup>7</sup>, Jessica Cohen<sup>4</sup>, Emily Gerth-Guyette <sup>4</sup>

<sup>1</sup>Shoklo Malaria Research Unit, Mahidol Oxford Tropical Medicine Research Unit, Faculty of Tropical Medicine, Mahidol University, Mae Sot, Thailand

<sup>2</sup>Centre for Tropical Medicine and Global Health, Nuffield Department of Medicine, University of Oxford, Oxford, UK

<sup>3</sup>The Ethox Centre and Wellcome Centre for Ethics and Humanities, Nuffield Department of Population Health, University of Oxford, Oxford, UK

<sup>4</sup>PATH, Seattle, Washington, USA

<sup>5</sup>Institute of Human Genetics, National Institutes of Health, University of the Philippines Manila, Manila, Philippines

<sup>6</sup>Philippine Genome Center, University of the Philippines System, Quezon City, Philippines

<sup>7</sup>Division of Chemical Pathology, Department of Pathology, Faculty of Medicine and Health Sciences, Stellenbosch University, Tygerberg Academic Hospital, Cape Town, South Africa

**v1** First published: 07 Feb 2020, 5:21  
<https://doi.org/10.12688/wellcomeopenres.15700.1>  
 Latest published: 07 Feb 2020, 5:21  
<https://doi.org/10.12688/wellcomeopenres.15700.1>

## Abstract

Safe access to the most effective treatment options for *Plasmodium vivax* malaria are limited by the absence of accurate point-of-care testing to detect glucose-6-phosphate dehydrogenase (G6PD) deficiency, the most common human genetic disorder. G6PD-deficient patients are at risk of life-threatening hemolysis when exposed to 8-aminoquinolines, the only class of drugs efficacious against *P. vivax* hypnozoites. Until recently, only qualitative tests were available in most settings. These accurately identify patients with severe G6PD deficiency (mostly male) but not patients with intermediate G6PD deficiency (always female). This has led to and reinforced a gap in awareness in clinical practice of the risks and implications of G6PD deficiency in females—who, unlike males, can have a heterozygous genotype for G6PD. Increasing recognition of the need for radical cure of *P. vivax*, first for patients' health and then for malaria elimination, is driving the development of new point-of-care tests for G6PD deficiency and their accessibility to populations in low-resource settings. The availability of simple, affordable, and accurate point-of-care diagnostics for the precise classification of the three G6PD phenotypes can reduce sex-linked disparities by ensuring safe and effective malaria treatment, providing opportunities to develop supportive counseling to enhance understanding of genetic test results, and improving the detection of all G6PD deficiency phenotypes in newborns and their family members.

## Open Peer Review

Reviewer Status

	Invited Reviewers	
	1	2
<b>version 1</b> 07 Feb 2020	 report	 report

- Chansuda Wongsrichanalai**, Independent Consultant, Bangkok, Thailand
- Manoj Menon**, Fred Hutchinson Cancer Research Center, Seattle, USA

Any reports and responses or comments on the article can be found at the end of the article.

## Keywords

G6PD deficiency, Plasmodium vivax, neonatal hyperbilirubinaemia, gender, sex, disparity, G6PD testing, primaquine, tafenoquine, genetic counselling, haemolysis, G6PD heterozygous females



This article is included in the [Mahidol Oxford Tropical Medicine Research Unit \(MORU\)](#) gateway.

**Corresponding author:** Cindy S Chu ([cindy@tropmedres.ac](mailto:cindy@tropmedres.ac))

**Author roles:** **Chu CS:** Formal Analysis, Investigation, Methodology, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Bancone G:** Formal Analysis, Investigation, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Kelley M:** Formal Analysis, Investigation, Validation, Writing – Original Draft Preparation, Writing – Review & Editing; **Advani N:** Investigation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Domingo GJ:** Conceptualization, Formal Analysis, Funding Acquisition, Investigation, Methodology, Project Administration, Supervision, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Cutiongo-de la Paz EM:** Investigation, Writing – Review & Editing; **van der Merwe N:** Investigation, Writing – Review & Editing; **Cohen J:** Investigation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Gerth-Guyette E:** Formal Analysis, Investigation, Methodology, Project Administration, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

**Competing interests:** PATH supports a portfolio of G6PD diagnostic test development efforts. PATH has no financial interests in the commercialisation of any resulting products.

**Grant information:** This work was supported by the Wellcome Trust through a Wellcome Trust Strategic Award [096527] and a Research Enrichment - Public Engagement Award to MK [200344]. CSC and GB are part of the Wellcome Trust Mahidol University Oxford Tropical Medicine Research Unit, funded by the Wellcome Trust (United Kingdom) [A150007]. This work was funded by the United Kingdom's Department for International Development (DFID) [204139], and the Bill & Melinda Gates Foundation [OPP1107113]. The findings and conclusions contained within are those of the authors and do not necessarily reflect the positions of DFID or the Bill & Melinda Gates Foundation.

*The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.*

**Copyright:** © 2020 Chu CS *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**How to cite this article:** Chu CS, Bancone G, Kelley M *et al.* **Optimizing G6PD testing for Plasmodium vivax case management: why sex, counseling, and community engagement matter [version 1; peer review: 2 approved]** Wellcome Open Research 2020, 5:21 <https://doi.org/10.12688/wellcomeopenres.15700.1>

**First published:** 07 Feb 2020, 5:21 <https://doi.org/10.12688/wellcomeopenres.15700.1>

**Box 1. Learning points**

1. Deficiency of the essential enzyme glucose-6-phosphate dehydrogenase (G6PD) in humans is caused by mutations in the *G6PD* gene located on the X-chromosome (Xq28). As such, males are either G6PD deficient or normal while females can be deficient, intermediate, or normal. The standard tests typically used to diagnose G6PD deficiency can identify deficient subjects but cannot reliably differentiate intermediate G6PD activity. Consequently, accurate assessment of G6PD status is more difficult in females, especially at the point of care where it is needed to inform *Plasmodium vivax* malaria case management.
2. Evidence for haemolysis associated with anti-malarials in G6PD-intermediate females has been documented, albeit infrequently, since 1958. Differentiating females with intermediate activity from those with normal activity has not been prioritised due to technical complexity of the testing. As a result, front-line health care providers are often unaware of the sex-related, oxidative drug-associated risk for female patients.
3. New point-of-care G6PD tests that provide accurate results for both males and females present important opportunities to address the sex-linked disparities related to safe and efficacious malaria treatment in women and girls. These tests can also be used to address other G6PD deficiency-associated medical indications, including improved management of neonatal hyperbilirubinemia in low-resource settings.
4. Realisation of these opportunities requires community engagement and improved counseling to enhance understanding of genetic test results and the implications for offspring and other family members.
5. Using G6PD testing beyond malaria treatment can further enhance the cost-effectiveness of the test, an important consideration for low-resource and malaria-endemic settings.

**Introduction**

A new, single-dose radical cure for *Plasmodium vivax*, tafenoquine has recently received registration in Australia (Kozenis®) and the United States (Krintafel®). This combined with the drive to eliminate malaria, is encouraging malaria-endemic countries to increase access to testing for glucose-6-phosphate dehydrogenase (G6PD) deficiency, the most common sex-linked genetic abnormality in humans, affecting more than 400 million people worldwide<sup>1,2</sup>, for the safe deployment of radical cure. The World Health Organization (WHO) estimates that there were 7.5 million cases of *P. vivax* in 2017 alone<sup>3</sup>, a large proportion of which occurred in populations with high G6PD deficiency prevalence<sup>4</sup>. For most countries approaching malaria elimination, *P. vivax* is now the main contributor to malaria disease burden, and there is recognition of the need for broader access to radical cure with either the current, standard, 7- to 14-day primaquine course or the new, single-dose tafenoquine, both of which are contraindicated in people with G6PD deficiency<sup>5-7</sup>. For G6PD-deficient patients, WHO recommends an 8-week course of primaquine (0.75mg/kg weekly), which is qualified in the WHO malaria treatment guideline as a conditional recommendation with very low quality evidence. Where G6PD testing is not available, WHO recommends “all females should be considered as potentially having intermediate G6PD

activity and given the 14-day regimen of primaquine, with counselling on how to recognise symptoms and signs of haemolytic anaemia”<sup>8</sup>. Tafenoquine is the first drug to be contraindicated additionally in females with intermediate G6PD activity<sup>9-13</sup>.

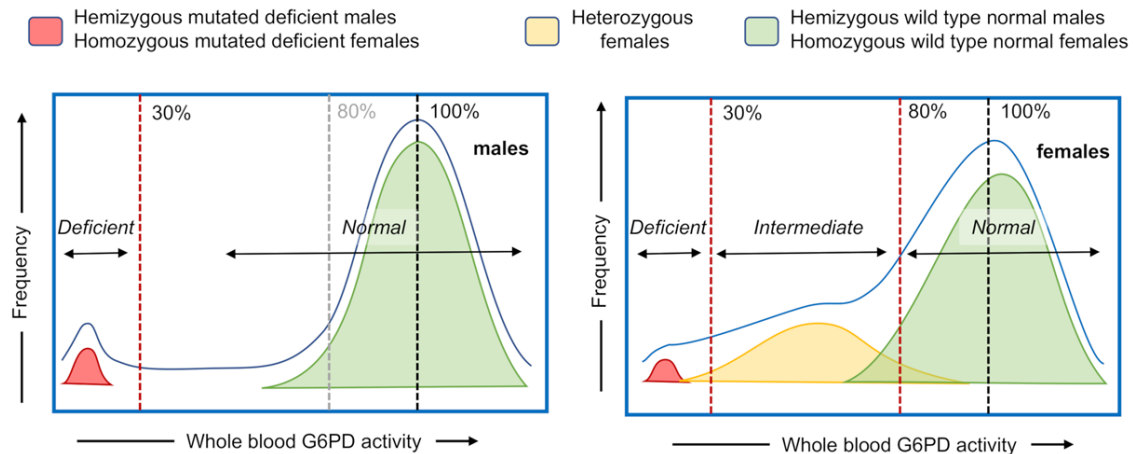
The *G6PD* gene is located on the X chromosome, so males have only one gene that expresses the G6PD enzyme and are either deficient in G6PD enzyme activity or normal, whereas females have two genes (but only one is expressed in each cell), and can have deficient, intermediate, or normal levels of G6PD activity (Figure 1)<sup>2,14</sup>.

Historically, it has been challenging to accurately identify and diagnose females with intermediate G6PD deficiency because existing quantitative diagnostic tests are complex and require good laboratory infrastructure<sup>15-17</sup>. While qualitative rapid tests and other near-patient methods, such as the fluorescent spot test, may be sufficient to differentiate gross deficiencies, they do not reliably assess moderate status. Instead, the tests misclassify females with intermediate enzyme activity as normal. As a result, males and females who are severely G6PD deficient can be identified correctly and treated with weekly primaquine (or no treatment in the case of other oxidative drugs), whereas females with an intermediate G6PD status remain under-diagnosed and thus inadvertently exposed to oxidative treatments.

As countries prepare to increase access to *P. vivax* radical cure with the old and new curative regimens of primaquine<sup>18,19</sup> and tafenoquine (Kozenis® and Krintafel®)<sup>11,12</sup>, respectively, there are several opportunities that should be considered within broader national health systems for expanding the acceptability, utility, and, concurrently, the cost-effectiveness of G6PD testing beyond malaria case management<sup>15</sup>. In this article, we discuss an important opportunity to reduce sex-related health disparities in an era of new tools and initiatives that improve the diagnosis of G6PD deficiency. This is in alignment with the need to recognise sex as a key determinant of health and its importance in health research to understand the impact on health outcomes<sup>20</sup>. We also propose a research agenda to investigate this opportunity (Table 1).

**G6PD deficiency in females**

In a given population with a mutated *G6PD* allele frequency of 10%, this same number will indicate the proportion of affected males. The proportion of females with the homozygous mutated genotype will be around 1%, while a large proportion of females (20%) will be heterozygous with a variable phenotype due to X-chromosome inactivation<sup>15,18,21</sup>. Of heterozygous females, around 60% have an intermediate phenotype<sup>22</sup>. Thus while males represent the highest proportion of individuals with severe G6PD deficiency, there is also a comparable proportion of females with intermediate G6PD deficiency. While this is based on well-established knowledge, the characterisation of G6PD phenotypes in heterozygous women has received little attention at the patient level, possibly because of the more labor-intensive laboratory techniques needed to characterise it



**Figure 1. Schematic of population histograms demonstrating the relationship between phenotype and genotype in G6PD deficiency in males (left panel) and females (right panel).** The *G6PD* gene is located on the X chromosome, such that females have two genes and males have only one. Males with a mutated *G6PD* allele (in red,  $G6PD_{DEF}$ ) that expresses a compromised (deficient) G6PD enzyme protein typically have a blood G6PD value of less than 30% of normal. Females with two mutated G6PD-deficient alleles (in red,  $G6PD_{DEF1, DEF2}$ ) also typically have a blood G6PD value of less than 30% of normal. Males with a wild type *G6PD* allele (in green,  $G6PD_{WT}$ ) that expresses a fully functional enzyme have G6PD activity in an approximate normal distribution around the 100% median, as do females with two wild type *G6PD* alleles (in green,  $G6PD_{WT1, WT2}$ ). Heterozygous females with both wild type and mutated *G6PD* alleles (in yellow,  $G6PD_{WT, DEF1}$ ) can express a spectrum of whole blood G6PD activity, ranging from severely deficient (<30%) to beyond the World Health Organization definition of normal for females (>80%), with the majority in the intermediate (30% to 80%) activity range. The colored zones indicate the distribution of enzymatic activities associated with the genotypes as described above; the blue line represents the cumulative G6PD activity-based histogram.

**Table 1. Research questions to investigate the implications of a new drive to increase access to safe radical cure of *Plasmodium vivax* malaria, including diagnosis of intermediate G6PD deficiency in females and availability of new point-of-care tests for G6PD deficiency.**

Research topics/agenda	Considerations
Studies to assess the feasibility of introducing point-of-care quantitative tests into health clinics in low-resource settings, including ethnographic/qualitative studies on a barriers/facilitators model.	A quantitative G6PD test may need to be integrated into dynamic and contextually specific malaria case management strategies, newborn screening policies, and other complex health services. Feasibility studies will help to ensure that this integration can be successfully scaled up across areas where G6PD deficiency is prevalent.
Ethnographic research to inform appropriate messaging and frontline staff training with respect to G6PD deficiency genetic counseling content and tools for target populations in low-resource, high disease burden settings.	Current practices vary widely and scant evidence exists regarding G6PD deficiency counseling best-practices. Ample lessons from other genetic conditions can be leveraged to design and validate such tools.
Studies comparing the costs of implementing current G6PD tests, including costs of inaccurate diagnosis of intermediate G6PD, and new point-of-care tests.	As new point-of-care tests become available, stakeholders within the health system will need clear guidance on the costs of various products. These costing studies will need to consider factors such as training, distribution, and product specifications such as shelf life.
Cost-effectiveness studies that take into consideration broader clinical benefits to the individual than those of <i>Plasmodium vivax</i> cure, in different G6PD prevalence settings.	Improve the value proposition of G6PD testing by considering other clinical benefits (not only as part of malaria case management) such as averting exposure to other oxidative agents, improved management of hyperbilirubinaemia in offspring and averting kernicterus.
Clinical studies to better define risk of clinical haemolysis in intermediate females given different anti-relapse regimens.	Clinical data focused on intermediate females will help inform downstream decisions regarding appropriate anti-relapse regimens. Currently, these decisions are often made based on a clinician's individual risk-benefit assessments. The need for this type of medical expertise can restrict 8-aminoquinoline usage to the highest tiers of the health system.

(Table 2). In clinical settings worldwide, the most commonly performed tests are qualitative tests (e.g., fluorescent spot test), which result in a systematic underestimation of the number of

females at risk (as intermediates are often classified as normal) and a perception among health workers that G6PD deficiency only concerns males<sup>23</sup>.

**Table 2. Scientific findings and technical advancement for the characterisation of women with intermediate G6PD activity.**

Reference	Year published	Main findings
Beutler <i>et al.</i>	1955	Development of the GSH test for sensitivity to primaquine
Beutler <i>et al.</i>	1955	Development of the Heinz Bodies test for sensitivity to primaquine
Alving, <i>et al.</i>	1958	Primaquine associated haemoglobin reduction observed in individuals with intermediate activity
Childs, <i>et al.</i>	1958	
Gross, <i>et al.</i>	1958	
Tarlov, <i>et al.</i>	1962	
Brewer, <i>et al.</i>	1960*	Development of the methaemoglobin reduction test (MRT) for sensitivity to primaquine
Stamatoyannopoulos, <i>et al.</i>	1967	Description of the enzymatic phenotypes in small samples of known G6PD heterozygous females
Panizon, <i>et al.</i>	1970	
Rinaldi, <i>et al.</i>	1976	
Beutler <i>et al.</i>	1977	Gold standard spectrophotometric assay
Van Noorden, <i>et al.</i>	1985	Description of new or improved cytochemical techniques for detection of G6PD in erythrocytes
Vives-Corrons, <i>et al.</i>	1986	
Vogels, <i>et al.</i>	1986	
Fanello, <i>et al.</i>	2008	Dapsone associated haemolysis in G6PD heterozygous females (no phenotype)
Premji, <i>et al.</i>	2009	
Tiono, <i>et al.</i>	2009	
Pamba, <i>et al.</i>	2012	
Shah, <i>et al.</i>	2012	Development of the cytofluorometric reading of MRT
Chu, <i>et al.</i>	2017	Primaquine and tafenoquine associated haemolysis in G6PD heterozygous females with intermediate activity
Rueangweerayut, <i>et al.</i>	2017	

\* For ease of visualization, this article is listed out of chronological order

Other than the malaria radical curative indication, quantitative screening for G6PD deficiency has additional clinical indications:

- (i) *Newborn screening to identify risk for pathologic hyperbilirubinaemia.* Evidence indicates that qualitative G6PD tests do not detect all clinically relevant cases of G6PD deficiency, particularly in female neonates<sup>24–28</sup>. This results in comparatively less emphasis on the clinical management of female neonates who have a risk of developing pathologic levels of serum bilirubin.
- (ii) *The use of dapsone through the antimalarial drug chloro-guanil-dapsone-artesunate (not used in malaria treatment anymore but is used for other medical indications).* Individual study analysis of phase 3 clinical trials with chloro-guanil-dapsone-artesunate did not identify risk of severe haemolysis in G6PD heterozygous females<sup>29,30</sup>. However, a complete analysis of the data, including 200 heterozygous females, demonstrated that heterozygous females showed a wide range of reactions, from large to imperceptible haemoglobin drops<sup>31</sup>. One published case

report describes a Greek female who tested G6PD normal by a screening test then experienced severe haemolysis to dapsone; quantitative testing was recommended<sup>32</sup>.

- (iii) *Guidelines for rasburicase therapy in the context of genotyping.* The Clinical Pharmacogenetics Implementation Consortium published guidelines that recognise the limitations of *G6PD* genotype results to inform the use of rasburicase therapy in females for management of tumor lysis syndrome as *G6PD* genotyping does not correlate with the three phenotypes in heterozygous females<sup>33,34</sup>.

With the introduction of new quantitative point-of-care G6PD diagnostics, the three G6PD phenotypes can be detected accurately and characterised precisely for the first time without the need for expensive, complicated assays that require good laboratory infrastructure. Immediate results can be obtained at the patient level in the clinic. Where testing is driven by malaria case management, often rural and lower socioeconomic populations will benefit, as these populations often experience

higher rates of malaria infection. Recent renewed attention to characterisation at a cellular level of heterozygous G6PD deficiency may help to raise awareness of the clinical implications of intermediate G6PD activity in females<sup>35–37</sup>.

#### *Plasmodium vivax* case management

A significant barrier to safe radical cure of *P. vivax* malaria with 8-aminoquinolines is the limited availability of G6PD tests. Until recently, primaquine has been administered either with no G6PD testing or with the use of qualitative tests<sup>38</sup>. Females with low-intermediate G6PD activity are at risk of potentially clinically significant haemolysis from high dose 14-day primaquine (0.5mg/kg daily) and tafenoquine (300mg single dose) (Kozenis® and Krintafel®)<sup>9,13,19,39,40</sup>. In a systematic review comparing treatment with chloroquine versus chloroquine and primaquine, G6PD “normal” (classified using qualitative tests) females taking primaquine had significantly greater haemoglobin reductions than males<sup>41</sup>. Recent studies exploring the efficacy of a high dose short course (7-day) primaquine regimen (1mg/kg) have shown consistently that females classified as normal by the fluorescent spot test are at risk for clinically significant primaquine-induced haemolysis<sup>19,40</sup>. One of the studies was carried out in an area with a high prevalence (15–18%) of G6PD deficiency; heterozygous females showed a significantly greater drop in haematocrit as compared to wild type homozygous females given the same treatments, and 2/16 females receiving dihydroartemisinin-piperazine with the 7-day primaquine regimen needed a blood transfusion<sup>18,40</sup>. In the second study, 2 females from the site in Hanura, Indonesia had clinically significant haemolysis (31–40% haemoglobin reduction from pre-treatment) where 229 participants were treated with the 7-day primaquine regimen<sup>19</sup>. Prevalence of G6PD deficiency in the area was unknown but previous reports in West Timor region gave a prevalence of 3.2% among males<sup>42</sup>. In contrast, in clinical studies using tafenoquine where females with intermediate G6PD activity were excluded, no haemolysis-related adverse events were observed<sup>11,12</sup>. The inability to diagnose intermediate G6PD activity can negatively impact the safe radical cure of *P. vivax* malaria with high dose 14 or 7-day primaquine regimens and tafenoquine 300mg single dose in girls and women. The risk of haemolysis in malaria-endemic locations is concerning because of low access to medical supervision and health facilities where haemolytic events can be detected with haemoglobin testing and managed with blood transfusion.

From a policy and clinical practice perspective if national malaria programs decide to support radical cure with primaquine in the absence of G6PD testing, important ethical considerations will be raised and difficult tradeoffs between ensuring patient safety and expanding access to critical treatments must be made. In places where no testing is done, more males are at risk of severe haemolysis simply because there are more G6PD deficient males than females. In places where only qualitative testing is done, malaria programs may not be comfortable treating women given the limitations of that platform, as explained above. As such, some policies may indicate that radical cure be given only to males after qualitative

G6PD testing at the point of care and all females referred to a health facility. In the absence of testing, practitioners may also assume all females (normal, intermediate and deficient) are G6PD deficient and they will be given the 8-week high dose primaquine regimen recommended for G6PD deficient individuals. These policies and practices generally are not standardized and can change quickly based on new information and varying perceptions of risk among key decision-makers. Decisions to include qualitative G6PD testing in clinical guidelines or use them as a matter of policy do provide broader access to G6PD testing and improve health disparities in malaria treatment. However, the benefit of this expanded access is mostly in males who have the most severe G6PD deficiency and are at highest risk of severe haemolysis. Alternatively, quantitative G6PD testing allows equal access to an accurate G6PD diagnosis, safe treatment and convenient health care delivery.

Anticipation of mandatory quantitative G6PD testing to support anti-relapse treatment of *P. vivax* with tafenoquine has spurred the development of new quantitative point-of-care G6PD diagnostic tests, which only recently have become commercially available. These tests demonstrate greater accuracy in identifying G6PD deficiency in females, including those with the intermediate phenotype<sup>43,44</sup>. A diagnosis of G6PD intermediate status presents an important opportunity to address disparities in appropriate treatment and care<sup>17</sup>, such as providers’ low understanding and recognition of haemolytic responses, patients’ low awareness of adverse symptoms and the need for prompt follow-up, and appropriate genetic counseling.

#### Increased newborn screening for G6PD deficiency

The diagnostic gap for G6PD testing extends to newborn screening globally. Newborn screening for G6PD deficiency has been recommended by the World Health Organization among populations where 3–5% of males are affected<sup>45</sup>. It has been recognised that even if the majority of G6PD deficient patients are asymptomatic as children and adults, they have an increased risk of kernicterus resulting from significant neonatal hyperbilirubinaemia<sup>46</sup>. Screening for G6PD deficiency is recommended in newborns with jaundice, especially when family history or background suggest the likelihood of G6PD deficiency, or when the response to phototherapy is poor<sup>45,47–50</sup>. Nonetheless, there is a high heterogeneity in practice among different countries and within countries between rural and urban areas, with urban and peri-urban areas having greater access to G6PD screening programs. The same heterogeneity applies to screening methods, with G6PD deficiency screening performed in some settings via high accuracy, gold-standard quantitative spectrophotometric methods, while in low-resource settings it is more commonly done using low-accuracy, qualitative diagnostic tests.

The diagnostic limitations of qualitative tests restrict the potential for downstream public health interventions to improve clinical care for all infants, particularly female infants. For example, a robust G6PD newborn screening program paired with health education programs implemented in Sassari District, Sardinia, Italy, resulted in a 75% reduction in G6PD deficiency-related

complications, showing that individual diagnosis helped prevent haemolytic triggers in the at-risk population of young male children. The benefit was observed disproportionately in boys, suggesting the intervention had been less effective in girls, in part because girls with low-intermediate G6PD activity were misclassified as normal and not “at risk.”<sup>51</sup>.

Quantitative point-of-care G6PD tests enable diagnosis of female newborns with the intermediate phenotype. This could improve management of neonatal hyperbilirubinaemia, including closer clinical follow-up with targeted early bilirubin testing, avoidance of haemolytic triggers, and focused parental support on signs and symptoms of hyperbilirubinaemia to prevent kernicterus<sup>26–28,52</sup>. Additional studies using new quantitative point-of-care tests for G6PD deficiency in the first year of life will show whether it is possible to use the results obtained at birth to provide a definitive diagnosis of the phenotype at least as G6PD deficient, or normal with intermediate perhaps requiring further follow up. This, coupled with the capacity to retain data throughout a patient’s life via personal or health system records, may enable once-per-lifetime testing of G6PD status.

As with adult management of malaria, newborn screening with only qualitative G6PD tests will identify males who are most frequently at the highest risk of developing G6PD related complications in the early neonatal period but will miss G6PD intermediate females. This means that largely avoidable G6PD related complications in female infants may be managed insufficiently, again introducing health related gender disparities from birth<sup>26–28,52</sup>.

### Counseling with G6PD testing

While G6PD deficiency testing has been conducted systematically in certain settings to support malaria treatment, it is not usually treated as a test requiring genetic counseling. Little guidance currently exists to help health care workers relay results to patients, explain the hereditary nature and autosomal dominant inheritance pattern of the condition, and describe the clinical implications. While genetic counseling is still emerging in its application in low-resource settings<sup>53–55</sup>, the malaria field could draw lessons from genetic counseling in other domains, such as sickle cell disease, thalassaemia, other haemoglobinopathies, prenatal testing, and cancer genetics<sup>56–61</sup>. In many regions, where malaria is highly prevalent, inherited blood disorders overlap in prevalence, such that investment in genetic counseling capacity-building in these areas may be leveraged<sup>22</sup>. This could be achieved by increasing the number of genetics professionals being trained annually and building such opportunities into genomics research projects, or by equipping other healthcare staff, including nurses and community healthcare workers to interpret genetic knowledge<sup>62</sup>.

While an individual can potentially go through life unaware of his or her G6PD deficiency, there are still lifelong benefits to being informed. Particularly for women, awareness of G6PD status and understanding of inheritance bring a direct clinical advantage, with the added benefit of prompting testing in their

newborns and other family members. In low-resource settings, screening for haemoglobinopathies often occurs too late to intervene, either during a pregnancy or after the birth of the severely sick child, bringing social stigma to the woman who “caused” the disease in the child<sup>63</sup>. In contrast, for G6PD deficiency, an opportunity exists to develop genetic counseling specifically designed to minimise stigma and maximise the importance of knowing one’s G6PD status in order to actively prevent haemolytic triggers and inform other decisions throughout an individual’s lifespan, such as food and environmental factors to avoid. Counseling may be particularly important to clarify potential gender biases given the x-linked inheritance and variable penetrance amongst male and female individuals, as well as population biases due to increased screening of certain populations<sup>64</sup>.

### Community engagement and sensitisation

Community engagement is essential for creating awareness and minimising stigma<sup>63</sup>. Extensive literature indicates the importance of community engagement and community leadership for the success of programs involving screening for genetic disorders<sup>56,59,61,65,66</sup>, including some targeting G6PD deficiency in Sardinia, Italy<sup>51</sup>. Lessons from studies performed at the community level for neglected tropical diseases suggest that coverage rates, adherence rates, and general acceptability of new interventions improve when communities and community leadership are involved<sup>67,68</sup>. For example, in a recent evaluation of rapid tests for onchocerciasis, participants reported that the manner in which new tests were introduced and results were provided influenced community perceptions of the acceptability of the tests and confidence in the test results<sup>69</sup>.

These same strategies can be adapted and applied to the expansion of G6PD diagnostics. Women who learn their G6PD status—and who understand the broader consequences of their condition through enhanced genetic counseling—may encourage their family members to get tested. This could have an important ripple effect within families and communities in malaria-endemic settings, whereby more individuals seek testing (possibly independent of a malaria episode), learn their G6PD status, and use this information for decisions beyond malaria treatment.

### Cost-effectiveness of G6PD testing

The health and economic advantages of introducing quantitative, point-of-care G6PD testing in malaria case management will be optimised if the clinical benefits of testing can be extended beyond the current primary indication, which is for malaria treatment. At a minimum, retention of the G6PD test result by the patient or the health system may eliminate the need to test again the next time a patient requires radical cure for *P. vivax* or develops a haemolytic crisis, during which testing would not be reliable<sup>70</sup>. At a higher level, knowing one’s G6PD status at subsequent clinical visits would allow the patient and clinician to prevent a severe haemolytic crisis by avoiding oxidative medication, or if such a prescription is unavoidable, to closely monitor for signs of haemolysis so interventions could be performed earlier. This benefit could expand even

further if, through appropriate community sensitisation and genetic counseling, a woman with intermediate G6PD activity seeks G6PD testing for her newborn. This could prompt closer follow-up care and avert hyperbilirubinaemia-related complications. For example, in Singapore, the implementation of universal screening for G6PD deficiency in the context of the Kernicterus Surveillance Programme has led to elimination of kernicterus in the country<sup>71,72</sup>.

Clearly, when G6PD status can be retained by the individual or medical facility, repeated testing is not necessary, which in turn generates cost savings. However, when G6PD status cannot be reliably retained (e.g., in migratory populations) or requires confirmatory testing (i.e., initial test during an acute illness), repeated G6PD testing will reduce the cost-effectiveness of any health-related program, such as malaria elimination.

Current rough estimates indicate that a quantitative reader machine for G6PD (i.e., biosensor) will be in the range of 50–300USD, with test kits in the range of 3–5USD per unit; this will likely be less expensive than gold-standard spectrophotometric assay in terms of equipment (reader vs. temperature-controlled spectrophotometer), reagents (temperature-stable and ready-to-use vs. refrigerated), and operational time and time to result (clinical staff vs. specialised laboratory), but more expensive than the widely used qualitative fluorescent spot test. Point-of-care quantitative tests could be used in secondary-level health care facilities and introduced in selected primary-level facilities.

With increased utilisation of and experience in quantitative G6PD testing, data will need to be captured to precisely estimate the associated costs and benefits across all performance domains. This will better inform future test development. It will also impact country-level decision-making in terms of cost trade-offs within limited health care budgets and acceptable costs for incorporating G6PD testing into health systems. The cost-effectiveness will be weighted in part by the prevalence of G6PD deficiency in a population. For example, if the prevalence is low or negligible, the benefits of testing for G6PD deficiency may be limited to the clinical benefit for which it was indicated, most likely radical cure of *P. vivax* malaria<sup>73–75</sup>. Deliberations over cost-benefit trade-offs at the country and

district level should also include careful consideration of the ethical and equity considerations discussed above.

## Conclusion

The sex-linked differences in G6PD deficiency and limitations of current G6PD diagnostic tools have led to a disparity in accurate G6PD diagnosis in females. The interaction of *P. vivax* malaria, G6PD status, 8-aminoquinolines, and treatment restrictions linked to pregnancy and the post-partum period results in a sex related inequity. Females are at higher risk for misclassification of a sex-linked disorder in women, low awareness of iatrogenic haemolysis of the intermediate phenotype, and no *P. vivax* anti-relapse vivax treatment for a large part of their reproductive life.

Standard high-dose primaquine and new anti-relapse treatment regimens against *P. vivax* malaria require G6PD testing. New, affordable, point-of-care G6PD diagnostics have been developed to support access to these drugs in malaria-endemic populations. In addition to the benefits at multiple levels of the health care system, these new diagnostic tools can help bring awareness to the front-line practitioner of the nuances of G6PD deficiency in females and the potential implications beyond malaria.

Furthermore, integration of G6PD test results across multiple clinical indications, such as hyperbilirubinaemia in neonates, will likely require improved genetic counseling, health systems strengthening, and improved record keeping and data management. In settings in which G6PD deficiency is prevalent, these efforts may result in greater cost-benefits beyond the use of G6PD tests for malaria treatment alone, and lead to more equitable malaria treatment.

## Data availability

### Underlying data

No data are associated with this article

## Acknowledgments

The authors would like to acknowledge Athena Anderle and Ingela Ziemek for editorial support in the production of the manuscript.

## References

- Howes RE, Battle KE, Satyagraha AW, *et al.*: **G6PD deficiency: global distribution, genetic variants and primaquine therapy.** *Adv Parasitol.* 2013; **81**: 133–201.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- Luzzatto L, Nannelli C, Notaro R: **Glucose-6-Phosphate Dehydrogenase Deficiency.** *Hematol Oncol Clin North Am.* 2016; **30**(2): 373–93.  
[PubMed Abstract](#) | [Publisher Full Text](#)
- World Health Organization: **World Malaria Report 2018.** 2018.  
[Reference Source](#)
- Howes RE, Piel FB, Patil AP, *et al.*: **G6PD deficiency prevalence and estimates of affected populations in malaria endemic countries: a geostatistical model-based map.** *PLoS Med.* 2012; **9**(11): e1001339.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Howes RE, Battle KE, Mendis KN, *et al.*: **Global Epidemiology of Plasmodium vivax.** *Am J Trop Med Hyg.* 2016; **95**(6 Suppl): 15–34.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- World Health Organization: **Control and elimination of Plasmodium vivax malaria—a technical brief.** 2015.  
[Reference Source](#)
- Robinson LJ, Wampfler R, Betuela I, *et al.*: **Strategies for understanding and**



- reducing the *Plasmodium vivax* and *Plasmodium ovale* hypnozoite reservoir in Papua New Guinean children: a randomised placebo-controlled trial and mathematical model. *PLoS Med.* 2015; 12(10): e1001891.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
8. World Health Organization: Guidelines for the treatment of malaria. Third edition. 2015.  
[Reference Source](#)
  9. Chu CS, Bancone G, Moore KA, *et al.*: Haemolysis in G6PD Heterozygous Females Treated with Primaquine for *Plasmodium vivax* Malaria: A Nested Cohort in a Trial of Radical Curative Regimens. *PLoS Med.* 2017; 14(2): e1002224.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  10. Chu CS, Freedman DO: Tafenoquine and G6PD: a primer for clinicians. *J Travel Med.* 2019; 26(4): pii: taz023.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  11. Lacerda MVG, Llanos-Cuentas A, Krudsood S, *et al.*: Single-Dose Tafenoquine to Prevent Relapse of *Plasmodium vivax* Malaria. *N Engl J Med.* 2019; 380(3): 215–228.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  12. Llanos-Cuentas A, Lacerda MVG, Hien TT, *et al.*: Tafenoquine versus Primaquine to Prevent Relapse of *Plasmodium vivax* Malaria. *N Engl J Med.* 2019; 380(3): 229–241.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  13. Rueangweerayut R, Bancone G, Harrell EJ, *et al.*: Hemolytic Potential of Tafenoquine in Female Volunteers Heterozygous for Glucose-6-Phosphate Dehydrogenase (G6PD) Deficiency (G6PD Mahidol Variant) versus G6PD-Normal Volunteers. *Am J Trop Med Hyg.* 2017; 97(3): 702–711.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  14. Luzzatto L: G6PD deficiency: a polymorphism balanced by heterozygote advantage against malaria. *Lancet Haematol.* 2015; 2(10): e400–1.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  15. Domingo GJ, Advani N, Satyagraha AW, *et al.*: Addressing the gender-knowledge gap in glucose-6-phosphate dehydrogenase deficiency: challenges and opportunities. *Int Health.* 2019; 11(1): 7–14.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  16. Domingo GJ, Satyagraha AW, Anvikar A, *et al.*: G6PD testing in support of treatment and elimination of malaria: recommendations for evaluation of G6PD tests. *Malar J.* 2013; 12: 391.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  17. von Seidlein L, Auburn S, Espino F, *et al.*: Review of key knowledge gaps in glucose-6-phosphate dehydrogenase deficiency detection with regard to the safe clinical deployment of 8-aminoquinoline treatment regimens: a workshop report. *Malar J.* 2013; 12: 112.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  18. Chu CS, Bancone G, Nosten F, *et al.*: Primaquine-induced haemolysis in females heterozygous for G6PD deficiency. *Malar J.* 2018; 17(1): 101.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  19. Taylor WRJ, Thriemer K, von Seidlein L, *et al.*: Short-course primaquine for the radical cure of *Plasmodium vivax* malaria: a multicentre, randomised, placebo-controlled non-inferiority trial. *Lancet.* 2019; 394(10202): 929–938.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  20. Sex and Gender Sensitive Research Call to Action Group: Sex and gender in health research: updating policy to reflect evidence. *Med J Aust.* 2019.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  21. Crow JF: Hardy, Weinberg and language impediments. *Genetics.* 1999; 152(3): 821–5.  
[PubMed Abstract](#) | [Free Full Text](#)
  22. Bancone G, Gilder ME, Chowwiwat N, *et al.*: Prevalences of inherited red blood cell disorders in pregnant women of different ethnicities living along the Thailand-Myanmar border [version 2; peer review: 2 approved]. *Wellcome Open Res.* 2017; 2: 72.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  23. van den Broek L, Heylen E, van den Akker M: Glucose-6-phosphate dehydrogenase deficiency: not exclusively in males. *Clin Case Rep.* 2016; 4(12): 1135–1137.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  24. Anderle A, Bancone G, Domingo GJ, *et al.*: Point-of-Care Testing for G6PD Deficiency: Opportunities for Screening. *Int J Neonatal Screen.* 2018; 4(4): 34.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  25. Kaplan M, Beutler E, Vreman HJ, *et al.*: Neonatal hyperbilirubinemia in glucose-6-phosphate dehydrogenase-deficient heterozygotes. *Pediatrics.* 1999; 104(1 Pt 1): 68–74.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  26. Kaplan M, Hammerman C, Vreman HJ, *et al.*: Acute hemolysis and severe neonatal hyperbilirubinemia in glucose-6-phosphate dehydrogenase-deficient heterozygotes. *J Pediatr.* 2001; 139(1): 137–40.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  27. Riskin A, Gery N, Kugelman A, *et al.*: Glucose-6-phosphate dehydrogenase deficiency and borderline deficiency: association with neonatal hyperbilirubinemia. *J Pediatr.* 2012; 161(2): 191–6.e1.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  28. Wang FL, Boo NY, Ainoon O, *et al.*: Comparison of detection of glucose-6-phosphate dehydrogenase deficiency using fluorescent spot test, enzyme assay and molecular method for prediction of severe neonatal hyperbilirubinaemia. *Singapore Med J.* 2009; 50(1): 62–7.  
[PubMed Abstract](#)
  29. Premji Z, Umeh RE, Owusu-Agyei S, *et al.*: Chlorproguanil-dapsone-artesunate versus artemether-lumefantrine: a randomized, double-blind phase III trial in African children and adolescents with uncomplicated *Plasmodium falciparum* malaria. *PLoS One.* 2009; 4(8): e6682.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  30. Tiono AB, Dicko A, Ndububa DA, *et al.*: Chlorproguanil-dapsone-artesunate versus chlorproguanil-dapsone: a randomized, double-blind, phase III trial in African children, adolescents, and adults with uncomplicated *Plasmodium falciparum* malaria. *Am J Trop Med Hyg.* 2009; 81(6): 969–78.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  31. Pamba A, Richardson ND, Carter N, *et al.*: Clinical spectrum and severity of hemolytic anemia in glucose 6-phosphate dehydrogenase-deficient children receiving dapsone. *Blood.* 2012; 120(20): 4123–33.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  32. Todd P, Samarantunga IR, Pembroke A: Screening for glucose-6-phosphate dehydrogenase deficiency prior to dapsone therapy. *Clin Exp Dermatol.* 1994; 19(3): 217–8.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  33. Relling MV, McDonagh EM, Chang T, *et al.*: Clinical Pharmacogenetics Implementation Consortium (CPIC) guidelines for rasburicase therapy in the context of G6PD deficiency genotype. *Clin Pharmacol Ther.* 2014; 96(2): 169–74.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  34. Robinson KM, Yang W, Haidar CE, *et al.*: Concordance between glucose-6-phosphate dehydrogenase (G6PD) genotype and phenotype and rasburicase use in patients with hematologic malignancies. *Pharmacogenomics J.* 2019; 19(3): 305–314.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  35. Bancone G, Kalnoky M, Chu CS, *et al.*: The G6PD flow-cytometric assay is a reliable tool for diagnosis of G6PD deficiency in women and anaemic subjects. *Sci Rep.* 2017; 7(1): 9822.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  36. Kalnoky M, Bancone G, Kahn M, *et al.*: Cytochemical flow analysis of intracellular G6PD and aggregate analysis of mosaic G6PD expression. *Eur J Haematol.* 2018; 100(3): 294–303.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  37. Peters AL, Van Noorden CJ: Glucose-6-phosphate dehydrogenase deficiency and malaria: cytochemical detection of heterozygous G6PD deficiency in women. *J Histochem Cytochem.* 2009; 57(11): 1003–11.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  38. Recht J, Ashley EA, White NJ: Use of primaquine and glucose-6-phosphate dehydrogenase deficiency testing: Divergent policies and practices in malaria endemic countries. *PLoS Negl Trop Dis.* 2018; 12(4): e0006230.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  39. Brito-Sousa JD, Santos TC, Avalos S, *et al.*: Clinical Spectrum of Primaquine-induced Hemolysis in Glucose-6-Phosphate Dehydrogenase Deficiency: A 9-Year Hospitalization-based Study From the Brazilian Amazon. *Clin Infect Dis.* 2019; 69(8): 1440–1442.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  40. Chu CS, Phyto AP, Turner C, *et al.*: Chloroquine Versus Dihydroartemisinin-Piperaquine With Standard High-dose Primaquine Given Either for 7 Days or 14 Days in *Plasmodium vivax* Malaria. *Clin Infect Dis.* 2019; 68(8): 1311–1319.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  41. Commons RJ, Simpson JA, Thriemer K, *et al.*: The haematological consequences of *Plasmodium vivax* malaria after chloroquine treatment with and without primaquine: a WorldWide Antimalarial Resistance Network systematic review and individual patient data meta-analysis. *BMC Med.* 2019; 17(1): 151.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  42. Tantular IS, Matsuoka H, Kasahara Y, *et al.*: Incidence and mutation analysis of glucose-6-phosphate dehydrogenase deficiency in eastern Indonesian populations. *Acta Med Okayama.* 2010; 64(6): 367–73.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  43. Alam MS, Kibria MG, Jahan N, *et al.*: Field evaluation of quantitative point of care diagnostics to measure glucose-6-phosphate dehydrogenase activity. *PLoS One.* 2018; 13(11): e0206331.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  44. Pal S, Bansil P, Bancone G, *et al.*: Evaluation of a Novel Quantitative Test for Glucose-6-Phosphate Dehydrogenase Deficiency: Bringing Quantitative Testing for Glucose-6-Phosphate Dehydrogenase Deficiency Closer to the Patient. *Am J Trop Med Hyg.* 2019; 100(1): 213–221.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
  45. Glucose-6-phosphate dehydrogenase deficiency. WHO Working Group. *Bull World Health Organ.* 1989; 67(6): 601–11.  
[PubMed Abstract](#) | [Free Full Text](#)
  46. Bhutani VK, Maisels MJ, Stark AR, *et al.*: Management of jaundice and prevention of severe neonatal hyperbilirubinemia in infants >or=35 weeks gestation. *Neonatology.* 2008; 94(1): 63–7.  
[PubMed Abstract](#) | [Publisher Full Text](#)
  47. Cunningham AD, Hwang S, Mochly-Rosen D: Glucose-6-Phosphate

- Dehydrogenase Deficiency and the Need for a Novel Treatment to Prevent Kernicterus.** *Clin Perinatol.* 2016; 43(2): 341–54.  
[PubMed Abstract](#) | [Publisher Full Text](#)
48. Kaplan M, Hammerman C: **Glucose-6-phosphate dehydrogenase deficiency and severe neonatal hyperbilirubinemia: a complexity of interactions between genes and environment.** *Semin Fetal Neonatal Med.* 2010; 15(3): 148–56.  
[PubMed Abstract](#) | [Publisher Full Text](#)
49. Olusanya BO, Emokpae AA, Zamora TG, *et al.*: **Addressing the burden of neonatal hyperbilirubinaemia in countries with significant glucose-6-phosphate dehydrogenase deficiency.** *Acta Paediatr.* 2014; 103(11): 1102–9.  
[PubMed Abstract](#) | [Publisher Full Text](#)
50. Olusanya BO, Ogunlesi TA, Kumar P, *et al.*: **Management of late-preterm and term infants with hyperbilirubinaemia in resource-constrained settings.** *BMC Pediatr.* 2015; 15: 39.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
51. Meloni T, Forteoloni G, Meloni GF: **Marked decline of favism after neonatal glucose-6-phosphate dehydrogenase screening and health education: the northern Sardinian experience.** *Acta Haematol.* 1992; 87(1–2): 29–31.  
[PubMed Abstract](#) | [Publisher Full Text](#)
52. Fu C, Luo S, Li Q, *et al.*: **Newborn screening of glucose-6-phosphate dehydrogenase deficiency in Guangxi, China: determination of optimal cutoff value to identify heterozygous female neonates.** *Sci Rep.* 2018; 8(1): 833.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
53. Abacan M, Alsubaie L, Barlow-Stewart K, *et al.*: **The Global State of the Genetic Counseling Profession.** *Eur J Hum Genet.* 2019; 27(2): 183–197.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
54. Marsh V, Kombe F, Fitzpatrick R, *et al.*: **Consulting communities on feedback of genetic findings in international health research: sharing sickle cell disease and carrier information in coastal Kenya.** *BMC Med Ethics.* 2013; 14: 41.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
55. Zhong A, Darren B, Loiseau B, *et al.*: **Ethical, social, and cultural issues related to clinical genetic testing and counseling in low- and middle-income countries: a systematic review.** *Genet Med.* 2018.  
[PubMed Abstract](#) | [Publisher Full Text](#)
56. Anie KA, Treadwell MJ, Grant AM, *et al.*: **Community engagement to inform the development of a sickle cell counselor training and certification program in Ghana.** *J Community Genet.* 2016; 7(3): 195–202.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
57. Culhane-Pera KA, Moua M, Vue P, *et al.*: **Leaves imitate trees: Minnesota Hmong concepts of heredity and applications to genomics research.** *J Community Genet.* 2017; 8(1): 23–34.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
58. Culhane-Pera KA, Straka RJ, Moua M, *et al.*: **Engaging Hmong adults in genomic and pharmacogenomic research: Toward reducing health disparities in genomic knowledge using a community-based participatory research approach.** *J Community Genet.* 2017; 8(2): 117–125.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
59. Green NS, Mathur S, Kiguli S, *et al.*: **Family, Community, and Health System Considerations for Reducing the Burden of Pediatric Sickle Cell Disease in Uganda Through Newborn Screening.** *Glob Pediatr Health.* 2016; 3: 2333794x16637767.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
60. He LQ, Njambi L, Nyamori JM, *et al.*: **Developing clinical cancer genetics services in resource-limited countries: the case of retinoblastoma in Kenya.** *Public Health Genomics.* 2014; 17(4): 221–7.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
61. Houston AJ, Abel RA, Lindsey T, *et al.*: **Feasibility of a Community-Based Sickle Cell Trait Testing and Counseling Program.** *J Health Dispar Res Pract.* 2016; 9(3): pii: 1.  
[PubMed Abstract](#) | [Free Full Text](#)
62. Wonkam A, de Vries J: **Returning incidental findings in African genomics research.** *Nat Genet.* 2020; 52(1): 17–20.  
[PubMed Abstract](#) | [Publisher Full Text](#)
63. Marsh VM, Kamuya DM, Molyneux SS: **'All her children are born that way': gendered experiences of stigma in families affected by sickle cell disorder in rural Kenya.** *Ethn Health.* 2011; 16(4–5): 343–59.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
64. Ley B, Luter N, Espino FE, *et al.*: **The challenges of introducing routine G6PD testing into radical cure: a workshop report.** *Malar J.* 2015; 14: 377.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
65. Cao A, Congiu R, Sollaino MC, *et al.*: **Thalassaemia and glucose-6-phosphate dehydrogenase screening in 13- to 14-year-old students of the Sardinian population: preliminary findings.** *Community Genet.* 2008; 11(3): 121–8.  
[PubMed Abstract](#) | [Publisher Full Text](#)
66. Hsu LL, Green NS, Donnell Ivy E, *et al.*: **Community Health Workers as Support for Sickle Cell Care.** *Am J Prev Med.* 2016; 51(1 Suppl 1): S87–98.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
67. Amazigo U, Crump A, Godal T: **Recognising the role of community-directed treatment and of women in the fight against NTDs.** *Lancet Glob Health.* 2017; 5(6): e569–e570.  
[PubMed Abstract](#) | [Publisher Full Text](#)
68. Tchounkeu YF, Onyeneho NG, Wanji S, *et al.*: **Changes in stigma and discrimination of onchocerciasis in Africa.** *Trans R Soc Trop Med Hyg.* 2012; 106(6): 340–347.  
[PubMed Abstract](#) | [Publisher Full Text](#)
69. Dieye Y, Storey HL, Barrett KL, *et al.*: **Feasibility of utilizing the SD BIOLINE Onchocerciasis IgG4 rapid test in onchocerciasis surveillance in Senegal.** *PLoS Negl Trop Dis.* 2017; 11(10): e0005884.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
70. Chu CS, Bancone G, Soe NL, *et al.*: **The impact of using primaquine without prior G6PD testing: a case series describing the obstacles to the medical management of haemolysis.** *Wellcome Open Res.* 2019; 4: 25.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
71. Ho NK: **Neonatal jaundice. A second 4-year experience in Toa Payoh Hospital (1986-1989).** *J Singapore Paediatr Soc.* 1991; 33(3–4): 149–55.  
[PubMed Abstract](#)
72. Shah VA, Yeo CL: **Identifying risk of neonatal hyperbilirubinaemia and early discharge for glucose-6-phosphate dehydrogenase deficient newborns in Singapore.** *Ann Acad Med Singapore.* 2007; 36(12): 1003–9.  
[PubMed Abstract](#)
73. Devine A, Parmiter M, Chu CS, *et al.*: **Using G6PD tests to enable the safe treatment of Plasmodium vivax infections with primaquine on the Thailand-Myanmar border: A cost-effectiveness analysis.** *PLoS Negl Trop Dis.* 2017; 11(5): e0005602.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
74. Peixoto HM, Brito MA, Romero GA, *et al.*: **Cost-effectiveness analysis of rapid diagnostic tests for G6PD deficiency in patients with Plasmodium vivax malaria in the Brazilian Amazon.** *Malar J.* 2016; 15: 82.  
[Publisher Full Text](#)
75. Xu JZ, Francis RO, Lerebours Nadal LE, *et al.*: **G6PD Deficiency in an HIV Clinic Setting in the Dominican Republic.** *Am J Trop Med Hyg.* 2015; 93(4): 722–9.  
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

# Open Peer Review

Current Peer Review Status:  

---

## Version 1

Reviewer Report 28 July 2020

<https://doi.org/10.21956/wellcomeopenres.17206.r38264>

© 2020 Menon M. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



### Manoj Menon

Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, WA, USA

In this review, "Optimizing G6PD testing for Plasmodium vivax case management: why sex, counseling, and community engagement matter", the authors advocate for the need for quantitative testing for G6PD - which could help identify an intermediate phenotype and potentially at risk of the sequelae of hemolysis. This would help foster both a clinical benefit as well as improve health equity.

1. Would consider specifying the danger of G6PD deficiency (as well as G6PD intermediate) earlier in the introduction.
2. Introduction, paragraph 1: "For most countries approaching malaria elimination, *P. vivax* is now the main contributor to malaria disease burden," Is this for coendemic countries or all countries? Please specify and source.
3. Introduction, paragraph 3: What percentage of intermediate test results are misclassified as normal?
4. Given that this review is meant to focus on G6PD testing for *P. vivax*, I wonder if there is too much emphasis on the newborn screening. It is obviously an important component, but not as related to the topic (as per the title of the review article)
5. "Guidelines for rasburicase therapy in the context of genotyping". This seems a bit out of place, the use of rasburicase for the management of tumor lysis syndrome (a relatively rare event) would be so uncommon in areas of malaria endemicity. There are other drugs which, I think, are more common: e.g. Co-trimoxazole, nitrofurantoin, possibly aspirin)
6. Counseling with G6PD testing, paragraph 1, 2nd sentence. Would clarify that G6PD is inherited in an x-linked pattern, not an "autosomal dominant inheritance pattern"
7. With regard to cost-effectiveness, would it be an effective strategy to use the less expensive qualitative test for males and the quantitative test for females?

8. The figure (population histogram) and table 1 (research agenda) are helpful. Consider discussing in the text.

Thank you for allowing me to review this interesting manuscript.

**Is the topic of the review discussed comprehensively in the context of the current literature?**

Yes

**Are all factual statements correct and adequately supported by citations?**

Yes

**Is the review written in accessible language?**

Yes

**Are the conclusions drawn appropriate in the context of the current research literature?**

Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** hematology, (previously malaria)

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**

Reviewer Report 15 July 2020

<https://doi.org/10.21956/wellcomeopenres.17206.r39288>

© 2020 Wongsrichanalai C. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



**Chansuda Wongsrichanalai**

Independent Consultant, Bangkok, Thailand

This review paper addresses the issues of access to effective radical treatment of *P. vivax* malaria. The danger of hemolysis associated with the use of 8-aminoquinoline drugs in G6PD-deficient malaria patients is emphasized. The authors provide an excellent review of the genetic basis of G6PD deficiency and the limitation of tools in the past to detect G6PD enzyme at Point-of-Care (PoC).

The recently-available PoC quantitative G6PD test is pointed out as an innovative tool that improves safe *P. vivax* radical cure by reporting the enzyme levels, which help to guide more precise 8-aminoquinoline treatment choices. As such, it offers safer primaquine treatment for heterozygous females. Additional benefit of the PoC quantitative G6PD tests for newborn screening to determine the risks of hyperbilirubinemia is also clearly discussed and possible research opportunities are suggested.

**Comments:**

1. In the Abstract “..... the availability of SIMPLE, affordable....”

By default, most readers working on malaria in the field would think of a SIMPLE test being somewhat similarly simple to malaria rapid test (RDT). Because the current PoC quantitative G6PD test is still not that simple, this sentence should be modified. The issue of test simplicity is considered by many field malaria workers as its constraint; this should also be mentioned somewhere in the paper. Probably the next generation of the test could be made simpler for users in public health services in the heart of remote malaria endemic areas?

Deployment of the current PoC quantitative G6PD tests will have limitations in some malaria settings depending on the levels of health/economic development of the country. The authors touch lightly on this matter (3<sup>rd</sup> paragraph of Cost-Effectiveness) indicating that the test “could be used in secondary-level health care facilities and introduced in selected primary-level facilities” but in practice it can be much more complicated. For example, in Cambodia, the test is not yet user-friendly enough to be operable by Village Malaria Workers or at many rural health centers (where a large percentage of malaria cases are detected). The national malaria statistics would show the number of patients served by these different health care levels (with different lab facilities/personnel capability), so it might be of interest to determine what percentage of *P. vivax* patients could potentially benefit from this new G6PD testing in such a country.

2. Cost-effectiveness

The authors made some good suggestions to apply PoC quantitative G6PD test beyond malaria to improve cost-effectiveness. However, to consider cost-effectiveness of *P. vivax* malaria treatment specifically, it would be useful to take into consideration parallel efforts to improve adherence to multi-days primaquine regimen. As failure to complete the required primaquine doses is known to be a key obstacle to *P. vivax* radical cure, detection of G6PD deficiency and the ability to prescribe primaquine safely alone would not be sufficient to contribute to treatment effectiveness; we need to enhance compliance to therapy.

3. Neonatal screening for the risk of hyperbilirubinemia

The authors should make it clear to the readers that this is not an indication of the current G6PD test products (as far as this reviewer has learnt). If any test that is designed for that indication is already available, please specify. The review of using the test for screening at birth is very informative but there is a chance of readers being misled to assume that the products available in the market now are meant for that purpose. Do we still need clinical studies to assess such an off-label use (i.e. for neonatal screening)? Or a new PoC product intended for newborn G6PD screening is being developed?

4. Under *Introduction*, the top line

If possible the text should be updated so readers will be aware that tafenoquine has also been registered in *P. vivax* endemic countries such as Brazil and Thailand.

**Is the topic of the review discussed comprehensively in the context of the current literature?**

Yes

**Are all factual statements correct and adequately supported by citations?**

Yes

**Is the review written in accessible language?**

Yes

**Are the conclusions drawn appropriate in the context of the current research literature?**

Partly

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** malaria epidemiology, drug-resistant malaria, malaria diagnosis

**I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.**

---