



Polymyositis and dermatomyositis – challenges in diagnosis and management



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ABSTRACT

Polymyositis (PM) and dermatomyositis (DM) are different disease subtypes of idiopathic inflammatory myopathies (IIMs). The main clinical features of PM and DM include progressive symmetric, predominantly proximal muscle weakness. Laboratory findings include elevated creatine kinase (CK), autoantibodies in serum, and inflammatory infiltrates in muscle biopsy. Dermatomyositis can also involve a characteristic skin rash. Both polymyositis and dermatomyositis can present with extramuscular involvement. The causative factor is agnogenic activation of immune system, leading to immunologic attacks on muscle fibers and endomysial capillaries. The treatment of choice is immunosuppression. PM and DM can be distinguished from other IIMs and myopathies by thorough history, physical examinations and laboratory evaluation and adherence to specific and up-to-date diagnosis criteria and classification standards. Treatment is based on correct diagnosis of these conditions.

1. Introduction

The idiopathic inflammatory myopathies (IIMs), also referred to generally as myositis, are classified into polymyositis (PM), dermatomyositis (DM), sporadic inclusion body myositis (sIBM) and necrotizing autoimmune myopathy (NAM). The classification is based on clinical and histological features [1,2]. The incidence of IIMs are fairly low as reported by different research groups. Both children and adults may suffer from DM while PM mainly affect adults. Manifestations of IIMs include chronic muscle weakness and fatigue, and skin exanthema in DM. The pathophysiology involves agnogenic inflammation-mediated muscular and/or connective tissue damage, along with other organ involvement including the heart, lung, joints and gastrointestinal tract. IIMs are therefore characterized as a systemic autoimmune disorder. The mainstay of therapy is immunosuppression with corticosteroids, steroid-sparing agents and other immunosuppressive drugs [1,3,4]. History, physical examination, and multiple laboratory examinations such as serological tests, neurological tests and most importantly, muscle

biopsy, help to discriminate between PM and DM, and also other autoimmune disorders or myopathies. Improved knowledge of the pathogenesis will help to more accurately classify disease and establish better diagnostic criteria, which will define treatment algorithms. Moreover, basic researches and clinical trials are necessary to find and develop potential target and therapies in order to improve the prognosis of patients [1–3,5,6].

2. The etiology of polymyositis and dermatomyositis

2.1. The epidemiology of PM and DM

PM is rare in childhood and presents mainly after the second decade of life. The most common time of presentation is between 45 and 60 years of age. DM affects both children and adults with an overall female/male ratio of about 2:1 [7]. In the last twenty years, epidemiological studies have shown a higher incidence and prevalence than historically reported. This may be due to more thorough diagnosis criteria and improved

Abbreviations: APC, antigen presenting cell; AZA, Azathioprine; CAM, cancer associated myositis; CK, creatine kinase; DM, dermatomyositis; EMG, electromyography; HLA, human leukocyte antigen; IIM, idiopathic inflammatory myopathies; ILD, interstitial lung disease; IV, intravenous; JDM, juvenile dermatomyositis; MAA, myositis associated antibody; MAC, membrane attack complex; MHC, major histocompatibility complex; MMF, mycophenolate mofetil; MRI, magnetic resonance imaging; MSA, myositis specific antibody; MTX, methotrexate; MUAP, motor unit action potential; NAM, necrotizing autoimmune myopathy; PM, polymyositis; sIBM, sporadic inclusion body myositis; TNF, tumor necrosis factor; Treg, regulatory T cell; UVR, ultraviolet radiation.

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patient access and services. Data from recent epidemiological studies are shown in Table 1. It should be noted that the reported prevalence and incidence of PM and DM are quite variable depending on differences in study methodology, but female gender and urban living appear to be consistent risk factors.

2.2. Hereditary susceptibility

It is well accepted that autoimmunity results from susceptible genes, environmental factors and a dysregulated/dysfunctional immune system [8]. Evidence from case reports, family studies, studies of animal models, candidate gene case-control studies, and whole genome investigations supports a role for genetic factors in the etiology of autoimmune disease [9]. The earliest known cases of familial IIM were reported in the 1950s [10,11], and there is a scarcity of affected sibling pairs and twins. To date, the major histocompatibility complex (MHC) has been shown to be the strongest genetic association for the IIMs, and candidate gene studies have identified shared genetic susceptibility with other autoimmune diseases [12–15]. In IIM, the strongest association is with the 8.1 ancestral haplotype (8.1 AH, HLA-A1-B8-Cw7-DRB1*0301-DQA1*0501-C4A*Q0), a large common haplotype in Caucasian populations in Northern and Western Europe [16,17]. This allele has also been found in other populations including African-American, Japanese, and Spanish but not in Korean or Mexican-American [18–21]. However, which gene or genes contribute to the pathogenesis is not clear. In addition, the non-HLA genes *UBE2L3*, *CD28*, *TRAF6*, *STAT4* [17], *TNF- α* [22], *IKBL (NFKBIL1)* [23,24], *ACTN3* [25], *BLK* [26,27], *IRF5* [28] and *PLCL1* [29] have also been reported as potential risk factors for IIM. Moreover, differences in candidate genes among clinical subgroups have also been identified. For example, *PTPN22*, *IL18R1*, *RGS1*, *IFN- γ* [30] and *IFIH1* [31] have been reported to be associated with PM, while *GSDMB* has been reported as a risk factor for DM [17]. Studies have also been able to correlate HLA with serology, complications and responses to therapy in IIMs.

2.3. Environmental factors

In recent years, evidence has shown that environmental factors play also play a role in the development of autoimmunity. Environmental factors include infection, gut microbiota, drugs, chemicals, pollutants and physical agents [32,33]. Animal models of myositis have been developed that are induced by viruses, drugs, or parasites, providing additional evidence for the likely role of environmental agents in the pathogenesis of IIMs [34].

An online survey of DM patients from the USA and Canada examined environmental factors in patients with or without disease flares over a

period of 6 months and found that sun exposure and nonsteroidal anti-inflammatory drug (NSAIDs) were significant factors. In addition, urinary tract infections, gastroenteritis, elevated blood pressure, use of antidepressants, mood changes and relocation were also risk factors for disease flares [35]. The association between ultraviolet radiation (UVR) and DM has been reported by several groups, who have demonstrated that UVR may modulate the clinical and immunologic expression of DM, including the levels of autoantibodies [36–38]. Infection is thought to be an important contributor to immune system activation, and it has been reported that there is a high frequency of opportunistic infections in PM/DM, which may lead to an increase in mortality [39]. An association of viral infections and IIM has also been reported. Coxsackie B virus is associated with increased muscle tropism and is considered to be a potential trigger for PM/DM [40]. Human immunodeficiency virus (HIV) infection has been reported to foster an environment favorable for the development of DM [41]. Notably, PM and DM are associated with a high risk of malignancy [42] and it has been proposed that hepatocellular carcinoma (HCC) and/or a chronic HBV infection may play a role in the pathogenesis of DM through a paraneoplastic mechanism [43,44]. Studies also suggest a possible interaction between tobacco smoking and autoantibody phenotypes of PM/DM [45].

3. The pathology of polymyositis and dermatomyositis

3.1. Animal models

Animal models are important tools for investigating the mechanisms of autoimmune diseases for a number of reasons that include low numbers of patients, an inability to acquire patient samples, ethical issues of doing particular types of studies on humans, variable phenotypes of the disease, non-compliance with study protocols and cost. Compared to other well-researched autoimmune disease such as rheumatoid arthritis and systemic lupus erythematosus, the development of animal model research in PM/DM has been lagging. Dogs and mice are the only two nonhuman species which have been reported to spontaneously develop myositis [46,47]. SJL/J mice spontaneously develop a chronic IIM resembling human myositis which presents as muscle inflammation, centralized nuclei, and muscle fiber necrosis [48–50]. There is limited similarity to human myositis. On the other hand, myositis may be induced in animals by injection with autologous or heterologous muscle homogenates or C protein, purified muscle antigens, viruses, drugs, and naked DNA constructs [34,47]. There are several other animal models which reveal new insights regarding the pathophysiology of IIM [47], but unfortunately, no single animal model fully reproduces the clinical and pathologic features of human IIM.

Table 1
Epidemiological reported data.

Reference	District	Year	Incidence (per 100000)	Prevalence (per 100000)	Gender
[255]	Olmsted County, Minnesota	1976–2007	DM: 0.96 (0.61–1.32)	21.42 (13.07–29.77)	Female: 76%
[256]	Buenos Aires	1999–2009	PM: 0.75 (0.35–1.16) DM: 0.32 (0.1–0.99)	PM: 7.2 (2.9–14.7) DM: 10.22 (4.9–18.8)	Female: 76%
[257]	Spain	1997–2004	PM: 0.89 (0.37–0.41) DM: 0.49 (0.47–0.52)	/	Female > male, p < 0.001
[258]	Japan	2003–2010	PM/DM: 10–13	PM/DM: 100–130	/
[259]	Korea	2004–2015	PM: 0.16–0.3 DM: 0.18–0.4	PM: 1.4–2.1 DM: 1.2–2.7	Female: 72.1%
[260]	Quebec, Canada	2003	/	PM/DM: 21.5 (19.4–23.9)	Female: Adjusted OR 2.52 (1.08–3.99)
[261]	USA (commercially insured and Medicare and Medicaid enrolled populations)	2004–2008	PM: 2.46 (2.33–2.59) and 3.53 (3.13–3.94) DM: 1.52 (1.42–1.63) and 1.70 (1.42–1.97)	IIM: 20.62 to 25.32 and 15.35 to 32.74	Female: 67%
[262]	USA (Managed care plan)	2003–2008	IIM: 5.8–7.9	IIM: 14–17.4	Female: 66.8%
[263]	Taiwan	2003–2007	PM: 0.44 (0.4–0.48) DM: 0.71 (0.66–0.76)	/	/

3.2. Immunological mechanisms

The immunological signaling pathways and immunopathogenesis involved in PM and DM have been extensively reviewed [3,5,51–53]. In PM, there is evidence of antigen-directed cytotoxic CD8⁺ T cells surrounding and attacking MHC-I-antigen expressing muscle fibers [52, 54–57]. Up-regulation of costimulatory molecules (BB1 and ICOSL) and their ligands (CD28, CTLA-4, and ICOS), as well as ICAM-1 or LFA-1, stabilizes the synaptic interaction between CD8⁺ T cells and MHC-I on muscle fibers, which means that these muscle fibers act as antigen-presenting cells (APCs) [5,58–60]. Upon activation, perforin granules are released by auto-aggressive CD8⁺ T cells and mediate muscle-fiber necrosis [61]. In DM, the main target is the vascular endothelium. Early activation of complement C3 by putative antibodies directed against endothelial cells leads to the formation and deposition of C3b, C3bNEO, C4b fragments and C5b–9 membrane attack complex (MAC) on the endothelial cells. These markers can be detected in the serum and muscle of patients in the early phases of the disease [62,63]. Sequentially, the complement deposits induce swollen endothelial cells, vacuolization, capillary necrosis, perivascular inflammation, ischemia, and destruction of muscle fibers, which results in the remaining capillaries developing dilated lumens to compensate for the ischemia [64,65]. At the same time, complement activation leads to the release of proinflammatory cytokines and chemokines [3]. As a result, both innate and acquired immune cells are recruited to the perimysial and endomysial spaces through higher expression of adhesion molecules on endothelial cells interacting with the integrins on immune cells, leading to aggravated immune attack and antibody production [8].

T cell infiltrates in the muscles of patients with PM/DM are dominated by CD28-null T Cells, which are long-lived, proinflammatory and terminally differentiated T cells lacking CD28 [66]. These T cells are linked to resistance against immunosuppression and poor clinical outcome [67]. CD8⁺ T cells play a critical role in muscle fiber damage especially in PM. T-cell lines expanded from muscle biopsy material of IIM patients consist predominantly of CD8⁺ T cells and are cytotoxic to autologous myotubes [68]. Clonal expansion of peripheral blood CD8⁺ T cells with activation of STAT and pZAP70 signaling [69] is more frequently seen in patients of PM than DM [70,71]. Analysis of T cell receptor (TCR) antigen-binding region sequences suggests that T-cell expansion is driven by a common antigen, possibly an autoantigen [56, 72].

In DM, complement activation induces capillary destruction and perivascular inflammation. This process is mediated by CD4⁺ T cells [73]. STAT, forkhead box transcription factor (FoxP3), and pZAP70 expression in peripheral CD4⁺ T cells is suppressed in active DM, but except for FoxP3, are improved during periods of remission [56]. Immunohistochemical analysis reveals that FoxP3⁺ regulatory T (Treg) cells predominantly locate in perivascular and perimysial infiltrates of DM muscle. In juvenile DM (JDM), Treg cells from peripheral blood show a lower expression of CTLA-4 and are functionally compromised as well [74].

B cells are detected in muscle biopsy specimens of IIM patients. B cells and plasma cells that infiltrate into the perivascularity of DM patients are also found in all subtypes of IIMs [75,76]. Upregulated BAFF signaling and Toll like receptor (TLR) expression [77–82], decreased Breg subset [83], and notably, multiple autoantibody production, demonstrate a highly activated humoral immune state in DM and PM. Researchers analyzed the molecular characteristics of the antigen (Ag) receptor on B cells from muscle and found that BCR affinity maturation and oligoclonal expansion occurred, suggesting a B cell Ag-specific response in the muscle tissue of patients with DM and PM [84,85]. Accordingly, rituximab, a B cell-depleting agent, has been proven to be helpful in some cases of DM and PM [86].

The innate immune system also plays an important role in the pathogenesis of DM and PM. Overexpression of interferon (IFN) -regulated proteins and cytokines have been found in the skin and muscle of patients

with DM and PM, suggesting an upregulated “type I IFN signature” [87–90]. Moreover, plasmacytoid dendritic cells (pDC) are a possible source of upregulated IFN in the muscle of DM patients [91]. Besides dendritic cells (DCs) [92], mast cells [93], neutrophils [94] and macrophages [95–97] are also involved in the development of IIMs, mainly acting as APCs and a source of pro-inflammatory cytokines to activate the T cell response and mediate tissue inflammation [98].

3.3. Changes in the target tissue

During active phases of PM/DM, muscle fibers and endomysial capillaries experience pathological alteration. Firstly, MHC-I is ubiquitously upregulated in polymyositis, even on muscle fibers that are remote from the site of inflammation [99], which is probably induced by cytokines secreted by activated T cells [51,55]. In addition, muscle fibers in PM also express co-stimulatory factors including ICOSL, CD40L, CD80 and to form tight immune synapses with T cells [100,101]. Overexpression of myxovirus resistance A (MxA), a type I interferon-inducible protein, is observed in a perifascicular distribution or sometimes diffusely in biopsied muscle specimens of DM [91], and may help with differential diagnosis [102]. On the other hand, Fas antigen on muscle fibers and FasL on autoinvasive CD8-positive T cells are identified, but the Fas-FasL-dependent apoptotic process is not functionally normal [103, 104]. Expression of the anti-apoptotic molecules BCL2, Fas associated death domain-like interleukin-1-converting enzyme inhibitory protein (FLICE), and human IAP-like protein (hILP) may confer resistance of muscle to Fas mediated apoptosis [103,105,106].

4. Clinical features

Both PM and DM present with a varying degree of muscle weakness, usually developing slowly over weeks to months, but acutely in rare cases [107]. The weakness is relatively symmetric, predominantly proximal and unassociated with sensory loss or ptosis with sparing of extraocular muscles which are characteristics of myasthenia [3,107]. In the late stages of PM and DM, distal muscle weakness which affects fine motor movements can occur. In contrast, this feature is an early and prominent finding in sIBM [108].

In PM and DM, the neck extensor muscles may also be involved, causing difficulty in holding up the head and rarely causing a dropped head syndrome (DHS) [109]. Primary weakness of the diaphragm and accessory muscles, or pharyngeal muscles in advanced cases, may contribute to respiratory insufficiency or dysphagia, nasal speech, hoarseness, nasal regurgitation, and aspiration pneumonia [110,111]. The tendon reflexes are usually preserved but may be absent in severely weakened or atrophied muscles. Myalgia occurs in less than 30% patient with polymyositis and dermatomyositis [112].

There are many diseases whose symptoms resemble PM or DM, increasing the possibility of misdiagnosis and introducing additional challenges in classification and management, especially in PM. Patients with anti-synthetase autoantibodies may carry a diagnosis of DM or PM. Presenting symptoms include myalgias, muscle weakness, and a combination of interstitial lung disease (ILD), Raynaud phenomenon, seronegative arthritis of the distal joints, fever, mechanic’s hands, and a skin rash that is different from the heliotrope erythema typically seen in DM [113–115]. PM may be diagnosed erroneously in DM patients who present with isolated proximal muscle weakness and develop the rash months later [116]. sIBM has been shown as the most common disease misdiagnosed as PM, whereby sIBM is suspected retrospectively in patients who do not respond to therapy for polymyositis. PM may also be diagnosed incorrectly in cases of NAM, overlap syndrome associated with a connective tissue disease, muscular dystrophies, myalgia syndromes, toxic and endocrine myopathies, and Kennedy’s disease (KD) [2,107, 117–119]. In chronic DM, patients may suffer from fasciitis and thickening of the skin, which can also occur in patients with eosinophilia-myalgia syndrome, eosinophilic fasciitis, or macrophagic

myofasciitis [120,121].

4.1. Polymyositis

PM is frequently misdiagnosed, as it lacks a unique clinical phenotype and remains a diagnosis of exclusion [3,122]. PM is best defined as a subacute proximal myopathy that evolves muscle weakness over weeks to months, affects adults but rarely children, and excludes those who have a rash, a family history of neuromuscular disease, exposure to myotoxic drugs (e.g., statins, penicillamine, and zidovudine), involvement of facial and extraocular muscles, endocrinopathy, or a clinical phenotype of sIBM [3,5]. PM mimics many other myopathies and may also be diagnosed incorrectly in cases of DM, sIBM, NAM, overlap syndrome associated with a connective tissue disease, muscular dystrophies, myalgia syndromes, or toxic and endocrine myopathies [118,122–124].

4.2. Dermatomyositis

DM is identified by a characteristic rash accompanying or preceding subacute muscle weakness [3]. About 6% of patients have no or poorly recognized skin involvement. However, histologic feature of the muscular biopsy sample may be helpful in diagnosing DM. In these cases, where there is no skin involvement, the condition is termed dermatomyositis sine dermatitis [125]. Up to 20% patients with cutaneous features of DM and typical histopathologic features on muscle biopsy but without clinical muscle weakness for more than 6 months are categorized as amyopathic dermatomyositis (ADM) [126,127]. The skin manifestations of DM include a violaceous eruption (Gottron's papules) on the knuckles, which is pathognomonic for DM; a characteristic periorbital heliotrope (blue–purple) rash with edema; an erythematous rash on the face, knees, elbows, malleoli, neck, anterior chest (in a V-sign), and back and shoulders (in a shawl sign); and, which may evolve into a scaly discoloration. Dilated capillary loops at the base of the fingernails, irregular and thickened cuticles, and cracked palmar fingertips ("mechanic's hands") are characteristic of DM. These lesions are photosensitive and are commonly pruritic.

4.3. JDM

In children, DM is the most frequent inflammatory myopathy while PM is very rare [128]. DM in children is referred to as Juvenile Dermatomyositis (JDM).

The average age of onset of JDM is approximately 7 years old and the female-to-male ratio is about 2:1 [129–131]. The pathogenesis and major clinical and autoantibody phenotypes in children are similar to those in adults. Skin rash, consisting of a heliotrope eyelid discoloration and Gottron's papules, and proximal muscle weakness are the most common manifestations of JDM, while lesions include cutaneous calcinosis, which develops over pressure points, occurs more commonly in JDM [132]. Other lesions that occur more commonly in JDM include subcutaneous calcifications, sometimes extruding to the surface of the skin and causing ulcerations and infections. Rare dermatologic findings include non-scarring alopecia, erythroderma, vesiculobullous lesions, leukocytoclastic vasculitis, and livedo reticularis [3,5,127,132].

Other organ systems including the gastrointestinal tract, lungs, heart, articular, visual, and nervous system can also be involved [131], although the prevalence of ILD and cancer is different between JDM and adult DM. An autoantibody to a 155 kDa protein with a second weaker 140 kDa band, called anti-p155/140, has been found in sera of 23–29% of JDM cases and is associated with a risk of more severe skin involvement and generalized lipodystrophy [133–135]. The serological and genetic differences between adult DM and JDM may provide insights into the pathogenic mechanisms that underlie their clinical differences [136].

4.4. Other extramuscular manifestations and involvements

IIM patients exhibit many other extramuscular manifestations. As a result, diagnosis and the evaluation for other organ system involvement is imperative towards determining the optimal treatment strategy for PM and DM.

4.4.1. Cardiac abnormalities

In adult onset IIM, cardiovascular complications represent a major cause of clinical deterioration and death [137]. Cardiac involvement may include inflammation of the myocardium, accelerated coronary atherosclerosis, angina, dysrhythmias and more [138–140]. Heart abnormalities may occur during any phase of PM/DM, even when PM/DM is in remission [141]. According to a retrospective analysis of adults PM/DM patients in British Columbia, there is an increased risk of myocardial infarction (MI) but not of stroke in patients with PM/DM compared with a control cohort [142]. Electrocardiograms (ECG), echocardiogram (ECHO), cardiac magnetic resonance (CMR) imaging and cardiac enzymes analysis may be helpful to diagnose subclinical heart complications [143–145]. In addition, anti-mitochondrial antibodies (AMA) may be associated with cardiovascular involvement in IIM [146]. The efficacy of glucocorticoids and immunosuppressants in the treatment of cardiac complications is undetermined [147,148].

4.4.2. Pulmonary symptoms

Interstitial lung disease (ILD) is a common extramuscular manifestation of myositis. PM/DM patients with accompanying ILD have poorer prognosis than those without [149,150] DM-ILD usually demonstrates a more severe course with a poorer prognosis that is more resistant to treatment than PM-ILD [151,152]. The characteristics of PM/DM-ILD include a non-productive inspiratory cough and dyspnea [153], lymphocyte or neutrophil alveolitis in bronchoalveolar lavage (BAL), and interstitial pneumonia in lung biopsy samples [154]. High-resolution computed tomography (HRCT) and pulmonary function tests (PFT) are important for diagnosis of lung involvement in PM and DM [155,156].

Recently, two types of myositis-specific autoantibodies (MSAs), anti-aminoacyl transfer RNA synthetases (ARS) and anti-CADM-140 (MDA-5/IFIH1) antibodies, have been shown to be associated with ILD in myositis, suggesting separate clinical and serological phenotypes [157]. For example, numerous reports have indicated a higher prevalence of rapidly progressive ILD with ADM and anti-MDA-5 antibodies, with many patients being refractory to immunosuppressive therapy [158–160]. In the case of anti-Jo-1 associated ILD, the presence of high levels of anti-Ro52 antibodies predicts a more severe acute-onset disease and non-responsiveness to immunosuppressive treatment [161,162].

4.4.3. Malignancy

An association between PM/DM and malignant disease has been reported and confirmed. Studies show that DM is strongly associated with ovarian, lung, pancreatic, stomach, colorectal cancers, and non-Hodgkin lymphoma [163], while PM is associated with an increased risk of non-Hodgkin lymphoma, lung and bladder cancers [164–168]. JDM has a 16-fold increased risk for hematopoietic or lymphoid malignancy [169]. In addition, malignant disease is more common in older patients (>50 years of age) and may occur before the onset of PM/DM, concurrently with PM/DM, or after the onset of PM/DM. However, the risk of malignant disease is highest shortly after myositis diagnosis in both DM and PM [164,169,170].

Scientists have assessed the diagnostic values of serum tumor markers for the detection of solid cancer in PM/DM patients and found that carcinoembryonic antigen CA125 and CA19-9 assessment may be useful markers [171]. On the other hand, there are several examples of tumor-associated immune responses that target seemingly unrelated tissues in a predictable fashion, which has been extensively described in the autoimmune paraneoplastic neurological disorders (PNDs) [172]. Compared to IIM without cancer, cancer associated myositis (CAM)

shows different clinical and immunological features, as well as antigen expression [173,174]. Absent myositis-specific/associated autoantibodies and positive anti-155/140 antibody, anti-SAE1, anti-TIF1- γ and anti-NXP2 antibodies are associated with a high risk of CAM [175–177]. Moreover, the gene expression profile of IIM with malignancy is similar to that of DM rather than PM, which suggests that humoral immunity plays a significant role in PM/DM [178].

4.4.4. Other manifestations

Several studies show an increased prevalence of celiac disease in IIM [179,180]. Renal involvement develops in about one fifth of IIM patients [181–183]. PM and DM are frequently associated with systemic sclerosis and mixed connective tissue disease in the context of an overlap syndrome [184,185]. It has also been reported that several cases of JDM have been complicated by systemic capillary leak syndrome (SCLS), a rare, life-threatening disorder characterized by severe hypotension, hypoalbuminaemia and hemoconcentration [186]. Lipodystrophy, hypertriglyceridemia and insulin resistance have also been associated with JDM [187,188].

5. Prognosis

A United States study in 2012 of 160 p.m./DM patients demonstrated a 10 year survival rate of 62% [189]. Deaths are mainly caused by cardiac (22%) and pulmonary (22%) complications, infections (15%), and cancer (11%) [189]. Prognostic factors including gender, age at time of diagnosis, presence of Raynaud phenomenon, ILD, dysphagia, respiratory muscle involvement and cardiac involvement at any time in the clinical course affect prognosis [137], while the prognostic role of autoantibodies needs further long-term investigation. The long-term data for JDM are still scarce. Compared to age-matched controls, adults who had JDM showed reduced quality of life and reduced fitness measured by maximal oxygen uptake as a measure of muscle function [190,191].

6. Diagnosis and classification

Bohan and Peter's diagnostic criteria, proposed in 1975, have been widely accepted [1,192]. Patients complaining about muscle weakness, fatigue and myalgia, with or without skin rash should be suspected as having PM or DM. Table 2 listed the Bohan and Peter's diagnostic criteria of PM/DM with the exclusion of family history of neuromuscular disorder, endocrine or neurogenic diseases, myotoxic drug exposure, muscular dystrophies and metabolic myopathies, sIBM, NAM or infection [1,192,193]. Because of studies with undersized patient cohort and potentially erroneous disease classification, these criteria are not perfect, and often fail to rule out IBM.

In 2003, Dalakas and Hohlfeld supplemented the existing criteria with pathologic features [3]. First, they reiterated the crucial role of the

muscle biopsy test, and proposed the following: primary inflammation with the CD8/MHC-I complex and no vacuoles for definite PM, ubiquitous MHC-I expression but no CD8⁺ cell infiltrates or vacuoles for probable PM, perifascicular, perimysial or perivascular infiltrates, perifascicular atrophy and rash present for definite DM; no rash present for probable DM. In addition, ADM is diagnosed when a rash is present but biopsy findings are nonspecific or are diagnostic for DM, and no weakness is present.

In the MSG and ENMC international workshop in 2003, many neurologists, rheumatologists, and statisticians worked together to propose classification criteria for IIMs [2], as shown in Table 3. More importantly, this workshop provides information on how to apply these criteria to each category of myositis, including definite and probable PM/DM, ADM, possible DM sine dermatitis, non-specific myositis and NAM. At the same time, this workshop also pointed out the unmet needs in treatment due to difficulties in the study design and the low incidence and prevalence of patients. The workshop also promoted the development of valid, sensitive, and reliable outcome measures for (randomized controlled trial) RCTs in myositis.

In recent decades, muscle immunopathology, myositis specific autoantibodies testing, and new techniques of muscle imaging such as contrast-enhanced ultrasound or Magnetic Resonance Imaging (MRI) have been used in the diagnosis of patients with IIM, contributing to improved diagnostic capability [125].

6.1. Clinical history and physical examinations

Obtaining a clinical history is crucial and should include information on disease onset, pattern of presentation, and possible inciting or environmental factors. Environmental factors including recent infections, drugs (over-the-counter, prescription or recreational), work exposures, diet and nutritional supplements. Family history can help determine the potential genetic contributors to the myopathy. In addition, a good history should also elicit information regarding pattern of weakness, including distal versus proximal, symmetric versus asymmetric, or bulbar involvement. This portion of the history can be confirmed or supported by physical findings. Validated patient/parent questionnaire of activities of daily living (Health Assessment Questionnaire, HAQ/childhood Health Assessment Questionnaire CHAQ) and validated observational tools of function, strength and endurance (Childhood Myositis Assessment Scale, CMAS) are recommended by the International Myositis Assessment and Clinical Studies Group (IMACS) [194,195].

6.2. Blood tests

6.2.1. Muscle enzymes

When the muscles are damaged, muscle enzyme elute from muscle fibers leading which can be detectable in plasma or serum. Serum

Table 2

Bohan and peter criteria for polymyositis and dermatomyositis.

Cretirion	Polymyositis		Dermatomyositis		Details
	Definite	Probable	Definite	Mild or early	
Muscle strength	Myopathic muscle weakness	Myopathic muscle weakness	Myopathic muscle weakness	Seemingly normal strength	Symmetrical and progressive weakness of the proximal limb-girdle muscles
Electromyographic findings	Myopathic	Myopathic	Myopathic	Myopathic or nonspecific	Short duration, low amplitude polyphasic units; fibrillations, complex repetitive discharges; positive sharp waves
Muscle enzymes	Elevated (up to 50-fold)	Elevated (up to 50-fold)	Elevated (up to 50-fold)	Elevated (up to 10-fold)	Creatine kinase, aspartate aminotransferase, alanine aminotransferase, lactate dehydrogenase, aldolase
Muscle-biology findings	Diagnostic for this type of inflammatory myopathy	Nonspecific myopathy without signs of primary inflammation	Diagnostic	Nonspecific or diagnostic	Perivascular, interfascicular or endomysial inflammatory infiltration; phagocytosis, necrosis, and regeneration of muscle fibers; capillary obliteration and endothelial damage
Rash or calcinosis	Absent	Absent	Present	Present	Heliotrope rash and Gottron's sign/papules

Table 3

Elements of the classification criteria for the IIM (except sIBM) approved by the MSG and the 119th ENMC workshop.

Clinical criteria	Inclusion criteria	a) Adult onset, or in childhood in DM and nonspecific myositis b) Subacute or insidious onset c) Pattern of weakness: symmetric proximal > distal, neck flexor > neck extensor d) Rash typical of DM: heliotrope periorbital edema; Gottron's papules/sign, scaly if chronic, at metacarpophalangeal and interphalangeal joints and other bony prominences; V-sign and shawl sign
	Exclusion criteria	a) Clinical features of IBM ^a b) Ocular weakness, isolated dysarthria, neck extensor > neck flexor weakness c) Toxic myopathy, active endocrinopathy, amyloidosis, family history of muscular dystrophy or proximal motor neuropathies
Electromyography	Inclusion criteria	a) Increased insertional and spontaneous activity in the form of fibrillation potentials, positive sharp waves, or complex repetitive discharges b) Morphometric analysis reveals the presence of short duration, small amplitude, polyphasic MUAPs
	Exclusion criteria	a) Myotonic discharges that would suggest proximal myotonic dystrophy or other channelopathy b) Morphometric analysis reveals predominantly long duration, large amplitude MUAPs c) Decreased recruitment pattern of MUAPs
Other laboratory criteria		a) MRI: diffuse or patchy increased signal within muscle tissue on STIR images b) Myositis-specific antibodies detected in serum c) Elevated serum creatine kinase level
Muscle biopsy		a) Endomysial inflammatory cell infiltrate (T-Cells) surrounding and invading non-necrotic muscle fibers b) Endomysial CD8 ⁺ T-cells surrounding, but not definitely invading non-necrotic muscle fibers, or ubiquitous MHC-I expression c) Perifascicular atrophy d) MAC depositions on small blood vessels, or reduced capillary density, or tubuloreticular inclusions in endothelial cells, or MHC-I expression of perifascicular fibers e) Perivascular, perimysial inflammatory cell infiltrate f) Scattered endomysial CD8 ⁺ T-cells infiltrate that does not clearly surround or invade muscle fibers g) Many necrotic muscle fibers as the predominant abnormal histological feature. Inflammatory cells are sparse or only slight perivascular; perimysial infiltrate is not evident. MAC deposition on small blood vessels or pipestem capillaries may be seen, but tubuloreticular inclusions in endothelial cells are uncommon or not evident. h) Rimmed vacuoles, ragged red fibers, cytochrome oxidase-negative fibers that would suggest IBM i) MAC deposition on the sarcolemma of nonnecrotic fibers and other indications of muscular dystrophies with immunopathology

Modified from Ref. [2].

^a Clinical features of IBM is reviewed in Ref. [264].

Creatine Kinase (CK) level is the most sensitive but not specific indicator. The CK level does not normally correlate with the severity of the symptoms among different patients, but can reflect changes in disease activity within an individual patient. Elevated CK levels range from 5 to 50-fold above normal in PM. 70–80% of DM patients will have up to 50-fold levels while the rest will have normal CK levels [196]. CK levels can be extremely high and reach 100-fold of normal in NAM, whereas they are often normal or only mildly elevated in sIBM patients [197]. Other identified elevated muscle enzymes include lactate dehydrogenase (LDH), aspartate aminotransferase (AST) and alanine aminotransferase (ALT), which are also markers of liver injury, and aldolase, which is a

marker of muscle cell degeneration or cell membrane damage. Other serum inflammatory biomarkers such as Erythrocyte Sedimentation Rate (ESR) and C-reactive protein may also be increased during the active phase [198]. Elevated Interleukin-1RA (IL-1RA) is considered to be a diagnostic clue in PM and DM, and can be found in most patients even in the absence of CK elevation [199].

6.2.2. Antibodies

Autoantibodies associated with IIM are subdivided into myositis specific autoantibodies (MSA) and myositis associated autoantibodies (MAA), as shown in Table 4. MSAs are found in approximately 50–60% of patients with IIM while MAAs are also found in other autoimmune diseases [200]. As is the case in other autoimmune diseases, the pathogenic role of the antibodies in PM and DM is unclear, although some appear to be specific for distinct clinical phenotypes and HLA-DR genotypes. The MSAs might be divided into three broad groups (1) anti-tRNA synthetases, (2) anti-signal recognition particle (anti-SRP), and (3) other antibodies against cytoplasmic or nuclear components involved in the regulation of protein synthesis and translocation, gene transcription, and viral recognition including anti-Mi-2 anti-PM-Scl and anti-CADM-140 [201,202].

6.3. Muscle imaging

6.3.1. Magnetic Resonance Imaging

MRI is a very useful imaging tool of choice for both assessment of disease activity and selection of the biopsy site, providing a detailed anatomic view of the extent of muscle involvement [203]. Muscle necrosis, degeneration, and inflammation can be detected by MRI and are characterized by increased signal intensity on short tau inversion recovery (STIR) [204]. T1-weighted images are useful for detecting muscle damage and for loss of volume and fatty replacement whereas T2-weighted images are useful for distinguishing the fatty infiltration and edema seen in active muscle inflammation. The latter correlates with disease activity [205–208]. It is recommended by new diagnostic criteria that MRI can be used to evaluate JDM in order to avoid electromyography (EMG) or muscle biopsy [209]. Although the edema in skeletal muscles on MRI is not specific for myositis, it is more commonly seen in myositis in comparison to non-inflammatory myopathies.

6.3.2. Muscular ultrasound

Ultrasound, specifically doppler sonography, contrast-enhanced ultrasound, and sonoelastography may be also be used to differentiate between normal and pathologic muscle [210], but its sensitivity and negative predictive value for diagnosis remain low compared to MRI. Acute muscular inflammation is characterized by normal or increased size, low echogenicity, and elevated perfusion of affected muscles, whereas in the chronic disease stage, muscle size and perfusion are reduced and echogenicity is increased. Moreover, being widely available and cheap, muscular ultrasound is a useful tool in the follow-up of muscle lesions especially when MRI is not available, and it can reveal complications such as fibrosis, cystic hematomas, or myositis ossificans [211].

6.4. EMG

The characteristic EMG features of myositis patients are: (1) increased insertional and spontaneous activity with fibrillation potentials, positive sharp waves, and occasionally pseudomyotonic or complex repetitive discharges, (2) polyphasic motor unit action potentials (MUAPs) of short duration and low amplitude, (3) early recruiting MUAPs [107]. Although it is nonspecific, abnormalities may be observed in 70–90% of patients. The additional value of EMG includes identifying the highest yield biopsy sites and assessing response to therapy.

Table 4
Autoantibodies in PM/DM.

Antibodies	Target antigen and function	Clinical association	References	
Myositis-specific autoantibodies (MSAs)	Anti-Mi-2	DNA helicase/Transcription regulation (component of the NuRD complex)	15–30% (DM, associated with a favorable prognosis and suggestions of environmental trigger in adult DM)	[265–267]
	Anti-CADM-140 (MDA5)	MDA5/Innate immune response against viruses	20% (DM, associated with ADM and aggressive ILD)	[268,269]
	Anti-SAE	SUMO-1 Activating Enzyme 1/Posttranslational modifications	3–8% (adult DM, associated with rash)	[162,270]
	Anti-p155/140	TIF1- γ /Nuclear transcription	20–30% (associated with malignancy in adult and calcinosis in JDM)	[271–273]
	Anti-MJ (NXP-2)	Nuclear matrix protein/p53-mediated cell senescence	2% (adult DM, associated with malignancy)	
	Anti-(HMGCR)	HMG-CoA reductase/Cholesterol biosynthesis	7% (adult PM, associated with poor therapy response in JDM)	[274,275]
	Anti-SRP	SRP/Protein translocation across the endoplasmic reticulum	4–6% (adult PM, associated with acute and severe proximal weakness, dilated cardiomyopathy, ILD, and steroid resistance)	[86,276]
	Anti-ARS	aminoacyl-tRNA synthetases/Protein synthesis	30–40% Predictors of antisynthetase syndrome	[277,278]
	Anti-Jo1	Histidyl	20–30%	
	Anti-PL-7	Threonyl	2–5%	
Anti-PL-12	Alanyl	2–5%		
Anti-EJ	Glycyl	2–5%		
Myositis-associated autoantibodies (MAAs)	Anti-KL6	Mucin-like glycoprotein (on alveoli or bronchial epithelial cells)	Patients with ILD	[3]
	Anti-Ro/SSA	Ro52 and RO60	9–19% (common in Sjögren syndrome)	[162]
	Anti-U1RNP	U1-small nuclear ribonucleic proteins/pre-messenger RNA processing	3–8% (common in overlap condition, good prognosis)	[278,279]
	Anti PM/Scl	75-kDa and 100-kDa subunits of the nucleolar exosome complex	15% (DM, associated with scleroderma in PM)	[280,281]
	Anti-Ku	70- and 80-kDa Ku heterodimers	1–3% (common in overlap syndrome, require of high-dose corticosteroids)	[282]

6.5. Muscle biopsy

Although the pivotal role that muscle biopsy plays in the diagnosis of IIM is agreed upon by many investigators, it was not until 1984 that histopathological features were clearly established by Arahata and Engel [212]. Although very specific and important, muscle biopsy is not regarded as obligatory for diagnosis when typical features such as skin changes or specific autoantibodies are present and are consistent with clinical manifestations. Muscle histology allows distinguishing 4 main subtypes of IIM on the basis of distinct immunopathologic features: DM, PM, s-IBM, and NAM.

In PM, perivascular inflammation is most typically concentrated in multiple foci within the endomysium. CD8⁺ T cells invading healthy-appearing, nonnecrotic muscle fibers expressing MHC class I antigen are typically involving the fascicles. In DM, histopathology typically shows perivascular inflammation which is most prominently located in the interfascicular septae or the periphery of the fascicles. The muscle fibers undergo necrosis and phagocytosis, leading to hypoperfusion and perifascicular atrophy, which is characterized by layers of atrophic fibers at the periphery of the fascicles with perivascular and interfascicular infiltrates. Capillary deposition of the complement C5b-9 MAC with the presence of endothelial tubuloreticular inclusions and microinfarcts can also be detected [64,107,122,213,214]. In contrast, vacuolated muscle fibers, degeneration/regeneration areas, necrotized/phagocytized fibers, β -pleated-sheet amyloid inclusions, and phosphorylated tau are typically found in biopsies of sIBM, while NAM is characterized by abundant necrotic and regenerative fibers that contrast with modest inflammation consisting of macrophage rather than T cells infiltration [6,215].

7. Treatment & management

A clinical misdiagnosis attributed to cursory examinations or erroneous interpretation of the biopsy usually leads to unnecessary, inappropriate or delayed therapies. For example, dermatomyositis responds better to conventional treatment than polymyositis, and some cases of

“polymyositis unresponsive to therapies” should be suspected as being sIBM, NAM or other myopathies [216]. Currently, the primary goal of therapy should be an objective increase in strength and daily activities, as well as an improvement in systemic manifestations. Decreased serum muscle enzymes may be observed after treatment in the absence of an improvement in muscle strength, so called “chemical improvement”. Unfortunately, some clinicians may fall into the habit of “chasing” or “treating” the CK concentration instead of the muscle weakness, ignoring the fact that the treatment they are using may be clinically ineffective. The main concern about drug therapy in IIMs is the lack of controlled trials and the absence of standardized outcome measures to capture meaningful changes to identify correlations between disability and quality of life [53,217,218]. When considering treatment options for patients with IIMs, great care should be taken to ensure that optimal therapies are being used which can positively impact patient quality of life.

The mainstay of therapy for DM and PM consists of immunosuppression, physical therapy, monitoring for adverse events from medications, and prevention of complications [53]. The most commonly used pharmacological therapies in clinical practice are listed in Table 5.

7.1. Immunosuppression medications

7.1.1. First-line therapy

Glucocorticoids remains the first-line therapy for IIMs with a standard oral prednisone dose of 1 mg per kilogram of body weight (high dose, up to 100 mg per day). It should be noted that this choice of drug is based on experience and not on placebo controlled trials [53,219]. A clinical trial found no difference in efficiency between pulsed high-dose oral dexamethasone and daily prednisolone as first line treatment of IIMs, although dexamethasone showed substantially fewer side-effects [220]. In patients with rapidly worsening disease, intravenous methylprednisolone administration is preferable at a dose of 1000 mg per day for 3–5 days before starting treatment with oral glucocorticoids. Depending on efficacy and side effects, the daily dose is slowly reduced or switched to

Table 5
Therapeutic regimens for PM/DM.

Patients condition	Agents	Typical dose	Side effect and toxicities
New-onset disease	Prednisone	1 mg per kilogram, up to 100 mg per day for 4–6 weeks (orally); taper to alternate days	Increased susceptibility to infection, osteoporosis, hypertension, diabetes, weight gain, steroid-induced myopathy, mood changes, skin fragility, avascular necrosis, glaucoma
Late or severe disease onset, rapidly worsening	Methylprednisolone	1000 mg per day for 3–5 days (IV), then switch to oral regimen	
Steroid refractory	Intravenous Immunglobulin	2 g/kg total dose given over 2–5 days (1 g/kg/day over 2 days or 0.4 g/kg/day over 5 days), repeated monthly for at least 3 months, intravenously	Flu-like symptoms, myalgias, fever, headache, fluid overload, rash, aseptic meningitis, caution in patients with cardiac conditions, risk of renal failure or thrombosis
Steroid-sparing regents	Azathioprine	2–3 mg/kg/day (orally)	Fever, abdominal pain, nausea, vomiting, anorexia; combination with allopurinol increased myelosuppression and hepatotoxicity; thiopurine methyltransferase activity may need to be checked in certain patients prior to initiation of medication or in those with poor reaction to treatment
	Methotrexate	15–25 mg/week (orally), should be given with folate 1 mg/day (IV);	Hepatotoxicity, ILD (caution in those with anti-Jo-1 antibody positivity), interstitial pneumonitis, myelosuppression, renal toxicity, alopecia, stomatitis, oncogenicity, teratogenicity
	Mycophenolate mofetil	1–1.5 mg twice daily, 500 mg twice daily in renal impairment (orally)	Diarrhea, myelosuppression, tremors, hypertension
	Tacrolimus	2–3 mg twice daily (orally)	Increased susceptibility to infection, lymphoma, alopecia, skin erythema, pruritis, constipation, diarrhea, nausea, anemia, leukocytosis, thrombocytopenia, headache, hypertension, paresthesia, tremor, renal failure
	Cyclophosphamide	1–2 mg/kg/day (orally), or 1 g/m ² monthly (intramuscularly)	Gastrointestinal upset, alopecia, risk of malignancy, hemorrhagic cystitis, teratogenicity, sterility, increased risk of infection
	Cyclosporine	3–4 mg/kg/day (orally) in two divided doses, then increase up to 6 mg/kg/day	Hypertension, renal failure, gingival hyperplasia, Gastrointestinal upset, hypertrichosis, oncogenicity, tremor, risk of infection
Steroid refractory	Rituximab	1 g (IV), repeat in 2 weeks; then subsequent doses 6–9 months after second dose	Fever, nausea, infection susceptibility, rare infusion reactions, rare progressive multifocal leukoencephalopathy
No objective improvement	If diagnosis is reconfirmed, recommend participation in a research trial (candidates include eculizumab, alemtuzumab, tocilizumab (anti-IL-6), anti-IL-17, and anti-IL-1 β and so on)		

Modified from Refs. [4,5].

an alternate-day program slowly over several weeks, until the lowest possible dose that controls the disease is reached.

Some clinicians claim that other steroid sparing agents also should be used [53,221]. Indeed, if the patient is unresponsive to steroids, and objective signs of increased strength and ability to perform activities in daily living in months are not observed, tapering should be accelerated so that treatment with an alternate agent can be initiated. Importantly, patients with other co-morbidities such as hypertension, diabetes, osteoporosis, and obesity, which will be exacerbated by corticosteroid use, should be started on a second-line agent early and subsequently have their prednisone tapered to a minimally effective dose. Adverse events need to be carefully monitored while on chronic high dose corticosteroids. Treatment for bone and liver complications and gastric mucosa protection should be considered to help minimize the adverse side effects of steroids.

7.1.2. Other immunosuppressive medications

Studies have demonstrated the efficacy of glucocorticoids in improving muscle strength and achieving prolonged treatment-free remissions [222–224]. However, there is still a high percentage of patients with IIM who fail to respond completely to glucocorticoids alone [225]. Intravenous immune globulin therapy has been shown to be effective in severe and rapidly progressive or refractory PM and DM in several clinical trials [219,226,227]. Azathioprine (AZA), a derivative of 6-mercaptopurine, is administered orally at a dose of 2–3 mg/kg daily and is usually effective after 4–8 months, according to a small number of case series or case studies [228–231]. Methotrexate (MTX), an antagonist of folate metabolism, has been reported to be used in steroid-refractory PM first in 1968 [232] and then in other studies [230,233,234]. MTX is often given orally, starting at 3 doses of 2.5 mg every 12 h weekly for the first 3

weeks, with a gradual increase by 2.5 mg per week up to a total of 15–20 mg weekly [235]. Mycophenolate mofetil (MMF), a morpholinoethyl ester of mycophenolic acid that blocks de novo purine synthesis, is administered orally at a dose of up to 3 g per day, but it can take up to 2–3 months to see the benefits of treatment. MMF is well tolerated although it may be expensive [218]. Cyclosporine, which affects T-cell-mediated immunity by inhibiting transcription of the *IL2* gene, is given at doses of 150 mg twice a day (not more than 5 mg/kg per day). Cyclosporine is useful in newly diagnosed PM and DM although it has significant side effects [127]. Each of these immunomodulating medicines exhibits efficacy at different stages of disease or in different complications and are associated with significant side effects, including bone marrow suppression and infections, so they need to be used under careful considering and monitoring.

7.2. New biological therapies

In refractory PM/DM, biological agents which have been approved for the treatment of other immune diseases may be considered as experimental treatment options.

7.2.1. Rituximab

Rituximab, an anti-CD20 antibody, causes depletion of the circulating B cells for at least 6 months [236]. It is reported that Rituximab administration can be effective for some patients with PM and DM who are resistant to other therapies [237–241]. It has also been reported that rituximab may be helpful in PM/DM-related ILD [242]. It follows that biologics directed against B cells should theoretically be helpful in treating autoimmune diseases with MSAs, and indeed there is research demonstrating that Rituximab is more effective in the presence of an

anti-synthetase, anti-Mi-2, or other autoantibody, with a shorter time to improvement, compared to the autoantibody negative subset [243].

7.2.2. Tumor necrosis factor inhibitors

TNF inhibitors (Infliximab, Adalimumab and Etanercept) have been approved to treat autoimmune disorders including rheumatoid arthritis (RA), juvenile idiopathic arthritis and psoriatic arthritis. Anecdotal reports suggest that TNF inhibitors can be helpful in the treatment of a subset of patients with PM or DM [244,245], although other reports show no such benefit and have even been reported to induce flares. It should be noted that treatment of autoimmune diseases with TNF inhibitors has been associated with the development of new autoimmune diseases, and there are numerous case reports of IIMs induced by anti-TNF agents as well [246,247]. In general, anti-TNF therapy is not routinely used in myositis and further studies are needed to obtain more data on efficacy and safety.

7.2.3. Newer agents

Other agents targeting cells or molecules that are involved in autoimmune diseases have been used in the treatment of PM and DM, and are described predominantly as case reports only. These regents and their mechanisms include Alemtuzumab, Efalizumab, tacrolimus and rapamycin, which target T-cell intracellular signaling pathways. Alemtuzumab is a humanized anti-CD52 monoclonal antibody, which interferes with T-cell signaling and has shown to be effective in improving muscle strength in a case of refractory PM [248] and has achieved comparable immune ablation compared to a pre-hematopoietic stem cell transplantation (HSCT) conditioning regimen in juvenile PM case [249]. Other strategies include targeting B-cell growth factors by inhibiting B-cell activating factor (BAFF) and a related ligand, APRIL, targeting complement with Eculizumab, a monoclonal antibody against C5, and targeting cellular adhesion and T-cell migration with Natalizumab, a monoclonal antibody directed against the $\alpha 4\beta 1$ integrin VLA4 [53,127, 218,250]. The involvement of activated complement, T cells, B cells, cytokines, adhesion molecules, and transmigration molecules in the pathogenesis of PM and DM justifies the use of several new biologic agents that target specific molecules, but there is still a need for further clinical studies to evaluate efficacy and safety.

7.3. Nonpharmacological treatment

Nonpharmacological therapies must also be integrated into the care of patients with myositis. Exercise and physical therapy are important components of treatment for patients with IIM. These therapies are safe and may improve aerobic capacity and muscle strength [251–253]. In addition, diet and lifestyle changes and modifications can have a positive effect. The role of dietary supplementation hitherto in the treatment of IIMs is limited. However it has been shown in a clinical trial that patients with DM or PM improved significantly with oral creatine supplements in conjunction with exercise as compared with exercise alone, based on functional performance times and better performance in laboratory examinations [254]. In addition, dietary modifications such as a low-fat, low-carbohydrate, and low-salt diet need to be undertaken by patients receiving corticosteroids to minimize effects of weight gain, hypertension, hyperglycemia and edema. Calcium (1 g/d) and vitamin D (400–800 IU/d) supplementation to decrease the risk of osteopenia is also recommended [53]. Thirdly, assistive devices, home modifications, precautions for aspiration in patients with severe dysphagia and emotional support may be helpful for those patients who suffer from more rapid disease progression and weakness [53].

8. Conclusion

Unmet needs or challenges in PM and DM include better diagnostic algorithms and more effective and safe treatment modalities. More specifically, how to improve diagnostic precision to avoid misdiagnosis or

delayed treatment due to the uncertainty of excluded diagnosis is critical. In addition, a better understanding of the disease pathogenesis and the progression of disease may help to guide future treatment and research strategies. Great strides have been made in advancing the diagnosis and treatment of other autoimmune diseases including rheumatoid arthritis, systemic lupus erythematosus and Crohn's disease, which regrettably have not yet been fully seen in patients with IIMs. The incidence and prevalence of PM/DM is fairly low, and as a result, the amount of basic and clinical research performed is much less than other autoimmune diseases, and unified and authoritative diagnosis criteria and management outlines are not frequently updated. In conclusion, a great deal of international cooperation still needs to be realized in order to improve the lives of patients suffering from IIMs.

9. Contributions

Shu-Han Yang and Christopher Chang wrote the manuscript. Shu-Han Yang made the tables. Christopher Chang and Zhe-Xiong Lian edited the manuscript.

Declaration of competing interest

The authors have declared that no conflict of interest exists.

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References

- [1] A. Bohan, J.B. Peter, Polymyositis and dermatomyositis (first of two parts), *N. Engl. J. Med.* 292 (7) (1975) 344–347.
- [2] J.E. Hoogendijk, et al., 119th ENMC international workshop: trial design in adult idiopathic inflammatory myopathies, with the exception of inclusion body myositis, 10–12 October 2003, Naarden, The Netherlands, *Neuromuscul. Disord.* 14 (5) (2004) 337–345.
- [3] M.C. Dalakas, R. Hohlfield, Polymyositis and dermatomyositis, *Lancet* 362 (9388) (2003) 971–982.
- [4] C. Castro, M. Gourley, Diagnosis and treatment of inflammatory myopathy: issues and management, *Ther Adv Musculoskelet Dis* 4 (2) (2012) 111–120.
- [5] M.C. Dalakas, Inflammatory muscle diseases, *N. Engl. J. Med.* 372 (18) (2015) 1734–1747.
- [6] M.C. Dalakas, Review: an update on inflammatory and autoimmune myopathies, *Neuropathol. Appl. Neurobiol.* 37 (3) (2011) 226–242.
- [7] M. Jakubaszek, B. Kwiatkowska, M. Maslinska, Polymyositis and dermatomyositis as a risk of developing cancer, *Reumatologia* 53 (2) (2015) 101–105.
- [8] A. Ceribelli, et al., The immune response and the pathogenesis of idiopathic inflammatory myositis: a critical review, *Clin. Rev. Allergy Immunol.* 52 (1) (2017) 58–70.
- [9] E.A. Shamim, F.W. Miller, Familial autoimmunity and the idiopathic inflammatory myopathies, *Curr. Rheumatol. Rep.* 2 (3) (2000) 201–211.
- [10] R.J. Wedgwood, C.D. Cook, J. Cohen, Dermatomyositis; report of 26 cases in children with a discussion of endocrine therapy in 13, *Pediatrics* 12 (4) (1953) 447–466.
- [11] H.B. CHRISTIANSON, L.A. BRUNSTING, H.O. PERRY, Dermatomyositis: unusual features, complications, and treatment, *JAMA Dermatol.* 74 (6) (1956) 581–589.
- [12] S. Rothwell, et al., Entering a new phase of immunogenetics in the idiopathic inflammatory myopathies, *Curr. Opin. Rheumatol.* 25 (6) (2013) 735–741.
- [13] S. Rothwell, J.A. Lamb, H. Chinoy, New developments in genetics of myositis, *Curr. Opin. Rheumatol.* 28 (6) (2016) 651–656.
- [14] H. Chinoy, et al., Recent advances in the immunogenetics of idiopathic inflammatory myopathy, *Arthritis Res. Ther.* 13 (3) (2011) 216.
- [15] H. Chinoy, et al., An update on the immunogenetics of idiopathic inflammatory myopathies: major histocompatibility complex and beyond, *Curr. Opin. Rheumatol.* 21 (6) (2009) 588–593.
- [16] F.W. Miller, et al., Genome-wide association study identifies HLA 8.1 ancestral haplotype alleles as major genetic risk factors for myositis phenotypes, *Genes Immun.* 16 (7) (2015) 470–480.
- [17] S. Rothwell, et al., Dense genotyping of immune-related loci in idiopathic inflammatory myopathies confirms HLA alleles as the strongest genetic risk factor and suggests different genetic background for major clinical subgroups, *Ann. Rheum. Dis.* 75 (8) (2016) 1558–1566.

- [18] T.P. O'Hanlon, et al., HLA polymorphisms in African Americans with idiopathic inflammatory myopathy: allelic profiles distinguish patients with different clinical phenotypes and myositis autoantibodies, *Arthritis Rheum.* 54 (11) (2006) 3670–3681.
- [19] F.C. Arnett, et al., Interrelationship of major histocompatibility complex class II alleles and autoantibodies in four ethnic groups with various forms of myositis, *Arthritis Rheum.* 39 (9) (1996) 1507–1518.
- [20] T.P. O'Hanlon, et al., Immunogenetic Risk and Protective Factors for the Idiopathic Inflammatory Myopathies: Distinct HLA-A, -B, -Cw, -DRB1 and -DQA1 Allelic Profiles and Motifs Define Clinicopathologic Groups in Caucasians, *Med. vol.* 84 (6) (2005) 338–349.
- [21] L.G. Rider, et al., Genetic risk and protective factors for idiopathic inflammatory myopathy in Koreans and American whites: a tale of two loci, *Arthritis Rheum.* 42 (6) (1999) 1285–1290.
- [22] V.P. Werth, et al., Associations of tumor necrosis factor α and HLA polymorphisms with adult dermatomyositis: implications for a unique Pathogenesis1, *J. Investig. Dermatol.* 119 (3) (2002) 617–620.
- [23] H. Chinoy, et al., Tumor necrosis factor-alpha single nucleotide polymorphisms are not independent of HLA class I in UK Caucasians with adult onset idiopathic inflammatory myopathies, *Rheumatology* 46 (9) (2007) 1411–1416.
- [24] H. Chinoy, et al., Genetic association study of NF-kappaB genes in UK Caucasian adult and juvenile onset idiopathic inflammatory myopathy, *Rheumatology* 51 (5) (2012) 794–799.
- [25] F. Sandoval-Garcia, et al., The ACTN3 R577X polymorphism is associated with inflammatory myopathies in a Mexican population, *Scand. J. Rheumatol.* 41 (5) (2012) 396–400.
- [26] T. Sugiura, et al., Association between a C8orf13-BLK polymorphism and polymyositis/dermatomyositis in the Japanese population: an additive effect with STAT4 on disease susceptibility, *PLoS One* 9 (3) (2014), e90019.
- [27] T. Sugiura, et al., Positive association between $\langle em \rangle$ STAT4 $\langle /em \rangle$ polymorphisms and polymyositis/dermatomyositis in a Japanese population, *Ann. Rheum. Dis.* 71 (10) (2012) 1646.
- [28] S. Chen, et al., Genetic association study of TNFAIP3, IFIH1, IRF5 polymorphisms with polymyositis/dermatomyositis in Chinese Han population, *PLoS One* 9 (10) (2014), e110044.
- [29] Q. Wang, et al., Positive association of genetic variations in the phospholipase C-like 1 gene with dermatomyositis in Chinese Han, *Immunol. Res.* 64 (1) (2016) 204–212.
- [30] H. Chinoy, et al., Interferon-gamma and interleukin-4 gene polymorphisms in Caucasian idiopathic inflammatory myopathy patients in UK, *Ann. Rheum. Dis.* 66 (7) (2007) 970.
- [31] T. Gono, et al., Interferon-induced helicase (IFIH1) polymorphism with systemic lupus erythematosus and dermatomyositis/polymyositis, *Mod. Rheumatol.* 20 (5) (2010) 466–470.
- [32] E. Generali, et al., Lessons learned from twins in autoimmune and chronic inflammatory diseases, *J. Autoimmun.* 83 (2017) 51–61.
- [33] E.C. Rosser, C. Mauri, A clinical update on the significance of the gut microbiota in systemic autoimmunity, *J. Autoimmun.* 74 (2016) 85–93.
- [34] K. Nagaraju, P.H. Plotz, Animal models of myositis, *Rheum. Dis. Clin. N. Am.* 28 (4) (2002) 917–933.
- [35] G. Mamyrova, et al., Environmental factors associated with disease flare in juvenile and adult dermatomyositis, *Rheumatology* 56 (8) (2017) 1342–1347.
- [36] S. Okada, et al., Global surface ultraviolet radiation intensity may modulate the clinical and immunologic expression of autoimmune muscle disease, *Arthritis Rheum.* 48 (8) (2003) 2285–2293.
- [37] L.A. Love, et al., Ultraviolet radiation intensity predicts the relative distribution of dermatomyositis and anti-Mi-2 autoantibodies in women, *Arthritis Rheum.* 60 (8) (2009) 2499–2504.
- [38] M. Shah, et al., Brief report: ultraviolet radiation exposure is associated with clinical and autoantibody phenotypes in juvenile myositis, *Arthritis Rheum.* 65 (7) (2013) 1934–1941.
- [39] I. Marie, et al., Opportunistic infections in polymyositis and dermatomyositis, *Arthritis Rheum.* 53 (2) (2005) 155–165.
- [40] N.E. Bowles, et al., Dermatomyositis, polymyositis, and Cocksackie-B-virus infection, *Lancet* 1 (8540) (1987) 1004–1007.
- [41] M.B. Carroll, R. Holmes, Dermatomyositis and HIV infection: case report and review of the literature, *Rheumatol. Int.* 31 (5) (2011) 673–679.
- [42] Y.F. Fang, et al., Malignancy in dermatomyositis and polymyositis: analysis of 192 patients, *Clin. Rheumatol.* 35 (8) (2016) 1977–1984.
- [43] S.Y. Yang, et al., Dermatomyositis associated with hepatitis B virus-related hepatocellular carcinoma, *Korean J. Intern. Med.* 29 (2) (2014) 231–235.
- [44] J.W. Chou, et al., Dermatomyositis induced by hepatitis B virus-related hepatocellular carcinoma: a case report and review of the literature, *Intern. Med.* 56 (14) (2017) 1831–1837.
- [45] A. Schiffenbauer, et al., The effect of cigarette smoking on the clinical and serological phenotypes of polymyositis and dermatomyositis, *Semin. Arthritis Rheum.* 48 (3) (2018) 504–512.
- [46] J.M. Evans, et al., Beyond the MHC: a canine model of dermatomyositis shows a complex pattern of genetic risk involving novel loci, *PLoS Genet.* 13 (2) (2017), e1006604.
- [47] Y. Katsumata, D.P. Ascherman, Animal models in myositis, *Curr. Opin. Rheumatol.* 20 (6) (2008) 681–685.
- [48] N.L. Rosenberg, Experimental models of inflammatory myopathies, *Bailliere. Clin. Neurol.* 2 (3) (1993) 693–715.
- [49] A.H. Weller, et al., Spontaneous myopathy in the SJL/J mouse: pathology and strength loss, *Muscle Nerve* 20 (1) (1997) 72–82.
- [50] N.L. Rosenberg, S.P. Ringel, B.L. Kotzin, Experimental autoimmune myositis in SJL/J mice, *Clin. Exp. Immunol.* 68 (1) (1987) 117–129.
- [51] M.C. Dalakas, Mechanisms of disease: signaling pathways and immunobiology of inflammatory myopathies, *Nat. Clin. Pract. Rheumatol.* 2 (4) (2006) 219–227.
- [52] M.C. Dalakas, Pathophysiology of inflammatory and autoimmune myopathies, *Presse Med.* 40 (4 Pt 2) (2011) e237–e247.
- [53] M.C. Dalakas, Immunotherapy of myositis: issues, concerns and future prospects, *Nat. Rev. Rheumatol.* 6 (3) (2010) 129–137.
- [54] J. Schmidt, M.C. Dalakas, Pathomechanisms of inflammatory myopathies: recent advances and implications for diagnosis and therapies, *Expert Opin. Med. Diagn.* 4 (3) (2010) 241–250.
- [55] H. Wiendl, R. Hohlfeld, B.C. Kieseier, Immunobiology of muscle: advances in understanding an immunological microenvironment, *Trends Immunol.* 26 (7) (2005) 373–380.
- [56] A. Bender, et al., T cell receptor repertoire in polymyositis: clonal expansion of autoaggressive CD8+ T cells, *J. Exp. Med.* 181 (5) (1995) 1863–1868.
- [57] M. Hofbauer, et al., Clonal tracking of autoaggressive T cells in polymyositis by combining laser microdissection, single-cell PCR, and CDR3-spectratype analysis, *Proc. Natl. Acad. Sci. U. S. A.* 100 (7) (2003) 4090–4095.
- [58] J. Schmidt, et al., Upregulated inducible co-stimulator (ICOS) and ICOS-ligand in inclusion body myositis muscle: significance for CD8+ T cell cytotoxicity, *Brain* 127 (Pt 5) (2004) 1182–1190.
- [59] B. De Paepe, K.K. Creus, J.L. De Bleecker, Role of cytokines and chemokines in idiopathic inflammatory myopathies, *Curr. Opin. Rheumatol.* 21 (6) (2009) 610–616.
- [60] H. Wiendl, et al., Muscle fibres and cultured muscle cells express the B7.1/2-related inducible co-stimulatory molecule, ICOSL: implications for the pathogenesis of inflammatory myopathies, *Brain* 126 (Pt 5) (2003) 1026–1035.
- [61] N. Goebels, et al., Differential expression of perforin in muscle-infiltrating T cells in polymyositis and dermatomyositis, *J. Clin. Investig.* 97 (12) (1996) 2905–2910.
- [62] J.T. Kissel, J.R. Mendell, K.W. Rammohan, Microvascular deposition of complement membrane attack complex in dermatomyositis, *N. Engl. J. Med.* 314 (6) (1986) 329–334.
- [63] A.M. Emslie-Smith, A.G. Engel, Microvascular changes in early and advanced dermatomyositis: a quantitative study, *Ann. Neurol.* 27 (4) (1990) 343–356.
- [64] A. Pestronk, Acquired immune and inflammatory myopathies: pathologic classification, *Curr. Opin. Rheumatol.* 23 (6) (2011) 595–604.
- [65] R. Lahoria, D. Selcen, A.G. Engel, Microvascular alterations and the role of complement in dermatomyositis, *Brain* 139 (Pt 7) (2016) 1891–1903.
- [66] A.E. Fasth, et al., T cell infiltrates in the muscles of patients with dermatomyositis and polymyositis are dominated by CD28null T cells, *J. Immunol.* 183 (7) (2009) 4792–4799.
- [67] J.M. Pandya, et al., Effects of conventional immunosuppressive treatment on CD244+ (CD28null) and FOXP3+ T cells in the inflamed muscle of patients with polymyositis and dermatomyositis, *Arthritis Res. Ther.* 18 (2016) 80.
- [68] R. Hohlfeld, A.G. Engel, Coculture with autologous myotubes of cytotoxic T cells isolated from muscle in inflammatory myopathies, *Ann. Neurol.* 29 (5) (1991) 498–507.
- [69] Y. Shimojima, et al., T-cell receptor-mediated characteristic signaling pathway of peripheral blood T cells in dermatomyositis and polymyositis, *Autoimmunity* 50 (8) (2017) 481–490.
- [70] J. Nishio, et al., Clonal biases of peripheral CD8 T cell repertoire directly reflect local inflammation in polymyositis, *J. Immunol.* 167 (7) (2001) 4051–4058.
- [71] O. Benveniste, et al., Severe perturbations of the blood T cell repertoire in polymyositis, but not dermatomyositis patients, *J. Immunol.* 167 (6) (2001) 3521–3529.
- [72] T.P. O'Hanlon, et al., Predominant TCR-alpha beta variable and joining gene expression by muscle-infiltrating lymphocytes in the idiopathic inflammatory myopathies, *J. Immunol.* 152 (5) (1994) 2569–2576.
- [73] A. Waschbisch, et al., FOXP3+ regulatory cells in idiopathic inflammatory myopathies, *J. Neuroimmunol.* 225 (1–2) (2010) 137–142.
- [74] Y. Vercoulen, et al., Increased presence of FOXP3+ regulatory T cells in inflamed muscle of patients with active juvenile dermatomyositis compared to peripheral blood, *PLoS One* 9 (8) (2014), e105353.
- [75] S.A. Greenberg, et al., Plasma cells in muscle in inclusion body myositis and polymyositis, *Neurology* 65 (11) (2005) 1782–1787.
- [76] D.X. Wang, et al., Clinical significance of peripheral blood lymphocyte subsets in patients with polymyositis and dermatomyositis, *Clin. Rheumatol.* 31 (12) (2012) 1691–1697.
- [77] O. Kryštůfková, et al., Increased serum levels of B cell activating factor (BAFF) in subsets of patients with idiopathic inflammatory myopathies, *Ann. Rheum. Dis.* 68 (6) (2009) 836.
- [78] A. Baek, et al., The expression of BAFF in the muscles of patients with dermatomyositis, *J. Neuroimmunol.* 249 (1) (2012) 96–100.
- [79] Q.-L. Peng, et al., B-cell activating factor as a serological biomarker for polymyositis and dermatomyositis, *Biomark. Med.* 8 (3) (2014) 395–403.
- [80] O. Kryštůfková, et al., Expression of BAFF receptors in muscle tissue of myositis patients with anti-Jo-1 or anti-Ro52/anti-Ro60 autoantibodies, *Arthritis Res. Ther.* 16 (5) (2014) 454.
- [81] A. Tourmadre, V. Lenief, P. Miossec, Expression of Toll-like receptor 3 and Toll-like receptor 7 in muscle is characteristic of inflammatory myopathy and is differentially regulated by Th1 and Th17 cytokines, *Arthritis Rheum.* 62 (7) (2010) 2144–2151.
- [82] A. Brunn, et al., Toll-like receptors promote inflammation in idiopathic inflammatory myopathies, *J. Neuropathol. Exp. Neurol.* 71 (10) (2012) 855–867.

- [83] Y. Kikuchi, et al., Difference in B cell activation between dermatomyositis and polymyositis: analysis of the expression of RP105 on peripheral blood B cells, *Ann. Rheum. Dis.* 60 (12) (2001) 1137.
- [84] E.M. Bradshaw, et al., A local antigen-driven humoral response is present in the inflammatory myopathies, *J. Immunol.* 178 (1) (2007) 547–556.
- [85] D. McIntyre, et al., The V(H) repertoire and clonal diversification of B cells in inflammatory myopathies, *Eur. J. Immunol.* 44 (2) (2014) 585–596.
- [86] R. Valiyil, et al., Rituximab therapy for myopathy associated with anti-signal recognition particle antibodies: a case series, *Arthritis Care Res.* 62 (9) (2010) 1328–1334.
- [87] T. Hornung, J. Wenzel, Innate immune-response mechanisms in dermatomyositis: an update on pathogenesis, diagnosis and treatment, *Drugs* 74 (9) (2014) 981–998.
- [88] J. Wenzel, et al., Type I interferon-associated skin recruitment of CXCR3+ lymphocytes in dermatomyositis, *Clin. Exp. Dermatol.* 31 (4) (2006) 576–582.
- [89] J. Wenzel, et al., Evidence for a role of type I interferons in the pathogenesis of dermatomyositis, *Br. J. Dermatol.* 153 (2) (2005) 462–463.
- [90] R.J. Walsh, et al., Type I interferon-inducible gene expression in blood is present and reflects disease activity in dermatomyositis and polymyositis, *Arthritis Rheum.* 56 (11) (2007) 3784–3792.
- [91] S.A. Greenberg, et al., Interferon- α/β -mediated innate immune mechanisms in dermatomyositis, *Ann. Neurol.* 57 (5) (2005) 664–678.
- [92] G. Page, G. Chevrel, P. Miossec, Anatomic localization of immature and mature dendritic cell subsets in dermatomyositis and polymyositis: interaction with chemokines and Th1 cytokine-producing cells, *Arthritis Rheum.* 50 (1) (2004) 199–208.
- [93] M. Yokota, et al., Roles of mast cells in the pathogenesis of inflammatory myopathy, *Arthritis Res. Ther.* 16 (2) (2014) R72.
- [94] S. Zhang, et al., Enhanced formation and impaired degradation of neutrophil extracellular traps in dermatomyositis and polymyositis: a potential contributor to interstitial lung disease complications, *Clin. Exp. Immunol.* 177 (1) (2014) 134–141.
- [95] K.M. Rostasy, et al., Monocyte/macrophage differentiation in dermatomyositis and polymyositis, *Muscle Nerve* 30 (2) (2004) 225–230.
- [96] M. Shimizu, et al., Role of activated macrophage and inflammatory cytokines in the development of calcinosis in juvenile dermatomyositis, *Rheumatology* 53 (4) (2014) 766–767.
- [97] Q.L. Peng, et al., Elevated serum levels of soluble CD163 in polymyositis and dermatomyositis: associated with macrophage infiltration in muscle tissue, *J. Rheumatol.* 42 (6) (2015) 979–987.
- [98] D.P. Ascherman, et al., Critical requirement for professional APCs in eliciting T cell responses to novel fragments of histidyl-tRNA synthetase (Jo-1) in Jo-1 antibody-positive polymyositis, *J. Immunol.* 169 (12) (2002) 7127–7134.
- [99] G. Karpati, Y. Pouliot, S. Carpenter, Expression of immunoreactive major histocompatibility complex products in human skeletal muscles, *Ann. Neurol.* 23 (1) (1988) 64–72.
- [100] T. Sugiura, et al., Increased CD40 expression on muscle cells of polymyositis and dermatomyositis: role of CD40-CD40 ligand interaction in IL-6, IL-8, IL-15, and monocyte chemoattractant protein-1 production, *J. Immunol.* 164 (12) (2000) 6593.
- [101] D. Xiaoyu, et al., Expression of B7-homolog 1 in polymyositis, *Ann. Clin. Lab. Sci.* 41 (2) (2011) 154–160.
- [102] A. Uruha, et al., Sarcoplasmic MxA expression: a valuable marker of dermatomyositis, *Neurology* 88 (5) (2017) 493–500.
- [103] L. Behrens, et al., Cytotoxic mechanisms in inflammatory myopathies. Co-expression of Fas and protective Bcl-2 in muscle fibres and inflammatory cells, *Brain* 120 (Pt 6) (1997) 929–938.
- [104] C. Schneider, et al., MHC class I-mediated cytotoxicity does not induce apoptosis in muscle fibers nor in inflammatory T cells: studies in patients with polymyositis, dermatomyositis, and inclusion body myositis, *J. Neuropathol. Exp. Neurol.* 55 (12) (1996) 1205–1209.
- [105] K. Nagaraju, et al., The inhibition of apoptosis in myositis and in normal muscle cells, *J. Immunol.* 164 (10) (2000) 5459–5465.
- [106] M. Li, M.C. Dalakas, Expression of human IAP-like protein in skeletal muscle: a possible explanation for the rare incidence of muscle fiber apoptosis in T-cell mediated inflammatory myopathies, *J. Neuroimmunol.* 106 (1–2) (2000) 1–5.
- [107] M.C. Dalakas, Polymyositis, dermatomyositis and inclusion-body myositis, *N. Engl. J. Med.* 325 (21) (1991) 1487–1498.
- [108] R.C. Griggs, et al., Inclusion body myositis and myopathies, *Ann. Neurol.* 38 (5) (1995) 705–713.
- [109] J. Finsterer, M. Frank, E. Krexner, Steroid-responsive dropped-head-syndrome due to polymyositis, *Jt. Bone Spine* 77 (5) (2010) 485–486.
- [110] E.C. Ebert, Review article: the gastrointestinal complications of myositis, *Aliment. Pharmacol. Ther.* 31 (3) (2010) 359–365.
- [111] P. de Merieux, et al., Esophageal abnormalities and dysphagia in polymyositis and dermatomyositis, *Arthritis Rheum.* 26 (8) (1983) 961–968.
- [112] S.A. Greenberg, Inflammatory myopathies: evaluation and management, *Semin. Neurol.* 28 (2) (2008) 241–249.
- [113] T. Mozaffar, A. Pestronk, Myopathy with anti-Jo-1 antibodies: pathology in perimysium and neighbouring muscle fibres, *J. Neurol. Neurosurg. Psychiatry* 68 (4) (2000) 472–478.
- [114] A.L. Mammen, Autoimmune myopathies: autoantibodies, phenotypes and pathogenesis, *Nat. Rev. Neurol.* 7 (6) (2011) 343–354.
- [115] E. Katzap, M.L. Barilla-LaBarca, G. Marder, Antisynthetase syndrome, *Curr. Rheumatol. Rep.* 13 (3) (2011) 175–181.
- [116] A.A. Amato, R.J. Barohn, Evaluation and treatment of inflammatory myopathies, *J. Neurol. Neurosurg. Psychiatry* 80 (10) (2009) 1060–1068.
- [117] N. Chahin, A.G. Engel, Correlation of muscle biopsy, clinical course, and outcome in PM and sporadic IBM, *Neurology* 70 (6) (2008) 418–424.
- [118] A.A. Amato, R.C. Griggs, Unicorns, dragons, polymyositis, and other mythological beasts, *Neurology* 61 (3) (2003) 288–289.
- [119] W. Shi, et al., A case of Kennedy's disease misdiagnosed as polymyositis, *Scand. J. Rheumatol.* 48 (2) (2019) 168–170.
- [120] R.K. Gherardi, et al., Macrophagic myofasciitis: an emerging entity. Groupe d'Etudes et Recherche sur les Maladies Musculaires Acquisies et Dysimmunitaires (GERMMAD) de l'Association Francaise contre les Myopathies (AFM), *Lancet* 352 (9125) (1998) 347–352.
- [121] P.A. Hertzman, et al., Association of the eosinophilia-myalgia syndrome with the ingestion of tryptophan, *N. Engl. J. Med.* 322 (13) (1990) 869–873.
- [122] M.F. van der Meulen, et al., Polymyositis: an overdiagnosed entity, *Neurology* 61 (3) (2003) 316–321.
- [123] D.R. Doyle, T.L. McCurley, J.S. Sergent, Fatal polymyositis in D-penicillamine-treated rheumatoid arthritis, *Ann. Intern. Med.* 98 (3) (1983) 327–330.
- [124] M.C. Dalakas, et al., Mitochondrial myopathy caused by long-term zidovudine therapy, *N. Engl. J. Med.* 322 (16) (1990) 1098–1105.
- [125] L. Iaccarino, et al., The clinical features, diagnosis and classification of dermatomyositis, *J. Autoimmun.* 48–49 (2014) 122–127.
- [126] E.E. Bailey, D.F. Fiorentino, Amyopathic dermatomyositis: definitions, diagnosis, and management, *Curr. Rheumatol. Rep.* 16 (12) (2014) 465.
- [127] A.R. Findlay, N.A. Goyal, T. Mozaffar, An overview of polymyositis and dermatomyositis, *Muscle Nerve* 51 (5) (2015) 638–656.
- [128] A.V. Ramanan, B.M. Feldman, Clinical features and outcomes of juvenile dermatomyositis and other childhood onset myositis syndromes, *Rheum. Dis. Clin. N. Am.* 28 (4) (2002) 833–857.
- [129] L.J. McCann, et al., The Juvenile Dermatomyositis National Registry and Repository (UK and Ireland)—clinical characteristics of children recruited within the first 5 yr, *Rheumatology* 45 (10) (2006) 1255–1260.
- [130] S. Compeyrot-Lacassagne, B.M. Feldman, Inflammatory myopathies in children, *Pediatr. Clin. N. Am.* 52 (2) (2005) 493–520 (vi-vii).
- [131] A.V. Ramanan, B.M. Feldman, Clinical features and outcomes of juvenile dermatomyositis and other childhood onset myositis syndromes, *Rheum. Dis. Clin. N. Am.* 28 (4) (2002) 833–857.
- [132] S. Khan, L. Christopher-Stine, Polymyositis, dermatomyositis, and autoimmune necrotizing myopathy: clinical features, *Rheum. Dis. Clin. N. Am.* 37 (2) (2011) 143–158 (v).
- [133] I.N. Targoff, et al., A novel autoantibody to a 155-kd protein is associated with dermatomyositis, *Arthritis Rheum.* 54 (11) (2006) 3682–3689.
- [134] A. Bingham, et al., Predictors of acquired lipodystrophy in juvenile-onset dermatomyositis and a gradient of severity, *Medicine (Baltim.)* 87 (2) (2008) 70–86.
- [135] H. Gunawardena, et al., Clinical associations of autoantibodies to a p155/140 kDa doublet protein in juvenile dermatomyositis, *Rheumatology* 47 (3) (2008) 324–328.
- [136] S.L. Tansley, N.J. McHugh, L.R. Wedderburn, Adult and juvenile dermatomyositis: are the distinct clinical features explained by our current understanding of serological subgroups and pathogenic mechanisms? *Arthritis Res. Ther.* 15 (2) (2013) 211.
- [137] K. Danko, et al., Long-term survival of patients with idiopathic inflammatory myopathies according to clinical features: a longitudinal study of 162 cases, *Medicine (Baltim.)* 83 (1) (2004) 35–42.
- [138] T. Schwartz, et al., Cardiac involvement in adult and juvenile idiopathic inflammatory myopathies, *RMD Open* 2 (2) (2016), e000291.
- [139] R. Gupta, et al., Clinical cardiac involvement in idiopathic inflammatory myopathies: a systematic review, *Int. J. Cardiol.* 148 (3) (2011) 261–270.
- [140] L.P. Diederichsen, Cardiovascular involvement in myositis, *Curr. Opin. Rheumatol.* 29 (6) (2017) 598–603.
- [141] L. Zhang, et al., Cardiac involvement in adult polymyositis or dermatomyositis: a systematic review, *Clin. Cardiol.* 35 (11) (2012) 686–691.
- [142] S.K. Rai, et al., Risk of myocardial infarction and ischaemic stroke in adults with polymyositis and dermatomyositis: a general population-based study, *Rheumatology* 55 (3) (2016) 461–469.
- [143] A. Rosenbohm, et al., Early diagnosis of cardiac involvement in idiopathic inflammatory myopathy by cardiac magnetic resonance tomography, *J. Neurol.* 262 (4) (2015) 949–956.
- [144] F. Chen, Y. Peng, M. Chen, Diagnostic approach to cardiac involvement in idiopathic inflammatory myopathies, *Int. Heart J.* 59 (2) (2018) 256–262.
- [145] M. Hughes, et al., Cardiac troponin testing in idiopathic inflammatory myopathies and systemic sclerosis-spectrum disorders: biomarkers to distinguish between primary cardiac involvement and low-grade skeletal muscle disease activity, *Ann. Rheum. Dis.* 74 (5) (2015) 795–798.
- [146] J. Albayda, et al., Inflammatory myopathy associated with anti-mitochondrial antibodies: a distinct phenotype with cardiac involvement, *Semin. Arthritis Rheum.* 47 (4) (2018) 552–556.
- [147] S. Mavrogeni, et al., Myocarditis during acute inflammatory myopathies: evaluation using clinical criteria and cardiac magnetic resonance imaging, *Int. J. Cardiol.* 164 (1) (2013) e3–4.
- [148] Y. Allanore, et al., Effects of corticosteroids and immunosuppressors on idiopathic inflammatory myopathy related myocarditis evaluated by magnetic resonance imaging, *Ann. Rheum. Dis.* 65 (2) (2006) 249–252.

- [149] S. Ye, et al., Adult clinically amyopathic dermatomyositis with rapid progressive interstitial lung disease: a retrospective cohort study, *Clin. Rheumatol.* 26 (10) (2007) 1647–1654.
- [150] J. Won Huh, et al., Two distinct clinical types of interstitial lung disease associated with polymyositis-dermatomyositis, *Respir. Med.* 101 (8) (2007) 1761–1769.
- [151] S. Hayashi, et al., High-resolution computed tomography characterization of interstitial lung diseases in polymyositis/dermatomyositis, *J. Rheumatol.* 35 (2) (2008) 260–269.
- [152] T. Fujisawa, et al., Differences in clinical features and prognosis of interstitial lung diseases between polymyositis and dermatomyositis, *J. Rheumatol.* 32 (1) (2005) 58–64.
- [153] L.A. Saketkoo, et al., Reconciling healthcare professional and patient perspectives in the development of disease activity and response criteria in connective tissue disease-related interstitial lung diseases, *J. Rheumatol.* 41 (4) (2014) 792–798.
- [154] I. Marie, et al., Interstitial lung disease in polymyositis and dermatomyositis, *Arthritis Rheum.* 47 (6) (2002) 614–622.
- [155] T. Fujisawa, et al., Prognostic factors for myositis-associated interstitial lung disease, *PLoS One* 9 (6) (2014), e98824.
- [156] I.J. Chen, et al., Interstitial lung disease in polymyositis and dermatomyositis, *Clin. Rheumatol.* 28 (6) (2009) 639–646.
- [157] T. Mimori, R. Nakashima, Y. Hosono, Interstitial lung disease in myositis: clinical subsets, biomarkers, and treatment, *Curr. Rheumatol. Rep.* 14 (3) (2012) 264–274.
- [158] F. Chua, et al., Idiopathic inflammatory myositis-associated interstitial lung disease: ethnicity differences and lung function trends in a British cohort, *Rheumatology* 51 (10) (2012) 1870–1876.
- [159] C.S. Lee, et al., Idiopathic inflammatory myopathy with diffuse alveolar damage, *Clin. Rheumatol.* 21 (5) (2002) 391–396.
- [160] T. Gono, et al., Brief report: association of HLA-DRB1*0101/*0405 with susceptibility to anti-melanoma differentiation-associated gene 5 antibody-positive dermatomyositis in the Japanese population, *Arthritis Rheum.* 64 (11) (2012) 3736–3740.
- [161] J. Bauhammer, et al., Rituximab in the treatment of Jo1 antibody-associated antisynthetase syndrome: anti-Ro52 positivity as a marker for severity and treatment response, *J. Rheumatol.* 43 (8) (2016) 1566.
- [162] Z. Betteridge, N. McHugh, Myositis-specific autoantibodies: an important tool to support diagnosis of myositis, *J. Intern. Med.* 280 (1) (2016) 8–23.
- [163] R. Buchbinder, et al., Incidence of malignant disease in biopsy-proven inflammatory myopathy. A population-based cohort study, *Ann. Intern. Med.* 134 (12) (2001) 1087–1095.
- [164] C.L. Hill, et al., Frequency of specific cancer types in dermatomyositis and polymyositis: a population-based study, *Lancet* 357 (9250) (2001) 96–100.
- [165] S.W. Lee, et al., Malignancies in Korean patients with inflammatory myopathy, *Yonsei Med. J.* 47 (4) (2006) 519–523.
- [166] W.K. Fung, H.L. Chan, W.M. Lam, Amyopathic dermatomyositis in Hong Kong – association with nasopharyngeal carcinoma, *Int. J. Dermatol.* 37 (9) (1998) 659–663.
- [167] Y.J. Chen, C.Y. Wu, J.L. Shen, Predicting factors of malignancy in dermatomyositis and polymyositis: a case-control study, *Br. J. Dermatol.* 144 (4) (2001) 825–831.
- [168] I. Marie, et al., Hematological malignancy associated with polymyositis and dermatomyositis, *Autoimmun. Rev.* 11 (9) (2012) 615–620.
- [169] Y.L. Huang, et al., Malignancies associated with dermatomyositis and polymyositis in Taiwan: a nationwide population-based study, *Br. J. Dermatol.* 161 (4) (2009) 854–860.
- [170] I. Marie, et al., Influence of age on characteristics of polymyositis and dermatomyositis in adults, *Medicine (Baltim.)* 78 (3) (1999) 139–147.
- [171] Z. Amoura, et al., Tumor antigen markers for the detection of solid cancers in inflammatory myopathies, *Cancer Epidemiol. Biomark. Prev.* 14 (5) (2005) 1279–1282.
- [172] M.L. Albert, R.B. Darnell, Paraneoplastic neurological degenerations: keys to tumour immunity, *Nat. Rev. Cancer* 4 (1) (2004) 36–44.
- [173] A. Ponyi, et al., Cancer-associated myositis: clinical features and prognostic signs, *Ann. N. Y. Acad. Sci.* 1051 (2005) 64–71.
- [174] S.M. Levine, Cancer and myositis: new insights into an old association, *Curr. Opin. Rheumatol.* 18 (6) (2006) 620–624.
- [175] A. Hida, et al., Anti-TIF1-gamma antibody and cancer-associated myositis: a clinicohistopathologic study, *Neurology* 87 (3) (2016) 299–308.
- [176] H. Yang, et al., Identification of multiple cancer-associated myositis-specific autoantibodies in idiopathic inflammatory myopathies: a large longitudinal cohort study, *Arthritis Res. Ther.* 19 (1) (2017) 259.
- [177] H. Chinoy, et al., The diagnostic utility of myositis autoantibody testing for predicting the risk of cancer-associated myositis, *Ann. Rheum. Dis.* 66 (10) (2007) 1345–1349.
- [178] T. Noda, et al., Gene expression profile of inflammatory myopathy with malignancy is similar to that of dermatomyositis rather than polymyositis, *Intern. Med.* 55 (18) (2016) 2571–2580.
- [179] O. Danielsson, et al., Increased prevalence of celiac disease in idiopathic inflammatory myopathies, *Brain Behav.* 7 (10) (2017), e00803.
- [180] A. Selva-O'Callaghan, et al., Celiac disease and antibodies associated with celiac disease in patients with inflammatory myopathy, *Muscle Nerve* 35 (1) (2007) 49–54.
- [181] T.H. Yen, et al., Renal involvement in patients with polymyositis and dermatomyositis, *Int. J. Clin. Pract.* 59 (2) (2005) 188–193.
- [182] G. Couvrat-Desvergnès, et al., The spectrum of renal involvement in patients with inflammatory myopathies, *Medicine (Baltim.)* 93 (1) (2014) 33–41.
- [183] D. Cucchiari, C. Angelini, Renal involvement in idiopathic inflammatory myopathies, *Clin. Rev. Allergy Immunol.* 52 (1) (2017) 99–107.
- [184] P.-J. Clements, et al., Muscle disease in progressive systemic sclerosis: diagnostic and therapeutic considerations, *Arthritis Rheum.* 21 (1) (1978) 62–71.
- [185] G.C. Sharp, et al., Mixed connective tissue disease—an apparently distinct rheumatic disease syndrome associated with a specific antibody to an extractable nuclear antigen (ENA), *Am. J. Med.* 52 (2) (1972) 148–159.
- [186] A. Meneghel, et al., Life-threatening systemic capillary leak syndrome in juvenile dermatomyositis, *Rheumatology* 56 (10) (2017) 1822–1823.
- [187] E. Quecedo, et al., Partial lipodystrophy associated with juvenile dermatomyositis: report of two cases, *Pediatr. Dermatol.* 13 (6) (1996) 477–482.
- [188] C. Huemer, et al., Lipodystrophy in patients with juvenile dermatomyositis—evaluation of clinical and metabolic abnormalities, *J. Rheumatol.* 28 (3) (2001) 610–615.
- [189] E. Schiopu, et al., Predictors of survival in a cohort of patients with polymyositis and dermatomyositis: effect of corticosteroids, methotrexate and azathioprine, *Arthritis Res. Ther.* 14 (1) (2012) R22.
- [190] A. Tollisen, et al., Quality of life in adults with juvenile-onset dermatomyositis: a case-control study, *Arthritis Care Res.* 64 (7) (2012) 1020–1027.
- [191] P.R. Mathiesen, et al., Aerobic fitness after JDM—a long-term follow-up study, *Rheumatology* 52 (2) (2013) 287–295.
- [192] A. Bohan, J.B. Peter, Polymyositis and dermatomyositis (second of two parts), *N. Engl. J. Med.* 292 (8) (1975) 403–407.
- [193] J.C. Milisenda, A. Selva-O'Callaghan, J.M. Grau, The diagnosis and classification of polymyositis, *J. Autoimmun.* 48–49 (2014) 118–121.
- [194] F.W. Miller, et al., Proposed preliminary core set measures for disease outcome assessment in adult and juvenile idiopathic inflammatory myopathies, *Rheumatology* 40 (11) (2001) 1262–1273.
- [195] L.G. Rider, Outcome assessment in the adult and juvenile idiopathic inflammatory myopathies, *Rheum. Dis. Clin. N. Am.* 28 (4) (2002) 935–977.
- [196] A. Bohan, M.E. Cronin, Computer-assisted analysis of 153 patients with polymyositis and dermatomyositis, *Medicine (Baltim.)* 56 (4) (1977) 255–286.
- [197] A. Malik, et al., Idiopathic inflammatory myopathies: clinical approach and management, *Front. Neurol.* 7 (2016) 64.
- [198] L. Yuan, et al., Serum levels of soluble ST2 and interleukin-33 in patients with dermatomyositis and polymyositis, *Clin. Exp. Rheumatol.* 31 (3) (2013) 428–432.
- [199] C. Gabay, et al., Elevated serum levels of interleukin-1 receptor antagonist in polymyositis/dermatomyositis. A biologic marker of disease activity with a possible role in the lack of acute-phase protein response, *Arthritis Rheum.* 37 (12) (1994) 1744–1751.
- [200] A. Ghirardello, et al., Clinical implications of autoantibody screening in patients with autoimmune myositis, *Autoimmunity* 39 (3) (2006) 217–221.
- [201] D.J. Gazeley, M.E. Cronin, Diagnosis and treatment of the idiopathic inflammatory myopathies, *Ther. Adv. Musculoskelet Dis* 3 (6) (2011) 315–324.
- [202] A. Ghirardello, et al., Autoantibodies in polymyositis and dermatomyositis, *Curr. Rheumatol. Rep.* 15 (6) (2013) 335.
- [203] J. Tomasova Studynkova, et al., The role of MRI in the assessment of polymyositis and dermatomyositis, *Rheumatology* 46 (7) (2007) 1174–1179.
- [204] E.M. Adams, et al., The idiopathic inflammatory myopathies: spectrum of MR imaging findings, *RadioGraphics* 15 (3) (1995) 563–574.
- [205] M. Schulze, et al., MRI findings in inflammatory muscle diseases and their noninflammatory mimics, *AJR Am. J. Roentgenol.* 192 (6) (2009) 1708–1716.
- [206] A.B. Kimball, et al., Magnetic resonance imaging detection of occult skin and subcutaneous abnormalities in juvenile dermatomyositis. Implications for diagnosis and therapy, *Arthritis Rheum.* 43 (8) (2000) 1866–1873.
- [207] S.M. Maillard, et al., Quantitative assessment of MRI T2 relaxation time of thigh muscles in juvenile dermatomyositis, *Rheumatology* 43 (5) (2004) 603–608.
- [208] F. Del Grande, et al., Magnetic Resonance Imaging of Inflammatory Myopathies 22 (2) (2011) 39–43.
- [209] V.E. Brown, et al., An international consensus survey of the diagnostic criteria for juvenile dermatomyositis (JDM), *Rheumatology* 45 (8) (2006) 990–993.
- [210] R.S. Adler, G. Garofalo, Ultrasound in the evaluation of the inflammatory myopathies, *Curr. Rheumatol. Rep.* 11 (4) (2009) 302–308.
- [211] M.-A. Weber, Ultrasound in the inflammatory myopathies, *Ann. N. Y. Acad. Sci.* 1154 (1) (2009) 159–170.
- [212] K. Arahata, A.G. Engel, Monoclonal antibody analysis of mononuclear cells in myopathies. I: quantitation of subsets according to diagnosis and sites of accumulation and demonstration and counts of muscle fibers invaded by T cells, *Ann. Neurol.* 16 (2) (1984) 193–208.
- [213] I.N. Lazarou, P.-A. Guerne, Classification, diagnosis, and management of idiopathic inflammatory myopathies, *J. Rheumatol.* 40 (5) (2013) 550.
- [214] Y.B. Luo, F.L. Mastaglia, Dermatomyositis, polymyositis and immune-mediated necrotising myopathies, *Biochim. Biophys. Acta* 1852 (4) (2015) 622–632.
- [215] W.K. Engel, V. Askanas, Inclusion-body myositis: clinical, diagnostic, and pathologic aspects, *Neurology* 66 (2 Suppl 1) (2006) S20–S29.
- [216] M.C. Dalakas, Inflammatory muscle diseases: a critical review on pathogenesis and therapies, *Curr. Opin. Pharmacol.* 10 (3) (2010) 346–352.
- [217] M.C. Dalakas, Therapeutic advances and future prospects in immune-mediated inflammatory myopathies, *Ther. Adv. Neurol. Disord.* 1 (3) (2008) 157–166.
- [218] M.C. Dalakas, Immunotherapy of inflammatory myopathies: practical approach and future prospects, *Curr. Treat. Options Neurol.* 13 (3) (2011) 311–323.
- [219] F.L. Mastaglia, P.J. Zilko, Inflammatory myopathies: how to treat the difficult cases, *J. Clin. Neurosci.* 10 (1) (2003) 99–101.
- [220] J. van de Vlekkert, et al., Oral dexamethasone pulse therapy versus daily prednisolone in sub-acute onset myositis, a randomised clinical trial, *Neuromuscul. Disord.* 20 (6) (2010) 382–389.

- [221] F.C. Ernste, A.M. Reed, Idiopathic inflammatory myopathies: current trends in pathogenesis, clinical features, and up-to-date treatment recommendations, *Mayo Clin. Proc.* 88 (1) (2013) 83–105.
- [222] G.S. Hoffman, et al., Presentation, treatment, and prognosis of idiopathic inflammatory muscle disease in a rural hospital, *Am. J. Med.* 75 (3) (1983) 433–438.
- [223] M.M. Joffe, et al., Drug therapy of the idiopathic inflammatory myopathies: predictors of response to prednisone, azathioprine, and methotrexate and a comparison of their efficacy, *Am. J. Med.* 94 (4) (1993) 379–387.
- [224] L.A. Drake, et al., Guidelines of care for dermatomyositis. American academy of dermatology, *J. Am. Acad. Dermatol.* 34 (5 Pt 1) (1996) 824–829.
- [225] Y. Troyanov, et al., Novel classification of idiopathic inflammatory myopathies based on overlap syndrome features and autoantibodies: analysis of 100 French Canadian patients, *Medicine (Baltim.)* 84 (4) (2005) 231–249.
- [226] M.G. Danieli, et al., Subcutaneous immunoglobulin in polymyositis and dermatomyositis: a novel application, *Autoimmun. Rev.* 10 (3) (2011) 144–149.
- [227] M.C. Dalakas, et al., A controlled trial of high-dose intravenous immune globulin infusions as treatment for dermatomyositis, *N. Engl. J. Med.* 329 (27) (1993) 1993–2000.
- [228] G. Ramirez, et al., Adult-onset polymyositis-dermatomyositis: description of 25 patients with emphasis on treatment, *Semin. Arthritis Rheum.* 20 (2) (1990) 114–120.
- [229] P. Hollingworth, et al., Intensive immunosuppression versus prednisolone in the treatment of connective tissue diseases, *Ann. Rheum. Dis.* 41 (6) (1982) 557–562.
- [230] J.A.L. Miller, Y. Walsh, S. Saminaden, Randomised Double Blind Controlled Trial of Methotrexate and Steroids Compared with Azathioprine and Steroids in the Treatment of Idiopathic Inflammatory Myopathy, vol. 199, 2002, p. S53.
- [231] T.W. Bunch, Prednisone and azathioprine for polymyositis: long-term followup, *Arthritis Rheum.* 24 (1) (1981) 45–48.
- [232] A.N. Malaviya, A. Many, R.S. Schwartz, Treatment of dermatomyositis with methotrexate, *Lancet* 2 (7566) (1968) 485–488.
- [233] M.E. Zieglschmid-Adams, et al., The value of methotrexate in dermatomyositis, *J. Am. Acad. Dermatol.* 38 (1) (1998) 130–132.
- [234] L. Villalba, et al., Treatment of refractory myositis: a randomized crossover study of two new cytotoxic regimens, *Arthritis Rheum.* 41 (3) (1998) 392–399.
- [235] M.C. Dalakas, Inflammatory myopathies: management of steroid resistance, *Curr. Opin. Neurol.* 24 (5) (2011) 457–462.
- [236] M.C. Dalakas, B cells as therapeutic targets in autoimmune neurological disorders, *Nat. Clin. Pract. Neurol.* 4 (10) (2008) 557–567.
- [237] T.D. Levine, Rituximab in the treatment of dermatomyositis: an open-label pilot study, *Arthritis Rheum.* 52 (2) (2005) 601–607.
- [238] L. Chung, M.C. Genovese, D.F. Fiorentino, A pilot trial of rituximab in the treatment of patients with dermatomyositis, *Arch. Dermatol.* 143 (6) (2007) 763–767.
- [239] K. Innami, et al., Successful treatment using rituximab in a patient with refractory polymyositis complicated by scleroderma renal crisis, *BMJ Case Rep.* 2017 (2017).
- [240] R. Aggarwal, et al., Cutaneous improvement in refractory adult and juvenile dermatomyositis after treatment with rituximab, *Rheumatology* 56 (2) (2017) 247–254.
- [241] L. Nalotto, et al., Rituximab in refractory idiopathic inflammatory myopathies and antisynthetase syndrome: personal experience and review of the literature, *Immunol. Res.* 56 (2–3) (2013) 362–370.
- [242] M. Sem, et al., Rituximab treatment of the anti-synthetase syndrome: a retrospective case series, *Rheumatology* 48 (8) (2009) 968–971.
- [243] R. Aggarwal, et al., Predictors of clinical improvement in rituximab-treated refractory adult and juvenile dermatomyositis and adult polymyositis, *Arthritis Rheum.* 66 (3) (2014) 740–749.
- [244] G.J. Hengstman, et al., Successful treatment of dermatomyositis and polymyositis with anti-tumor-necrosis-factor-alpha: preliminary observations, *Eur. Neurol.* 50 (1) (2003) 10–15.
- [245] A. Schiffenbauer, et al., A randomized, double-blind, placebo-controlled trial of infliximab in refractory polymyositis and dermatomyositis, *Semin. Arthritis Rheum.* 47 (6) (2018) 858–864.
- [246] M. Dastmalchi, et al., A high incidence of disease flares in an open pilot study of infliximab in patients with refractory inflammatory myopathies, *Ann. Rheum. Dis.* 67 (12) (2008) 1670–1677.
- [247] A randomized, pilot trial of etanercept in dermatomyositis, *Ann. Neurol.* 70 (3) (2011) 427–436.
- [248] B. Thompson, et al., Alemtuzumab (Campath-1H) for treatment of refractory polymyositis, *J. Rheumatol.* 35 (10) (2008) 2080–2082.
- [249] A. Reiff, et al., Anti-CD52 antibody-mediated immune ablation with autologous immune recovery for the treatment of refractory juvenile polymyositis, *J. Clin. Immunol.* 31 (4) (2011) 615–622.
- [250] I. Marie, Therapy of polymyositis and dermatomyositis, *Presse Med.* 40 (4 Pt 2) (2011) e257–e270.
- [251] H. Alexanderson, Exercise in inflammatory myopathies, including inclusion body myositis, *Curr. Rheumatol. Rep.* 14 (3) (2012) 244–251.
- [252] G.F. Wiesinger, et al., Improvement of physical fitness and muscle strength in polymyositis/dermatomyositis patients by a training programme, *Br. J. Rheumatol.* 37 (2) (1998) 196–200.
- [253] L.G. Johnson, et al., Improvement in aerobic capacity after an exercise program in sporadic inclusion body myositis, *J. Clin. Neuromuscul. Dis.* 10 (4) (2009) 178–184.
- [254] Y.L. Chung, et al., Creatine supplements in patients with idiopathic inflammatory myopathies who are clinically weak after conventional pharmacologic treatment: six-month, double-blind, randomized, placebo-controlled trial, *Arthritis Rheum.* 57 (4) (2007) 694–702.
- [255] M.J. Bendewald, et al., Incidence of dermatomyositis and clinically amyopathic dermatomyositis: a population-based study in Olmsted County, Minnesota, *Arch. Dermatol.* 146 (1) (2010) 26–30.
- [256] J. Rosa, et al., Incidence and prevalence of polymyositis and dermatomyositis in a Health Management Organization in Buenos Aires 19 (6) (2013) 303–307.
- [257] H. Vargas-Legas, et al., [Polymyositis-dermatomyositis: incidence in Spain (1997–2004)], *Med. Clínica* 129 (19) (2007) 721–724.
- [258] A. Ohta, et al., Prevalence and incidence of polymyositis and dermatomyositis in Japan, *Mod. Rheumatol.* 24 (3) (2014) 477–480.
- [259] S.-K. Cho, et al., Incidence and prevalence of idiopathic inflammatory myopathies in Korea: a nationwide population-based study, *J. Korean Med. Sci.* 34 (8) (2019).
- [260] S. Bernatsky, et al., Estimating the prevalence of polymyositis and dermatomyositis from administrative data: age, sex and regional differences, *Ann. Rheum. Dis.* 68 (7) (2009) 1192–1196.
- [261] K.E. Smoyer-Tomic, A.A. Amato, A.W. Fernandes, Incidence and prevalence of idiopathic inflammatory myopathies among commercially insured, Medicare supplemental insured, and Medicaid enrolled populations: an administrative claims analysis, *BMC Musculoskelet. Disord.* 13 (2012) 103.
- [262] D.E. Furst, et al., Epidemiology of adult idiopathic inflammatory myopathies in a U.S. managed care plan, *Muscle Nerve* 45 (5) (2012) 676–683.
- [263] C.F. Kuo, et al., Incidence, cancer risk and mortality of dermatomyositis and polymyositis in Taiwan: a nationwide population study, *Br. J. Dermatol.* 165 (6) (2011) 1273–1279.
- [264] M.R. Rose, 188th ENMC international workshop: inclusion body myositis, 2–4 december 2011, naarden, The Netherlands, *Neuromuscul. Disord.* 23 (12) (2013) 1044–1055.
- [265] L.A. Love, et al., Ultraviolet radiation intensity predicts the relative distribution of dermatomyositis and anti-Mi-2 autoantibodies in women, *Arthritis Rheum.* 60 (8) (2009) 2499–2504.
- [266] Y. Hamaguchi, et al., Clinical correlations with dermatomyositis-specific autoantibodies in adult Japanese patients with dermatomyositis: a multicenter cross-sectional StudyDM-specific autoantibodies in patients with DM, *JAMA Dermatol.* 147 (4) (2011) 391–398.
- [267] M.M. Joffe, et al., Drug therapy of the idiopathic inflammatory myopathies: predictors of response to prednisone, azathioprine, and methotrexate and a comparison of their efficacy, *Am. J. Med.* 94 (4) (1993) 379–387.
- [268] K. Hoshino, et al., Anti-MDA5 and anti-TIF1-γ antibodies have clinical significance for patients with dermatomyositis, *Rheumatology* 49 (9) (2010) 1726–1733.
- [269] I. Kobayashi, et al., Anti-melanoma differentiation-associated gene 5 antibody is a diagnostic and predictive marker for interstitial lung diseases associated with juvenile dermatomyositis, *J. Pediatr.* 158 (4) (2011) 675–677.
- [270] Y. Ge, et al., Clinical characteristics of anti-SAE antibodies in Chinese patients with dermatomyositis in comparison with different patient cohorts, *Sci. Rep.* 7 (1) (2017) 188.
- [271] H. Chinoy, et al., The diagnostic utility of myositis autoantibody testing for predicting the risk of cancer-associated myositis, *Ann. Rheum. Dis.* 66 (10) (2007) 1345.
- [272] L. Casciola-Rosen, A.L. Mammen, Myositis autoantibodies 24 (6) (2012) 602–608.
- [273] H. Gunawardena, et al., Autoantibodies to a 140-kd protein in juvenile dermatomyositis are associated with calcinosis, *Arthritis Rheum.* 60 (6) (2009) 1807–1814.
- [274] L. Christopher-Stine, et al., A novel autoantibody recognizing 200-kd and 100-kd proteins is associated with an immune-mediated necrotizing myopathy, *Arthritis Rheum.* 62 (9) (2010) 2757–2766.
- [275] S.L. Tansley, et al., Anti-hmgcr autoantibodies in juvenile idiopathic inflammatory myopathies identify a rare but clinically important subset of patients, *J. Rheumatol.* 44 (4) (2017) 488–492.
- [276] A.H. Kao, et al., Anti-signal recognition particle autoantibody in patients with and patients without idiopathic inflammatory myopathy, *Arthritis Rheum.* 50 (1) (2004) 209–215.
- [277] I.N. Targoff, et al., Classification criteria for the idiopathic inflammatory myopathies, *Curr. Opin. Rheumatol.* 9 (6) (1997) 527–535.
- [278] J.C. Lega, et al., The clinical phenotype associated with myositis-specific and associated autoantibodies: a meta-analysis revisiting the so-called antisynthetase syndrome, *Autoimmun. Rev.* 13 (9) (2014) 883–891.
- [279] P. Coppo, et al., Inflammatory myositis associated with anti-U1-small nuclear ribonucleoprotein antibodies: a subset of myositis associated with a favourable outcome, *Rheumatology* 41 (9) (2002) 1040–1046.
- [280] L. Plestilova, et al., THU0253ANTI-PM-SCL autoantibodies in polymyositis and dermatomyositis, *Ann. Rheum. Dis.* 71 (Suppl 3) (2013) 240.
- [281] I. Marie, et al., Long-term outcome of patients with polymyositis/dermatomyositis and anti-PM-Scl antibody, *Br. J. Dermatol.* 162 (2) (2010) 337–344.
- [282] A. Rigolet, et al., Inflammatory myopathies with anti-Ku antibodies: a prognosis dependent on associated lung disease, *Medicine (Baltim.)* 91 (2) (2012) 95–102.