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## West Nile Virus-Induced Neurologic Sequelae—Relationship to Neurodegenerative Cascades and Dementias

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### Abstract

**Purpose of Review**—West Nile virus (WNV) emerged from Central Africa in the 1990s and is now endemic throughout much of the world. Twenty years after its introduction in the USA, it is becoming apparent that neurological impairments can persist for years following infection. Here, we review the epidemiological data in support of such long-term deficits and discuss possible mechanisms that drive these persistent manifestations.

**Recent Findings**—Focusing on the recently discovered antimicrobial roles of amyloid and alpha-synuclein, we connect WNV late pathology to overlapping features encountered in neurodegenerative diseases such as Alzheimer’s disease and Parkinson’s disease. We also summarize new research on microglial activation and engulfment of neural synapses seen in recovered WNV as well as in neurodegenerative diseases, and discuss how loss of integrity of the blood-brain barrier (BBB) may exacerbate this process.

**Summary**—Neuroinvasive viral infections such as WNV may be linked epidemiologically and mechanistically to neurodegeneration. This may open doors to therapeutic options for hitherto untreatable infectious sequelae; additionally, it may also shed light on the possible infectious etiologies of age-progressive neurodegenerative dementias.

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## Keywords

West Nile virus; Neurological sequelae; Neurodegenerative disease; Antimicrobial peptide; Amyloid beta; Alpha-synuclein; Alzheimer's disease; Parkinson's disease

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## Introduction

The neuroinvasive, mosquito-borne West Nile virus (genus *Flavivirus*, family *Flaviviridae*) is a small, positive-sense, single-stranded RNA virus originating from Central Africa. Its range has extended significantly in recent decades, and it now occurs throughout the Americas, Europe, Australia, Africa, Asia, and the Middle East [1]. Since its introduction to North America in 1999, it is estimated that 3 to 5 million people have been infected by WNV in the USA alone [2]. Approximately 1% of WNV infections result in neuroinvasive disease, including meningitis, encephalitis, or acute flaccid paralysis/poliomyelitis [3]. While clinically apparent neuroinvasive disease is relatively rare, even mild and asymptomatic WNV infections have been associated with a significant prevalence of sequelae such as memory loss, confusion, and fatigue years later [4, 5]. This has led to some intriguing epidemiological and animal-model studies of the acute and late CNS pathology of WNV, revealing features that overlap with neurodegenerative dementias such as Alzheimer's disease (AD) and Parkinson's disease (PD).

In this review, we discuss epidemiological studies on WNV neurological sequelae and delve into some key mechanisms that may be driving these long-term symptoms. The literature on neurological impairment, following neuroinvasive infections, and research on the etiology of neurodegenerative diseases are converging on the role of the innate immune system [6]. In both instances, production of antimicrobial peptides and activation of glial cells precede and may possibly underlie the ensuing CNS dysfunction. Comorbidities, such as diabetes mellitus and hypertension, appear to predispose individuals to both conditions, possibly by their vasculopathies or by weakening the blood-brain barrier (BBB). Considering these relationships and possibilities opens the door to many new questions. First, do these pathogens contribute to age-dependent neurodegeneration and if so, what are the molecular pathways affected? Which host factors—environmental as well as genetic—confer not only risk but also protection following infection by a neuroinvasive microbe? While we have definitely observed high levels of cognitive impairment following alphaviral infection in endemic regions (Carrera and Vittor, unpublished data), we will limit our discussion here to WNV since the literature is most robust for this pathogen. We will then link the WNV findings to recent developments in the two most common neurodegenerative dementias, namely AD and PD.

## Neurological Sequelae of WNV

WNV infection results in a self-limited febrile illness in 25% of exposed individuals, while neuroinvasive disease (meningitis, encephalitis acute flaccid paralysis) occurs in less than 1% [3]. Following the initial waves of WNV outbreaks throughout North America in the early 2000s, it became increasingly clear to clinicians taking care of these patients that the acute phase of illness was often followed by a prolonged phase characterized by excessive

fatigue, confusion, memory loss, muscle weakness, and occasionally, tremors. The first studies to systematically examine the possibility of such long-term sequelae demonstrated that indeed, 20 to 60% of patients reported difficulties with these symptoms even after 6 to 18 months after the initial infection [7–9]. While it was initially hypothesized that patients with neuroinvasive disease (encephalitis or meningitis) would report higher rates of ongoing symptoms, most studies have shown that even patients with non-neuroinvasive disease (WNV fever) reported similar sequelae at equal frequencies [10]. Due to a paucity of data on long-term outcomes in people asymptomatically infected with WNV, the fate of this population is unknown.

As more time elapsed since WNV first arrived in North America, it became possible to study these sequelae over longer periods of time. Though the studies are still few in number, it appears that significant portions of patients with WNV have ongoing symptoms years after the acute phase of illness [4, 11]. For example, Murray et al. [4] note that at 8 years from initial infection (neuroinvasive and febrile WNV), 40% of patients still reported sequelae [4]. Fatigue, weakness, and depression were most common, and interestingly, 9% of these patients reported memory loss and confusion. Poor recovery was associated with older age and diabetes.

These self-reported symptoms were further substantiated in a series of studies utilizing various batteries of neuropsychological tests [5, 11–13, 14••, 15, 16]. Most of these studies have included two groups of subjects—those who initially suffered from WNV neuroinvasive disease, and others who had WNV fever. The emerging themes from these studies are that psychomotor speed, memory, and executive functions are impacted significantly. In addition, as with the self-reported symptoms, the objective measures of cognitive impairment do not appear to differ significantly between the neuroinvasive and non-neuroinvasive groups [5, 12, 13]. Assessments are mostly performed 1 year after the onset of the initial illness. Samaan et al. [11], however, evaluated subjects whose time since infection ranged from less than 1 month up to 4 years. Not only did the authors demonstrate that a sizeable proportion of patients had ongoing motor function deficits (69%), executive function deficits (30%), and verbal learning and memory deficits (30%), but they showed that these impairments were more pronounced in subjects with longer intervals since their initial illness.

A caveat of these studies is that only few studies include healthy, or normative controls in their analyses [15, 17]. In a study that included age- and sex-matched controls, Balakrishnan et al. [17] followed up patients 1 year after illness due to WNV encephalitis. Cognitive impairment was determined using the Mini-Mental Status Exam (MMSE; probable dementia MMSE score < 21). The authors report a 57% (95% CI 22.5–76.1%) attributable risk of WNV encephalitis for probable dementia, with a crude relative risk of 2.3 (95% CI 1.3–4.2). Notably, of the 40 cases initially identified as having had WNV encephalitis, 10 had died within a year. On the other hand, Sejvar et al. [15] found that motor deficits persisted 1.5 years after illness onset, but did not find a significant difference in neurocognitive performance in WNV patients compared to normative controls using the Cambridge Neuropsychological Testing Automated Battery (CANTAB [18]).

A recent study extended these findings further by conducting magnetic resonance imaging (MRI) on a subset of a cohort of patients with WNV neuroinvasive disease or WNV fever [14••]. The population included subjects at 0 to 11 years following initial infection, 25–31% of whom demonstrated memory impairments on cognitive testing. Compared to age- and sex-matched controls, significant bilateral frontal and limbic cortical thinning was observed in WNV subjects. A review of MRI findings (Table 1) reveals that acute illness is characterized predominantly by abnormalities in the thalami and basal ganglia, though many other structures can be affected. Late findings include cortical thinning in structures within the frontal, temporal, and limbic cortices and regional atrophy in the thalami, basal ganglia, cerebellum, and brain stem. Interestingly, several of these affected structures are also atrophied in AD [39] (e.g. posterior cingulate cortex and insular cortex, and the hippocampus and entorhinal regions in the temporal lobe) and PD [40] (e.g., substantia nigra within the basal ganglia and thalamic connections).

Taken together, the preponderance of epidemiological evidence suggests that cognitive impairment following WNV infection is common, and that its occurrence does not correlate with severity of the initial presentation (neuroinvasive cases and febrile WNV infections). On neuropsychological testing, the domains most affected in WNV patients include psychomotor, memory, and executive function. While these studies have definitely advanced our understanding of long-term clinical and neuropsychiatric outcomes of WNV, the lack of healthy controls and asymptotically infected individuals in most studies hampers our ability to accurately define the extent of neurological sequelae following WNV exposure.

## CNS Infection in WNV Patients

Following peripheral infection, WNV accesses the CNS via one or more of four pathways: free virus crosses the BBB via passive transport through the endothelium [41], peripheral inflammation (e.g., due to tumor necrosis factor- $\alpha$  (TNF- $\alpha$ )) weakens the tight junctions of the BBB allowing the virus to “leak” in [42], infected virus leukocytes are trafficked across the BBB in a “trojan horse” fashion [41], or it infects peripheral nerves with retrograde axonal flow into the CNS [43]. Once the virus enters the brain, it can replicate in astrocytes, microglia, and neurons [44, 45], which produce pro-inflammatory cytokines such as type I interferons (IFN- $\alpha$ , IFN- $\beta$ ), type III interferons (IFN- $\lambda$ ), TNF- $\alpha$ , and interleukin-1 $\beta$  (IL-1 $\beta$ ). Direct neuronal injury is incurred via apoptosis of infected cells [46] or by secondary insults due to the ensuing inflammation [47]. The innate immune response (i.e., pro-inflammatory cytokines, antimicrobial peptides, phagocytes, natural killer cells, and  $\gamma\delta$  T cells), complement, and the adaptive immune response are all critical for clearance of virus and survival [48]. WNV persistence in the CNS has been studied in mice, revealing that in immunocompetent mice, infectious virus is typically found up to 1 month after infection [49]. Thereafter, WNV RNA remains detectable for up to 6 months post infection (p.i.). However, Appler et al. demonstrated that infectious virus appears to reactivate beyond 1 month p.i. upon treatment with cyclophosphamide, suggesting that lymphocyte function to restrict the replication of WNV [49]. Detailed analysis of these lymphocyte subpopulations showed that activated, virus-specific CD4+ and CD8+ T cells as well as plasma cells remained in the CNS even at 4 months p.i. [50]. Importantly, regulatory T cells

(Tregs) were also present at elevated levels, likely preventing neuronal injury due to ongoing inflammation.

## Misfolded Protein or Antimicrobial Peptide?

As it has become clear that WNV is associated with long-term neurological deficits, a growing body of research indicates that neurodegenerative diseases may in turn be associated with an infectious trigger. The most prevalent neurodegenerative dementia, AD, currently affects 5.7 million Americans [51]. Hallmark pathological features of AD include amyloid (neuritic) plaques, which are extracellular  $\beta$  amyloid ( $A\beta$ ) aggregates and intraneuronal neurofibrillary tangles (NFTs), which are comprised of hyperphosphorylated and aberrantly folded tau protein [52]. The progressive accumulation of these proteinopathies in selectively vulnerable brain regions, such as the hippocampus and cortex, are associated with increased brain inflammation, neuritic pathology, neurodegeneration, and brain atrophy. The prevailing paradigm for late-onset sporadic forms of AD (the most common manifestation), the Amyloid Cascade Hypothesis, posits that deposition of  $A\beta$  leads to harmful  $A\beta$  aggregates and to the pathological tau alterations that result in NFT which cause neuronal dysfunction and cell death [53]. Clinical trials targeting extracellular  $A\beta$  have successfully reduced levels of  $A\beta$  in the brain, but this has not correlated with an improvement or slowing of clinical progression of AD in most cases, albeit in already symptomatic subjects [54–58]. Though  $A\beta$  is posited to be the initial trigger for AD, most, if not all studies agree that  $A\beta$  is present even in brains of neurologically healthy aged individuals. Also interestingly,  $A\beta$  is conserved in organisms across kingdoms over 400 million years, including prokaryotes [59].

New studies suggest that  $A\beta$  may actually serve a *protective*, innate immune function role as an antimicrobial peptide (AMP) [60]. In vitro, the  $A\beta_{42}$  and  $A\beta_{40}$  (differently cleaved forms of  $A\beta$  peptide) oligomers were shown to potently inhibit the growth of the yeast *Candida albicans*, the gram-negative bacteria *Escherichia coli*, and gram-positive bacteria *Staphylococcus epidermidis*, *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Streptococcus agalactiae*, *Enterococcus faecalis*, and *Listeria monocytogenes*. No inhibitory activity was detected against *Streptococcus mitis*, *Streptococcus pyogenes*, *Streptococcus salivarius*, or *Pseudomonas aeruginosa* [61]. The same group then examined the antimicrobial activity of  $A\beta$  in vivo in mouse and nematode models [62]. They found that  $A\beta$  oligomers bound to microbial cell wall carbohydrates and ultimately entrapped *Salmonella typhimurium* in the mouse model, and similarly interacted with *Candida albicans* in the nematode model.

A separate group went on to demonstrate that  $A\beta$  has antiviral properties as well [63, 64]. Infected neuroglioma H4 cells with herpes simplex virus-1 (HSV-1) resulted in the secretion of  $A\beta_{42}$  and the production of the inflammatory cytokines TNF- $\alpha$  and IL-1 $\beta$ . When they transferred the conditioned media from the HSV-1 infected cells to a de novo neuroglioma culture, the cells were refractory to HSV-1 infection. Subsequently, Eimer et al. went on to show that  $A\beta$  oligomers bound HSV-1 and human herpesvirus 6A and 6B surface glycoproteins, which caused the rapid seeding and deposition of additional  $A\beta$  [65]. Amyloid has been known to be elevated in HIV-infected brains as well and was recently

shown to bind HIV-1 [66]. Interestingly, HIV-1 gag protein promotes secretase-dependent cleavage of the amyloid precursor protein (APP) into A $\beta$ 40 and A $\beta$ 42 isoforms. Viruses that are not known to establish latency in the CNS also are recognized by A $\beta$ . In vitro, respiratory syncytial virus (RSV) was shown to bind amyloid [67], and A $\beta$ 42 was demonstrated to inhibit influenza A viral replication [68]. No studies on A $\beta$  and WNV infection have been reported, but given the broad antimicrobial activity of A $\beta$  and the neuroinvasive nature of WNV, it can be reasonably deduced that there may be increased A $\beta$  production due to WNV infection, as a protective means for viral containment.

Evidence is also mounting for a potential antimicrobial role of  $\alpha$ -synuclein (Asyn), the protein associated with PD. Asyn has been shown to have antimicrobial activity against bacteria (*E. coli*, *S. aureus*) and fungi (*Aspergillus flavus*, *A. fumigatus*, *Rhizoctonia solani*) [69], and infection of Asyn-knockout mice with WNV resulted in dramatically increased viral titers in the brain compared to wild-type control mice, with a 95% mortality rate (vs. 20% in controls) [70••]. The authors replicated these findings with Venezuelan equine encephalitis (VEE) virus (family *Togaviridae*, genus *Alphavirus*) vaccine strain TC-83 in the Asyn-knockout mice; TC-83 is not neuroinvasive in wild-type mice, demonstrating that Asyn restricts neuroinvasion. The authors further linked this to humans by showing increased Asyn expression in primary neuronal cell cultures infected with WNV.

CD-1 outbred mice, when infected with Western equine encephalitis virus (WEEV) and rescued with passive immunotherapy (WEEV E1 antibodies) [71], provided additional evidence of a link between viral infection and neurodegenerative proteinopathies. This served as a model of recovered alphaviral infection, with demonstration of complete viral clearance from the CNS 8 weeks p.i. The recovered mice had Asyn aggregates in the hippocampus, cortex, substantia nigra and mammillary bodies, and deficits consistent with parkinsonism (tremors, dyskinesia, rigidity) as well. The mice also had a 4- to 6-fold upregulation of genes linked to AD (APP, phospholipase D1 and D2, Akt1, and Pten) and PD (Park7, Aldh1a1, Irgm1). Chronic microglial and astrocyte activation, also seen in AD [72] and PD [73], were observed in the recovered brains.

While A $\beta$  and Asyn may function as antimicrobial peptides, when produced in excess, or if clearance mechanisms fail, these proteins can trigger a neurotoxic cascade. Therapeutic interventions to clear these proteins have not yet produced clinical improvement or slowing of decline in AD, though some argue that patients need to undergo therapy earlier, in the presymptomatic state [74]. Anti- $\alpha$ -synuclein therapies (e.g., monoclonal antibodies) are currently in clinical trials [75, 76]; it remains to be seen if these will be successful.

## Microglial Activation and Synaptic Engulfment

The cellular endpoint along the path of neurodegeneration constitutes the loss of neurons and synapses. Direct neuronal injury and death result from the acute phase of neuroinvasive West Nile disease [77]. Following viral clearance, however, progressive neurodegeneration is caused by complement-mediated microglial synaptic engulfment [78••] that shares similarity to the neurotoxic cascades seen in AD [72, 79–82] and PD [73]. Complement activation plays an important role in limiting WNV replication, though it can also lead to excessive

inflammation [83]. All three complement pathways (classical, lectin, and alternative pathways) are engaged during WNV infection and lead to neutralization of virions and lysis of WNV-infected cells [84]. Mice depleted of microglia prior to WNV infection have increased mortality and elevated CNS viral titers following infection, demonstrating that microglia are crucial in limiting WNV disease [85]. However, Vasek et al. recently demonstrated that in mice infected with an attenuated WNV strain, complement C1q in concert with activated microglia led to neurodegeneration [78••]. The authors noted error-prone slower learning in the infected animals when submitted to spatial memory testing, which was attributed to persistent phagocytic microglia that drove complement-mediated (C1qA) CA3 synaptic terminal engulfment and remodeling in the hippocampus. An accompanying examination of postmortem brain tissue from patients with WNV neuroinvasive disease also showed reduced CA3 presynaptic terminals in the hippocampus and in the entorhinal cortex, compared to their age-matched controls [78••].

IFN- $\gamma$  also plays an important role in hippocampal neurogenesis and synapse formation [86, 87]. Garber et al. demonstrated that hippocampus-resident memory WNV-specific T cells secrete IFN- $\gamma$ , which activate microglia; this led to a loss of presynaptic terminals and spatial learning defects in the infected mice [88•]. These studies build on the mounting evidence of the importance of microglia, the brain's resident macrophages, in AD and PD [89–91]. Indeed, it is now appreciated that there is a complex interplay between the immune system and neurodegeneration [92]. Microglia are crucial for pathogen host defense as well as neural circuit pruning [79]. Microglial activation, mediated by the pro-inflammatory cytokines IL-6, TNF- $\alpha$ , and IFN- $\gamma$ , is also critical for clearance of A $\beta$  deposits [93–95], whereas anti-inflammatory cytokines IL-4 and IL-10 actually promote the deposition of A $\beta$  [96, 97]. Proper microglial function is therefore protective in AD, but prolonged activation may lead to synaptotoxicity and neuronal injury [92]. Based on murine models, excessive accumulation of A $\beta$  and hyperphosphorylated tau may induce microglia into this pathological activation [98]. Furthermore, in murine AD models, microglia can induce astrocytes into a neurotoxic state via release of IL-1 $\alpha$ , TNF- $\alpha$ , and C1q complement [98]. In WNV, reactive changes in microglia and astrocytes persist for weeks after viral clearance, which may result in induction of such a pathological state as well, [78••, 99•]. Interestingly, in mice, antiviral CD8+ T cells have been shown to persist for months in the CNS in clusters; these are presumed to be prior infection hot spots, irrespective of persisting antigen [100]. Though controversial in AD, T cells and antigen presentation have been thought to play an important role in the pathogenesis of PD [101].

In PD, pro-inflammatory microglia have also been implicated in dopaminergic neuron loss, mediated by aberrant activation of the complement system [73, 102]. Bodea et al. [102] repeatedly injected mice with bacterial LPS, resulting in elevated plasma levels of TNF- $\alpha$  and IL-113. This, in turn, elevated the levels of these cytokines in the brain and stimulated microglia into a prolonged activated state. Expression of complement C1q, C3, and C4b was upregulated, and there was a 40% loss of dopaminergic neurons in the substantia nigra. In C3-deficient mice, no loss of dopaminergic neurons occurred, suggesting that neuron loss was mediated by complement-mediated glial phagocytosis. This is strikingly similar to the process described in AD and in WNV-infected mice above. Microglia have also been shown to engulf A $\alpha$ syn, though aggregated forms of A $\alpha$ syn may actually inhibit phagocytosis by

blocking FcγR signaling leading to accumulation of extracellular Asyn [103]. In addition, the importance of microglia in PD as well as AD has emerged from genomewide association studies (GWAS), in which the triggering receptor expressed on myeloid cells 2 (TREM2) gene has been identified as a genetic risk factor for both diseases [104, 105]. TREM2, a cell surface receptor expressed on myeloid cells, promotes microglial survival, activation, cytokine release, and phagocytosis [106]. It plays a crucial role in synaptic pruning during postnatal development, but TREM2 signaling may also serve as a switch between a homeostatic and neurodegenerative microglial phenotype [107]. While research on TREM2 in WNV is nascent, Garber et al. noted increased microglial expression of TREM2 isolated from mouse brain infected with an attenuated strain of WNV [99•].

Healthy brain function has been compared to the planetary science concept of a “Goldilocks zone,” in that it may also require an immune environment that is balanced and just right [92]. CNS infections such as WNV cause a sizeable inflammatory response [108], and as such, are highly likely to disturb this delicate balance.

### Disruption of the BBB

The BBB is comprised of vascular endothelial cells and basal lamina lining the microvasculature of the CNS and the pericytes and astrocytic foot processes that surround them. Inter-endothelial tight junctions, lack of fenestration, and specialized transport mechanisms are key features of the BBB [109]. These serve to minimize the entry of pathogens, fluid, and entrained molecules into the CNS. A compromised BBB, therefore, can lead to an influx of pathogens and neurotoxic agents with trigger CNS inflammation and possibly neurodegeneration [110, 111]. Murine studies have demonstrated that amyloid protein upregulates the gelatinase matrix metalloproteinase-9 (MMP-9), which in turn degrades the basal lamina and the tight junction in acute injury (though MMPs also play a role in angiogenesis and remodeling) [112]. A detailed examination was provided in a recent study using a 3D AD BBB model with neurons expressing mutations in the amyloid precursor protein (APP) and PSEN1 genes [113]. This model demonstrated increased BBB permeability due to diminished expression of tight junction proteins, resulting from increased levels of matrix metalloproteinase-2, reactive oxygen species (ROS), interferon- $\gamma$  (IFN- $\gamma$ ), and aggregation of A $\beta$  onto the endothelium [113].

The BBB plays a central role in WNV infection as well. Early host responses are triggered when WNV activates pattern recognition receptors (PRRs), including Toll-like receptors (TLR 3, TLR 7), RIG-I-like receptors (RLRs), and NODlike receptors [114]. This results in the production of type I and III interferons (IFNs) as well as other antiviral cytokines. In mice, type I and III IFNs promote the integrity of the BBB by enhancing tight junction formation, thereby restricting the neuroinvasive capacity of WNV [115]. However, type III IFN (IFN- $\gamma$ ) secreted by T cells during WNV infection compromises the BBB by promoting transendothelial migration of CD4<sup>+</sup> T cells [116] as well as by causing junctional breakdown and cell-cell separation [117]. This is a double-edged sword, in that T cells are crucial for WNV clearance in the CNS [118].



In addition to IFNs, TNF- $\alpha$  mediates BBB permeability in WNV infection. In a murine model of WNV CNS infection,  $\gamma\delta$  T cells proliferate by day 2 after WNV infection. These cells reduce viral burden and inhibit viral CNS infiltration by secretion of IFN- $\gamma$  and TGF- $\beta$ , respectively, but simultaneously increase BBB permeability by producing TNF- $\alpha$  [119]. These pro-inflammatory and anti-inflammatory effector functions are mediated by different  $\gamma\delta$  T cell subsets. In aged mice, V $\gamma$ 4+ cells comprise a larger proportion compared to younger mice, and it is these cells that secrete TNF- $\alpha$ , which compromises the BBB [119]. TNF- $\alpha$  induces BBB endothelial cells to increase expression of adhesion molecules (i.e., selectins, vascular endothelial cell adhesion molecule-1 (VCAM-1), intercellular cell adhesion molecule-1 (ICAM-1)) [120]. This leads to endothelial adhesion of circulating leukocytes, which in turn produce reactive oxygen species and proteases that damage the microvessels [121]. Activated microglia also produce TNF- $\alpha$ , as well as IL-1 $\beta$ , reactive oxygen species, and nitric oxide, all of which contribute to enhancing BBB permeability [122].

Thus, WNV infection leads to the production of cytokines that promote BBB integrity as well as permeability, and host factors, such as age, modulate the balance of the two. Indeed, normal aging in itself appears to be associated with BBB breakdown [123]. Compared to young mice, aged mice demonstrated significantly higher levels of inflammation (TNF- $\alpha$ ), leading to extravasation of circulating IgG in the cortex and hippocampus—mostly attributable to diminished tight junction complex expression [123]. Hypertension is also a significant factor in BBB breakdown, mediated by angiotensin II-driven downregulation of key BBB protein constituents (endothelial barrier antigen, transferrin receptor) [124]. Diabetes mellitus also impairs the BBB integrity [125]; in vitro models of the BBB under hyperglycemic conditions demonstrated the expression of numerous pro-inflammatory cytokines (TNF- $\alpha$ , IL-6, IL-1, and IL-4) in astrocytes [125], which may also act to compromise the tight junction complex. It is no coincidence, then, that WNV neuroinvasive disease occurs predominantly in the elderly, and that hypertension and diabetes mellitus are prominent risk factors as well [126–128].

From this, we can surmise that a compromised BBB will be more permissive to the entry of WNV as well as other microbes and injurious molecules, which in turn activate microglia and lead to the upregulation of A $\beta$  and/or A $\gamma$  as antimicrobial peptides. Persistent microglial activation and excess production, or lack of clearance of A $\beta$  and/or A $\gamma$  may lead to complement-mediated synaptotoxicity, and ultimately neurodegeneration.

## Conclusion

The frequency with which cognitive and psychomotor deficits persist long after WNV exposure, coupled with the continued transmission of WNV in the USA and worldwide, underscores the importance of understanding the pathways underlying these impairments. Furthermore, the world is experiencing the emergence of ongoing outbreaks due to other neuroinvasive arboviruses such as Zika virus, eastern equine encephalitis virus, VEE virus, and Japanese encephalitis virus, to name just a few. Many of these are also associated with long-term neurological sequelae [129,130]. While there are no therapies nor prognoses

available at present, critical progress has been made in recent years to assess the relationship between neuroinvasive viral infection and neurodegenerative diseases.

It is becoming clear that there are convergent mechanisms resulting in neuronal loss [131]. The data thus far suggest that hosts who are older, have diabetes, hypertension, or other disorders associated with BBB-damage are more susceptible to neuroinvasive WNV infection and subsequent neurological sequelae, as well as to neurodegenerative diseases. In the case of WNV, this may be mediated by enhanced viral entry into the CNS, but additional factors play an important role in residual cognitive and psychomotor impairments. Abnormal BBB permeability may lead to CNS entry of peripheral antigens that activate microglia, which then precipitates a neurodegenerative phenotype that results in synaptotoxicity [132, 133]. Infection stimulates production of A $\beta$  and A $\alpha$ syn, which aggregate in the setting of a viral or bacterial brain infection [59, 60]. The inciting infection and initial host response lead to an aberrant inflammatory response, excess elevation of A $\beta$  and/or A $\alpha$ syn, and in time, neurodegenerative changes [59] (Box 1, Fig. 1).

This proposed sequence is not limited to WNV. Early work connecting an infectious agent to AD was performed by Itzhaki et al. in the 1990s, who examined the role of herpes simplex virus-1 (HSV-1) in AD [134, 135]. Her group found that the presence of HSV-1 DNA, coupled with the presence of an apolipoprotein E4 allele (apoE; apoE4 is strongly associated with AD), was correlated with AD. A subsequent population-based study examined the association between HSV-1 and AD, reporting twice the risk for AD in reactivated HSV-1 (defined by the presence of HSV-1 IgM) as well as an interaction with ApoE4 [136, 137]. An expanding suite of pathogens has since been identified that promotes this pattern of neurodegeneration, including spirochetes [138], HIV [139–141], *Chlamydia pneumoniae* [142], *Porphyromonas gingivalis* [143], human herpesvirus-6A (HHV-6A), and HHV-7 [144]. Post-encephalitic parkinsonism and encephalitis lethargica, conditions that significantly overlap with PD, have been associated with infectious etiologies for a century [145]. In this sense, an infectious association with PD has long been entertained. Specific pathogens that have been considered include influenza A virus [146], *Porphyromonas gingivalis* [147], *Helicobacter pylori* [148], hepatitis C virus [149], and gram-negative bacterial gastrointestinal infections [150].

Moving forward, studies that further our mechanistic understanding are crucial. ApoE is of particular interest in its potential ability to modify the effect of a neurotropic virus. Apolipoprotein E (ApoE) is a cholesterol carrier that binds to cell surface receptors to deliver lipids as well as the A $\beta$  peptide in the CNS [151]. The apolipoprotein E E4 allele, which confers a strong risk for AD, has also been associated with higher CNS viral loads [152]. In a murine model of acute HSV-1 infection, apoE4 mice had a 10-fold increased number of HSV-1 genomic copies compared to apoE3 mice, and apoE3 mice had in turn a 10-fold higher number of genomic copies than mice lacking apoE [152]. The HSV-1 genomic copy number is directly linked to the risk of reactivation [153], thus explaining how apoE may influence HSV-1 reactivation. While no such connection has been made to WNV, the capsid protein of another flavivirus, dengue virus, has been shown to bind to the apoE component of very low-density lipoprotein (VLDL) and thought to thus “hitch a ride” to distant organs [154].

Looking to modify disease outcomes, could we learn about neuroprotective factors and resilience from people fully recovered from neuroinvasive pathogens? What mechanisms promote a return to microglial homeostasis in such individuals? And while there is epidemiological and MRI evidence for cognitive and other neurological deficits in WNV-exposed patients suggesting syndromes similar to those seen in neurodegenerative diseases, data on WNV exposure in people with known AD or PD are lacking. In AD as well as psychiatric diseases, efforts to study the “brain microbiome” are expanding with the increasing recognition that microbes are permanent residents of the CNS and can also distally influence a wide range of neurological functions [155]. Such studies will help to establish the role played by latent microbes such as HSV-1, HHV-6A/B, and HHV-7. However, these studies will not address infections that are transient but leave a lasting imprint, as described for WNV. To understand the impact of these types of pathogens, carefully designed epidemiological studies will prove important in assessing the magnitude of the link between neuroinvasive viral exposure and subsequent neurodegeneration. Bringing together experts in infectious disease, neuroscience and neurology will help deepen our understanding of the intricacies of how pathogens may trigger neurodegenerative cascades, ultimately with the aim of improving outcomes for people living with chronic sequelae of infectious diseases as well as patients devastated by dementias.

## References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
  - Of major importance
1. Chancey C, Grinev A, Volkova E, Rios M. The global ecology and epidemiology of West Nile virus. *Biomed Res Int*. 2015;2015: 376230. [PubMed: 25866777]
  2. Petersen LR, Carson PJ, Biggerstaff BJ, Custer B, Borchardt SM, Busch MP. Estimated cumulative incidence of West Nile virus infection in US adults, 1999-2010. *Epidemiol Infect*. 2013;141(3):591–5. [PubMed: 22640592]
  3. Petersen LR, Brault AC, Nasci RS. West Nile virus: review of the literature. *JAMA*. 2013;310(3):308–15. [PubMed: 23860989]
  4. Murray KO, Garcia MN, Rahbar MH, Martinez D, Khuwaja SA, Arafat RR, et al. Survival analysis, long-term outcomes, and percentage of recovery up to 8 years post-infection among the Houston West Nile virus cohort. *PLoS One*. 2014;9(7):e102953. [PubMed: 25054656]
  5. Carson PJ, Konewko P, Wold KS, Mariani P, Goli S, Bergloff P, et al. Long-term clinical and neuropsychological outcomes of West Nile virus infection. *Clin Infect Dis*. 2006;43(6):723–30. [PubMed: 16912946]
  6. Heneka MT, Golenbock DT, Latz E. Innate immunity in Alzheimer’s disease. *Nat Immunol*. 2015;16(3):229–36. [PubMed: 25689443]
  7. Sejvar JJ, Haddad MB, Tierney BC, Campbell GL, Marfin AA, Van Gerpen JA, et al. Neurologic manifestations and outcome of West Nile virus infection. *JAMA*. 2003;290(4):511–5. [PubMed: 12876094]
  8. Klee AL, Maidin B, Edwin B, Poshni I, Mostashari F, Fine A, et al. Long-term prognosis for clinical West Nile virus infection. *Emerg Infect Dis*. 2004;10(8):1405–11. [PubMed: 15496241]
  9. Ou AC, Ratard RC. One-year sequelae in patients with West Nile Virus encephalitis and meningitis in Louisiana. *J La State Med Soc*. 2005;157(1):42–6. [PubMed: 15887668]

10. Watson JT, Pertel PE, Jones RC, Siston AM, Paul WS, Austin CC, et al. Clinical characteristics and functional outcomes of West Nile fever. *Ann Intern Med.* 2004;141(5):360–5. [PubMed: 15353427]
11. Samaan Z, McDermid Vaz S, Bawor M, Potter TH, Eskandarian S, Loeb M. Neuropsychological impact of West Nile virus infection: an extensive neuropsychiatric assessment of 49 cases in Canada. *PLoS One.* 2016;11(6):e0158364. [PubMed: 27352145]
12. Kathleen Y, Haaland JS, Pergam S, Echevarria LA, Davis LE, Goade D, et al. Mental status after West Nile virus infection. *Emerg Infect Dis.* 2006;12(8):1260–2. [PubMed: 16965710]
13. Sadek JR, Pergam SA, Harrington JA, Echevarria LA, Davis LE, Goade D, et al. Persistent neuropsychological impairment associated with West Nile virus infection. *J Clin Exp Neuropsychol.* 2010;32(1):81–7. [PubMed: 19513920]
14. Murray KO, Nolan MS, Ronca SE, Datta S, Govindarajan K, Narayana PA, et al. The neurocognitive and MRI outcomes of West Nile Virus infection: preliminary analysis using an external control group. *Front Neurol.* 2018;9:111 [PubMed: 29636722] This study examines neuropsychological testing and MRI findings in patients an average of 4 years after their initial illness onset. Normative controls are used as a comparison group for the MRI portion of the study. The authors report significant frontal, temporal, and limbic cortical thinning in the WNV participants.
15. Sejvar JJ, Curns AT, Welburg L, Jones JF, Lundgren LM, Capuron L, et al. Neurocognitive and functional outcomes in persons recovering from West Nile virus illness. *J Neuropsychol.* 2008;2(Pt 2):477–99. [PubMed: 19824176]
16. Sheppard DP, Woods SP, Hasbun R, Salazar L, Nolan MS, Murray KO. Does intra-individual neurocognitive variability relate to neuroinvasive disease and quality of life in West Nile Virus? *J Neuro-Oncol.* 2018;24(4):506–13.
17. Balakrishnan A, Thekkekara RJ, Tandale BV. Outcomes of West Nile encephalitis patients after 1 year of West Nile encephalitis outbreak in Kerala, India: a follow-up study. *J Med Virol.* 2016;88(11):1856–61. [PubMed: 27061922]
18. Fray PJ, Robbins TW. CANTAB battery: proposed utility in neurotoxicology. *Neurotoxicol Teratol.* 1996;18(4):499–504. [PubMed: 8866544]
19. Rabadi MH. Brought down by a mosquito? West Nile Virus encephalitis. *Am J Med.* 2018;131(9):1064–6. [PubMed: 29571980]
20. Pradhan S, Anand S, Choudhury SS. Cognitive behavioural impairment with irreversible sensorineural deafness as a complication of West Nile encephalitis. *J Neurovirol.* 2019;25(3):429–33. [PubMed: 30903400]
21. Lyons JL, Schaefer PW, Cho TA, Azar MM. Case 34-2017. A 76-year-old man with fever, weight loss, and weakness. *N Engl J Med.* 2017;377(19):1878–86. [PubMed: 29117489]
22. Guth JC, Futterer SA, Hijaz TA, Liotta EM, Rosenberg NF, Naidech AM, et al. Pearls & oysters: bilateral thalamic involvement in West Nile virus encephalitis. *Neurology.* 2014;83(2):e16–7. [PubMed: 25002571]
23. Brilla R, Block M, Geremia G, Wichter M. Clinical and neuroradiologic features of 39 consecutive cases of West Nile Virus meningoencephalitis. *J Neurol Sci.* 2004;220(1–2):37–40. [PubMed: 15140603]
24. Racsa L, Gander R, Chung W, Southern P, Le J, Beal S, et al. Clinical features of West Nile virus epidemic in Dallas, Texas, 2012. *Diagn Microbiol Infect Dis.* 2014;78(2):132–6. [PubMed: 24316017]
25. Ocal M, onder H, Arsava EM, Alp S, Ozkul A, Ergunay K. A case of central nervous system infection due to West Nile Virus lineage-1 in Ankara province, Turkey. *Mikrobiyol Bul.* 2013;47(1):164–72. [PubMed: 23390915]
26. Mainali S, Afshani M, Wood JB, Levin MC. The natural history of West Nile virus infection presenting with West Nile virus meningoencephalitis in a man with a prolonged illness: a case report. *J Med Case Rep.* 2011;5:204. [PubMed: 21612601]
27. Abraham A, Ziv S, Drory VE. Posterior reversible encephalopathy syndrome resulting from Guillain-Barre-like syndrome secondary to West Nile virus infection. *J Clin Neuromuscul Dis.* 2011;12(3): 113–7. [PubMed: 21321488]

28. Jain N, Fisk D, Sotir M, Kehl KS. West Nile encephalitis, status epilepticus and West Nile pneumonia in a renal transplant patient. *Transpl Int.* 2007;20(9):800–3. [PubMed: 17630998]
29. Bosanko CM, Gilroy J, Wang AM, Sanders W, Dulai M, Wilson J, et al. West Nile Virus encephalitis involving the substantia nigra: neuroimaging and pathologic findings with literature review. *Arch Neurol.* 2003;60(10):1448–52. [PubMed: 14568817]
30. DeBiasi RL, Parsons JA, Grabert BE. West Nile virus meningoencephalitis in an immunocompetent adolescent. *Pediatr Neurol.* 2005;33(3):217–9. [PubMed: 16139740]
31. Ali M, Safriel Y, Sohi J, Llave A, Weathers S. West Nile virus infection: MR imaging findings in the nervous system. *AJNR Am J Neuroradiol.* 2005;26(2):289–97. [PubMed: 15709126]
32. Rosas H, Wippold FJ 2nd. West Nile virus: case report with MR imaging findings. *AJNR Am J Neuroradiol.* 2003;24(7):1376–8. [PubMed: 12917131]
33. Brener ZZ, et al. Acute renal failure in a patient with West Nile viral encephalitis. *Nephrol Dial Transplant.* 2007;22(2):662–3. [PubMed: 17012739]
34. Shepherd JC, Subramanian A, Montgomery RA, Samaniego MD, Gong G, Bergmann A, et al. West Nile virus encephalitis in a kidney transplant recipient. *Am J Transplant.* 2004;4(5):830–3. [PubMed: 15084182]
35. Farnaes L, Schiff D, McElroy A, Coufal NG, Crawford JR, Cannavino C. Encephalitis and thalamic injury from neuroinvasive West Nile Virus in children on treatment for acute lymphoblastic leukemia. *Pediatr Neurol.* 2018;80:84–7. [PubMed: 29398166]
36. Petropoulou KA, Gordon SM, Prayson RA, Ruggieri PM. West Nile virus meningoencephalitis: MR imaging findings. *AJNR Am J Neuroradiol.* 2005;26(8):1986–95. [PubMed: 16155147]
37. Hwang J, Ryu H-S, Kim H, Lee S-A. The first reported case of West Nile encephalitis in Korea. *J Korean Med Sci.* 2015;30(3):343–5. [PubMed: 25729260]
38. Kadkhoda K, Embil JM, McKibbin LR, McEachern J, Michael A, Drebot West Nile Virus infection in a renal transplant recipient resulting in polioencephalomyelitis, quadriplegia, and global brain atrophy. *IDCases.* 2019;17:e00551. [PubMed: 31193054]
39. Ledig C, Schuh A, Guerrero R, Heckemann RA, Rueckert D. Structural brain imaging in Alzheimer's disease and mild cognitive impairment: biomarker analysis and shared morphometry database. *Sci Rep.* 2018;8(1):11258. [PubMed: 30050078]
40. Helmich RC, Vaillancourt DE, Brooks DJ. The future of brain imaging in Parkinson's disease. *J Park Dis.* 2018;8(s1):S47–51.
41. Verma S, Lo Y, Chapagain M, Lum S, Kumar M, Gurjav U, et al. West Nile virus infection modulates human brain microvascular endothelial cells tight junction proteins and cell adhesion molecules: transmigration across the in vitro blood-brain barrier. *Virology.* 2009;385(2):425–33. [PubMed: 19135695]
42. Diamond MS, Shrestha B, Mehlhop E, Sitati E, Engle M. Innate and adaptive immune responses determine protection against disseminated infection by West Nile encephalitis virus. *Viral Immunol.* 2003;16(3):259–78. [PubMed: 14583143]
43. Samuel MA, Wang H, Siddharthan V, Morrey JD, Diamond MS. Axonal transport mediates West Nile virus entry into the central nervous system and induces acute flaccid paralysis. *Proc Natl Acad Sci U S A.* 2007;104(43):17140–5. [PubMed: 17939996]
44. Shrestha B, Gottlieb D, Diamond MS. Infection and injury of neurons by West Nile encephalitis virus. *J Virol.* 2003;77(24): 13203–13. [PubMed: 14645577]
45. Hayes EB, Sejvar JJ, Zaki SR, Lanciotti RS, Bode AV, Campbell GL. Virology, pathology, and clinical manifestations of West Nile virus disease. *Emerg Infect Dis.* 2005;11(8):1174–9. [PubMed: 16102303]
46. Diniz JA, da Rosa AP, Guzman H, Xu F, Xiao SY, Popov VL, et al. West Nile virus infection of primary mouse neuronal and neuroglial cells: the role of astrocytes in chronic infection. *Am J Trop Med Hyg.* 2006;75(4):691–6. [PubMed: 17038696]
47. Lesteborg KE, Beckham JD. Immunology of West Nile virus infection and the role of alpha-synuclein as a viral restriction factor. *Viral Immunol.* 2019;32(1):38–47. [PubMed: 30222521]
48. Suthar MS, Diamond MS, Gale M Jr. West Nile virus infection and immunity. *Nat Rev Microbiol.* 2013;11(2):115–28. [PubMed: 23321534]

49. Appler KK, Brown AN, Stewart BS, Behr MJ, Demarest VL, Wong SJ, et al. Persistence of West Nile virus in the central nervous system and periphery of mice. *PLoS One*. 2010;5(5):e10649.
50. Stewart BS, Demarest VL, Wong SJ, Green S, Bernard KA. Persistence of virus-specific immune responses in the central nervous system of mice after West Nile virus infection. *BMC Immunol*. 2011;12:6. [PubMed: 21251256]
51. Matthews KA, Xu W, Gaglioti AH, Holt JB, Croft JB, Mack D, et al. Racial and ethnic estimates of Alzheimer's disease and related dementias in the United States (2015-2060) in adults aged  $\geq$ 65 years. *Alzheimers Dement*. 2019;15(1):17–24. [PubMed: 30243772]
52. Nelson PT, Alafuzoff I, Bigio EH, Bouras C, Braak H, Cairns NJ, et al. Correlation of Alzheimer disease neuropathologic changes with cognitive status: a review of the literature. *J Neuroopathol Exp Neurol*. 2012;71(5):362–81. [PubMed: 22487856]
53. Selkoe DJ. The molecular pathology of Alzheimer's disease. *Neuron*. 1991;6(4):487–98. [PubMed: 1673054]
54. Doody RS, Farlow M, Aisen PS, Alzheimer's Disease Cooperative Study Data Analysis and Publication Committee. Phase 3 trials of solanezumab and bapineuzumab for Alzheimer's disease. *N Engl J Med*. 2014;370(15):1460.
55. Doody RS, Raman R, Farlow M, Iwatsubo T, Vellas B, Joffe S, et al. Alzheimer's Disease Cooperative Study Steering Committee, Siemers E, Sethuraman G, Mohs R; Semagacestat Study Group. A phase 3 trial of semagacestat for treatment of Alzheimer's disease. *N Engl J Med*. 2013;369(4):341–50. [PubMed: 23883379]
56. Doody RS, Thomas RG, Farlow M, Iwatsubo T, Vellas B, Joffe S, et al. Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease. *N Engl J Med*. 2014;370(4):311–21. [PubMed: 24450890]
57. LaMotte S Another promising Alzheimer's drug trial ends in failure: This one hurts: *CNN Health*; 2019.
58. Salloway S, Sperling R, Gilman S, Fox NC, Blennow K, Raskind M, et al. A phase 2 multiple ascending dose trial of bapineuzumab in mild to moderate Alzheimer disease. *Neurology*. 2009;73(24): 2061–70. [PubMed: 19923550]
59. Gosztyla ML, Brothers HM, Robinson SR. Alzheimer's amyloid-beta is an antimicrobial peptide: a review of the evidence. *J Alzheimers Dis*. 2018;62(4):1495–506. [PubMed: 29504537]
60. Moir RD, Lathe R, Tanzi RE. The antimicrobial protection hypothesis of Alzheimer's disease. *Alzheimers Dement*. 2018;14(12):1602–14. [PubMed: 30314800]
61. Socia SJ, Kirby JE, Washicosky KJ, Tucker SM, Ingelsson M, Hyman B, et al. The Alzheimer's disease-associated amyloid beta-protein is an antimicrobial peptide. *PLoS One*. 2010;5(3):e9505. [PubMed: 20209079]
62. Kumar DK, Choi SH, Washicosky KJ, Eimer WA, Tucker S, Ghofrani J, et al. Moir RD Amyloid-beta peptide protects against microbial infection in mouse and worm models of Alzheimer's disease. *Sci Transl Med*. 2016;8(340):340–72.
63. Bourgade K, Dupuis G, Frost EH, Fulop T. Anti-viral properties of amyloid-beta peptides. *J Alzheimers Dis*. 2016;54(3):859–78. [PubMed: 27392871]
64. Bourgade K, Le Page A, Bocti C, Witkowski JM, Dupuis G, Frost EH, et al. Protective effect of amyloid-beta peptides against herpes simplex virus-1 infection in a neuronal cell culture model. *J Alzheimers Dis*. 2016;50(4):1227–41. [PubMed: 26836158]
65. Eimer WA, Vijaya Kumar DK, Navalpur Shanmugam NK, Rodriguez AS, Mitchell T, Washicosky KJ, et al. Alzheimer's disease-associated beta-amyloid is rapidly seeded by herpesviridae to protect against brain infection. *Neuron*. 2018;100(6):1527–32. [PubMed: 30571943]
66. Chai Q, Jovasevic V, Malikov V, Sabo Y, Morham S, Walsh D, et al. HIV-1 counteracts an innate restriction by amyloid precursor protein resulting in neurodegeneration. *Nat Commun*. 2017;8(1):1522. [PubMed: 29142315]
67. Ezzat K, Pernemalm M, Palsson S, Roberts TC, Jarver P, Dondalska A, et al. The viral protein corona directs viral pathogenesis and amyloid aggregation. *Nat Commun*. 2019;10(1): 2331. [PubMed: 31133680]

68. White MR, Kandel R, Tripathi S, Condon D, Qi L, Taubenberger J, et al. Alzheimer's associated beta-amyloid protein inhibits influenza A virus and modulates viral interactions with phagocytes. *PLoS One*. 2014;9(7):e101364. [PubMed: 24988208]
69. Park SC, Moon JC, Shin SY, Son H, Jung YJ, Kim NH, et al. Functional characterization of alpha-synuclein protein with antimicrobial activity. *Biochem Biophys Res Commun*. 2016;478(2): 924–8. [PubMed: 27520375]
- 70••. Beatman EL, Massey A, Shives KD, Burrack KS, Chamanian M, Morrison TE, et al. Alpha-synuclein expression restricts RNA viral infections in the brain. *J Virol*. 2015;90(6):2767–82 [PubMed: 26719256] This murine study shows that neuronal alpha-synuclein inhibited WNV infection in the CNS, and that deletion of the gene for alpha-synuclein gene resulted in decreased survival and increased CNS viral burden and neuronal injury. This article speaks directly to the hypothesis that asyn is an antimicrobial peptide.
71. Bantle CM, Phillips AT, Smeyne RJ, Rocha SM, Olson KE, Tjalkens RB. Infection with mosquito-borne alphavirus induces selective loss of dopaminergic neurons, neuroinflammation and widespread protein aggregation. *NPJ Parkinsons Dis*. 2019;5:20. [PubMed: 31531390]
72. Hong S, Beja-Glasser VF, Nfonoyim BM, Frouin A, Li S, Ramakrishnan S, et al. Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science*. 2016;352(6286):712–6. [PubMed: 27033548]
73. Tremblay ME, Cookson MR, Civiero L. Glial phagocytic clearance in Parkinson's disease. *Mol Neurodegener*. 2019;14(1):16. [PubMed: 30953527]
74. Golde TE, DeKosky ST, Galasko D. Alzheimer's disease: the right drug, the right time. *Science*. 2018;362(6420):1250–1. [PubMed: 30545877]
75. Jankovic J, Goodman I, Safirstein B, Marmon TK, Schenk DB, Koller M, et al. Safety and tolerability of multiple ascending doses of PRX002/RG7935, an anti-alpha-synuclein monoclonal antibody, in patients with Parkinson disease: a randomized clinical trial. *JAMA Neurol*. 2018;75(10):1206–14. [PubMed: 29913017]
76. Brys M, Fanning L, Hung S, Ellenbogen A, Penner N, Yang M, et al. Randomized phase I clinical trial of anti-alpha-synuclein antibody BIIB054. *Mov Disord*. 2019;34(8):1154–63. [PubMed: 31211448]
77. Chambers TJ, Diamond MS. Pathogenesis of flavivirus encephalitis. *Adv Virus Res*. 2003;60:273–342. [PubMed: 14689697]
- 78••. Vasek MJ, Garber C, Dorsey D, Durrant DM, Bollman B, Soung A, et al. A complement-microglial axis drives synapse loss during virus-induced memory impairment. *Nature*. 2016;534(7608):538–43 [PubMed: 27337340] This murine study found impaired spatial learning, attributable to persistent phagocytic microglia that drove complement-mediated synaptic loss in the hippocampus following infection with an attenuated WNV strain. An accompanying examination of human postmortem brain tissue showed reduced CA3 presynaptic terminals in the hippocampus and entorhinal cortex associated with WNV neuroinvasive disease, compared to age-matched controls.
79. Schafer DP, Lehrman EK, Kautzman AG, Koyama R, Mardinly AR, Yamasaki R, et al. Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron*. 2012;74(4):691–705. [PubMed: 22632727]
80. Wu T, Dejanovic B, Gandham VD, Gogineni A, Edmonds R, Schauer S, et al. Complement C3 is activated in human AD brain and is required for neurodegeneration in mouse models of amyloidosis and tauopathy. *Cell Rep*. 2019;28(8):2111–23 e6. [PubMed: 31433986]
81. Lian H, Yang L, Cole A, Sun L, Chiang AC, Fowler SW, et al. NFκB-activated astroglial release of complement C3 compromises neuronal morphology and function associated with Alzheimer's disease. *Neuron*. 2015;85(1):101–15. [PubMed: 25533482]
82. Canchi S, Raaouf B, Masliah D, Rosenthal SB, Sasik R, Fisch KM, et al. Integrating gene and protein expression reveals perturbed functional networks in Alzheimer's disease. *Cell Rep*. 2019;28(4):1103–16 e4. [PubMed: 31340147]
83. Conde JN, Silva EM, Barbosa AS, Mohana-Borges R. The complement system in Flavivirus infections. *Front Microbiol*. 2017;8:213. [PubMed: 28261172]

84. Mehlhop E, Whitby K, Oliphant T, Marri A, Engle M, Diamond MS, et al. Complement activation is required for induction of a protective antibody response against West Nile virus infection. *J Virol.* 2005;79(12):7466–77. [PubMed: 15919902]
85. Seitz S, Clarke P, Tyler KL. Pharmacologic depletion of microglia increases viral load in the brain and enhances mortality in murine models of flavivirus-induced encephalitis. *J Virol.* 2018;92(16).
86. Kim IJ, Beck HN, Lein PJ, Higgins D. Interferon gamma induces retrograde dendritic retraction and inhibits synapse formation. *J Neurosci.* 2002;22(11):4530–9. [PubMed: 12040060]
87. Li L, Walker TL, Zhang Y, Mackay EW, Bartlett PF. Endogenous interferon gamma directly regulates neural precursors in the noninflammatory brain. *J Neurosci.* 2010;30(27):9038–50. [PubMed: 20610738]
- 88•. Garber C, Soung A, Vollmer LL, Kanmogne M, Last A, Brown J, et al. Tcells promotemicroglia-mediated synaptic elimination and cognitive dysfunction during recovery from neuropathogenic flaviviruses. *NatNeurosci.* 2019;22(8):1276–88In a murine model of West Nile neuroinvasive disease (WNND)-induced cognitive dysfunction, the authors demonstrate that T cell-derived IFN- $\gamma$  signaling in microglia gives rise to spatial-learning defects and presynaptic termini elimination with lack of repair.
89. Nott A, Holtman IR, Coufal NG, CM SJ, Yu M, Hu R, et al. Brain cell type-specific enhancer-promoter interactome maps and disease risk association. *Science.* 2019.
90. Labzin LI, Heneka MT, Latz E. Innate immunity and neurodegeneration. *Annu Rev Med.* 2018;69:437–49. [PubMed: 29106805]
91. Salter MW, Stevens B. Microglia emerge as central players in brain disease. *Nat Med.* 2017;23(9):1018–27. [PubMed: 28886007]
92. Golde TE. Harnessing immunoproteostasis to treat neurodegenerative disorders. *Neuron.* 2019;101(6):1003–15. [PubMed: 30897353]
93. Chakrabarty P, Ceballos-Diaz C, Beccard A, Janus C, Dickson D, Golde TE, et al. IFN-gamma promotes complement expression and attenuates amyloid plaque deposition in amyloid beta precursor protein transgenic mice. *J Immunol.* 2010;184(9):5333–43. [PubMed: 20368278]
94. Chakrabarty P, Herring A, Ceballos-Diaz C, Das P, Golde TE. Hippocampal expression of murine TNF $\alpha$  results in attenuation of amyloid deposition in vivo. *Mol Neurodegener.* 2011;6:16. [PubMed: 21324189]
95. Chakrabarty P, Jansen-West K, Beccard A, Ceballos-Diaz C, Levites Y, Verbeeck C, et al. Massive gliosis induced by interleukin-6 suppresses A $\beta$  deposition in vivo: evidence against inflammation as a driving force for amyloid deposition. *FASEB J.* 2010;24(2):548–59. [PubMed: 19825975]
96. Chakrabarty P, Li A, Ceballos-Diaz C, Eddy JA, Funk CC, Moore B, et al. IL-10 alters immunoproteostasis in APP mice, increasing plaque burden and worsening cognitive behavior. *Neuron.* 2015;85(3):519–33. [PubMed: 25619653]
97. Chakrabarty P, Tianbai L, Herring A, Ceballos-Diaz C, Das P, Golde TE. Hippocampal expression of murine IL-4 results in exacerbation of amyloid deposition. *Mol Neurodegener.* 2012;7:36. [PubMed: 22838967]
98. Hansen DV, Hanson JE, Sheng M. Microglia in Alzheimer’s disease. *J Cell Biol.* 2018;217(2):459–72. [PubMed: 29196460]
- 99•. Garber C, Vasek MJ, Vollmer LL, Sun T, Jiang X, Klein RS. Astrocytes decrease adult neurogenesis during virus-induced memory dysfunction via IL-1. *Nat Immunol.* 2018;19(2):151–61 [PubMed: 29292385] In a murine model of WNND-induced cognitive dysfunction, the authors demonstrate that there are fewer neuroblasts and increased astrogenesis without recovery of hippocampal neurogenesis at 30 days, mediated by IL-1 secreted predominantly by pro-inflammatory astrocytes.
100. Wakim LM, Woodward-Davis A, Bevan MJ. Memory T cells persisting within the brain after local infection show functional adaptations to their tissue of residence. *Proc Natl Acad Sci U S A.* 2010;107(42):17872–9. [PubMed: 20923878]
101. Sulzer D, Alcalay RN, Garretti F, Cote L, Kanter E, Agin-Liebes J, et al. T cells from patients with Parkinson’s disease recognize alpha-synuclein peptides. *Nature.* 2017;546(7660):656–61. [PubMed: 28636593]



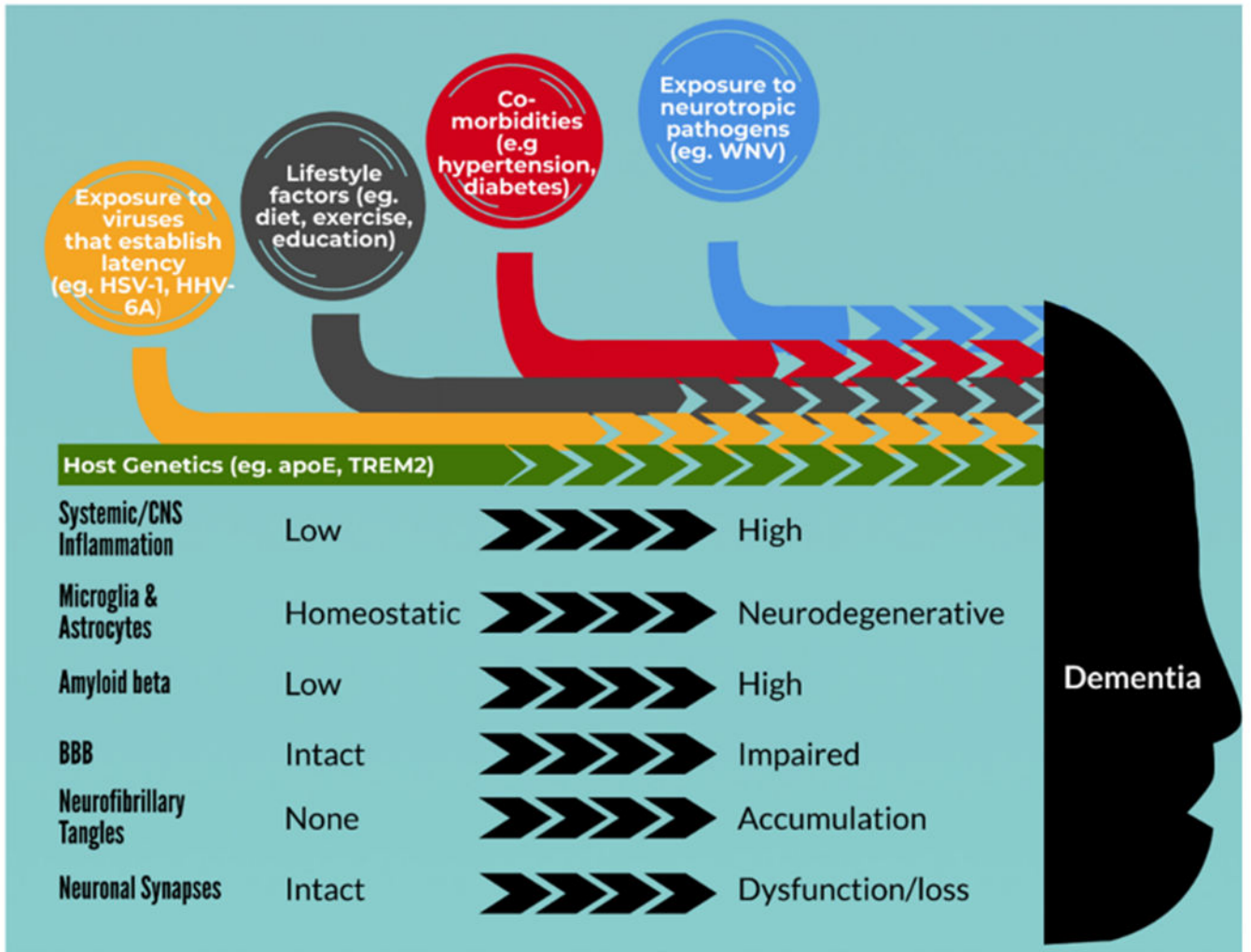
102. Bodea LG, Wang Y, Linnartz-Gerlach B, Kopatz J, Sinkkonen L, Musgrove R, et al. Neurodegeneration by activation of the microglial complement-phagosome pathway. *J Neurosci*. 2014;34(25):8546–56. [PubMed: 24948809]
103. Choi YR, Kang SJ, Kim JM, Lee SJ, Jou I, Joe EH, et al. FcyRIIB mediates the inhibitory effect of aggregated  $\alpha$ -synuclein on microglial phagocytosis. *Neurobiol Dis*. 2015;83:90–9. [PubMed: 26342897]
104. Lill CM, Rengmark A, Pihlstrom L, Fogh I, Shatunov A, Sleiman PM, et al. The role of TREM2 R47H as a risk factor for Alzheimer's disease, frontotemporal lobar degeneration, amyotrophic lateral sclerosis, and Parkinson's disease. *Alzheimers Dement*. 2015;11(12):1407–16. [PubMed: 25936935]
105. Rayaprolu S, Mullen B, Baker M, Lynch T, Finger E, Seeley WW, et al. TREM2 in neurodegeneration: evidence for association of the p.R47H variant with frontotemporal dementia and Parkinson's disease. *Mol Neurodegener*. 2013;8:19. [PubMed: 23800361]
106. Colonna M, Wang Y. TREM2 variants: new keys to decipher Alzheimer disease pathogenesis. *Nat Rev Neurosci*. 2016;17(4): 201–7. [PubMed: 26911435]
107. Krasemann S, Madore C, Cialic R, Baufeld C, Calcagno N, el Fatimy R, et al. The TREM2-APOE pathway drives the transcriptional phenotype of dysfunctional microglia in neurodegenerative diseases. *Immunity*. 2017;47(3):566–81 e9. [PubMed: 28930663]
108. Schafer A, Brooke CB, Whitmore AC, Johnston RE. The role of the blood-brain barrier during Venezuelan equine encephalitis virus infection. *J Virol*. 2011;85(20):10682–90. [PubMed: 21849461]
109. Obermeier B, Verma A, Ransohoff RM. The blood-brain barrier. *Handb Clin Neurol*. 2016;133:39–59. [PubMed: 27112670]
110. Sweeney MD, Sagare AP, Zlokovic BV. Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders. *Nat Rev Neurol*. 2018;14(3):133–50. [PubMed: 29377008]
111. Sweeney MD, Kisler K, Montagne A, Toga AW, Zlokovic BV. The role of brain vasculature in neurodegenerative disorders. *Nat Neurosci*. 2018;21(10):1318–31. [PubMed: 30250261]
112. Seo JH, Guo S, Lok J, Navaratna D, Whalen MJ, Kim KW, et al. Neurovascular matrix metalloproteinases and the blood-brain barrier. *Curr PharmDes*. 2012;18(25):3645–8.
113. Shin Y, Choi SH, Kim E, Bylykbashi E, Kim JA, Chung S, et al. Blood-brain barrier dysfunction in a 3D in vitro model of Alzheimer's disease. *Adv Sci (Weinh)*. 2019;6(20):1900962. [PubMed: 31637161]
114. Luo H, Wang T. Recent advances in understanding West Nile virus host immunity and viral pathogenesis. *F1000Res*. 2018;7:338. [PubMed: 29623197]
115. Daniels BP, Holman DW, Cruz-Orengo L, Jujavarapu H, Durrant DM, Klein RS. Viral pathogen-associated molecular patterns regulate blood-brain barrier integrity via competing innate cytokine signals. *MBio*. 2014;5(5):e01476–14. [PubMed: 25161189]
116. Sonar SA, Shaikh S, Joshi N, Atre AN, Lal G. IFN-gamma promotes transendothelial migration of CD4(+) T cells across the blood-brain barrier. *Immunol Cell Biol*. 2017;95(9):843–53. [PubMed: 28682305]
117. Bonney S, Seitz S, Ryan CA, Jones KL, Clarke P, Tyler KL, et al. Gamma interferon alters junctional integrity via Rho kinase, resulting in blood-brain barrier leakage in experimental viral encephalitis. *MBio*. 2019;10(4).
118. Sitati EM, Diamond MS. CD4+ T-cell responses are required for clearance of West Nile virus from the central nervous system. *J Virol*. 2006;80(24):12060–9. [PubMed: 17035323]
119. Wang T, Welte T. Role of natural killer and Gamma-delta T cells in West Nile virus infection. *Viruses*. 2013;5(9):2298–310. [PubMed: 24061543]
120. Frijns CJ, Kappelle LJ. Inflammatory cell adhesion molecules in ischemic cerebrovascular disease. *Stroke*. 2002;33(8):2115–22. [PubMed: 12154274]
121. Wang F, Zou Z, Gong Y, Yuan D, Chen X, Sun T. Regulation of human brain microvascular endothelial cell adhesion and barrier functions by memantine. *J Mol Neurosci*. 2017;62(1):123–9. [PubMed: 28429235]

122. Dudvarski Stankovic N, Teodorczyk M, Ploen R, Zipp F, Schmidt MHH. Microglia-blood vessel interactions: a double-edged sword in brain pathologies. *Acta Neuropathol.* 2016;131(3):347–63. [PubMed: 26711460]
123. Elahy M, Jackaman C, Mamo JC, Lam V, Dhaliwal SS, Giles C, et al. Blood-brain barrier dysfunction developed during normal aging is associated with inflammation and loss of tight junctions but not with leukocyte recruitment. *Immun Ageing.* 2015;12:2. [PubMed: 25784952]
124. Biancardi VC, Son SJ, Ahmadi S, Filosa JA, Stern JE. Circulating angiotensin II gains access to the hypothalamus and brain stem during hypertension via breakdown of the blood-brain barrier. *Hypertension.* 2014;63(3):572–9. [PubMed: 24343120]
125. Prasad S, Sajja RK, Naik P, Cucullo L. Diabetes mellitus and blood-brain barrier dysfunction: an overview. *Aust J Pharm.* 2014;2(2):125.
126. Badawi A, Velummailum R, Ryoo SG, Senthinathan A, Yaghoubi S, Vasileva D, et al. Prevalence of chronic comorbidities in dengue fever and West Nile virus: a systematic review and meta-analysis. *PLoS One.* 2018;13(7):e0200200. [PubMed: 29990356]
127. Hart J Jr, Tillman G, Kraut MA, Chiang HS, Strain JF, Li Y, et al. NIAID Collaborative Antiviral Study Group West Nile Virus 210 Protocol Team. West Nile virus neuroinvasive disease: neurological manifestations and prospective longitudinal outcomes. *BMC Infect Dis.* 2014;14:248. [PubMed: 24884681]
128. Jean CM, Honarmand S, Louie JK, Glaser CA. Risk factors for West Nile virus neuroinvasive disease, California, 2005. *Emerg Infect Dis.* 2007;13(12):1918–20. [PubMed: 18258047]
129. Ronca SE, Dineley KT, Paessler S. Neurological sequelae resulting from encephalitic alphavirus infection. *Front Microbiol.* 2016;7:959. [PubMed: 27379085]
130. Lebov JF, Brown LM, MacDonald P, Robertson K, Bowman NM, Hooper SR, et al. Review: evidence of neurological sequelae in children with acquired Zika virus infection. *Pediatr Neurol.* 2018;85:16–20. [PubMed: 30343688]
131. Liddel SA, Barres BA. Reactive astrocytes: production, function, and therapeutic potential. *Immunity.* 2017;46(6):957–67. [PubMed: 28636962]
132. Clarke LE, Liddel SA, Chakraborty C, Munch AE, Heiman M, Barres BA. Normal aging induces A1-like astrocyte reactivity. *Proc Natl Acad Sci USA.* 2018;115(8):E1896–905. [PubMed: 29437957]
133. Liddel SA, Guttenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, et al. Neurotoxic reactive astrocytes are induced by activated microglia. *Nature.* 2017;541(7638):481–7. [PubMed: 28099414]
134. Itzhaki RF, Lin WR, Shang D, Wilcock GK, Faragher B, Jamieson GA. Herpes simplex virus type 1 in brain and risk of Alzheimer's disease. *Lancet.* 1997;349(9047):241–4. [PubMed: 9014911]
135. Jamieson GA, Maitland NJ, Wilcock GK, Craske J, Itzhaki RF. Latent herpes simplex virus type 1 in normal and Alzheimer's disease brains. *J Med Virol.* 1991;33(4):224–7. [PubMed: 1649907]
136. Lovheim H, Gilthorpe J, Adolfsson R, Nilsson LG, Elgh F. Reactivated herpes simplex infection increases the risk of Alzheimer's disease. *Alzheimers Dement.* 2015;11(6):593–9. [PubMed: 25043910]
137. Lovheim H, Norman T, Weidung B, Olsson J, Josefsson M, Adolfsson R, et al. Herpes simplex virus, APOEε4, and cognitive decline in old age: results from the Betula cohort study. *J Alzheimers Dis.* 2019;67(1):211–20. [PubMed: 30636735]
138. Miklossy J, Khalili K, Gem L, Ericson RL, Darekar P, Bolle L, et al. *Borrelia burgdorferi* persists in the brain in chronic lyme neuroborreliosis and may be associated with Alzheimer disease. *J Alzheimers Dis.* 2004;6(6):639–49 discussion 673–81. [PubMed: 15665404]
139. Stanley LC, Mrak RE, Woody RC, Perrot LJ, Zhang S, Marshak DR, et al. Glial cytokines as neuropathogenic factors in HIV infection: pathogenic similarities to Alzheimer's disease. *J Neuropathol Exp Neurol.* 1994;53(3):231–8. [PubMed: 8176406]
140. Esiri MM, Biddolph SC, Morris CS. Prevalence of Alzheimer plaques in AIDS. *J Neurol Neurosurg Psychiatry.* 1998;65(1):29–33. [PubMed: 9667557]
141. Green DA, Masliah E, Vinters HV, Beizai P, Moore DJ, Achim CL. Brain deposition of beta-amyloid is a common pathologic feature in HIV positive patients. *AIDS.* 2005;19(4):407–11. [PubMed: 15750394]

142. Balin BJ, Gérard HC, Arking EJ, Appelt DM, Branigan PJ, Abrams JT, et al. Identification and localization of *Chlamydia pneumoniae* in the Alzheimer's brain. *Med Microbiol Immunol*. 1998;187(1):23–42. [PubMed: 9749980]
143. Dominy SS, Lynch C, Ermini F, Benedyk M, Marczyk A, Konradi A, et al. *Porphyromonas gingivalis* in Alzheimer's disease brains: evidence for disease causation and treatment with small-molecule inhibitors. *Sci Adv*. 2019;5(1):eaau3333. [PubMed: 30746447]
144. Readhead B, Haure-Mirande JV, Funk CC, Richards MA, Shannon P, Haroutunian V, et al. Multiscale analysis of independent Alzheimer's cohorts finds disruption of molecular, genetic, and clinical networks by human herpesvirus. *Neuron*. 2018;99(1): 64–82 e7. [PubMed: 29937276]
145. Vilensky JA, Gilman S, McCall S. Does the historical literature on encephalitis lethargica support a simple (direct) relationship with postencephalitic Parkinsonism? *Mov Disord*. 2010;25(9):1124–30. [PubMed: 20629127]
146. Takahashi M, Yamada T. Viral etiology for Parkinson's disease—a possible role of influenza a virus infection. *Jpn J Infect Dis*. 1999;52(3):89–98. [PubMed: 10507986]
147. Adams B, Nunes JM, Page MJ, Roberts T, Carr J, Nell TA, et al. Parkinson's disease: a systemic inflammatory disease accompanied by bacterial inflammagens. *Front Aging Neurosci*. 2019;11:210. [PubMed: 31507404]
148. Shen X, Yang H, Wu Y, Zhang D, Jiang H. Meta-analysis: association of *Helicobacter pylori* infection with Parkinson's diseases. *Helicobacter*. 2017;22(5).
149. Wijarnpreecha K, Chesdachai S, Jaruvongvanich V, Ungprasert P. Hepatitis C virus infection and risk of Parkinson's disease: a systematic review and meta-analysis. *Eur J Gastroenterol Hepatol*. 2018;30(1):9–13. [PubMed: 29049127]
150. Matheoud D, Cannon T, Voisin A, Penttinen AM, Ramet L, Fahmy AM, et al. Intestinal infection triggers Parkinson's disease-like symptoms in *Pink1*( $-/-$ ) mice. *Nature*. 2019;571(7766):565–9. [PubMed: 31316206]
151. Liu CC, Liu CC, Kanekiyo T, Xu H, Bu G. Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nat Rev Neurol*. 2013;9(2):106–18. [PubMed: 23296339]
152. Burgos JS, Ramirez C, Sastre I, Valdivieso F. Effect of apolipoprotein E on the cerebral load of latent herpes simplex virus type 1 DNA. *J Virol*. 2006;80(11):5383–7. [PubMed: 16699018]
153. Hoshino Y, Pesnicak L, Cohen JI, Straus SE. Rates of reactivation of latent herpes simplex virus from mouse trigeminal ganglia ex vivo correlate directly with viral load and inversely with number of infiltrating CD8+ T cells. *J Virol*. 2007;81(15):8157–64. [PubMed: 17522198]
154. Martins AS, Carvalho FA, Faustino AF, Martins IC, Santos NC. West Nile virus capsid protein interacts with biologically relevant host lipid systems. *Front Cell Infect Microbiol*. 2019;9:-8.
155. Mayer EA, Knight R, Mazmanian SK, Cryan JF, Tillisch K. Gut microbes and the brain: paradigm shift in neuroscience. *J Neurosci*. 2014;34(46):15490–6. [PubMed: 25392516]

**Box 1.****Proposed pathways underlying WNV-mediated reduction in neural resilience and cognitive damage leading to neurodegenerative dementias:**

- Neuroinvasion → complement mediated neuronal damage and synaptotoxicity
- White matter injury → impaired cognition
- Chronic inflammation → dysfunctional immunity → reduced A $\beta$  clearance increased hyperphosphorylated tau
- Impaired BBB → vascular instability → impaired brain metabolism
- Selective neuronal vulnerability and progressive brain organ failure



**Fig. 1.** Conceptual model of the role of WNV and the multifactorial cascade leading to neurodegenerative dementia

**Table 1**

MRI findings in acute and chronic WNV neuroinvasive disease. *Italic structures are also abnormal in AD, and bold structures are abnormal in PD*

<b>Acute vs chronic</b>	<b>Time after illness onset</b>	<b>MRI findings</b>	<b>Ref.</b>
Acute	1–30 days, 113 cases combined	T2 and FLAIR hyperintensities in the thalami, basal ganglia (i.e., substantia nigra, caudate nucleus, putamen), temporal lobes, parieto-occipital region, temporo-parietal region, fronto-parietal region, <i>insula</i> , <i>hippocampus</i> , periventricular white matter, corona radiata, internal capsule, corticospinal tracts, cerebellum, corpus callosum, pons	[7, 19–36]
Chronic	2 months, 1 case	Temporal lobes, <b>basal ganglia</b> , internal capsule posterior limb, and corona radiata	[37]
Chronic	5 months, 1 case	Periventricular and subcortical white matter of the bilateral frontal lobes, basal ganglia and thalami. Generalized volume loss all levels of cortex	[38]
Chronic	Average of 4 years, 30 subjects	Cortical thinning: frontal, temporal and limbic cortices (i.e., <i>posterior cingulate cortex</i> , superior and inferior frontal cortices, medial-orbito frontal region, anterior cingulate cortex, cuneus, parahippocampal region, middle and inferior temporal cortex, supramarginal region, <i>insular cortex</i> ). Also atrophy of: cerebellum, brain stem, thalamus, <b>basal ganglia</b> .	[14]